

SUPPLEMENTAL FIGURES

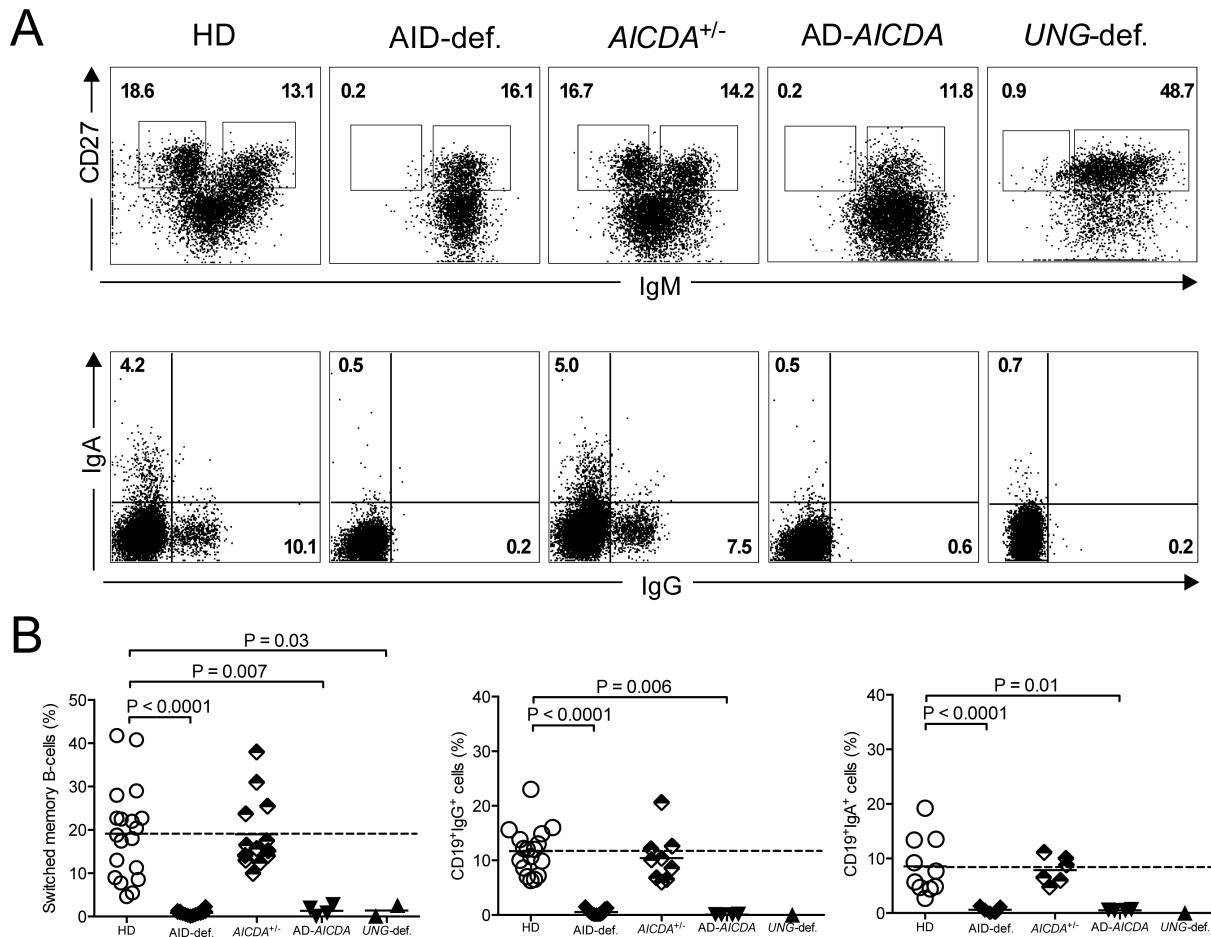
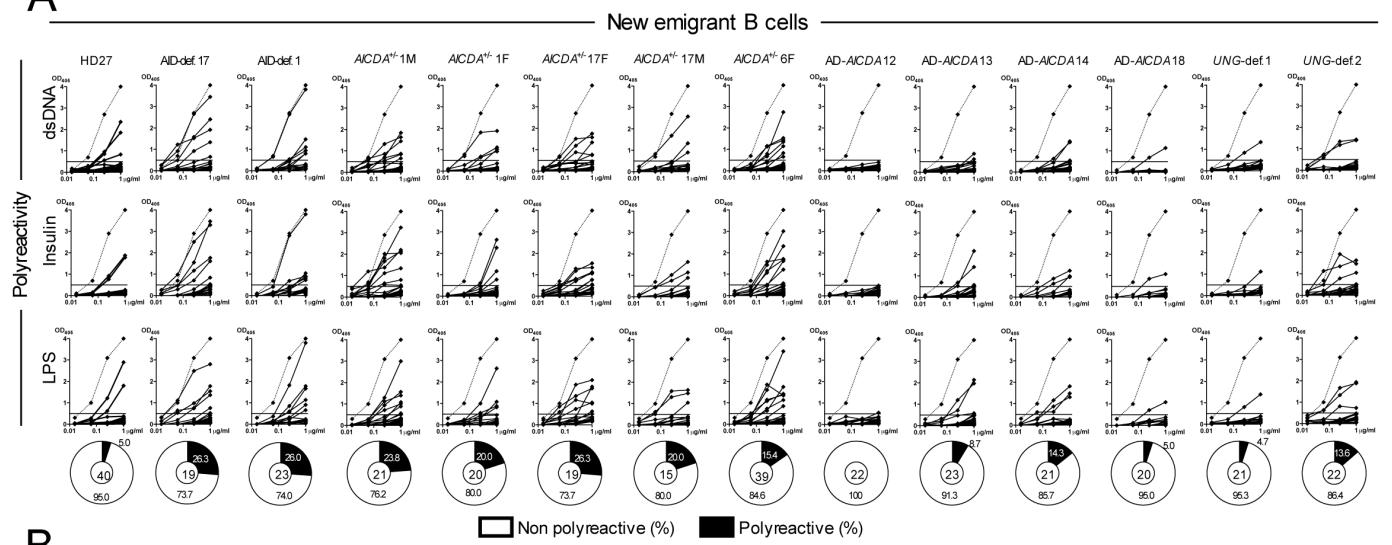
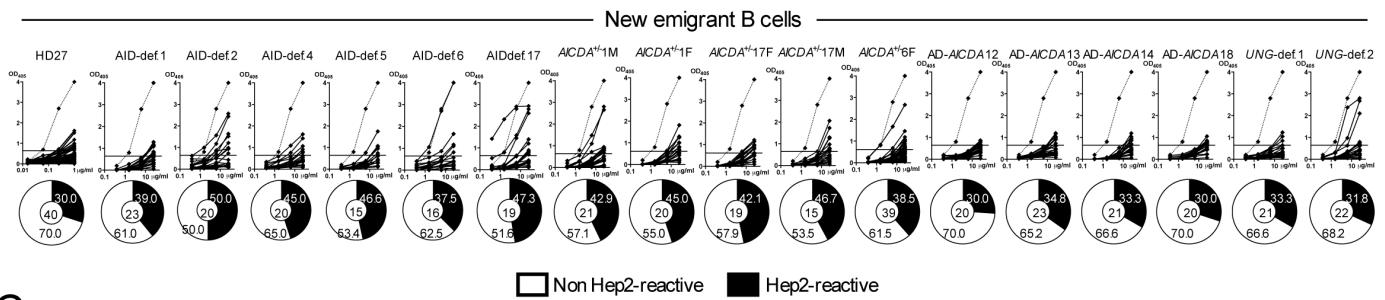


Figure S1, related to Figure 1. Absence of class-switched memory B cells in AD-AICDA and UNG-deficient patients resembles that in AID-deficient patients. (A) Representative dot plots for CD27 and IgM (upper row) or IgG and IgA (lower row) staining on purified CD20⁺ B cells. (B) Summary of the data shows absence of switched B cells in autosomal recessive AICDA-deficient (AID-def.) patients (n=17), autosomal dominant (AD)-AICDA patients (n=4) and UNG-deficient (UNG-def) patients (n=2), compared to healthy donors (HD, n=20) and asymptomatic healthy heterozygotes (AICDA^{+/−}, n=13).

A



B



C

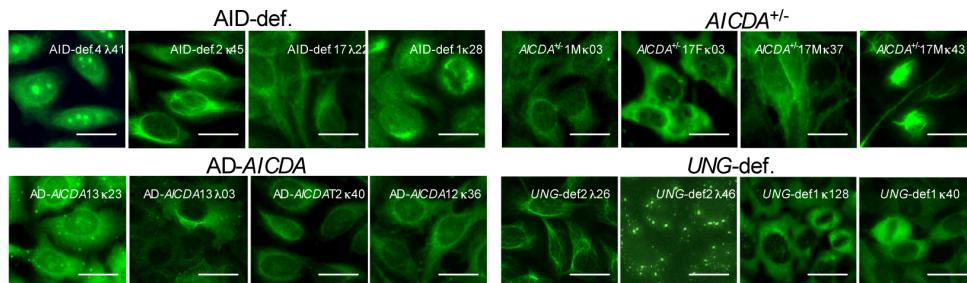


Figure S2, related to Figure 1. Increased frequencies of autoreactive new emigrant B cells in AID-def. patients and A/CDA^{+/−} healthy carriers. Recombinant antibodies expressed by CD19⁺CD21^{lo}CD10⁺IgM^{hi}CD27[−] new emigrant B cells from AID-def. patients, asymptomatic healthy heterozygotes (A/CDA^{+/−}), AD-A/CDA and UNG-def. patients were tested by ELISA for polyreactivity against dsDNA, insulin and LPS (A) or HEp-2 reactivity (B). Solid lines show binding for each cloned recombinant antibody. Dotted lines show ED38-positive control. Horizontal lines show cutoff OD405 for positive

reactivity. Antibodies were considered polyreactive when they recognized all three antigens. For each individual, the frequency of polyreactive or HEp-2-reactive B cells are summarized in pie charts, with the number of antibodies tested shown in the center. (C) Autoreactive antibodies show various patterns of HEp-2 staining. Original magnification 40X, scale bar: 10 μ m. Please see also Figure 1.

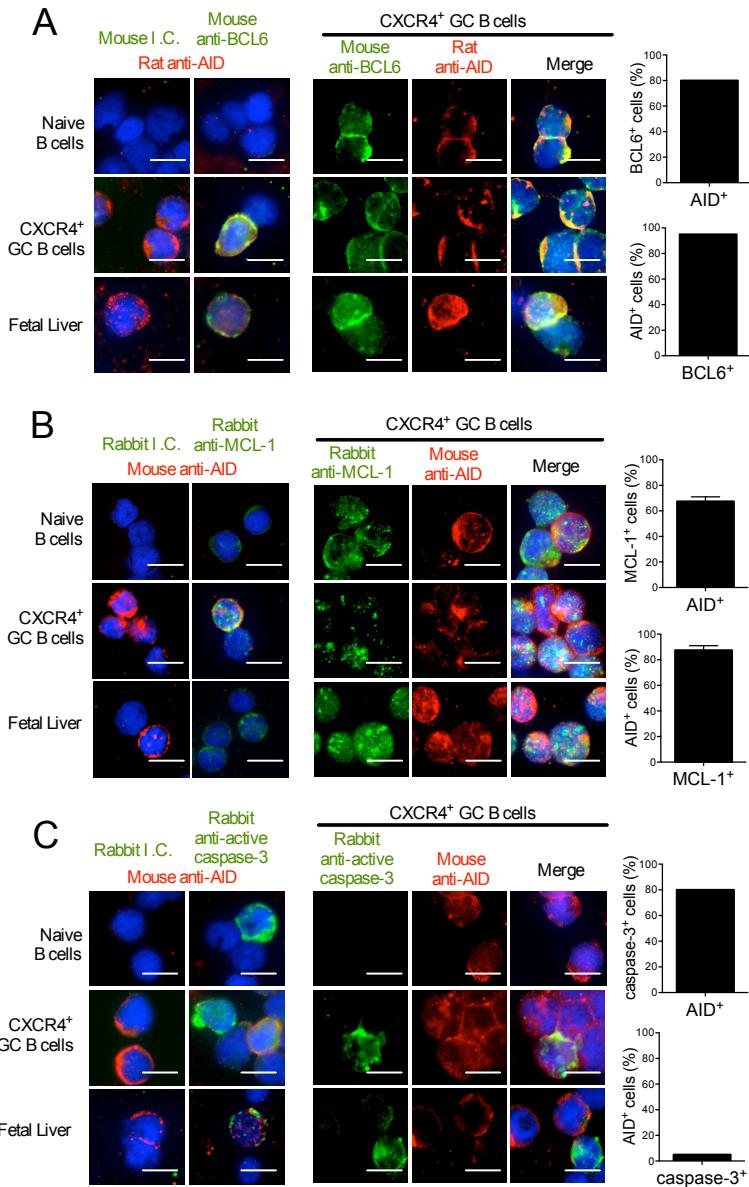


Figure S3, related to Figure 5. Assessment of anti-BCL6, anti-MCL-1 and anti-active caspase-3 antibody specificity. Cytospin slides of sorted CD19⁺IgD⁺CD38⁻CXCR4⁻ naïve B cells, CD19⁺IgD⁻CD38⁺CXCR4⁺ GC B cells and CD34⁻CD19⁺ fetal liver B cells were stained for (A) BCL-6, (B) MCL-1, (C) activated caspase-3 or concentration matched isotype control (green) and AID (red) and co-staining was quantified. Data is representative of 2 independent experiments. Original magnification 40X, scale bar: 10 μ m, Please see also Figure 5.

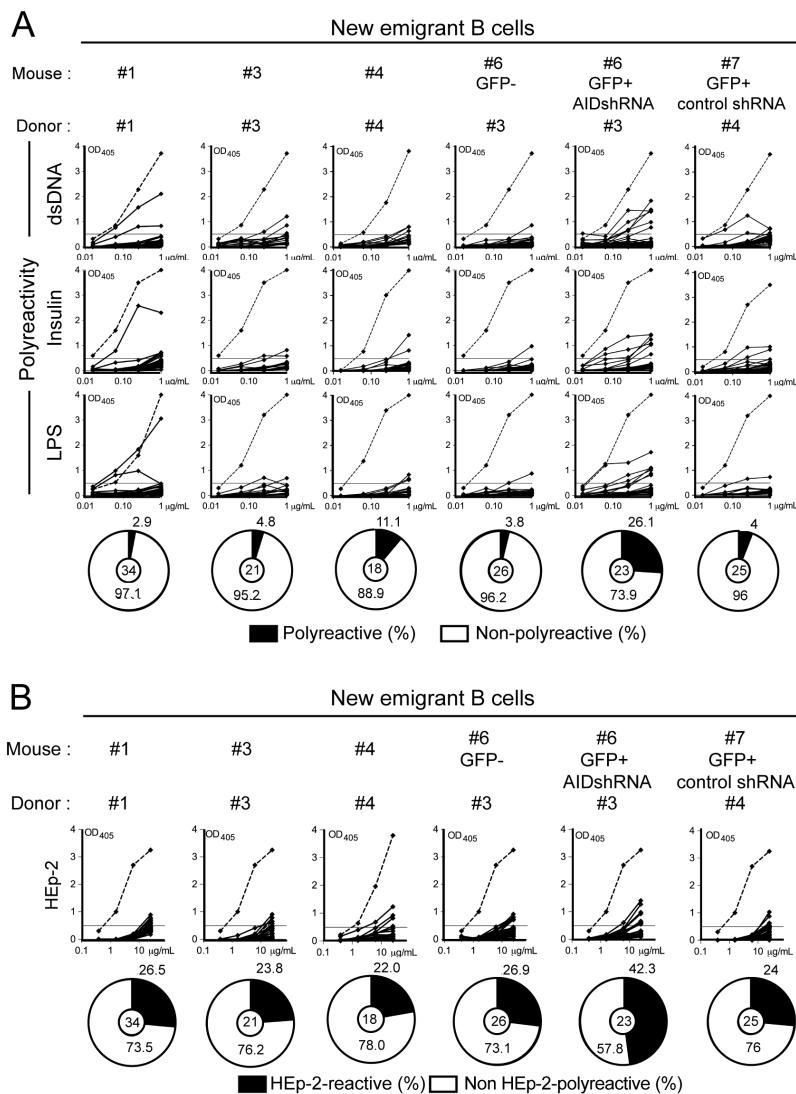


Figure S4, related to Figures 6 and 7. Central B-cell tolerance requires B-cell intrinsic AID expression. Antibodies from new emigrant B cells isolated from control humanized mice and sorted GFP- as well as GFP+ fractions expressing AID shRNA or control shRNA were tested by ELISA for reactivity against dsDNA, insulin and LPS (A) and HEp-2 cell lysates (B). Polyreactive antibodies reacted against all 3 antigens. Dotted lines show ED38-positive control. Horizontal lines show cutoff OD405 for positive reactivity. For each mouse, the frequency of reactive and non-reactive clones is summarized in pie charts, with the number of antibodies tested indicated in the center. Please see also Figures 6 and Figure 7.

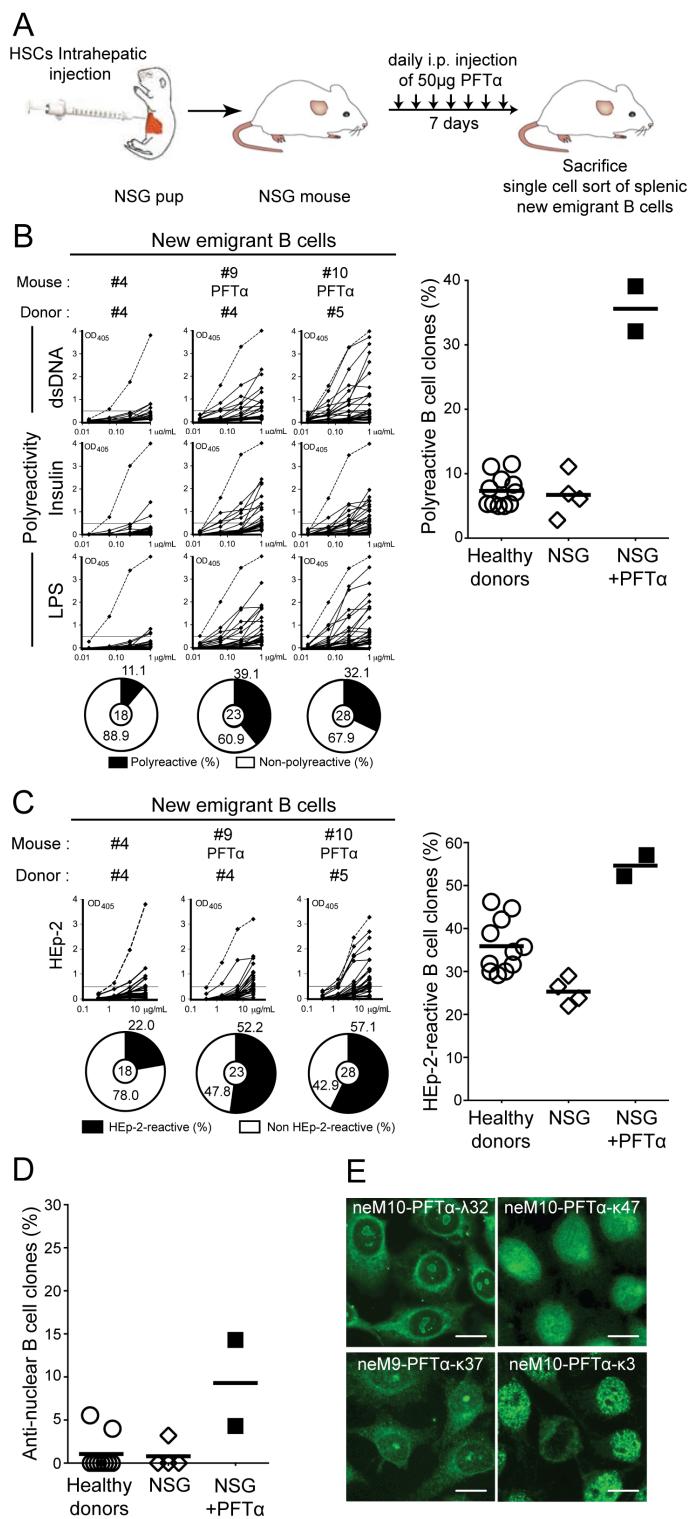


Figure S5, related to Figures 6 and 7. Defective central B-cell tolerance in PFT α -treated mice. (A) Schematic diagram depicting PFT α injections in NSG humanized

mice. CD34⁺ hematopoietic stem cells (HSCs) were injected in recipients NOD.Cg-Prkdcscid Il2rgtm1Wjl/SzJ mice. Before sacrifice mice were injected with 50µg of PFTα daily for 7 days. Antibodies from new emigrant B cells isolated from one control and two PFTα-treated humanized NSG mice were tested by ELISA for polyreactivity (B) and HEp2-reactivity (C). Dotted lines show ED38-positive control. Horizontal lines show cutoff OD405 for positive reactivity. For each mouse, the frequency of reactive and non-reactive clones is summarized in pie charts, with the number of antibodies tested indicated in the center. Polyreactive antibodies reacted against all 3 antigens (dsDNA, insulin and LPS). The frequencies of polyreactive or HEp2-reactive new emigrant B cells in healthy donors and humanized mice treated or not with PFTα are summarized in the right panels. (D) The frequencies of anti-nuclear new emigrant B cells are compared between healthy donors and humanized mice treated or not with PFTα. (E) Anti-nuclear antibodies expressed by new emigrant B cells isolated from the PFTα-treated mice show various chromatin reactive and non-reactive nuclear reactivity. Original magnification 40X, scale bar 10µm. Please see also Figures 6 and Figure 7.

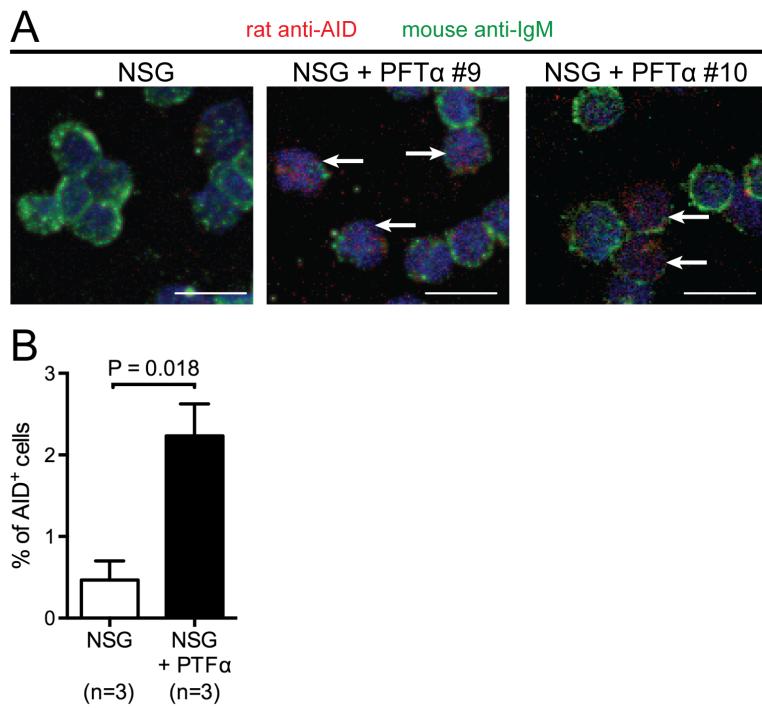


Figure S6, related to Figures 6 and 7. Increased frequency of AID⁺ bone marrow B cells in PFT α -treated mice. (A) Cytospin slides of CD19⁺ purified bone marrow cells were stained for IgM (green) and AID (red) and quantified for co-staining. White arrows show IgM⁺AID⁺ cells. Data are representative of 3 controls and 3 PFT α -treated mice. Original magnification 40X. scale bar 50 μ m (B) 300 CD19⁺ B cells (on 3 distinct fields) were analyzed for IgM and AID co-staining in control and PFT α -treated NSG humanized mice. Please see also Figures 6 and Figure 7.

Table S1, related to Figure 1. Characteristics of patients with *AICDA* or *UNG* mutations and heterozygote asymptomatic individuals

Patient number	Ethnic group	Sex	Age at diagnosis (yr)	Current age (yr)	Serum immunoglobins in mg/dl			Clinical Manifestations	Mutation in <i>AICDA</i> gene	Mutation in AID	Inheritance
					IgM	IgA	IgG				
AID-def.1*	Moroccan	M	5	22	240	<7	40	lymphoid hyperplasia, recurrent infections	203G>A / 175-184 del	W68X / H25-E58 del insV	AR
AID-def.2*	Moroccan	M	1	16	150	<7	<6	recurrent infections	203G>A / 175-184 del	W68X / H25-E58 del insV	AR
AID-def.4	Moroccan	M	1	20	100	<7	<6	lymphoid hyperplasia, recurrent infections	203G>A / del	W68X / 0	AR
AID-def.5	Turkish	M	11	21	1150	<5.8	264	lymphoid hyperplasia, recurrent infections	415A>G / 415A>G	M139V / M139V	AR
AID-def.6	Pakistani	M	4	4	4000	<7	<7	recurrent infections	394G>C / 394G>C	A132P / A132P	AR
AID-def.8	French Canadian	F	28	60	8830	<60	na	lymphoid hyperplasia, recurrent infections	334C>T / 334C>T	R112C / R112C	AR
AID-def.9	French Canadian	F	5	26	451	<30	178	lymphoid hyperplasia, recurrent infections	334C>T / 334C>T	R112C / R112C	AR
AID-def.10	French Canadian	F	3	22	621	40	60	recurrent infections	334C>T / 334C>T	R112C / R112C	AR
AID-def.11	French Canadian	M	2	50	6800	<60	na	recurrent infections	334C>T / 334C>T	R112C / R112C	AR
AID-def.17	Turkish	M		18					415A>G / 415A>G	M139V / M139V	AR
AID-def.19	Kuwait	M	1	5	223	<6	<33	lymphoid hyperplasia, recurrent infections	254G>A / 254G>A	S85N / S85N	AR
AID-def.20	Kuwait	M	1.5	16	408	40	<30	lymphoid hyperplasia, recurrent infections	254G>A / 254G>A	S85N / S85N	AR
AID-def.22*	Kuwait	M	3	14	1800	<7	<7	lymphoid hyperplasia, recurrent infections	254G>A / 254G>A	S85N / S85N	AR
AID-def.23*	Kuwait	M	8.5	13	677	<6	<33	lymphoid hyperplasia, recurrent infections	254G>A / 254G>A	S85N / S85N	AR
AID-def.26	French	M	6	10	140	<7	<7			C87R / C147X	AR
AID-def.27	French Canadian	M							334C>T / 334C>T	R112C / R112C	AR
AD-AICDA 12	French	F	71	76	203	<7	130	recurrent infections	568C>T / Normal	R190X / Normal	AD
AD-AICDA 13	Japan	M							568C>T / Normal	R190X / Normal	AD
AD-AICDA 14	Japan	F		58					568C>T / Normal	R190X / Normal	AD
AD-AICDA 18	Turkish	M		17					559del / Normal	V186X / Normal	AD
<i>AICDA</i> ** 1M	Moroccan	F		adult				healthy	203G>A / Normal	W68X / Normal	HET
<i>AICDA</i> ** 1F	Moroccan	M		adult				healthy	175-184 del	H25-E58 del insV / Normal	HET
<i>AICDA</i> ** 6F	Pakistani	M		adult				adult	394G>C / Normal	A132P / Normal	HET
<i>AICDA</i> ** 6M	Pakistani	F		adult				adult	394G>C / Normal	A132P / Normal	HET
<i>AICDA</i> ** 17F	Turkish	M		adult				healthy	415A>G / Normal	M139V / Normal	HET
<i>AICDA</i> ** 17M	Turkish	F		adult				healthy	415A>G / Normal	M139V / Normal	HET
<i>AICDA</i> ** 19F	Kuwait	M		35				healthy	254G>A / Normal	S85N / Normal	HET
<i>AICDA</i> ** 20F	Kuwait	M		48				healthy	254G>A / Normal	S85N / Normal	HET
<i>AICDA</i> ** 22M	Kuwait	F		33				healthy	254G>A / Normal	S85N / Normal	HET
<i>AICDA</i> ** 26M	French	F		41				healthy		C87R / Normal	HET
<i>AICDA</i> ** 27B	French Canadian	M		57				healthy	334C>T / Normal	R112C / Normal	HET
<i>AICDA</i> ** 27S	French Canadian	F		47				healthy	334C>T / Normal	R112C / Normal	HET
UNG-def.1	USA	M	39	40	785	<7	209	lymphoid hyperplasia, recurrent infections	497,498 AT del/497,498 AT del	frameshift	
UNG-def.2	Japan	F	3	12	267	25	<50	HIGM	T822C/T822C	F251S/F251S	
UNG-def.3	French	M	7	26	740	48	50	HIGM	462 C del/639,640 TA del	frameshift	

* siblings

AD: autosomal dominant

HET: heterozygote

Supplementary Table 2, related to Figure 1. Repertoire and reactivity of antibodies from new emigrant B cells of HD30

Ig	HEAVY					LIGHT					REACTIVITY		
	VH	D	RF	JH	CDR3(aa)	Length	Vκ	Jκ	CDR3(aa)	Length	Poly	HEp-2	Staining
neHD30 K3	4-39	5-12	2	4	PSLSGYDYSLDY	12	3-20	3	QQYGSSPLFT	10	-	-	-
neHD30 K4	3-15	/	/	3	GGGVGAFDI	9	2-28	5	MQALQTPT	8	-	-	c
neHD30 K5	3-30-3	1-26	2	6	QSSPTGGGMDV	11	1-5	2	QQYNNSYSYT	9	-	+	-
neHD30 K10	4-4	3-9	3	5	VSNHILTGNRLFDP	14	3-20	1	QQYGSSPRT	9	-	+	-
neHD30 K12	1-2	2-15	2	3	HPIGYCSGGSCYGGAFDI	18	1-33	4	QQYDNLPT	8	-	-	-
neHD30 K14	4-34	3-10	3	2	KGTMVRGVIIIPHWYFDL	18	2-28	1	MQALQTPQT	9	-	+	-
neHD30 K16	3-30	/	/	6	EGVSHYYGMDV	12	2-28	4	MQALQTPPT	9	-	-	-
neHD30 K20	3-30	4-17	2	6	DIHRDYGDYETPNYYYYGMDV	21	1-9	1	QQLNSYPLA	9	-	+	-
neHD30 K24	3-72	/	/	6	DNRGMVD	7	4-1	4	QQYYSTPLT	9	-	-	-
neHD30 K26#	1-18	3-16	2	4	NYDYIWGSYFTRGGY	15	1-33	3	QQYDNLPLFT	10			
neHD30 K43	3-9	3-22	2	2	VDSSGLFGWYFDL	13	3-20	1	QQYGSSPQT	9	-	-	-
neHD30 K45#	3-15	2-15	3	4	DPLIVVVVAATRDFDY	16	1-39	2	QQSYSTPYT	9			
	VH	D	RF	JH	CDR3(aa)	Length	Vλ	Jλ	CDR3(aa)	Length	Poly	HEp-2	Staining
neHD30 L7	4-59	/	/	4	DRNFDY	6	3-1	2	QAWDSSSTAV	9	-	-	-
neHD30 L11	3-33	/	/	4	GGGGGDY	7	2-14	2	SSFTSITYVV	10	-	-	-
neHD30 L19	3-11	1-7	2	4	DRGNNGRNPYNWNYFFDY	18	2-14	1	SSYTSSSTLLYV	12	-	+	-
neHD30 L21#	3-53	4-17	2	4	VGRYGDYAYTITSRVYYFDY	20	2-14	2	SSYTSSSTLGV	11			
neHD30 L28	3-15	1-26	1	6	AGITKFWELPGYYYYGMDV	19	1-44	3	AAWDDSLNVVV	11	-	-	-
neHD30 L30	3-23	2-21	3	6	GEGGVVTTQALMDV	15	1-44	2	AAWDDSLNGVV	11	-	-	-
neHD30 L42	1-69	4-4	2	6	SDYSNYVLYDYYYYYGMVD	19	2-8	2	SSYAGSNNLV	10	+	-	-
	VH	D	RF	JH	CDR3(aa)	Length							
neHD30 H2	4-34	3-10	2	4	GQDYYGSGSTADY	13							
neHD30 H29	3-23	6-13	2	4	DEVSSSWLFGY	11							
neHD30 H38	3-72	3-10	1	6	AGSWWFGETRYGMDV	15							
neHD30 H41	3-23	5-12	2	4	VRGRPDSGYRFDY	13							
neHD30 H44	1-69	2-15	2	6	GSYCSGGSCYSSFYYYYGMDV	21							
neHD30 H46	3-21	/	/	6	DDNLPIDYGMVD	12							

RF, reading frame; #, antibody failed to be expressed

-, non-reactive; +, reactive;

c, diffuse cytoplasmic staining; N, nuclear staining; F, cytoplasmic fibers; V, vesicles

Inventory Data for Figure 1: Equipment and supplies that were present at the time of the survey									
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1	151	0	0	0	0	0	0	0	0
1	152	0	0	0	0	0	0	0	0
1	153	0	0	0	0	0	0	0	0
1	154	0	0	0	0	0	0	0	0
1	155	0	0	0	0	0	0	0	0
1	156	0	0	0	0	0	0	0	0
1	157	0	0	0	0	0	0	0	0
1	158	0	0	0	0	0	0	0	0
1	159	0	0	0	0	0	0	0	0
1	160	0	0	0	0	0	0	0	0
1	161	0	0	0	0	0	0	0	0
1	162	0	0	0	0	0	0	0	0
1	163	0	0	0	0	0	0	0	0
1	164	0	0	0	0	0	0	0	0
1	165	0	0	0	0	0</td			

month and year	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec

or leaving blank if no relevant information

for example, "none" or "not applicable".

Supplementary Table 4, related to Figure 6 and 7. Repertoire and reactivity of antibodies from new emigrant B cells of NSG mice

Ig	HEAVY							LIGHT				REACTIVITY			
	VH	D	RF	JH	CDR3(aa)		Length	Vκ	Jκ	CDR3(aa)	Length	Poly	HEp2	Staining	
neM1 K2	1-3	1-7	2	4	DLELYYFDY		9	3-20	1	QQYGSSPT	8	-	-	-	
neM1 K5	4-30-2	3-10	2	4	ISGSYYNY		8	1-39	1	QQSYSTPWT	9	-	+	-	
neM1 K8	4-59	3-9	2	4	SPPFDWIYYFDY		12	3-20	1	QQYGSSPWT	9	-	-	-	
neM1 K9	3-73	4-23	2	4	RYGGNYYFDY		10	3-11	4	QQRSNWPLT	9	-	+	-	
neM1 K10	3-15	1-26	1	3	QTEWELDAFDI		11	3-20	4	QQYGSSPPLT	10	-	-	-	
neM1 K11	3-74	3-10	2	6	DPGKGYYGSGSYYYYGMDV		20	1-6	2	LQDYNYPYT	9	-	-	-	
neM1 K16	4-30-2	/	/	3	TNEPNAFDI		9	2-28	4	MQALQTPLT	9	-	-	-	
neM1 K22	4-4	3-22	2	3	DRADYYDSSGYYYAFDI		17	1-17	4	LQHNSYPPT	9	-	-	-	
neM1 K23	3-7	1-26	2	6	NRPPGAINYYGMDV		14	3-11	1	QQRSNWPLT	9	-	-	-	
neM1 K25	4-34	1-26	3	4	GGGATEY		7	1-17	1	LQHNSYLWT	9	-	-	-	
neM1 K32	4-59	/	/	3	THPQSDAFDI		10	2-30	4	MQGTHWPLT	9	-	+	-	
neM1 K33#	3-7	3-10	3	3	ILGGITMVRGAEDAFDI		17	3-20	1	QQYGSSPRT	9				
neM1 K37	3-30	3-10	1	4	EKGWFGELEGLAIDY		15	3-15	1	QQYNNWPQT	9	-	-	-	
neM1 K38	4-34	7-27	2	2	GLTGDSGTDWYFDL		14	1-39	4	QQSYSTPLT	9	-	-	-	
neM1 K40	4-39	6-13	2	4	HPYSSSFDY		9	3-20	2	QQYGSSYT	8	-	-	-	
neM1 K42	3-33	3-10	2	6	DPRLGSGSYYYYGMDV		16	1-39	3	QQSYSTPFT	9	-	+	-	
neM1 K43#	4-30-4	7-27	2	4	NKLNWGLDY		9	1-5	1	QQYNSYSPWT	10				
neM1 K45	4-4	7-27	2	5	GWGFGNWFDP		10	3-20	2	QQYGSSPPYT	10	-	-	-	
neM1 K46	4-59	/	/	4	SFSRLASFY		10	3-20	1	QQYGSSLGT	9	-	-	-	
neM1 K48	4-59	/	/	4	VGGRRGGDFDY		10	1-33	3	QQYDNLPRVT	10	-	-	-	
	VH	D	RF	JH	CDR3(aa)		Length	Vλ	Jλ	CDR3(aa)	Length	Poly	Hep2	Staining	
neM1 L1	3-30	6-13	2	4	SGSSWYYFDY		10	2-14	2	SSYTSSSTV	9	-	-	-	
neM1 L3	1-69	/	/	4	VMAVYYFDY		9	1-40	1	QSYDSSLGKV	11	+	+	-	
neM1 L6	3-30-3	3-16	2	4	GPSRGGEVFDY		11	3-21	3	QVWDSSSDQNWV	12	-	-	-	
neM1 L7	5-51	3-22	2	4	LRIPDYDSSGYYFDY		15	2-8	3	SSYAGSNNLV	10	-	-	-	
neM1 L12	4-4	/	/	4	GSIPSDYYFDY		11	3-25	2	QSADSSGTPV	11	-	-	-	
neM1 L13#	1-69	6-13	2	5	TIYSSSWYWFDP		12	5-37	2	MIWPSNAYVV	10				
neM1 L14	3-30	3-22	2	2	DYYDSSGYYYWYFDL		15	1-44	2	AAWDDSLNGPV	11	-	+	+	
neM1 L24	3-13	6-13	2	3	GGRDSSWYIAFDI GQTTNLGMGPNWFDP DPLSRYCSGGSCYSGAFDI		13	2-23	1	CSYAGSSTYV	10	-	+	-	
neM1 L26	1-3	7-27	2	5			15	1-40	2	QSYDSSLGVV	11	-	-	-	
neM1 L27	3-23	2-15	2	3			19	1-44	2	AAWDDSLNGHVV	12	-	-	-	
neM1 L28	1-18	6-19	3	4			10	2-8	2	SSYAGSNNLV	10	-	-	-	
neM1 L30	3-7	/	/	4	DRLGSFDY		8	1-47	3	AAWDDSLSGWV	11				
neM1 L31	3-33	3-9	2	4	DAALRYFDWLDDY		13	2-14	1	SSYTSSSTLV	10	-	-	-	
neM1 L34	3-7	/	/	4	DLVGIRATDY		10	1-51	1	GTWDSSLSAYV	11	-	+	-	
neM1 L35	3-30	6-13	3	1	DVNAAGNRAYFQH		14	3-21	1	QVWDSSSDHYV	11	-	-	-	
neM1 L36	3-30	1-26	2	4	DGDSGSYFDY		10	2-23	2	CSYAGSSTYVV	11	-	-	-	
neM1 L39	3-30-3	7-27	1	3	VCSPELGGQWIDI		12	3-21	2	QVWDSSSDHVV	11	-	+	-	
neM1 L41	3-15	3-16	2	3	DWGTRAFDI		9	1-51	3	GTWDSSLSAGV	11	-	-	-	
	VH	D	RF	JH	CDR3(aa)		Length								
neM1 H4	3-64	6-19	3	4	GIVAVAGNLDY		11								

Supplementary Table 5, related to Figure 6 and 7. Repertoire and reactivity of antibodies from new emigrant B cells of AIDshRNA treated I

Ig	HEAVY						LIGHT				REACTIVITY			
	VH	D	RF	JH	CDR3(aa)		Length	Vκ	Jκ	CDR3(aa)	Length	Poly	HEp2	Staining
neGFP+M5 K3	1-46	4-23	2	1	KPNYGANQYFQH		12	3-20	3	QQYGSSPLT	9	+	+	n
neGFP+M5 K10	3-7	5-5	2	3	DSVRYSYGTDAFDI		14	1-37	4	QRTYNAPLT	9	-	-	-
neGFP+M5 K11	3-30	5-5	3	5	DRDTAMVTWFDP		12	1-5	1	QQYNSYSWT	9	+	-	-
neGFP+M5 K14	3-7	6-19	2	4	APLRWEYSSGWYGDY		15	3-15	1	QQYNNWPPWT	10	-	+	-
neGFP+M5 K16#	1-2	6-13	2	4	SPRSWYDY		8	3-20	5	QQYGSSPTT	9			
neGFP+M5 K18	5-51	6-13	2	3	YSSSWWGAFDI		11	2-30	1	MQGTHWPPT	9	+	-	-
neGFP+M5 K19	3-30-3	1-26	1	6	DRENGGAWERLYGMVD		16	3-20	1	QQYGSSPT	8	+	+	-
neGFP+M5 K20	3-15	1-7	2	2	GPNYPNQSRYFDL		13	3D-15	4	QQYNNWPPAPFT	12	+	+	n
neGFP+M5 K21	3-33	2-21	2	4	EAYCGGDCYFPWDY		14	1-5	2	QQYNSYSHT	9	-	+	c
neGFP+M5 K22	5-a	6-6	2	3	PHEYSSSDAFDI		13	1-5	1	QQYNSYST	8	-	+	-
neGFP+M5 K23	4-61	4-17	2	4	ESGDYYFDY		9	4-1	2	QQYYSTPYT	9	-	-	-
neGFP+M5 K25#	4-39	6-13	3	3	WYGAFDI		7	3-15	1	QQYNNWLTWT	10			
neGFP+M5 K27	4-61	/	/	3	KPNIPDAFDI		10	1-39	1	QQSystPGP	9	-	-	-
neGFP+M5 K28	1-69	3-22	2	4	DYYDSSGYYPFDY		13	3-20	2	QQYGSSPYT	9	+	-	-
neGFP+M5 K29	3-30	4-17	2	4	DLSKVRTYDYGDYNNY		16	1-17	1	LQHNSYPWT	9	-	+	-
neGFP+M5 K31	4-34	3-22	2	3	GVYYDSSGYSNDAFDI		16	3-11	2	QQRSNWYT	8	-	-	-
neGFP+M5 K33	4-34	6-13	3	5	GKGIAAAGYWNFDP		14	3-15	1	QQYNNWLTWT	10	+	+	-
neGFP+M5 K34	3-33	3-22	2	4	DAHYYDSSGYPPAYYFDY		18	1-5	3	QQYNSYLFT	9	-	+	-
neGFP+M5 K38	3-33	/	/	4	GQTVF DY		7	3-15	2	QQYNNWPTYT	9	-	+	n
neGFP+M5 K42	4-34	3-16	1	4	GPAGELSLDY		10	3-11	5	QQRSNWLPI T	10	-	+	-
neGFP+M5 K48	3-33	3-16	3	4	DLYRVTFGGAI DY		13	1-5	2	QQYNSYPWT	9	-	+	-
	VH	D	RF	JH	CDR3(aa)		Length	Vλ	Jλ	CDR3(aa)	Length	Poly	Hep2	Staining
neGFP+M5 L5	3-23	4-17	2	2	DGYGDYYWYFDL		12	1-40	2	QSYDSSLGSV	11	-	-	-
neGFP+M5 L7	3-64	/	/	4	WAGLGSYYFDY		11	3-21	2	QVWDSSSDHHV	11	-	-	-
neGFP+M5 L8	3-15	6-13	2	6	DRDSSS WTYYYYYMDV		18	2-14	1	SSYTSSSTLV	10	-	-	n
neGFP+M5 L9#	3-7	6-19	2	3	GRSSGWFDADI		12	3-21	2	QVWDSSSDHPV	11			
neGFP+M5 L12	3-7	3-10	2	6	DTTYYYGSGSYSDYYYYYMDV		21	1-51	2	GTDSSSLAGV	11	-	+	-
neGFP+M5 L15	4-34	3-22	2	4	GYS DSSGV DY		10	2-14	3	SSYTSSSTLG	10	-	-	n
neGFP+M5 L24	3-64	3-3	2	2	DSSKDFWSGPWYFDL		15	2-14	2	SSYTSSSTLV	11	-	-	-
neGFP+M5 L30	1-3	7-27	2	4	DSLGI DL PFDY		11	2-11	2	CSYAGSYTLV	10	-	+	-
neGFP+M5 L32#	3-30	6-19	2	4	VTSYSSGWYGDY		13	2-14	2	SSYTSSSTLV	10			
neGFP+M5 L37	5-51	6-13	3	4	GIAAAGPFDY		10	1-44	1	AAWDDSHYV	9	-	-	-
neGFP+M5 L38					see Kappa			3-25	2	QSADSSGTYVV	11	-	+	-
neGFP+M5 L39	3-30	/	/	6	FHGDYYGMDV		10	3-21	1	QVWDSSSDHYV	11	+	+	-
neGFP+M5 L40	3-30	6-19	3	4	DHHRIAVADLF DY		13	3-21	2	QVWDSSSDHHV	11	-	-	-
neGFP+M5 L41	4-34	2-8	2	4	HRNCTNGVCYFDY		13	2-8	2	SSYAGSNNVV	10	-	-	-
neGFP+M5 L43	4-4	3-3	1	3	RFLEWLECAF DI		12	1-51	2	GTWDSSLSAVV	11	-	+	-
neGFP+M5 L46#	1-3	6-13	3	6	DGELAAAAGYYYYGMDV		18	1-47	3	AAWDDSLSGWV	11			
neGFP+M5 L13								2-14	3	SSYTSSSIWV	10			
	VH	D	RF	JH	CDR3(aa)		Length							
neGFP+M5 H6	3-11	6-13	2	4	VYDSSTYYFDY		11							
neGFP+M5 H45	1-18	6-13	3	4	DLAAAGHGGY		10							

Supplementary Table 6, related to Figure 6 and 7. Repertoire and reactivity of antibodies from new emigrant B cells of PFTα treated NSG mice

Ig	HEAVY							LIGHT					REACTIVITY		
	VH	D	RF	JH	CDR3(aa)	Length	Vκ	Jκ	CDR3(aa)	Length	Poly	Hep2	Staining		
neM9-PFTa K3	5-10	2-2	2	4	HVGYCSTSCDDY	13	1-17	1	LQHNSYPWT	9	+	-	-		
neM9-PFTa K8	3-7	2-8	2	2	VVDLPILGYCTGGVCDKRGWYFDL	24	3-20	2	QQYGSSPQYT	10	+	+	-		
neM9-PFTa K16	4-39	3-10	2	5	TWRGSGSYNNATTRKDWFDP	20	3-15	1	QQYNNWPQT	9	+	+	-		
neM9-PFTa K17	3-23	2-2	2	4	EDCSSTSCFDYFDY	14	3-20	3	QQYGSSPLFT	11	-	-	-		
neM9-PFTa K20	3-23	6-13	3	3	DRSIAAAGTGAAFDI	15	1-5	1	QQYNSYFWT	9	-	+	-		
neM9-PFTa K27	3-48	/	/	4	SFLGLKWVDY	10	3-15	1	QQYNNWPQT	9	+	+	c		
neM9-PFTa K36	4-34	7-27	2	6	EVTNWGSYYYMDV	13	1-8	4	QQYYSYPRS	9	+	+	-		
neM9-PFTa K37	3-7	2-15	2	4	SSGGYCSGGSCYTLPPYYFDY	20	3-20	2	QQYGSSPRYT	10	-	+	n		
neM9-PFTa K41	1-69	2-21	2	3	GAYCGGDCNDAFDI	14	3-20	2	QQCGSSPYT	9	-	-	-		
neM9-PFTa K42	4-39	6-13	3	2	DVLAAAGNDWYFDL	15	1-5	2	QQYNSYSRT	9	+	-	-		
neM9-PFTa K44	3-7	5-12	3	4	DGGRVATRATSFDY	14	3-15	2	QQYNNWPPMYT	11	-	-	-		
neM9-PFTa K45	3-23	/	/	4	DSGWGAFDY	9	1-39	4	QQSYSTPQRDRLT	13	+	+	-		
	VH	D	RF	JH	CDR3(aa)	Length	Vλ	Jλ	CDR3(aa)	Length	Poly	Hep2	Staining		
neM9-PFTa L1	3-30	4-17	2	4	DEGGLAYFDDYGDYCPGY	19	1-47	3	AAWDDSLSGWV	11	-	+	-		
neM9-PFTa L3					see κ-chain	11	2-11	3	CSYAGSYTWV	10					
neM9-PFTa L6	1-2	6-6	2	4	DRAPFRSSSPHDY	14	3-21	2	QVWDSSSDHV	11	-	+	-		
neM9-PFTa L7	1-18	/	/	4	ASTRYGADY	9	1-51	3	GTWDSSLASAWV	11	-	+	-		
neM9-PFTa L9	3-23	/	/	4	VPYPQYYFDY	10	3-25	2	QSADSSGTSVV	11	-	-	-		
neM9-PFTa L10	3-23	2-2	3	3	LVVPAAMRDAFDI	13	2-8	2	SSYAGSNIVV	10	-	-	-		
neM9-PFTa L11	3-23	2-21	2	5	GAYCGGDCLLDWFDP	15	2-11	1	CSYAGSYTYV	10	+	+	-		
neM9-PFTa L12#	4-34	3-10	2	4	GGDYYGSGSYLFNRKIQGSKGFDYYFDY	28	2-23	1	CSYAGSYV	8					
neM9-PFTa L18	3-23	6-13	3	4	QPAAGDY	7	2-14	2	SSYTSSSTVV	10	-	-	-		
neM9-PFTa L22	1-18	7-27	1	4	VLLPIGLGMDY	11	1-51	1	GTWDSSLASAYV	11					
neM9-PFTa L24	4-61	3-10	3	4	EARVRRQGVITSPYFDY	18	1-44	1	AAWDDSLNGPV	11	-	+	-		
neM9-PFTa L25#	3-53	6-13	2	6	SGSSSSLHYYYYMDV	15	2-23	1	CSYAGSSPNVV	11					
neM9-PFTa L30#	1-3	6-6		6	GTDIAARPSEGYYYYGMDV	19	2-14	1	SSYTTSSSTYV	10					
neM9-PFTa L33#	4-34	2-2	3	4	VTDIVVVPAAPRSPPCFDY	19	1-51	2	GTWDSSLASAVV	11					
neM9-PFTa L45					see κ-chain	13	1-40	2	QSYDSSLGGSV	11	-	-	-		
neM9-PFTa L46	3-30	4-4	2	4	EGDYSNLAWRRYFDY	15	2-8	1	SSYAGSNNYV	10	-	-	-		
neM9-PFTa L47	3-11	3-10	1	5	GGGVLFRELLNNWFDP	17	2-14	2	SSYTSSSTVV	10	+	+	-		
	VH	D	RF	JH	CDR3(aa)	Length	Vκ	Jκ	CDR3(aa)	Length	Poly	HEp2	Staining		
neM10-PFTa K3	3-21	1-26	2	4	EQNSGSYYY	9	1-39	1	QQSYSTTWT	9	+	+	n		
neM10-PFTa K8	3-30-	3	1-26	2	VCVGSYYFDY	10	1-39	3	QQSYSTPFT	9	+	+	-		
neM10-PFTa K9	1-2	6-13	3	3	DLRIAAGDAFDI	13	3-15	5	QQYNNWPIT	9	-	+	-		