

FIGURE S1 (related to Figure 1)

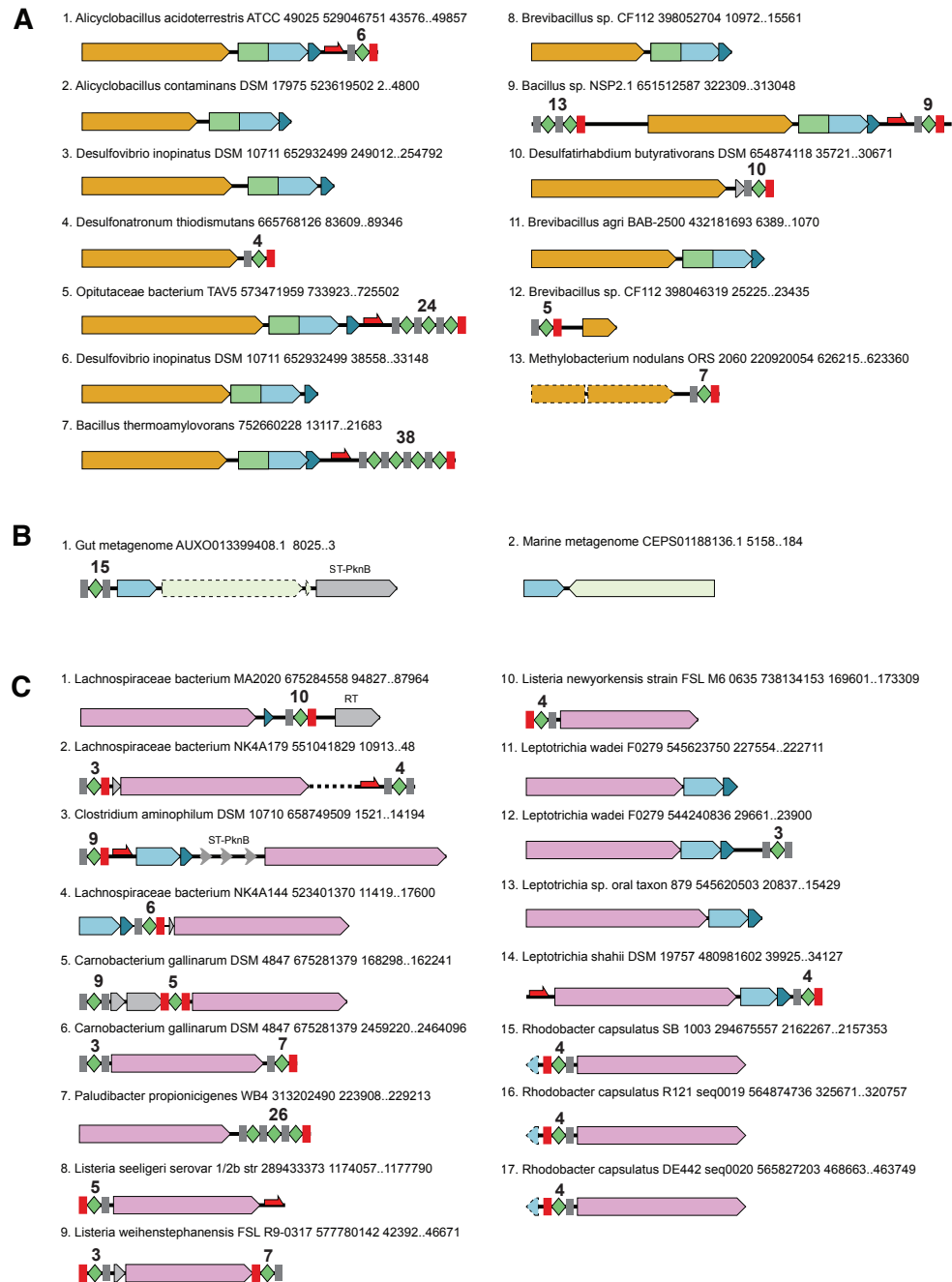


Figure S1 | Genomic architectures of all identified candidate novel CRISPR-Cas loci.

(A) The C2c1 loci (subtype V-B). (B) The C2c3 loci (subtype V-C). (C) The C2c2 loci (type VI). Genes are colored according to the schematic in Figure 2. The number of repeats in CRISPR arrays is indicated. For each genomic contig, Genbank numeric ID and the coordinates of the locus are indicated. Additional designations: ST-PknB, serine/threonine protein kinase; RT, reverse transcriptase.

FIGURE S3 (related to Figure 2)

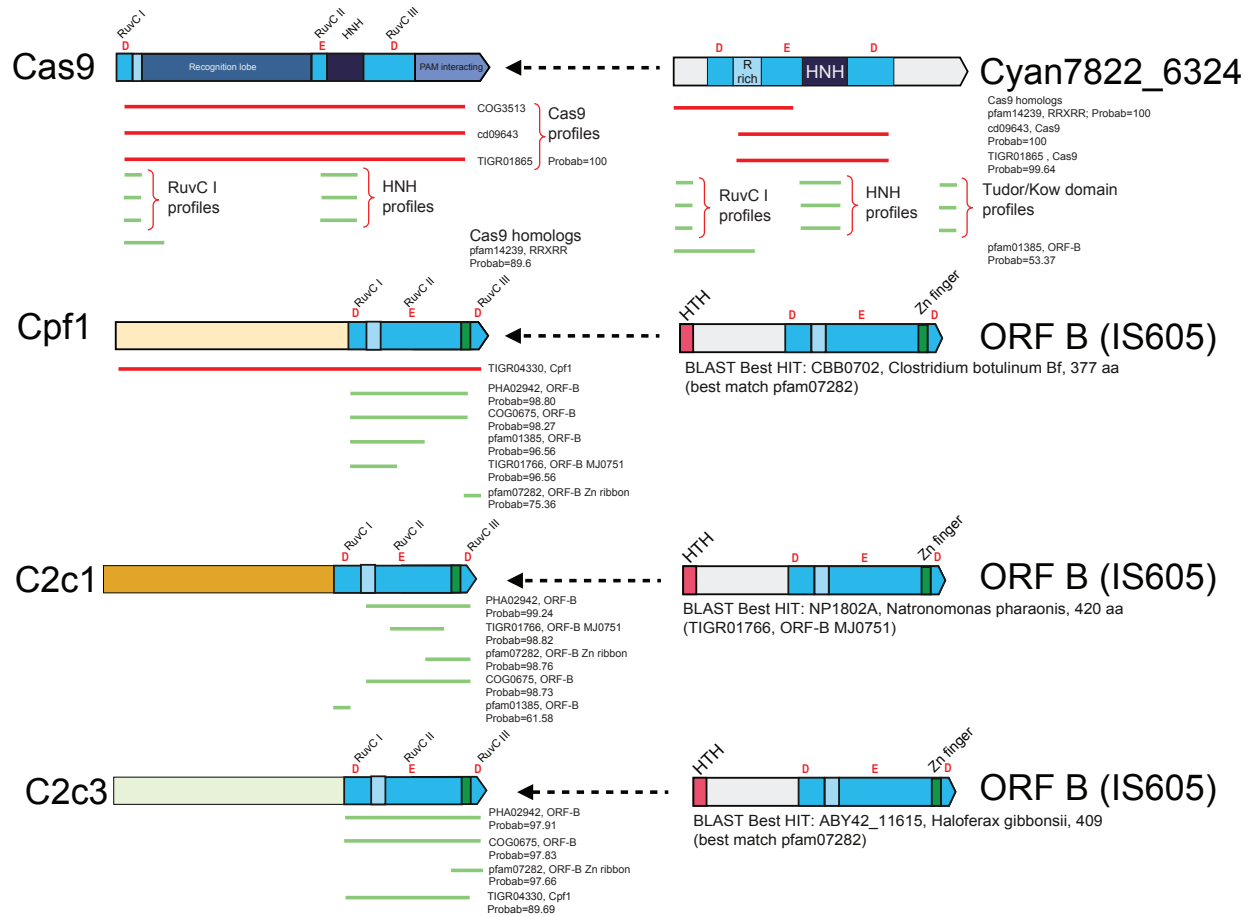


Figure S3 | The closest homologs of the new type V effector proteins among the transposon-encoded proteins: non-overlapping sets of homologs.

FIGURE S4 (con't)

```
544884152_Alicyclobacillus_acidoterrestriis ---HFTGEFGGA---KIQCRRDQLA-----HMHR--RRGARDVYLNVSVRVQSQSE--ARGERRPYYAAVFRVLG---DNHRAFFVHF
652589596_Alicyclobacillus_contaminans ---LVFVAFGGA---KIQFNRAHLS-----AGYRKGVLASAGGGGSIYFNVTLDVQVNE-----RDVSKTFSFSD
652932497_Desulfovibrio_inopinatus ---QFAGLAGGA---EVLFRPYME-----HDERSEESLLERPQAVWFKLTLDVATQAP-----PNWLDGKGRVTR
667765471_Desulfonatronum_thiodismutans ---QFEGLLGGA---EILFDRKRIA-----NEQHGATDLASKPGHVVFKLTLDRVQAP-----QGWLDGKGRPAL
497199019_Opitutaceae_bacterium_TAV5 APLPTLPQDLTG---MPVFLMPDVT-----LSGERRILLNLPVTLPEAALVEQLGNAGRWQNFPGF-----SREDPFALR
654153037_Tuberibacillus_calidus ---ELKGTLLGGA---RIQFDREHLL---R-RQGVK--AGNVGRIFLNVTLNIEPMQPFPSRGNLQTSVQKALKVYV--DGYPKVNVFKPK
754485389_Bacillus_thermoamylovorans ---PLKGTLLGGA---RVQFDRDHLR---RYPHKVE--SGNVGRIYFNMTVNIETP-----ESPVSRSKLIHR--DDFPKVNFKPK
495056180_Brevibacillus_sp_Cf112 ---TFNHLGGA---KIQLERGDLE-----KEEKNFE--DGEIGSVYLNVDIFEPL-GEVKNRQVQAPYQGVQLLIRRPNEFPKVTYKSE
651512544_Bacillus_sp_NSP2-1 ---TFNHLGGA---KIQLERGDLE-----KEEKNFE--DGEIGSVYLNVDIFEPL-GEVKNRQVQAPYQGVQLLIRRPNEFPKVTYKSE
Secondary_structure_for_651512544_(Jpred) -----EEEEEE--E-----HH-----HHH-----EEEEEEEEEE--HH-----HHHHHHHHHH-----HHH
654874074_Desulfatirhabdium_butyraivorans ---NITGLFEQADWNGRLQAPRQQLAIAVVRDNPRLSEQERNLRMCGMIEHIRLWLVTFVSKLQPC-----GPWCAYAEQ-----HGLNTNPFQY
652569729_Alicyclobacillus_herbarius -----
652589403_Alicyclobacillus_contaminans -----
411770298_Citrobacter_freundii_ATCC_8090 -----
696372964_Citrobacter_freundii -----
492410745_Brevibacillus_agri -----
492410748_Brevibacillus_agri -----
495062547_Brevibacillus_sp_Cf112 -----
506407588_Methylobacterium_nodulans -----
219945206_Methylobacterium_nodulans_ORS_2060 -----
760065057_Methylobacterium_nodulans -----
CONSENSUS_0.8 -----
RuvC-like_motifs -----
544884152_Alicyclobacillus_acidoterrestriis DKLSDYLAEHPPDGKLGSEGLLSGLRVMSVDLGLRTSASISVFRVARKDLK-----PNSKGRVFFFFIKGND
652589596_Alicyclobacillus_contaminans RDLVSLKAEELKRYMETKPLQMPGVYRVMSVDLGVRYGAAISVFEVFKPFAEVR-----KDK-----LHYPTGCE
652932497_Desulfovibrio_inopinatus PVEVHFHTALSNSKSKHRTTLPQGLRVLSVDLQMRFTFACSVFELIEGKPEFG-----RAFFPADGR
667765471_Desulfonatronum_thiodismutans PPEAKHFHTALSNSKSFADQVRPGLRVLSVDLQMRFTFACSVFELVIRGGPDQ-----G-----TYFFPADGR
497199019_Opitutaceae_bacterium_TAV5 WPADGAVTKAGKTHIPWHQDRDHTYVLGVDLGRDAGALALLNVTAQKPAK-----PVHRI I
654153037_Tuberibacillus_calidus ELTEHIKESKNTLTLGVESLPTGLRVMSVDLQMRQAAASISFEVVEKPD-----DNK-----LFYVKDD
754485389_Bacillus_thermoamylovorans ELTEWIKDSKGGKLSKESLELIGLRVMSIDLQGRQAAASISFEVVEKPD-----EGK-----LFFPKGTE
495056180_Brevibacillus_sp_Cf112 QLVEWIKASPGHSA--GVESLASGFRVMSIDLGLRAAATSISFVSESSDKN-----AAD-----FSYWIEGTP
651512544_Bacillus_sp_NSP2-1 QLVEWIKASPGHSA--GVESLASGFRVMSIDLGLRAAATSISFVSESSDKN-----AAD-----FSYWIEGTP
Secondary_structure_for_651512544_(Jpred) HHHHHHH-----HHHHHHHHHH-----HHH-----EEEE-----EEEE
654874074_Desulfatirhabdium_butyraivorans WPHADTNRDRKHARLILPLPL-GLRVLSVDLGHRYAAACAVWEAVNTETVKEACQNVGRDMPKHEHDYLIHKVKKGIKQTEVDKTTIYRRI GADTLP
652569729_Alicyclobacillus_herbarius -----
652589403_Alicyclobacillus_contaminans -----
411770298_Citrobacter_freundii_ATCC_8090 -----
696372964_Citrobacter_freundii -----
492410745_Brevibacillus_agri -----
492410748_Brevibacillus_agri -----
495062547_Brevibacillus_sp_Cf112 -----
506407588_Methylobacterium_nodulans -----
219945206_Methylobacterium_nodulans_ORS_2060 -----
760065057_Methylobacterium_nodulans -----
CONSENSUS_0.8 -----
RuvC-like_motifs -----
544884152_Alicyclobacillus_acidoterrestriis N-----LVAVHERSQQLLKPGET-----ESKDLRAIEERQRTLRQLRTOQLAYLRLVRCGS-EDVGRR---ERSWAKIIEQPV
652589596_Alicyclobacillus_contaminans -----FVAHERSVLILKPGCE-----GVTAGQSQSERKQALAAIRAEMSILRKLWLRVSVQVTEEDRA-----KAVRGLLEDERG
652932497_Desulfovibrio_inopinatus SMDSPNKLMAKHERSFKLTLPGET-----PSRKEEERSIARAEYALKRDIQRKLSLRGLGEDNDNRR-----DALLEQFFKQWG
667765471_Desulfonatronum_thiodismutans TVDDEKTLMAKHERSFKTLPGEN-----PSRKEEIRARAAMEELRSIMGDIRRKLAILRLSVLQEDDPR-----THELRLPMAEIV
497199019_Opitutaceae_bacterium_TAV5 GEADGRTWYASLADAMIRLPGEDARLTVRQKLVQEPYGERGNASLEWEDARNIILRLGQNPDELGDPRHRSYPIENDKLLVALRAQARLARLQ
654153037_Tuberibacillus_calidus -----LEAVHRTSFNKLKPGKE-----RTERRMLEQKQKQDQATRLSRKLFKLVNVMQKLEKTDTER-----EKRVNRWIKDRE
754485389_Bacillus_thermoamylovorans -----LVAVHRSFNIKLKPGET-----LVKSREVLIRKAREDNKLMNQKLNFRNVHLFPQGFEDITER-----EKRVTKWISROE
495056180_Brevibacillus_sp_Cf112 -----LVAVHRSYMLRLKPGEQ-----VEKQVMEKRDREDFQLHQRVQFQIRVLAQIMRMAN-KQYGRD---WDELDSIKQAVE
651512544_Bacillus_sp_NSP2-1 -----LVAVHRSYMLRLKPGEQ-----VEKQVMEKRDREDFQLHQRVQFQIRVLAQIMRMAN-KQYGRD---WDELDSIKQAVE
Secondary_structure_for_651512544_(Jpred) -----EEEEEE-EEEE-----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHHHHHH-----HHH-----HHHHHHHHHHHH
654874074_Desulfatirhabdium_butyraivorans DGRPHAPAPMARLDRQLIFLKLQGE-----EKDAREASNEEIALWQHMECKLDRTKPLIDRLIASWGGLL---KRQMARLDALKE
652569729_Alicyclobacillus_herbarius -----
652589403_Alicyclobacillus_contaminans -----
411770298_Citrobacter_freundii_ATCC_8090 -----
696372964_Citrobacter_freundii -----
492410745_Brevibacillus_agri -----
492410748_Brevibacillus_agri -----
495062547_Brevibacillus_sp_Cf112 -----
506407588_Methylobacterium_nodulans -----
219945206_Methylobacterium_nodulans_ORS_2060 -----
760065057_Methylobacterium_nodulans -----
CONSENSUS_0.8 -----
RuvC-like_motifs -----
544884152_Alicyclobacillus_acidoterrestriis DAANHMTDPDWEAFENELQK-----LKSLHGICSDKE-----W
652589596_Alicyclobacillus_contaminans GGWTMDPGEDSDHQPLQQL-----HEARLAVGELVNLVHLSFAEW-----
652932497_Desulfovibrio_inopinatus EEDVVPQQA-----FPRSLFQGLGAAPFRSTPELW-----
667765471_Desulfonatronum_thiodismutans DDPAKSA-----LNAEFLKGGGDDFRSTPELW-----
497199019_Opitutaceae_bacterium_TAV5 RSWRLDLAESDKALDEIHAERAGE-----KPSPLPLPLARDDAIKSTDEA-----
654153037_Tuberibacillus_calidus REENEP-----VYQDEFEM-----ISKVLY-SPHSV-----W
754485389_Bacillus_thermoamylovorans NSDVFL-----VYQDELQ-----IRELMY-KPYKD-----W
495056180_Brevibacillus_sp_Cf112 QKKSFLDQDRTFWEGIVCD-----LTKVLP-RNEAD-----W
651512544_Bacillus_sp_NSP2-1 QKKSFLDQDRTFWEGIVCD-----LTKVLP-RNEAD-----W
Secondary_structure_for_651512544_(Jpred) -----EEEE-----H-----
654874074_Desulfatirhabdium_butyraivorans LGWIPAPDSSENLSREDGEAKDYRESLAVDDLMFSAVRTLRLLALQRHGNPARIAYYLSSEVRIIPGGIQEKLDENGRIDLLQDALALWHELFPSPGWRDE
652569729_Alicyclobacillus_herbarius -----
652589403_Alicyclobacillus_contaminans -----
411770298_Citrobacter_freundii_ATCC_8090 -----
696372964_Citrobacter_freundii -----
492410745_Brevibacillus_agri -----
492410748_Brevibacillus_agri -----
495062547_Brevibacillus_sp_Cf112 -----
506407588_Methylobacterium_nodulans -----
219945206_Methylobacterium_nodulans_ORS_2060 -----
760065057_Methylobacterium_nodulans -----
CONSENSUS_0.8 -----
RuvC-like_motifs -----
```

FIGURE S4 (con't)

544884152_Alicyclobacillus_acidoterrestris -----MDAVESVRRVWRHMGKQVRDRKDVRSGERPKIRGYADV-----V
652589596_Alicyclobacillus_contaminans -----ERAVIERHRRLERITASHIRVFQTMKRVWGRNRNEDAAH-----T
652932497_Desulfovibrio_inopinatus -----RQHCTYYDKAEACLAKHISDWRKRTRPRPTSREMYKTRSY-----H
667765471_Desulfonatronum_thiodismutans -----KQCHFFHKDAEKVVAERFSRWRTRPKSSSWQDWRREGRYA-----
497199019_Opitutaceae_bacterium_TAV5 -----LLSQRDIIRRSFVQIANLILPLRGRWRWRPHVEVPDCHILA-----QSDPGTDDTKRLVAGO
654153037_Tuberibacillus_calidus -----VDQLKSIHRKLEQLGKIEISKWRQSIQ--GRQG-----V
754485389_Bacillus_thermoamylovorans -----VAFKQLHKKRLEVEIGKEVKHWRKSLSD--GRKG-----L
495056180_Brevibacillus_sp_Cf112 -----EQAVVQIHRKAEYVGVKAVQAWKRFAA--DERKG-----I
651512544_Bacillus_sp_NSP2-1 -----EQAVVQIHRKAEYVGVKAVQAWKRFAA--DERKG-----I
Secondary_structure_for_651512544_(Jpred) -----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH-----
654874074_Desulfatirhabdium_butyraetivorans AAKQLWDSRIATLAGYKPAEENGDNVSDVAYRKKQVYERQLRNVAKTLSGDVITCKELSDAWKERWEDEQRWKLRLWFKDWLPSGTQANNATIRNV
652569729_Alicyclobacillus_herbarius -----
652589403_Alicyclobacillus_contaminans -----
411770298_Citrobacter_freundii_ATCC_8090 -----
696372964_Citrobacter_freundii -----
492410745_Brevibacillus_agri -----EQAVVQIHRKAEYVGVKAVQAWKRFAA--DERKG-----I
492410748_Brevibacillus_agri -----
495062547_Brevibacillus_sp_Cf112 -----
506407588_Methylobacterium_nodulans -----QDTCRAARLYRTEFGAVVSEWRSRTRSREDRK-----Y
219945206_Methylobacterium_nodulans_ORS_2060 -----
760065057_Methylobacterium_nodulans -----
CONSENSUS_0.8 -----
RuvC-like_motifs -----
544884152_Alicyclobacillus_acidoterrestris GGNSEIQIEYLERQYKFLKSWSPFGYVSGQ-----VIRAEGKSR--FAITREHIDHAKEDRLKLLADRIIMEALGVYVALDERGKQKW-----
652589596_Alicyclobacillus_contaminans GGISLAHIEHILQQRKLFIRKSWTHARTYGE-----VRLPKHEG--FAKRLQKHTNHVKEDRIKKLADMIVMAARGYRF--LDRKRAM-----V
652932497_Desulfovibrio_inopinatus GGRKSWMLLEYLDVAVRLLKLSWLRGRTYGA-----INRQDTRFSGSLASRLHHSINSKEDRIKTGADSVIQAARGYIP--LPHGKGM-----
667765471_Desulfonatronum_thiodismutans GGRKSWAVTYLEAVRGLILRWNRMRGTYGE-----VNRQDKQFGTVASALLHHINQLKEDRIKTGADMIQAARGYIP--RKNAGAV-----
497199019_Opitutaceae_bacterium_TAV5 RGISHERIEQIEKTRRQCSLNRLRHKPGERPVLGRPAKGEIADPCALLEKINRLRQDRVQDTAHAI LAAALGVRRLRAPSCKDRAER--RHRDIGHYI
654153037_Tuberibacillus_calidus YGILSKNIEDIEKTRRLLFRWSMRPEPGE-----VRQLQPER--FAIDQNHNLHDKDRIKKLANQIIMVMTALGYRY--DGKRRKW-----C
754485389_Bacillus_thermoamylovorans YGILSKNIDEIDTRKFLRWSLRPTPEGE-----VRLQPER--FAIDQNHNLHDKDRIKKLANQIIMVMTALGYRY--DGKRRKW-----C
495056180_Brevibacillus_sp_Cf112 AGLSMWNIIELEGRLKLLISWSRRSRNPQE-----VNRFERGHT--SHQRLTHIQNVKEDRLKQLSHAVMTALGYVY--DERKQEW-----Q
651512544_Bacillus_sp_NSP2-1 AGLSMWNIIELEGRLKLLISWSRRSRNPQE-----VNRFERGHT--SHQRLTHIQNVKEDRLKQLSHAVMTALGYVY--DERKQEW-----C
Secondary_structure_for_651512544_(Jpred) -----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH-----
654874074_Desulfatirhabdium_butyraetivorans GGLSLSLRATITTEFRKVVQGVFF--TRLRDP-----GTRHEIGEQ--FGQKTLDALELLRQRVQLASRIAEALGIG--SEGKQWGGGRPRQRIN
652569729_Alicyclobacillus_herbarius -----
652589403_Alicyclobacillus_contaminans -----
411770298_Citrobacter_freundii_ATCC_8090 -----
696372964_Citrobacter_freundii -----
492410745_Brevibacillus_agri AGLSMWNIIELEGRLKLLISWSRRSRNPQE-----VNRFERGHT--SHQRLTHIQNVKEDRLKQLSHAVMTALGYVY--DERKQEW-----
492410748_Brevibacillus_agri -----
495062547_Brevibacillus_sp_Cf112 -----
506407588_Methylobacterium_nodulans AGKSMWSVQHITDVRRLQSWSLAGRASGD-----IRRLDRERGGVFAKDLDDHIDALKDDRLLKTGADLIVQAARGF--QRNEFGY-----W
219945206_Methylobacterium_nodulans_ORS_2060 -----
760065057_Methylobacterium_nodulans -----
CONSENSUS_0.8 -----
RuvC-like_motifs -----
544884152_Alicyclobacillus_acidoterrestris VAKYPCQLIILLEELSEYQFNDRPSENNQLMWSHRGVFQELINQAQVHDLI-----VGTWYAASFSSFDARTGAPGIRCKRVF
652589596_Alicyclobacillus_contaminans KTHAPCDLILFEDLSRYRFTMDRPPENSQLMWWSHRLEKVTYKMQAALFGIG-----VGTVPAFTSFDATGAPGIRCKRVF
652932497_Desulfovibrio_inopinatus EORVEPCQLILFEDLSRYRFTMDRPPENSQLMWWSHRLEKVTYKMQAALFGIG-----VGTVPAFTSFDATGAPGIRCKRVF
667765471_Desulfonatronum_thiodismutans VQVHEPCRLILFEDLSRYRFTMDRPPENSQLMWWSHRLEKVTYKMQAALFGIG-----VGTVPAFTSFDATGAPGIRCKRVF
497199019_Opitutaceae_bacterium_TAV5 ERFRAPADVVIENLSRYLSQDRASENTRLMQWCHRQIQVQLRQLCETYGIP-----VLAVPAAYSRSFSSRDGAGFRVHIT
654153037_Tuberibacillus_calidus A-KHPACQIILFEDLSRYRFTMDRPPENSQLMWWSHRLEKVTYKMQAALFGIG-----VGTVPAFTSFDATGAPGIRCKRVF
754485389_Bacillus_thermoamylovorans A-KNFAQCIILFEDLSRYRFTMDRPPENSQLMWWSHRLEKVTYKMQAALFGIG-----VGTVPAFTSFDATGAPGIRCKRVF
495056180_Brevibacillus_sp_Cf112 A-EYFACQIILFENLSQYRSNLRDSTKENSTLMKWAHRSIPKYVHMQAEPYGIQ-----IGDVRAEYSSRFYAKTGTGPGIRCKRVF
651512544_Bacillus_sp_NSP2-1 A-EYFACQIILFENLSQYRSNLRDSTKENSTLMKWAHRSIPKYVHMQAEPYGIQ-----IGDVRAEYSSRFYAKTGTGPGIRCKRVF
Secondary_structure_for_651512544_(Jpred) -----EEEEH--H-----HHHHHHHHHHHHHHHHHHHHHHHHHHHHHH-----EEEE-----E
654874074_Desulfatirhabdium_butyraetivorans DSRFAPCHAVVIENLANYRDETRTRLENRLMWTASAKVHKYLSAQCINGLY-----LCTVSAWYTSRQDSRTGAPGIRCKRVF
652569729_Alicyclobacillus_herbarius -----
652589403_Alicyclobacillus_contaminans -----
411770298_Citrobacter_freundii_ATCC_8090 -----
696372964_Citrobacter_freundii -----
492410745_Brevibacillus_agri CAEYFACQIILFENLSQYRSNLRDSTKENSTLMKWAHRSIPKYVHMQAEPYGIQ-----IGDVRAEYSSRFYAKTGTGPGIRCKRVF
492410748_Brevibacillus_agri -----
495062547_Brevibacillus_sp_Cf112 -----
506407588_Methylobacterium_nodulans VQKHAPCHVILFEDLSRYRMTDRPPRENSQLMWAHRGVPMVGMQGEIYQIDRRDPDSARKHARQPLAAFLDTPAAFSSRYHASTMTGPIRCHPLR
219945206_Methylobacterium_nodulans_ORS_2060 -----
760065057_Methylobacterium_nodulans -----
CONSENSUS_0.8 -----
RuvC-like_motifs -----
544884152_Alicyclobacillus_acidoterrestris ARCTQEHNPFPFWLNLKVF-----VEHTLDACPLRADD-----LIPTGEGEIVFSP--FSAEEDGFQIHADLNAAQLQRRRL
652589596_Alicyclobacillus_contaminans KQDKKPTFWLI-----QFAEITGVNVTNVEPQ-----LIPVDGGEVFPSPKGPRAADGLKCVHADINAAHNLQRRF
652932497_Desulfovibrio_inopinatus ERDFDNDLKPYPYLLRELSWMLGNKTV-----ESEEEKLRLSEKIRPGS-----LVPWDGGEQFATL--HPKRTQLCVHADNAAQLQRRF
667765471_Desulfonatronum_thiodismutans EEDFDGDLPGMHLVGLDNLWLPKDKD-----RTANEARKLLGGMVRPGM-----LVPWDGGEQFATL--NAASQLHVIHADNAAQLQRRF
497199019_Opitutaceae_bacterium_TAV5 PDHRRMFWRSRI LARKLAHEEDGKRLKTVLDEARAVRGLFDRDRFNAGHVPKGPWRLLAPLFGPVFVL-----GDATPMQADLNAAIINALGI
654153037_Tuberibacillus_calidus EHELYITEGGQVNRKFLD-----SL-----VENNIIEPDDARLEPGD-----LIRDQGGKQFATL--DERGELVITHADNAAQLQRRF
754485389_Bacillus_thermoamylovorans K-----EKIQDNRFK-----NL-----QREGRLTDKIAVLKEDG-----LVPDKGGEKIFSL--SKDRKLVTHADNAAQLQRRF
495056180_Brevibacillus_sp_Cf112 GQDL-----GRRFENLQK-----RL-----VNEQFLTEEQVKLRPGD-----IVPDDSGELFMTLTDGSGSKEVFLQADINAAHNLQRRF
651512544_Bacillus_sp_NSP2-1 GQDL-----GRRFENLQK-----RL-----VNEQFLTEEQVKLRPGD-----IVPDDSGELFMTLTDGSGSKEVFLQADINAAHNLQRRF
Secondary_structure_for_651512544_(Jpred) -----HH-----HH-----EEEE-----EEEEHHHHHHHHHHHHHHHHHHHHHHHHHHHHHH-----
654874074_Desulfatirhabdium_butyraetivorans VREFMQSPFWKQVQKAEAKHDENK-----DARERFLCELNKTWAKTAPAEWKAGFVRIPLRGEVFSVA--DSKSPSARGIHADLNAAANIGLRAL
652569729_Alicyclobacillus_herbarius -----
652589403_Alicyclobacillus_contaminans -----
411770298_Citrobacter_freundii_ATCC_8090 -----
696372964_Citrobacter_freundii -----
492410745_Brevibacillus_agri GQDL-----GRRFENLQK-----RL-----VNEQFLTEEQVKLRPGD-----IVPDDSGELFMTLTDGSGSKEVFLQADINAAHNLQRRF
492410748_Brevibacillus_agri -----
495062547_Brevibacillus_sp_Cf112 -----
506407588_Methylobacterium_nodulans KREFEDQGFLELLKREM-----EGLDLNGYKPGD-----LVPLPGGEVFCVL--NANGLSRIHADINAAQLQRRF
219945206_Methylobacterium_nodulans_ORS_2060 -----
760065057_Methylobacterium_nodulans -----
CONSENSUS_0.8 -----
RuvC-like_motifs -----
P...G...F.....AD.NAA.N...R...
-----D-----

FIGURE S4 (con't)

```

544884152_Alicyclobacillus_acidoterrestris          SDFDISQIRLRCWGEVDGELVLPLRLTGKRT-----ADSYSNKVFYNTGVTYERERGKRRKRVFAQEKLESEEAELLVEADE
652589596_Alicyclobacillus_contaminans             IP---RLPSVKRCRRVVEAEGFAAVPSSAFMKVHGKG-----AFVSDVDFEYEQKGRVAV---NRADRTSSTLDEDEGDI GEEMLVSN
652932497_Desulfovibrio_inopinatus                 GRCG-EAFRLVCPHGDDVLRLASTPGARLLGALQQL-----ENGQGAFLVDRMGSTSQMNRVFMKLGKKIKPLQDNNGDDELDVLSVLPPE
667765471_Desulfonatronum_thiodismutans           GRCG-EAIRIVCNQLSDVGSSTRYEMAKAPKARLLGALQQLKNGDAPPHLTSIPNSQKPNYSVMTPTNAGKKYRAGPGEKSSGEE--DELALDIVEQAE
497199019_Opitutaceae_bacterium_TAV5              AAP--DRHDIHHRRAENKRLSLR-----LGTQREKARWPGGAPAVTLSTPNNGASPEDSDALPERVSNLFVDIAGV
654153037_Tuberibacillus_calidus                   TRTH-GLYRIRCESREIKDVAVLPSSDKDQKEMENL-----FGIGYLQPFKQENDVYKVVGKEKIG--KKTSSQSDKELV-SEILQEASVMADE
754485389_Bacillus_thermoamylovorans              TRTH-GFYKVVYCKAVQVDGQTVYIPESKDKQKIIE-----FGEGY--FILKDGVEWGNAGKLI--KKGSSQSSSELVSDILKDSFDLASE
495056180_Brevibacillus_sp_Cf112                  QRYN-ELFKVSCRIVRDEEYLVPKTKSVQAKLKGK-----LFVKKS--DTAWKDVYVWDSQAKLKG--KTTFTTEESPEQ-LEDFQEIIEEAE
651512544_Bacillus_sp_NSP2-1                      QRYN-ELFKVSCRIVRDEEYLVPKTKSVQAKLKGK-----LFVKKS--DTAWKDVYVWDSQAKLKG--KTTFTTEESPEQ-LEDFQEIIEEAE
Secondary_structure_for_651512544_(Jpred)         H-----EE--EEEE--EEEE--HHHHH-----EEE-----EEEE--HH-----HEHH-----HH-HHHHHHHHHHHH
654874074_Desulfatirhabdium_butyrativorans       TDP-----DWPQKVVVPCD-----PVSESKMDYVKCAAVKVGQPLRPAQTADGAASKIRKGGKNTAG
652589403_Alicyclobacillus_contaminans           -----
652589403_Alicyclobacillus_contaminans           -----
411770298_Citrobacter_freundii_ATCC_8090          -----
696372964_Citrobacter_freundii                    -----
492410745_Brevibacillus_agri                      -----
492410745_Brevibacillus_agri                      -----
495062547_Brevibacillus_sp_Cf112                  -----
506407588_Methylobacterium_nodulans                -----
219945206_Methylobacterium_nodulans_ORS_2060     TQHG-DAFRLPCGSAVQGIIRWAPLSMGKRQAGALG-----GFGYLEPTGHDSSGCRKRTAEAWRRLSQAQKDRDEAAAAAEEDEELQGLEEELE
760065057_Methylobacterium_nodulans              -----
CONSENSUS_0.8                                     .....
RuvC-like_motifs                                  .....

544884152_Alicyclobacillus_acidoterrestris          AREKSVVLMRDPGGIINRNG---WTRQKEFWSM/NQRIEGLVQIIRSVPLQDSACENTGDI-----
652589596_Alicyclobacillus_contaminans             GAGEFVRMFYDESQVYGYG---RWMSKVFQKVRQIVHRAIQDQVEKRAAARGENGATSSR-----
652932497_Desulfovibrio_inopinatus                 DDTGRIITVFRDSSGIFPCPN---VWIPAKQFPAWRAMIKVMASHLSG-----
667765471_Desulfonatronum_thiodismutans           LAQGRKTFTRDPSGVFFPAD---RWLPSEIYWSRIRRRIRWQVTLERNSSGRQERAEEMDEMPY-----
497199019_Opitutaceae_bacterium_TAV5              ANFERVTIEGVSQ-----KPAFRGLWASVKQRAWNRVARLNTVTDNNRNEEEDDIPM-----
654153037_Tuberibacillus_calidus                   LKGNRKTFRDPSGVVFPKD---RWYTGGRYFGLTEHLKRLAER---RLFDGSSRRGLFNGTDSNTNVE
754485389_Bacillus_thermoamylovorans              LKGEKMLFRDPSGNVFPD---KWAAGVYFGKLERLILSKLTNQYSISTIEDSSKQSM-----
495056180_Brevibacillus_sp_Cf112                  AKGTYRTLFRDPSGVFFPES---WVYQKDFWGEVKKRKYGKLRERFLTKAR-----
651512544_Bacillus_sp_NSP2-1                      AKGTYRTLFRDPSGVFFPES---WVYQKDFWGEVKKRKYGKLRERFLTKAR-----
Secondary_structure_for_651512544_(Jpred)         ---EEEEEE---EEE-----HHHHHHHHHHHHHHHHHHHH
654874074_Desulfatirhabdium_butyrativorans       TSKEKVLWRDISAPPLESNEIGEWKETSAYQNDVQYRIVRLKEHIKSLDNRGTGDNVEG-----
652589403_Alicyclobacillus_contaminans           -----
652589403_Alicyclobacillus_contaminans           -----
411770298_Citrobacter_freundii_ATCC_8090          -----
696372964_Citrobacter_freundii                    -----
492410745_Brevibacillus_agri                      -----
492410745_Brevibacillus_agri                      -----
495062547_Brevibacillus_sp_Cf112                  -----
506407588_Methylobacterium_nodulans                -----
219945206_Methylobacterium_nodulans_ORS_2060     RSGERVVFRDPSGVVLPD---LWFPAAFWIVRAKTVGRILRSHLDAQEASAYAAAGL-----
760065057_Methylobacterium_nodulans              -----
CONSENSUS_0.8                                     .....
RuvC-like_motifs                                  .....

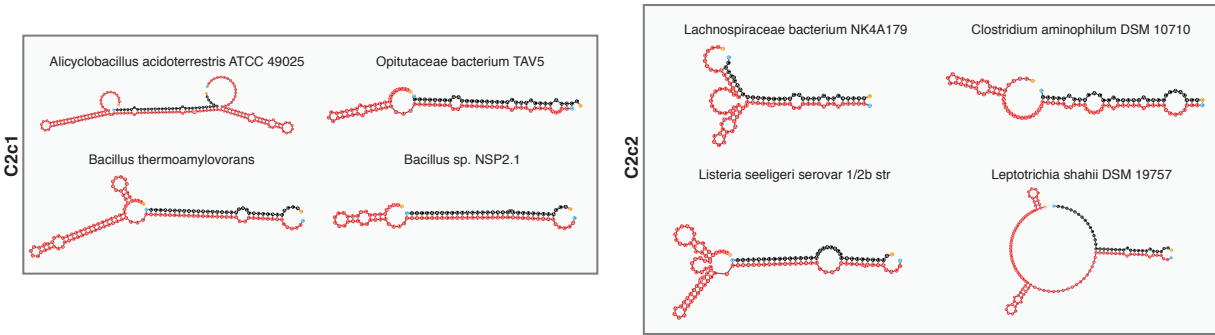
```

Figure S4. Multiple alignment of C2c1 protein family.

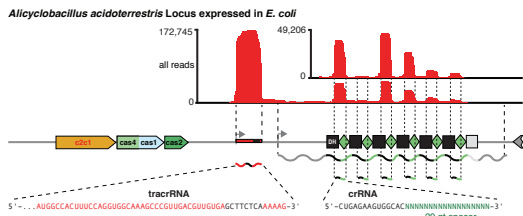
The alignment was built using MUSCLE program and modified manually on the basis of local PSI-BLAST pairwise alignments. Each sequence is labelled with GenBank Identifier (GI) number and systematic name of an organism. Secondary structure was predicted by Jpred and shown underneath the sequence which was used as a query (designations: H- alpha helix, E – beta strand). CONSENSUS was calculated for each alignment column by scaling the sum-of-pairs score within the column between those of a homogeneous column (the same residue in all aligned sequences) and a random column with homogeneity cutoff 0.8. Active site motifs of RuvC-like domain are shown below alignment.

FIGURE S7 (Related to Figures 3 and 4)

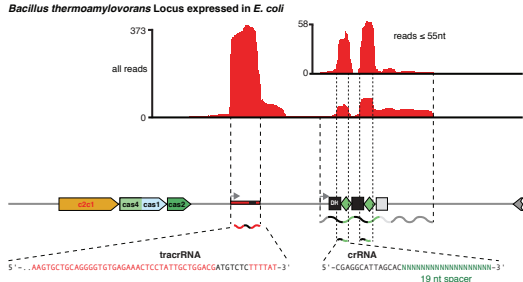
A



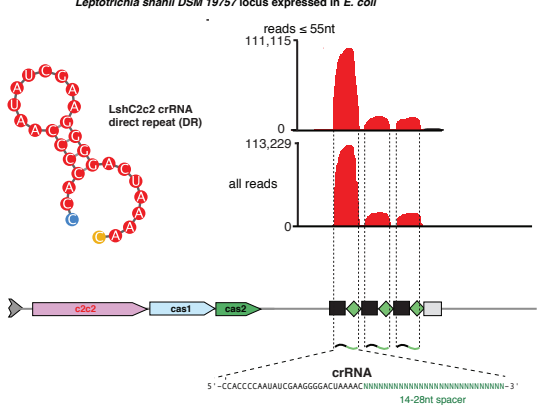
B



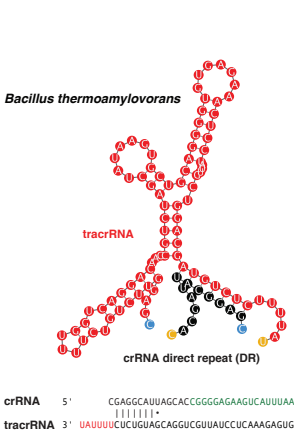
C



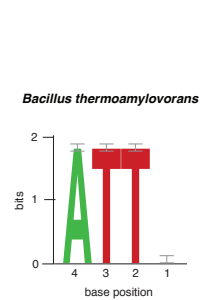
F



D



E



G

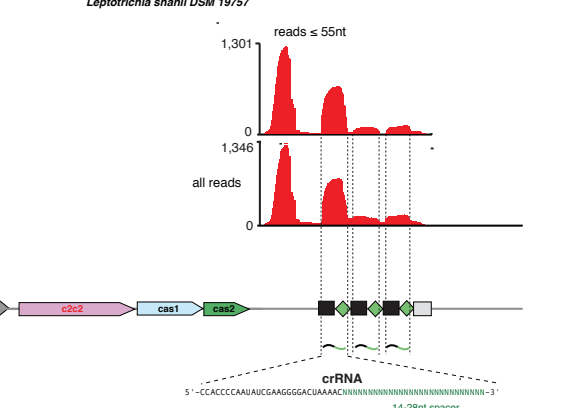


Figure S7. Additional functional validation off type V-B (C2c1 loci) and type VI (C2c2 loci) CRISPR-Cas systems.

(A) Predicted structures of tracrRNAs base-paired with the repeats. TracrRNA for *Alicyclobacillus acidoterrestris* was identified using RNAseq. For the remaining loci, putative tracrRNAs were identified based on presence of an anti-direct repeat (DR) sequence. Anti-DRs were identified using Geneious (www.geneious.com) by searching for sequences within each respective CRISPR locus that are highly homologous to DR. The 5' and 3' ends of each putative tracrRNA was determined through computational prediction of bacterial transcription start and termination sites using BPROM (www.softberry.com) and ARNOLD (rna.igmors.u-psud.fr/toolbox/arnold/) respectively. Co-folding predictions were generated using Geneious. 5' ends are colored blue and 3' ends are colored orange.

(B) Heterologous expression of the *Alicyclobacillus acidoterrestris* C2c1 locus in pACYC-184 transformed into *E. coli* shows identical results to the expression observed in the endogenous strain (Fig. 4A). Processed crRNAs have a 5' 14-nt DR and 20-nt spacer and a putative 79-nt tracrRNA is expressed robustly.

(C) The *Bacillus thermoamylovorans* locus was heterologously expressed in *E. coli*. The putative tracrRNA is significantly expressed and is processed to 91 nt. Processed crRNAs are also present with a 5' 14 nt DR and 19nt spacer.

(D) In silico co-folding of the crRNA direct repeat and putative tracrRNA shows stable secondary structure and complementarity between the two RNAs. 5' bases are colored blue and 3' bases are colored orange.

(E) Depletion from the 5' left PAM library reveals a 5' ATTN PAM. Depletion is measured as the negative log₂ fold ratio and PAMs above a threshold of 3.5 are used to calculate the entropy score at each position.

(F) RNA-sequencing of the *Leptotrichia shahii* DSM 19757 locus expressed in *E. coli* shows processing of the CRISPR array in the 3' to 5' direction (direction of the locus). crRNAs are processed to have a 5' DR that is 28nt in length and spacers with lengths 14-28nt. *Inset:* In silico folding of the *L. shahii* crRNA DR predicts stable secondary structure.

(G) RNA-sequencing of the endogenous *Leptotrichia shahii* DSM 19757 C2c2 locus shows similar results to (A).

Table S1 (related to Figure 1)

The discovered Class 2 CRISPR-Cas loci. Please see accompanying Excel spreadsheet.

SUPPLEMENTAL EXPERIMENTAL PROCEDURES

Computational sequence analysis

The multiple alignments of the C2c1, C2c2, C2c3 and Cas1 proteins in the Fasta format and the complete phylogenetic tree for Cas1 proteins in the Newick format are available at ftp://ftp.ncbi.nih.gov/pub/wolf/_suppl/Class2/.

Growth of Bacterial Strains

For RNA-sequencing, *Alicyclobacillus acidoterrestris*(ATCC) was cultured using ATCC Medium 1655 in suspension at 50°C and 300 rpm. *E. coli* containing heterologous constructs were cultured in Luria broth supplemented with appropriate antibiotics in suspension at 37°C and 300rpm. Both strains were grown in aerobic conditions and harvested in stationary growth phase.

For the PAM screen, *E. coli* were grown on LB-agar plates supplemented with ampicillin and the appropriate antibiotics (chloramphenicol or kanamycin) for maintenance of the C2c1 locus plasmid at 37°C.

Generation of heterologous plasmids

To generate the AacC2c1 locus for heterologous expression, genomic DNA was purified from *Alicyclobacillus acidoterrestris* using the Qiagen DNeasy Blood and Tissue kit and the C2c1 locus was PCR amplified for cloning into pACYC-184. The BthC2c1 and LseC2c2 locus was synthesized by Genscript into a pET-28 vector. Cells harboring plasmids were made competent using the Z-competent kit (Zymo).

RNA extraction and Northern blotting

The procedure was performed essentially as described previously (Pougach and Severinov, 2012). *E. coli* BL21 AI cells were transformed with plasmids pACYCduet-1 containing, under inducible T7 promoters, *A. ac.* or *L. sh.* cas operons and plasmids pCDF-1b containing the respective minimal CRISPR cassettes with single spacers. Total RNA was extracted from 5 mL of *E. coli* cells induced with 1 mM arabinose/0.2 mM IPTG and grown until OD₆₀₀ 0.8 – 1.0. The cells were lysed by 5-minute treatment using Max Bacterial Enhancement Reagent followed by RNA purification with the TRIzol reagent (Thermo Fisher Scientific). 15 mg of total RNA were

separated on a denaturing 8 M urea - 12% polyacrylamide gel and electrophoretically transferred to Hybond-XL membrane (GE Healthcare) using a Mini Trans-Blot Electrophoretic Transfer Cell (Bio-Rad). The membrane was dried and then UV cross-linked. ExpHyb hybridization solution (Clontech) was used for hybridization according to manufacturer's instructions for 1 hour at 40 °C with ³²P-end labeled oligonucleotide probes.

Preparation of C2c1 Protein Lysate

C2c1 proteins codon optimized for human expression and designed with an N-terminal nuclear localization tag. The protein sequences were synthesized and cloned into the pcDNA3.1 expression plasmid by Genscript. Using Lipofectamine 2000 reagent (Life Technologies), 2,000ng of the expression plasmids were transfected into HEK293FT cells (6-well plates at 90% confluence). After 48 hours, cells were washed with DPBS (Left Technologies), lysed using lysis buffer [20mM Hepes pH 7.5, 100mM KCl, 5mM MgCl₂, 1 mM DTT, 5% glycerol, 0.1% Triton X-100, 1X cComplete Protease Inhibitor Cocktail Tablets (Roche)]. After sonicating for 10 minutes in a Biorupter sonicators (Diagenod) (50% duty cycle on high), the lysate was centrifuged and the supernatant was frozen at -80°C for later use in cleavage assays.

Primers used in this work

Primer Description	Sequence (5' to 3')
<i>PAM library generation</i>	
Oligo for 5' AacC2c1 PAM library (forward)	ggccagtgaattcgagctcggtagccgggNNNNNNNtgttggtaaagg taaaaagacgaatgatgcatccagcttggcgtaatcatggatagctg tt
Oligo for 3' AacC2c1 PAM library (forward)	ggccagtgaattcgagctcggtagccgggtgttggtaaaggtaaaaag acgaatgatgcatccNNNNNNNagcttggcgtaatcatggatagctg tt
Oligo for 5' BsC2c1 PAM library (forward)	ggccagtgaattcgagctcggtagccgggNNNNNNNcggggagaagtca ttaaataaggccactgttaaaaagcttggcgtaatcatggatagctg tt
Oligo for 3' BsC2c1 PAM library (forward)	ggccagtgaattcgagctcggtagccggggagaagtcatttaaataaggc cactgttaaaaNNNNNNNNagcttggcgtaatcatggatagctgttt
Oligo for reverse strand synthesis of PAM library (reverse)	AAACAGCTATGACCATGATTACGCCAA
<i>Northern Blot Probes</i>	

Aac_tracrRNA_probe1	tctttttgagaagctcaacggg
Aac_crRNA_probe2	cctttaccaaacagtgccacttctcagatcgctc
Aac_crRNA_probe3	tcagtgatccgacggatgcatcattcgctttttacc
Lsh_crRNA_probe1	ggtagaacagcaatctagtttttagtccccttcgat
Lsh_crRNA_probe2	attgggggtggtctatatcaatatggattacttggg
Lsh_crRNA_probe3	atcgaaggggactaaaactagattgctggttctacc
Lsh_crRNA_probe4	accaagtaatccatattgatatagaccacccaat
<i>Cloning</i>	
Aac_cas_Nco_d	acgtccatggcgtcaaataccatcaaagtg
Aac_cas_Kpn_r	cgtaggtacctcaaaataccaatcccttcaaataccaatgg
Aac_cas_Nde_d	cagctggctcatatgcaccgacg
Aac_cas_Nde_r	cgtcggtgcatatgagccagctg
AacC2c1 PAM Validation Target TTT Forward	ctagTTttgtttggttaaaggtaaaaagacgaatgatgcatcc
AacC2c1 PAM Validation Target TTA Forward	ctagTtatgtttggttaaaggtaaaaagacgaatgatgcatcc
AacC2c1 PAM Validation Target TTC Forward	ctagTTctgtttggttaaaggtaaaaagacgaatgatgcatcc
AacC2c1 PAM Validation Target TTG Forward	ctagTTgtgtttggttaaaggtaaaaagacgaatgatgcatcc
AacC2c1 PAM Validation Target GGA Forward	ctagGgatgtttggttaaaggtaaaaagacgaatgatgcatcc
AacC2c1 PAM Validation Target TTT Reverse	agctggatgcatcattcgctctttttacctttaccaaacaaaa
AacC2c1 PAM Validation Target TTA Reverse	agctggatgcatcattcgctctttttacctttaccaaacataa
AacC2c1 PAM Validation Target TTC Reverse	agctggatgcatcattcgctctttttacctttaccaaacagaa
AacC2c1 PAM Validation Target TTG Reverse	agctggatgcatcattcgctctttttacctttaccaaacacaa
AacC2c1 PAM Validation Target GGA Reverse	agctggatgcatcattcgctctttttacctttaccaaacatcc

<i>crRNA/tracrRNA IVT</i>	
AacC2c1 crRNA targeting EMX1 target 1	gaaatTAATACGACTCACTATAGGGCGAGCGATCTGAGAAGTGGCACCA GAACCGGAGGACAAAGTA
AacC2c1 crRNA targeting AacC2c1 spacer 1	gaaatTAATACGACTCACTATAGGGCGAGCGATCTGAGAAGTGGCACTG TTTGGTAAAGGTAAAAAG
AacC2c1 tracrRNA 151nt	GTCTAGAGGACAGAATTTTTCAACGGGTGTGCCAATGGCCACTTTCCAG GTGGCAAAGCCCCTTGAGCTTCTCAAAAAGAACGCTCGCTCAGTGTCT GACCTTTTCGAGCGCCTGTTTCAGGGCGAAAACCTGGGAGGCGCTCGAAT CATA
AacC2c1 tracrRNA 96nt	GTCTAGAGGACAGAATTTTTCAACGGGTGTGCCAATGGCCACTTTCCAG GTGGCAAAGCCCCTTGAGCTTCTCAAAAAGAACGCTCGCTCAGTGT
AacC2c1 tracrRNA 90nt	GTCTAGAGGACAGAATTTTTCAACGGGTGTGCCAATGGCCACTTTCCAG GTGGCAAAGCCCCTTGAGCTTCTCAAAAAGAACGCTCGCTC
AacC2c1 tracrRNA 78nt	GTCTAGAGGACAGAATTTTTCAACGGGTGTGCCAATGGCCACTTTCCAG GTGGCAAAGCCCCTTGAGCTTCTCAAAA
AacC2c1 tracrRNA 67nt	GTCTAGAGGACAGAATTTTTCAACGGGTGTGCCAATGGCCACTTTCCAG GTGGCAAAGCCCCTTGAG
AacC2c1 sgRNA targeting EMX1 target 1	gaaatTAATACGACTCACTATAGGGGTCTAGAGGACAGAATTTTTCAAC GGGTGTGCCAATGGCCACTTTCCAGGTGGCAAAGCCCCTTGAGCTTCTC AAATCTGAGAAGTGGCACCAGAACCGGAGGACAAAGTA
AacC2c1 sgRNA targeting AacC2c1 spacer 1	gaaatTAATACGACTCACTATAGGGGTCTAGAGGACAGAATTTTTCAAC GGGTGTGCCAATGGCCACTTTCCAGGTGGCAAAGCCCCTTGAGCTTCTC AAATCTGAGAAGTGGCACTGTTTGGTAAAGGTAAAAAG

Plasmid and codon-optimized C2c1/C2c2 protein-coding sequences used in this work

Constructs used for RNA-sequencing, Bacterial Interference, and Lysate Cleavage	
<i>C2c1 locus constructs</i>	<i>Permanent Sequence Links</i>
AacC2c1 locus cloned in pACYC184	https://benchling.com/s/YfJMfbu2/edit
BtC2c1 locus in pET28a	https://benchling.com/s/7SJzEUPG/edit
<i>C2c2 locus constructs</i>	
LshC2c2 locus cloned in pACYC184	https://benchling.com/s/gco8u7H8/edit
LseC2c2 locus cloned in pET28a	https://benchling.com/s/3NojkRem/edit

<i>C2c1 human codon optimized constructs</i>	
AacC2c1 Human Codon Optimized Protein	https://benchling.com/s/WivDaYfG/edit
Constructs used for Northern Blotting	
AacC2c1 and Cas genes in pACYCduet	https://benchling.com/s/wgbYgHeZ/edit
AacC2c1 CRISPR array in pCDF	https://benchling.com/s/kjfElooT/edit
LshC2c2 and Cas genes in pACYCduet	https://benchling.com/s/mFzf1E7u/edit
LshC2c2 CRISPR array in pCDF	https://benchling.com/s/v26xY1KC/edit

SUPPLEMENTAL REFERENCES

Pougach, K., and Severinov, K. (2012). Use of semi-quantitative Northern blot analysis to determine relative quantities of bacterial CRISPR transcripts. *Methods Mol Biol* 905, 73-86.