The PARP inhibitor olaparib enhances the sensitivity of Ewing sarcoma to trabectedin

Supplementary Material



Supplementary Figure S1: (A) IC50 of proliferation after treatment with Olaparib: 72 hours versus 6 days. (B) Grafic representation of IC50 of ES cell lines after treatment with Olaparib with regard to ES cell lines1q status. (C) Grafic representation of IC50 of ES cell lines with regard to their p53 status.



Supplementary Figure S2: (A). Graphics represent data obtained in the apoptosis assay determined by caspase 3 and 7 DEVDase activity. RLU stands for Relative Light Units. Treatment with Olaparib increased apoptosis as compared to the controls (141% in RM82 and 174% in TC71 cell lines). Treatment with Trabectedin elicited a higher apoptosis induction (240% and 242% in RM82 and TC71 cell lines respectively). The combination of Olaparib and Trabectedin in TC71 cells induced an increase in apoptosis as compared with the effects induced by drugs alone (299%). (B) PARP cleavage was evaluated by Western Blot in TC71 cells after treatment with the drugs alone/combined in short (8 hours) and longer (24 hours) administration. The combination induced an increase in PARP cleavage, especially when compared to Trabectedin. (C) Graphics representing data from cell cycle analysis after 24 hours of treatment in the RM82 and TC71 cell lines. Trabectedin induced S-phase accumulation, whereas Olaparib induced strong G2/M accumulation. The combination induced accumulation in the S-phase and G2/M and a significant decrease or even the abrogation of G0/G1 phase. In all cases, combination refers to 250pM Trabectedin and 5 μ M Olaparib. OLA refers to Olaparib.



Supplementary Figure S3: Migration assays performed by wound healing in the RM82 cell line after treatment. No differences in cell migration were observed after drug exposure at 24, 48, and 72 hours as compared to the controls. Combination stands for 250pM Trabectedin and 5μ M Olaparib.



Supplementary Figure S4: Quantification of p-H2AX foci in the TC71 cell line after treatment. The number of pH2AX foci per cell and the total number of foci as measured by CTCF were higher in cells treated with the combination of drugs. CTCF stands for Corrected Total Cell Fluorescence, normalized to background subtraction. Trabectedin (250pM), Olaparib (5 μ M). Combination refers to 250pM Trabectedin and 5 μ M Olaparib. * p<0,05 Student's T-Test.



Supplementary figure S5: A) EWSR1-FLI1 target expression after treatments in the TC71 cell line. The expression of EWSR1-FLI1 targets was not affected after treatment with the drug combination. Only *NKX2.2*, a known upregulated indirect target of EWSR1-FLI, was slightly down-regulated after exposure to the combination of drugs. Combination refers to 250pM Trabectedin and 5μ M Olaparib. B) Quantification of DNA damage induction shown in figure 3 C using Image J software. Relative intensity is shown as compared to control condition (DMSO).



Supplementary Figure S6: IPA analysis in the condition of Trabectedin alone.

Nro11		
Multit	Molecules	Exp. Value
Złęcce 6	EXO1	1.927
RADOR	BRIP1	1.904
XPC RA050	TOP3A	1.903
MSH2	RAD54L	1.886
ERCC6 PRKDC MSH6	RFC1	1.700
	PARP3	1.625
ERGES CORT	RAD51B	1.613
BRCAT	XRCC1	1.609
	POLD3	1.597
	RAD18	1.550
CCNH mediator		
	Molecules	Exp. Value
	RAD23B	-1.302
BRCA2 MMMS19	NEIL2	-1.089
ERCES	NEIL1	-1.046
KPA ATT PUST	DMC1	-1.044
	ATXN3	-1.044
ERCC4	NTHL1	-1.037
	XPC	-1.027
RPAT	PARP1	-1.017
	ХРА	-1.005
	PMS1	-1.003

Supplementary Figure S7: IPA analysis in the condition of Olaparib alone.



Supplementary Figure S8: Quantification of the histological findings found in tumors treated in the *in vivo* study. (A) Tumors treated with Trabectedin or Olaparib, and especially with the combination of both drugs, showed a higher % of necrosis. (B) Tumors treated with Trabectedin or combinations showed a smaller % of proliferating cells. (C) Tumors treated with combinations showed a higher number of pH2AX-positive cells. (D) Tumors treated with combinations or Trabectedin showed a higher number of BRCA2-positive cells. Student's T-test *p<0.05. Combination 1 refers to 100mg/kg Olaparib and 0.15mg/kg Trabectedin, which started to be administered 7 days after the start of administration of Olaparib. Combination 2 refers to 100mg/Kg Olaparib and 0.15mg/kg Trabectedin. The Dotslide analysis program (Olympus) was used to quantify pH2AX-, BRCA2- and Ki67-positive cells.



Supplementary Figure S9: γ H2AX immunostaining performed in tumor sections derived from the *in vivo* study. Comb 1 refers to Combination1. Comb 2 refers to Combination2



Supplementary Figure S10: BRCA2 immunostaining performed in tumor sections derived from the *in vivo* study. Comb 1 refers to Combination1. Comb 2 refers to Combination2.

Cell line	Pathology	Fusion type	<i>TP 53</i> status	1q Status
RDES	Ewing Sarcoma	EWSR1-FLI 1, type 2	arg273stop	Gained
WE68	Ewing Sarcoma	EWSR1-FLI 1, type 1	Wild type	Gained
STAET- 2.1	Ewing Sarcoma	EWSR1-FLI 1, type 3	cys277tyr	Normal
RM82	Ewing Sarcoma	EWSR1-ERG	arg273his	Gained
TC71	Ewing Sarcoma	EWSR1-FLI 1, type 1	arg213stop	Gained
SK-N-MC	Ewing Sarcoma	EWSR1-FLI 1, type 1	del EX2-4	Normal
CADO	Ewing Sarcoma	EWSR1-ERG	Wild type	Gained
A673	Ewing Sarcoma	EWSR1-FLI 1, type 1	2 bp ins.codon 119	Normal
A4573	Ewing Sarcoma	EWSR1-FLI 1, type 3	-	Gained
STAET -1	Ewing Sarcoma	EWSR1-FLI 1, type 1	Wild type	Normal
SJRH	Rhabdomyosarcoma	-	-	-

Supplementary Table S1. Molecular characteristics of ES cell lines.

Supplementary Table S2. Expression changes in DNA damage repair genes in ES cells from the TC71 cell line. The table depicts the most significantly up/down-expressed genes in each treatment condition. Numerical data refer to levels of expression in comparison with the controls.

Up-Regulated							
Trabectedin	Exp value	Olaparib	Exp value	Combination	Exp value		
BRCA1	1.644	EXO1	1.927	RAD54L	2.548		
POLD3	1.394	BRIP1	1.904	ТОРЗА	2.501		
BRIP1	1.382	TOP3A	1.903	EXO1	2.466		
RAD18	1.347	RAD54L	1.886	BRIP1	2.338		
NEIL3	1.346	RFC1	1.700	POLD3	2.160		
RFC1	1.247	PARP3	1.625	RFC1	2.159		
OGG1	1.229	RAD51B	1.613	PARP2	1.899		
RAD52	1.205	XRCC1	1.609	BRCA1	1.869		
XRCC1	1.166	POLD3	1.597	RAD18	1.819		
UNG	1.160	RAD18	1.550	XRCC1	1.810		

Down-Regulated

Trabectedin	Exp value	Olaparib	Exp value	Combination	Exp value
PMS2	-625.000	RAD23B	-1.302	MSH4	-3.358
RAD51B	-44.444	NEIL2	-1.089	XRCC4	-2.477
RAD51	-10.846	NEIL1	-1.046	RAD51B	-1.892
MSH5	-8.795	DMC1	-1.044	MGMT	-1.873
ТОРЗВ	-8.532	ATXN3	-1.044	RAD23B	-1.423
PMS1	-8.019	NTHL1	-1.037	ATXN3	-1.294
LIG1	-5.952	XPC	-1.027	SMUG1	-1.193
XPA	-4.653	PARP1	-1.017	NEIL2	-1.159
FEN1	-4.137	XPA	-1.005	APEX1	-1.153
XRCC6	-4.050	PMS1	-1.003	NTHL1	-1.143

		Fold Change		ange	
		(comparing to control group)		ontrol group)	Gene Description
Symbo	Unige	Trabect	Olap	Combinati	
Ī	ne	edin	arib	on	
	Ho 72		1 01		ADEX puelesses (multifunctional DNA repair
	HS.73	0.04	1.01	0.0070	APEX nuclease (multifunctional DNA repair
APEAT	122	0.04	o	0.0072	enzyme) i
	Hs 65		1 70		
APEX2	9558	0 7434	92	1 8451	endonuclease) 2
	0000	011 101	02	novor	
	Hs.36		1.18		
ATM	7437	0.8223	76	1.069	Ataxia telangiectasia mutated
	Hs.27		1.34		
ATR	1791	0.9984	14	1.3968	Ataxia telangiectasia and Rad3 related
	11 50		0.05		
	Hs.53	0 7057	0.95	0 7700	
ATXN3	2632	0.7057	11	0.7729	Ataxin 3
	He 10		1 49		
BRCA1	/1/3	1 6442	13	1 8691	Breast cancer 1 early onset
BROAT	115	1.0442	10	1.0001	Dreast cancer 1, carry onset
	Hs.34		1.39		
BRCA2	012	1.0234	9	1.449	Breast cancer 2, early onset
					· · ·
	Hs.53		1.90		BRCA1 interacting protein C-terminal helicase
BRIP1	2799	1.3821	45	2.3385	1
			4 4 4		
	HS.29	0 0000	1.14	1 2009	Cycolin H
CONH	2524	0.9000	43	1.2090	Сусшін
	Hs.30		1.28		
CCNO	41	0.6936	28	1.4174	Cvclin O
	Hs.18		1.38		
CDK7	4298	0.3131	44	1.6298	Cyclin-dependent kinase 7
	11 00		4.05		
0004	HS.29	0.0044	1.05	4 4 4 7 4	Damage-specific DNA binding protein 1,
DDRJ	0758	0.3241	67	1.1474	127KDa
	Hs 70		1 26		Damage-specific DNA binding protein 2
DDB2	0338	0 7444	94	1 4446	48kDa
	0000	•	•		londu
					DMC1 dosage suppressor of mck1 homolog,
	Hs.33		0.95		meiosis-specific homologous recombination
DMC1	9396	0.5471	77	1.4324	(yeast)
					Excision repair cross-complementing rodent
	Hs.43	0.5300	1.14	4 0 4 5 0	repair deficiency, complementation group 1
ERCC1	5981	0.5789	04	1.2456	(includes overlapping antisense sequence)
FRCC2		0 859		1 0561	
211002	Hs.48	0.000	1.08	1.0001	Excision repair cross-complementing rodent

Supplementary Table S3. Expression changes in DNA damage repair genes in ES cells from the TC71 cell line. Data compared to the control (condition treated with DMSO).

	7294		32		repair deficiency, complementation group 2
ERCC3	Hs.46 9872	0.9135	1.14 49	1.1692	Excision repair cross-complementing rodent repair deficiency, complementation group 3 (xeroderma pigmentosum group B complementing)
ERCC4	Hs.56 7265	1.079	1.21 99	1.2171	Excision repair cross-complementing rodent repair deficiency, complementation group 4
ERCC5	Hs.25 8429	1.0688	1.13 8	1.3476	Excision repair cross-complementing rodent repair deficiency, complementation group 5
ERCC6	Hs.65 4449	0.7418	1.35 66	1.1502	Excision repair cross-complementing rodent repair deficiency, complementation group 6
ERCC8	Hs.43 5237	0.703	1.05 84	0.9856	Excision repair cross-complementing rodent repair deficiency, complementation group 8
EXO1	Hs.49 8248	0.6029	1.92 73	2.4661	Exonuclease 1
FEN1	Hs.40 9065	0.2417	1.28 77	1.7279	Flap structure-specific endonuclease 1
LIG1	Hs.17 70	0.168	1.02 5	1.1166	Ligase I, DNA, ATP-dependent
LIG3	Hs.10 0299	0.5081	1.25 77	0.9574	Ligase III, DNA, ATP-dependent
LIG4	Hs.16 6091	0.8789	1.49 08	1.6593	Ligase IV, DNA, ATP-dependent
MGMT	Hs.50 1522	0.568	1.05 37	0.534	O-6-methylguanine-DNA methyltransferase
MLH1	Hs.19 5364	1.0108	1.04 15	0.9544	MutL homolog 1, colon cancer, nonpolyposis type 2 (E. coli)
MLH3	Hs.43 6650	1.0298	1.48 32	1.2214	MutL homolog 3 (E. coli)
MMS19	Hs.50 0721	0.8353	1.22 65	1.2222	MMS19 nucleotide excision repair homolog (S. cerevisiae)
MPG	Hs.45 9596	0.8467	1.19 12	1.0777	N-methylpurine-DNA glycosylase
MRE11 A	Hs.19 2649	0.8344	1.08 33	1.236	MRE11 meiotic recombination 11 homolog A (S. cerevisiae)
MSH2	Hs.59 7656	0.7699	1.23 99	1.4563	MutS homolog 2, colon cancer, nonpolyposis type 1 (E. coli)
MSH3	Hs.28 0987	0.321	1.31 93	0.9042	MutS homolog 3 (E. coli)
MSH4	Hs.21 6639	0.3384	1.23 4	0.2978	MutS homolog 4 (E. coli)

MSH5	Hs.64 7011	0.1137	1.49 45	1.4797	MutS homolog 5 (E. coli)
MSH6	Hs.44 5052	0.7723	1.04 11	1.1494	MutS homolog 6 (E. coli)
MUTY H	Hs.27 1353	0.7336	1.15 51	1.1608	MutY homolog (E. coli)
NEIL1	Hs.51 2732	0.816	0.95 57	0.8762	Nei endonuclease VIII-like 1 (E. coli)
NEIL2	Hs.29 3818	0.6822	0.91 84	0.8625	Nei endonuclease VIII-like 2 (E. coli)
NEIL3	Hs.40 5467	1.3464	1.11 8	1.2745	Nei endonuclease VIII-like 3 (E. coli)
NTHL1	Hs.66 196	0.3407	0.96 4	0.875	Nth endonuclease III-like 1 (E. coli)
OGG1	Hs.38 0271	1.2294	1.20 9	1.0776	8-oxoguanine DNA glycosylase
PARP1	Hs.17 7766	0.5906	0.98 34	0.9337	Poly (ADP-ribose) polymerase 1
PARP2	Hs.40 9412	0.711	1.16 88	1.8986	Poly (ADP-ribose) polymerase 2
PARP3	Hs.27 1742	0.4062	1.62 48	1.4864	Poly (ADP-ribose) polymerase family, member 3
PMS1	Hs.11 1749	0.1247	0.99 75	0.9253	PMS1 postmeiotic segregation increased 1 (S. cerevisiae)
PMS2	Hs.63 2637	0.0016	1.07 41	1.0127	PMS2 postmeiotic segregation increased 2 (S. cerevisiae)
PNKP	Hs.78 016	0.9887	1.31 08	1.3773	Polynucleotide kinase 3'-phosphatase
POLB	Hs.65 4484	1.123	1.15 87	1.3479	Polymerase (DNA directed), beta
POLD3	Hs.82 502	1.3943	1.59 69	2.1602	Polymerase (DNA-directed), delta 3, accessory subunit
POLL	Hs.52 3230	0.2838	1.14 24	1.3329	Polymerase (DNA directed), lambda
PRKD C	Hs.49 1682	1.1391	1.32 54	1.1871	Protein kinase, DNA-activated, catalytic polypeptide
RAD18	Hs.37 5684	1.3467	1.55 02	1.8189	RAD18 homolog (S. cerevisiae)
RAD21	Hs.81 848	0.7353	1.02 63	0.9866	RAD21 homolog (S. pombe)
RAD23	Hs.64	0.4415	1.12	1.3637	RAD23 homolog A (S. cerevisiae)

Α	3267		59		
RAD23	Hs 52		0.76		
B	1640	0.4044	79	0.7029	RAD23 homolog B (S. cerevisiae)
_				••=•	·
	Hs.65		1.34		
RAD50	5835	0.5595	4	1.1334	RAD50 homolog (S. cerevisiae)
	Hs.63		1.03		
RAD51	1709	0.0922	08	1.2539	RAD51 homolog (S. cerevisiae)
DADEA	11 47		4.04		
RAD51	HS.17	0 0005	1.61	0 5000	
В	2587	0.0225	29	0.5286	RAD51 nomolog B (S. cerevisiae)
	Нс /1		1 1 3		
C	2587	0 5584	62	1 1026	RAD51 homolog C (S. cerevisiae)
Ŭ	2007	0.0004	02	111020	
RAD51	Hs.63		1.11		
D	1757	0.9553	58	0.9528	RAD51 homolog D (S. cerevisiae)
					,
	Hs.70		1.25		
RAD52	9202	1.2049	17	1.4908	RAD52 homolog (S. cerevisiae)
			1.00		
RAD54	HS.64	0.0544	1.88	0 5 4 7 0	
L	2042	0.6544	58	2.5478	RAD54-IIKe (S. cerevisiae)
	He 50		1 70		
RFC1	7475	1 2467	01	2 1594	Replication factor C (activator 1) 1 145kDa
	1415	1.2407	01	2.1334	
	Hs.46		1.01		
RPA1	1925	0.8569	46	1.0292	Replication protein A1, 70kDa
	Hs.48		1.02		
RPA3	7540	0.6118	5	1.1033	Replication protein A3, 14kDa
			1 1 5		
SIK	1022	0 /088	78	1 2302	STE20-like kinase
OLIX	1522	0.4000	10	1.2502	OT EZO IIKE KIIdase
SMUG	Hs.63		1.13		Single-strand-selective monofunctional uracil-
1	2721	0.463	86	0.8381	DNA glycosylase 1
	Hs.58		1.09		
TDG	4809	0.8917	34	1.1476	Thymine-DNA glycosylase
			1 00		
TOD2A	HS.39	0 2001	1.90	2 5012	Tanaiaamaraaa (DNA) III alaba
TUFJA	2115	0.3331	21	2.5015	Topolsomerase (DNA) in alpha
	Hs.43		1.39		
TOP3B	6401	0.1172	49	1.5978	Topoisomerase (DNA) III beta
				_	· · · · · · · · · · · · · · · · · · ·
	Hs.70		1.49		
TREX1	7026	0.8717	47	1.6449	Three prime repair exonuclease 1
			4.40		
	⊓S.19 1224	1 1605	61.10 64	1 5070	Uracil DNA ducaculase
DNO	1334	1.1005	04	1.5270	UTACII-DINA GIYCOSYIASE
	Hs.98		1.32		
XAB2	22	1.126	67	1.6688	XPA binding protein 2
	Hs.65		0.99		Xeroderma pigmentosum, complementation
XPA	4364	0.2149	54	1.1093	group A

ХРС	Hs.47 5538	0.4052	0.97 39	0.973	Xeroderma pigmentosum, complementation group C
XRCC1	Hs.98 493	1.1662	1.60 94	1.8095	X-ray repair complementing defective repair in Chinese hamster cells 1
XRCC2	Hs.64 7093	0.9138	1.38 56	1.2881	X-ray repair complementing defective repair in Chinese hamster cells 2
XRCC3	Hs.59 2325	0.3498	1.03 26	1.143	X-ray repair complementing defective repair in Chinese hamster cells 3
XRCC4	Hs.56 7359	0.33	1.04 01X-ray repair complementing defective rep010.4037Chinese hamster cells 4		X-ray repair complementing defective repair in Chinese hamster cells 4
XRCC5	Hs.38 8739	0.5948	1.20 94	1.1488	X-ray repair complementing defective repair in Chinese hamster cells 5 (double-strand-break rejoining)
XRCC6	Hs.29 2493	0.2469	1.14 74	1.085	X-ray repair complementing defective repair in Chinese hamster cells 6
XRCC6 BP1	Hs.61 188	0.2786	1.25 61	1.286	XRCC6 binding protein 1

Supplementary Table S4. Top pathways covered by the array. Major pathways affected after treatment with the combination of Olaparib and Trabectedin in ES cells from the TC71 cell line are involved in the repair of DSB. The ratio indicates the number of affected genes of a pathway/total number of genes of a pathway. The p-value is an estimation of the likelihood that the correlation between the number of affected genes in our study that participate in a pathway and the total number of genes that are known to be associated with that pathway will be due to random association. p-values<0.05 indicate statistically significant differences.

Pathways	Ratio	p-value
NER	14/35 (0.4)	1.02E-23
NHEJ	11/16 (0.688)	1.81E-23
HR	9/16 (0.562)	4.41E-18
BRCA1	13/63 (0.206)	4.52E-18
HBCS	15/119 (0.126)	2.24E-17