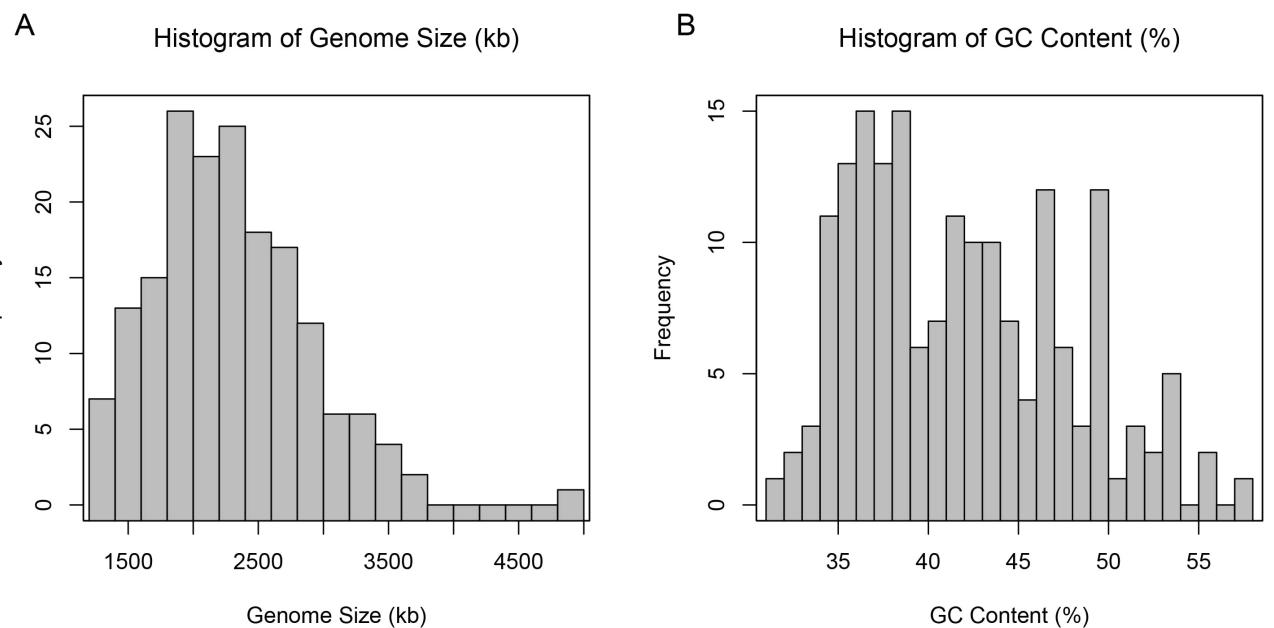
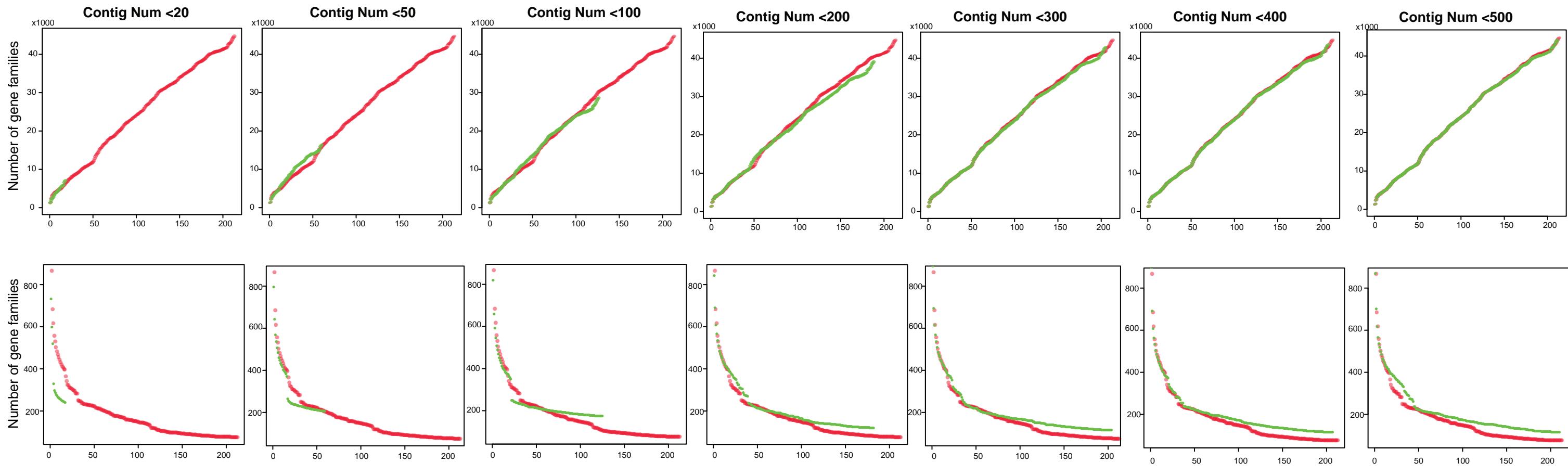


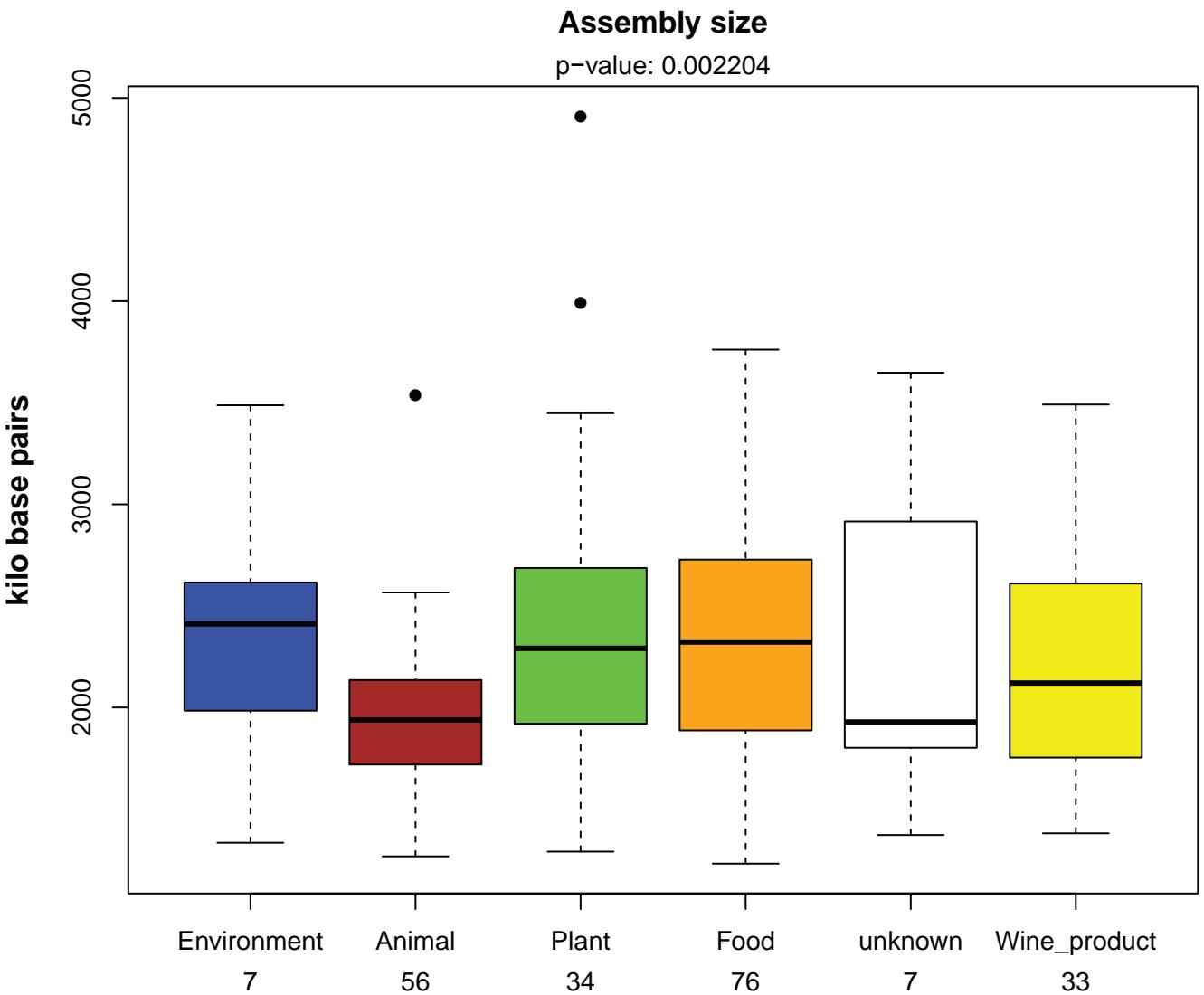
Supplementary Figures



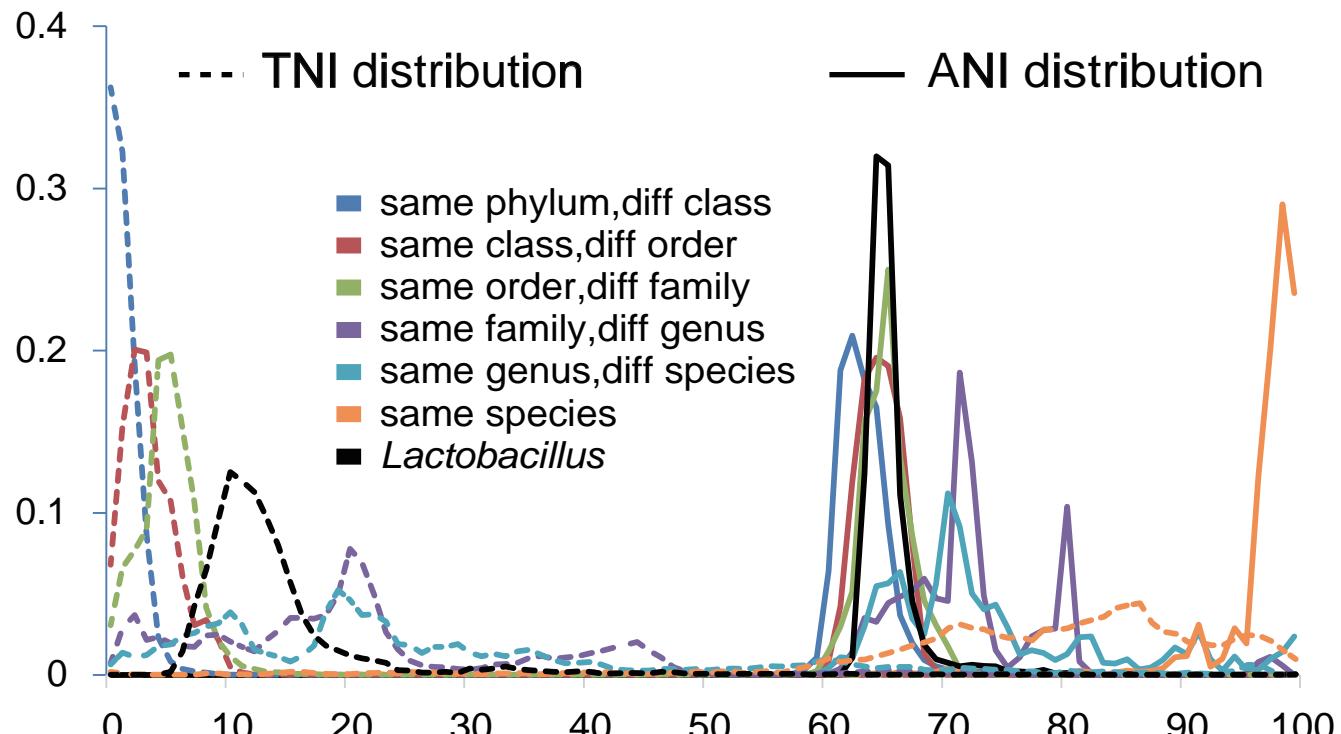
Supp. Figure 1. Histograms of genome size distribution (A) and GC% (B) for 175 *Lactobacillus* genomes.



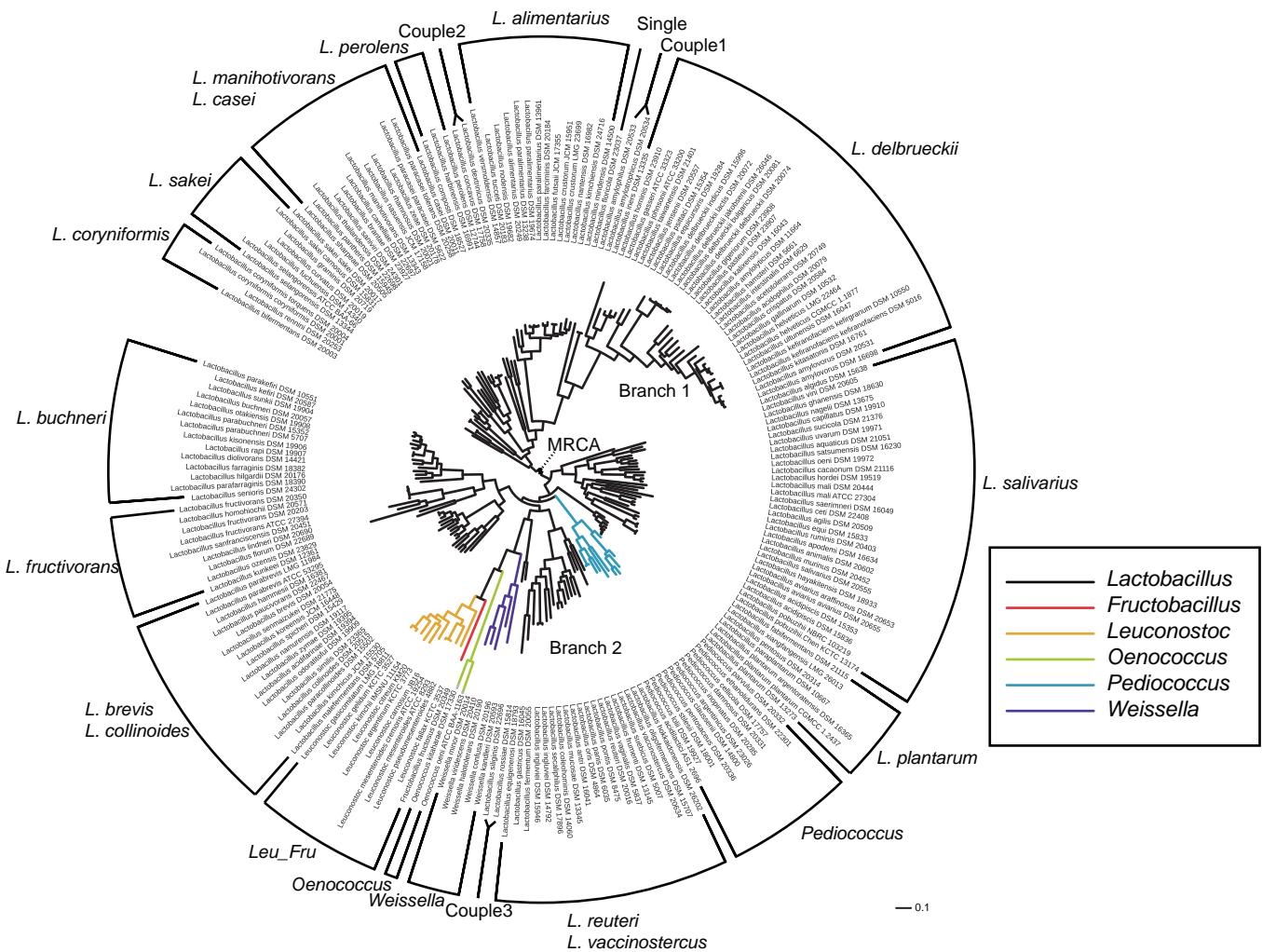
Supp. Figure 2. Sizes of the pan-genome (top) and core-genomes (bottom) in all 213 genomes (red) and in all genomes with less than 20, 50, 100, 200, 300, 400 and 500 contigs (green).



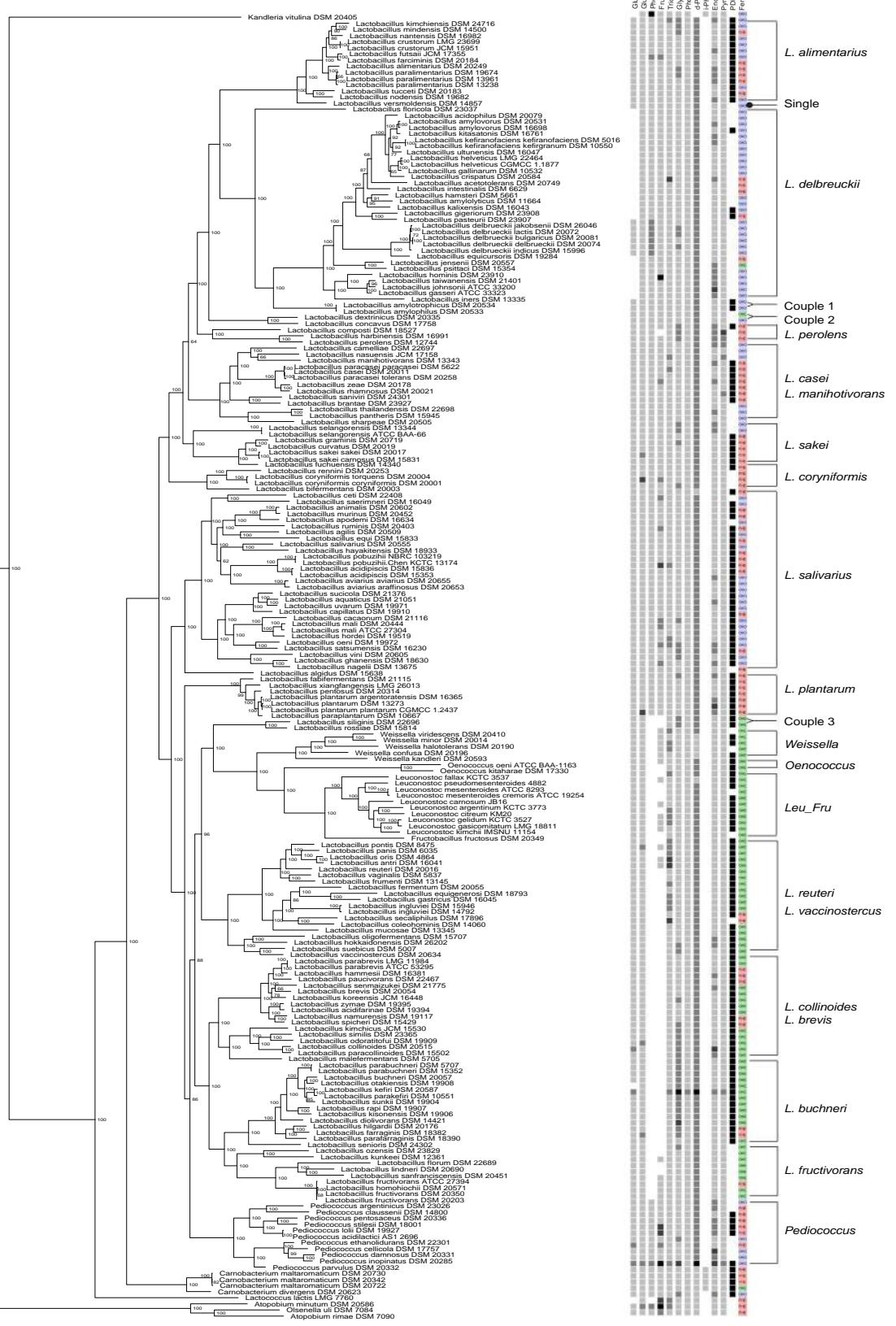
Supp. Figure 3. Analysis of genome assembly size as a function of niche. Niche categories are plotted on the x-axis and genome assembly size in kilobase pairs is plotted on the y-axis. Box-plots represent a five-point summary of the data in the following order (from bottom to top); minimum, first quartile, median, third quartile and maximum. Outliers are represented as individual points above or below the boxplot.



Supp. Figure 4. Frequency distribution of ANI and TNI values for the *Lactobacillus* species compared to those of traditionally defined taxonomic units. The black lines indicate the frequency distribution of values for the lactobacilli, which revealed lower values for both ANI and TNI than the majority of strains within the same family but in different genera.

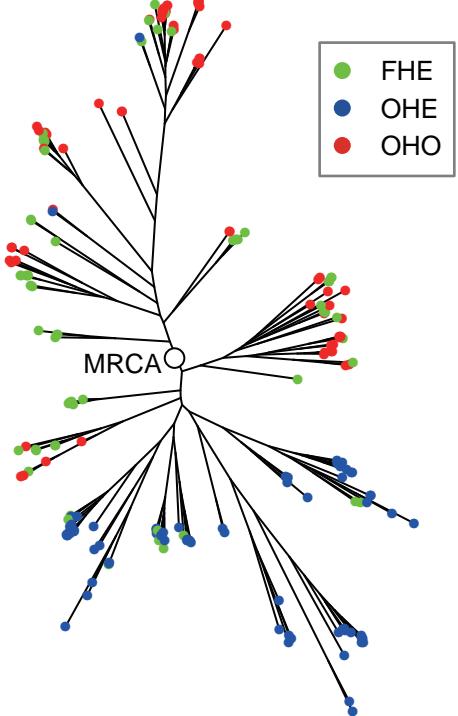


Supp. Figure 6. Maximum likelihood tree of strains of the *Lactobacillus* Genus Complex based on 73 core genes. The branch colors indicate different genera.

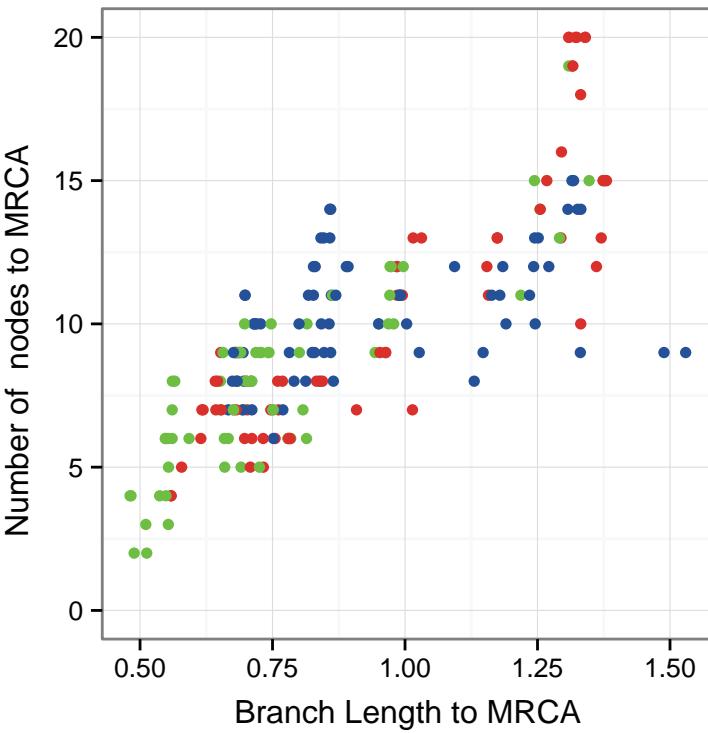


Supp. Figure 7. Distribution of glycolytic and pyruvate dehydrogenase genes across 213 lactobacilli and related species. The distribution of phosphoglycerate mutase is discriminated by the presence of genes encoding the cofactor-dependent (d) or the cofactor-independent (i) isofunctional enzymes. For all 10 core glycolytic enzymes, gene distribution is indicated in grey-scale from absence (white) to presence of 4 gene copies (black). For the pyruvate dehydrogenase operon (4 genes), presence of a functional complex is indicated in black, and absence of a functional complex in white. The fermentation metabolism phenotype is indicated as OHO: obligately homofermentative (purple), FHE: facultatively heterofermentative (pink) or OHE: obligately heterofermentative (green).

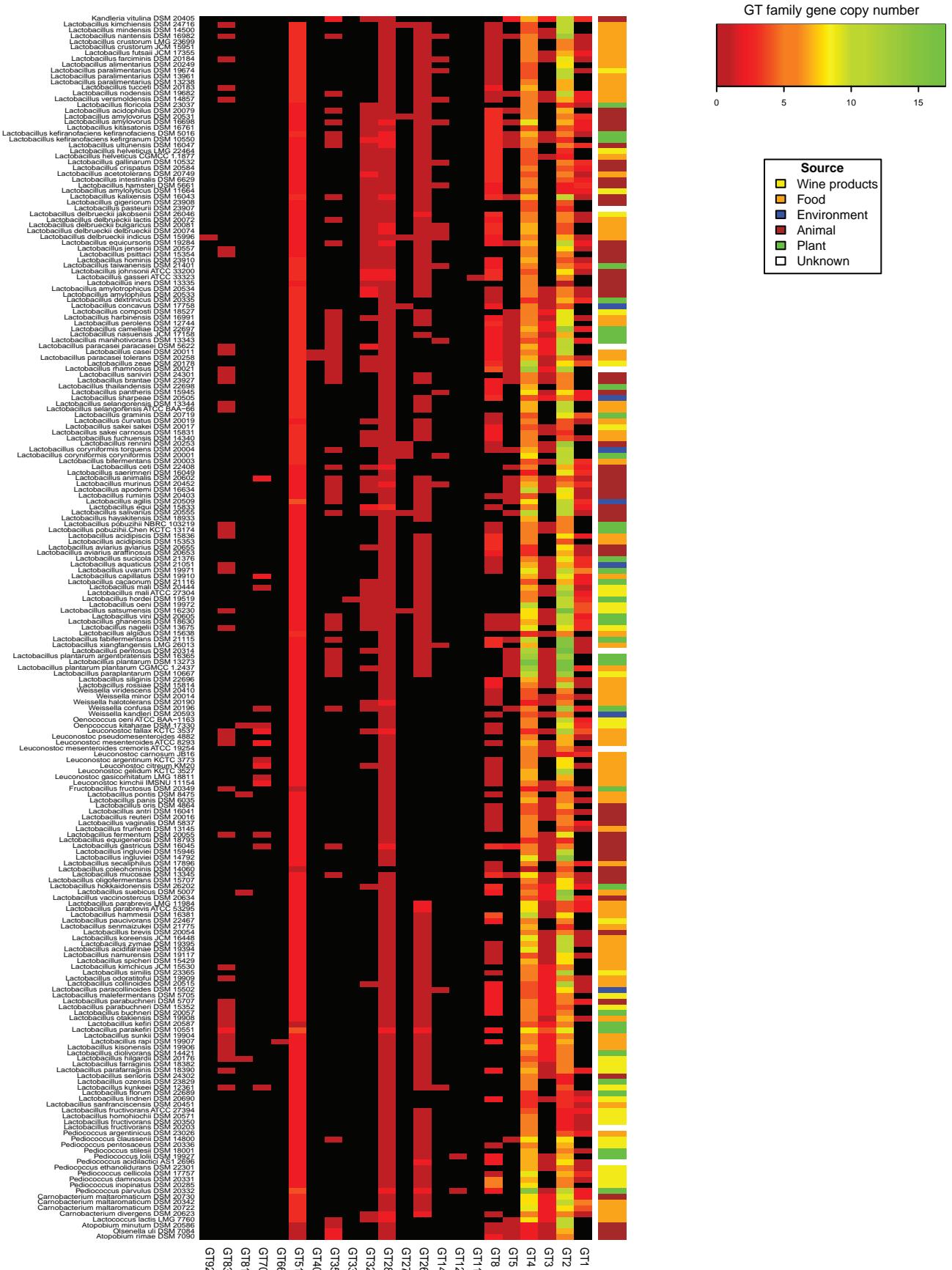
A



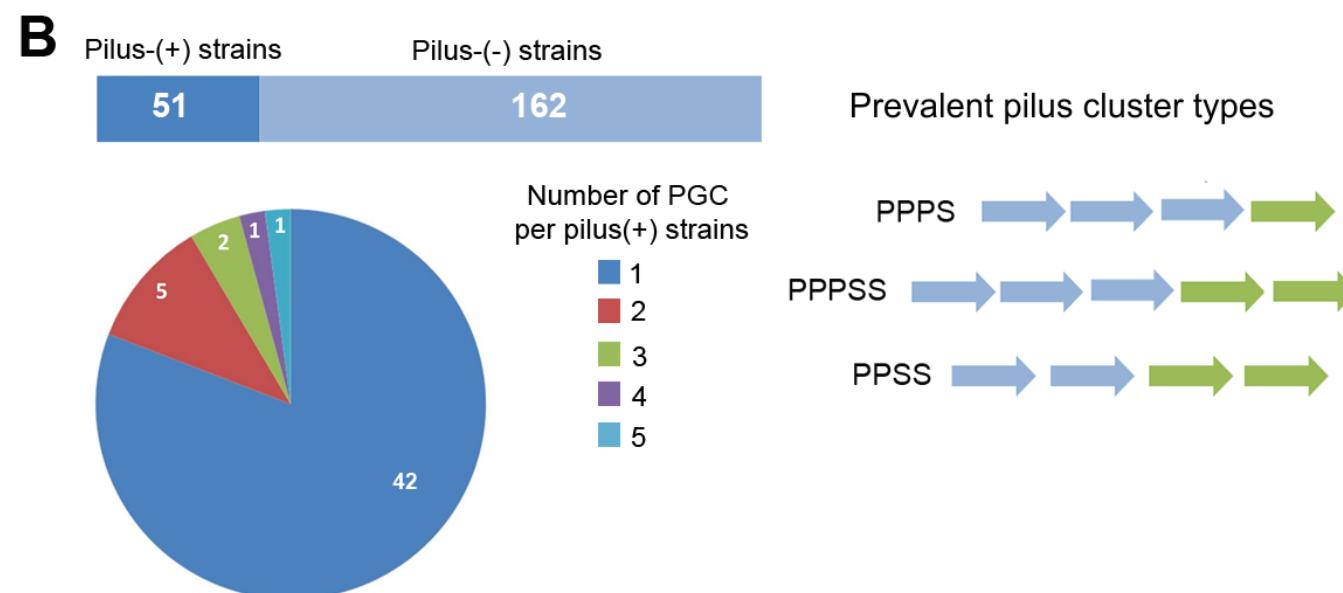
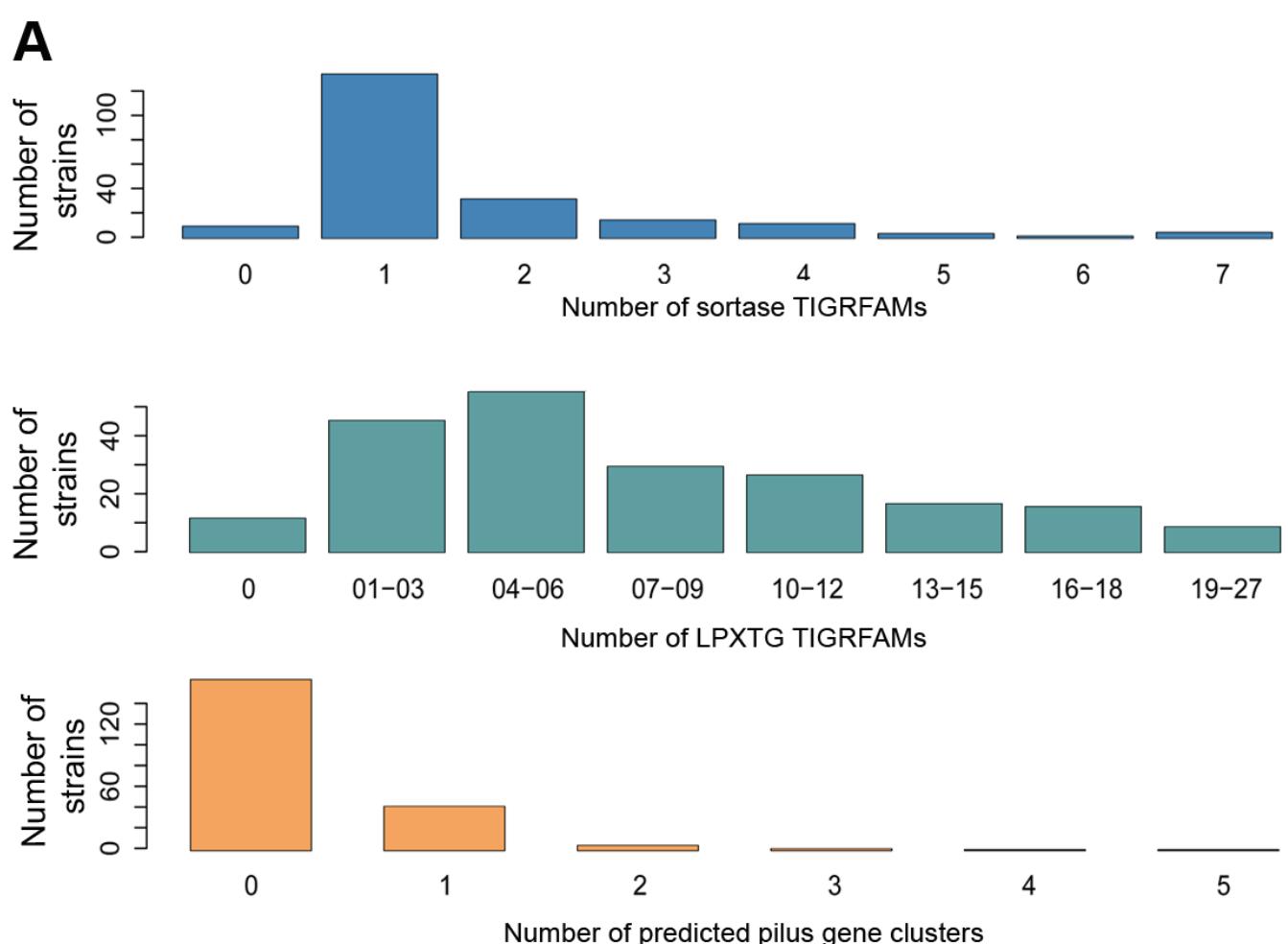
B



Supp. Figure 8. Evolution of carbohydrate metabolism in the *Lactobacillus* Genus Complex. A) Maximum likelihood tree of 204 strains of the *Lactobacillus* Genus Complex based on concatenated amino acid sequence of 73 core genes. The tree was built using RAxML with 100 bootstrap iterations. B) The number of nodes and the branch lengths to the MRCA for each strain/genome. The color of the branches in panel A and the dots in panel B indicate different fermentation types, with green representing FHE, blue representing OHE and red representing OHO.

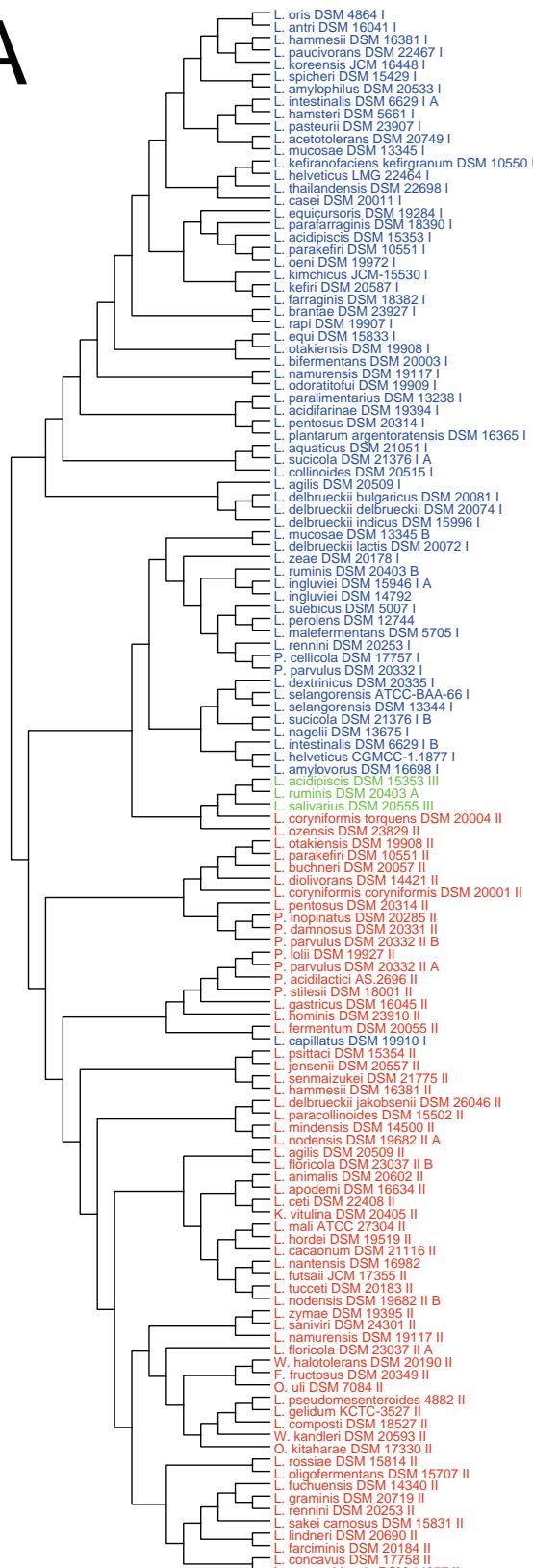


Supp. Figure 9. Heatmap of the distribution and abundance of glycosyl transferase families across the *Lactobacillus* Genus Complex and associated genera. Gene copy number of each of the 22 represented GT family members is indicated by the color key ranging from black (absent) to green. Strains are graphed in the same order from top to bottom as they appear top to bottom in the phylogeny (Fig. 2) with the isolation source of each strain indicated by the color bar at the top of the heat-map.

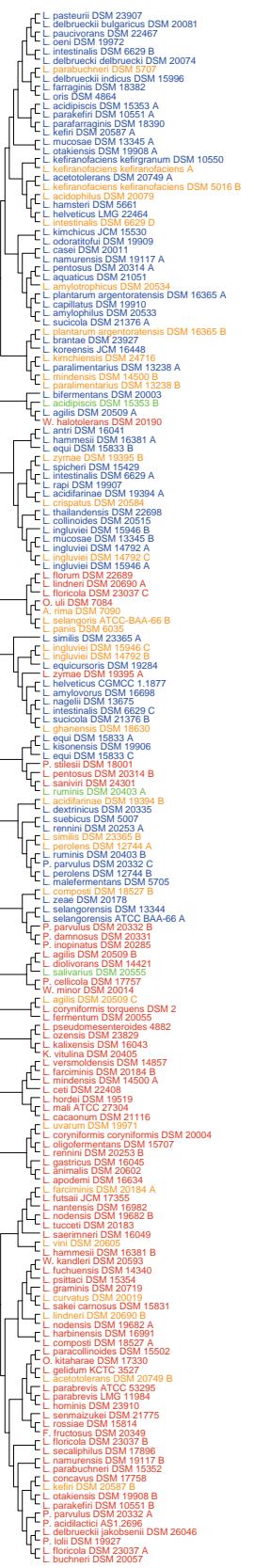


Supp. Figure 10. Distribution of LPXTG proteins, sortases and pilus gene clusters among the 213 genomes analysed. Panel A shows the pilus gene clusters (PGCs) that were found in 24% of all analyzed genomes and had prevalently one of the four types illustrated in Panel B. Legend: green arrow = sortase gene; blue arrow = pilin gene.

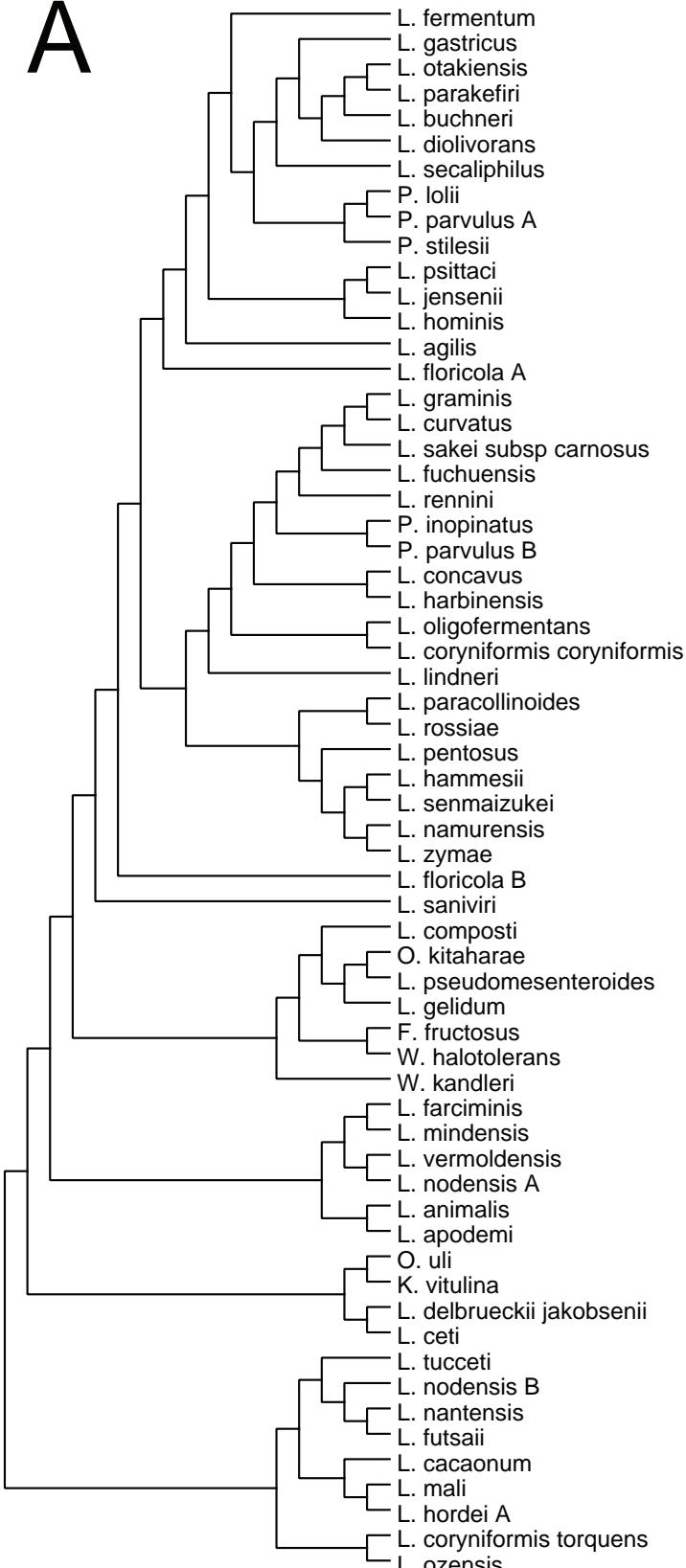
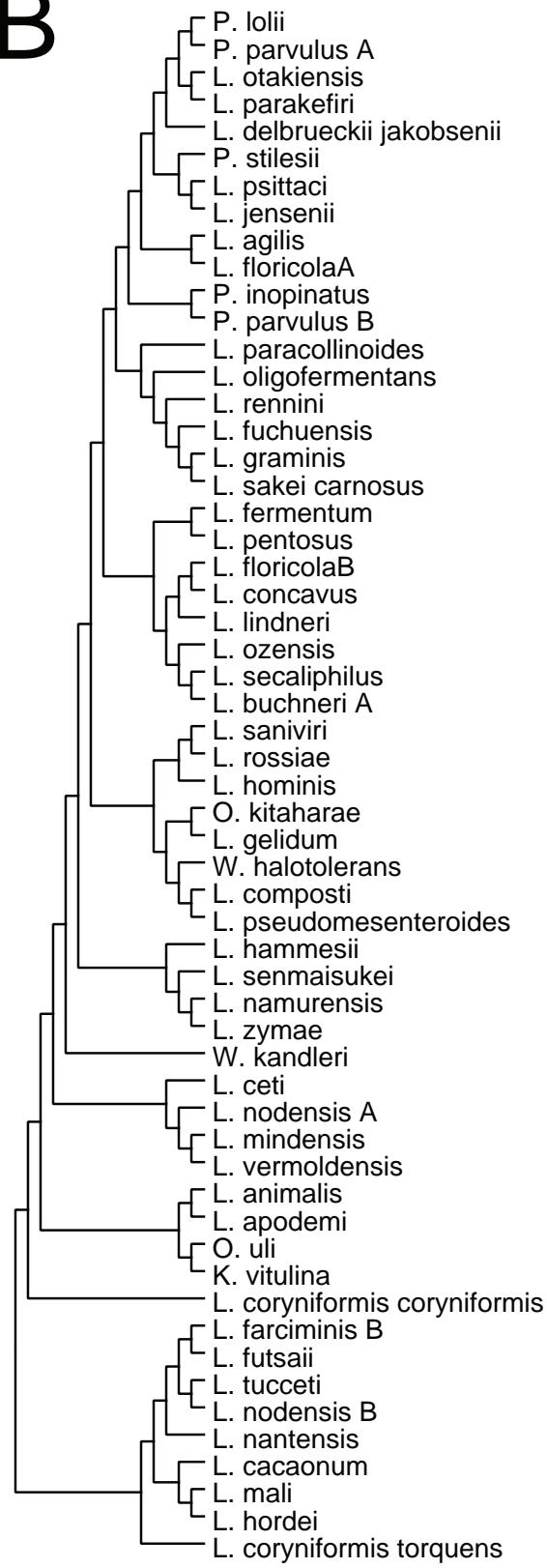
A



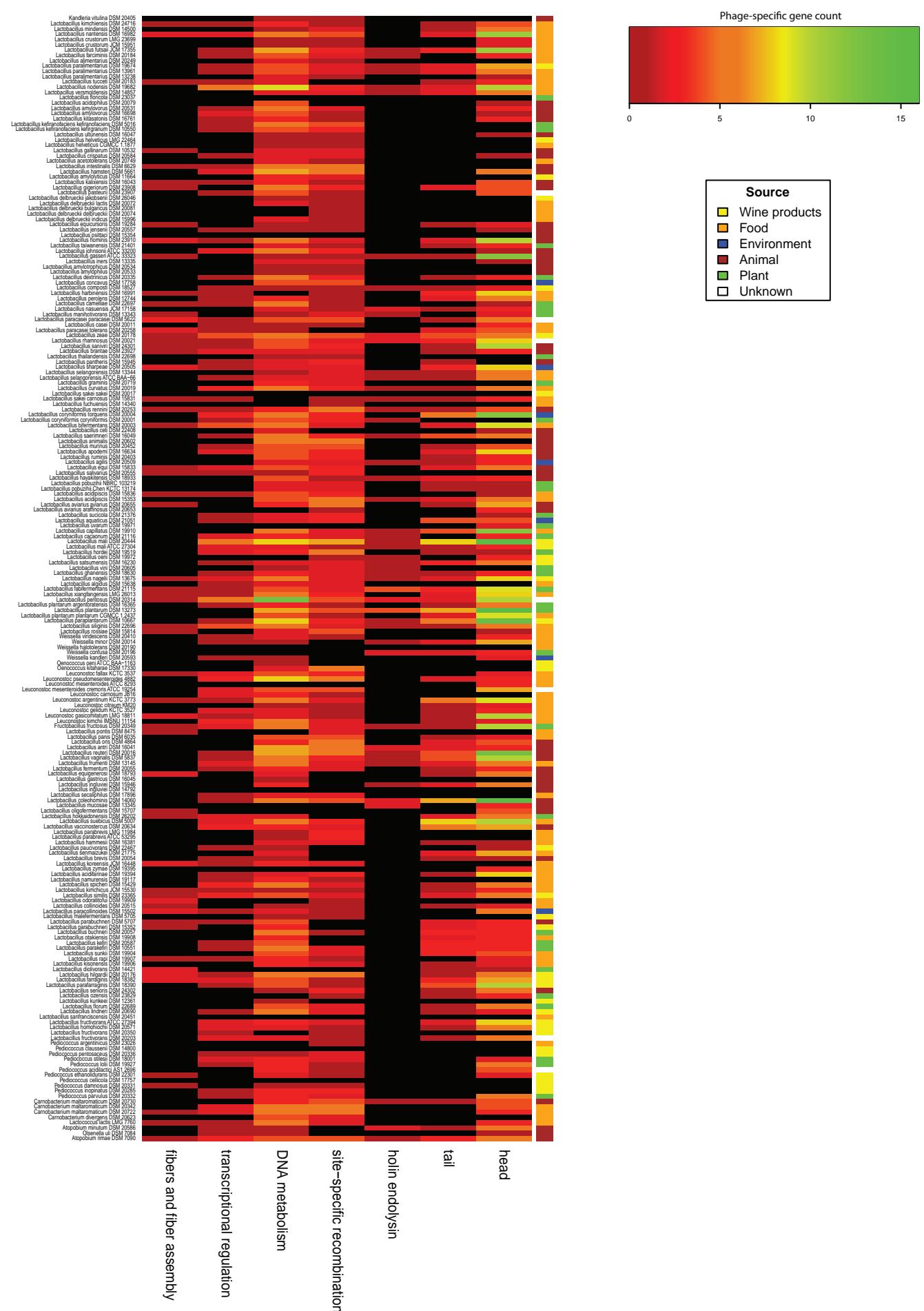
B



Supp. Figure 11. Comparative analysis of core CRISPR elements. The tree in panel A is derived from an alignment of the sequence of the universal Cas protein, Cas1, to create a phylogenetic tree showing the relatedness of all CRISPR-Cas systems in lactobacilli and closely related organisms (see Fig. 5A). The strain designation is followed by I, II, or III, corresponding to the respective CRISPR-Cas system type, using blue, red and green for Type I, II and III systems, respectively. Undefined systems are represented in orange. When multiple Cas1 proteins were found within a genome, they were differentiated by a letter. The tree in panel B is derived from an alignment of the CRISPR repeat sequences. All strain names correlate with the master CRISPR table (Supplementary Table 7). When a strain had multiple CRISPR repeats, they were given different letters to distinguish the repeats.

A**B**

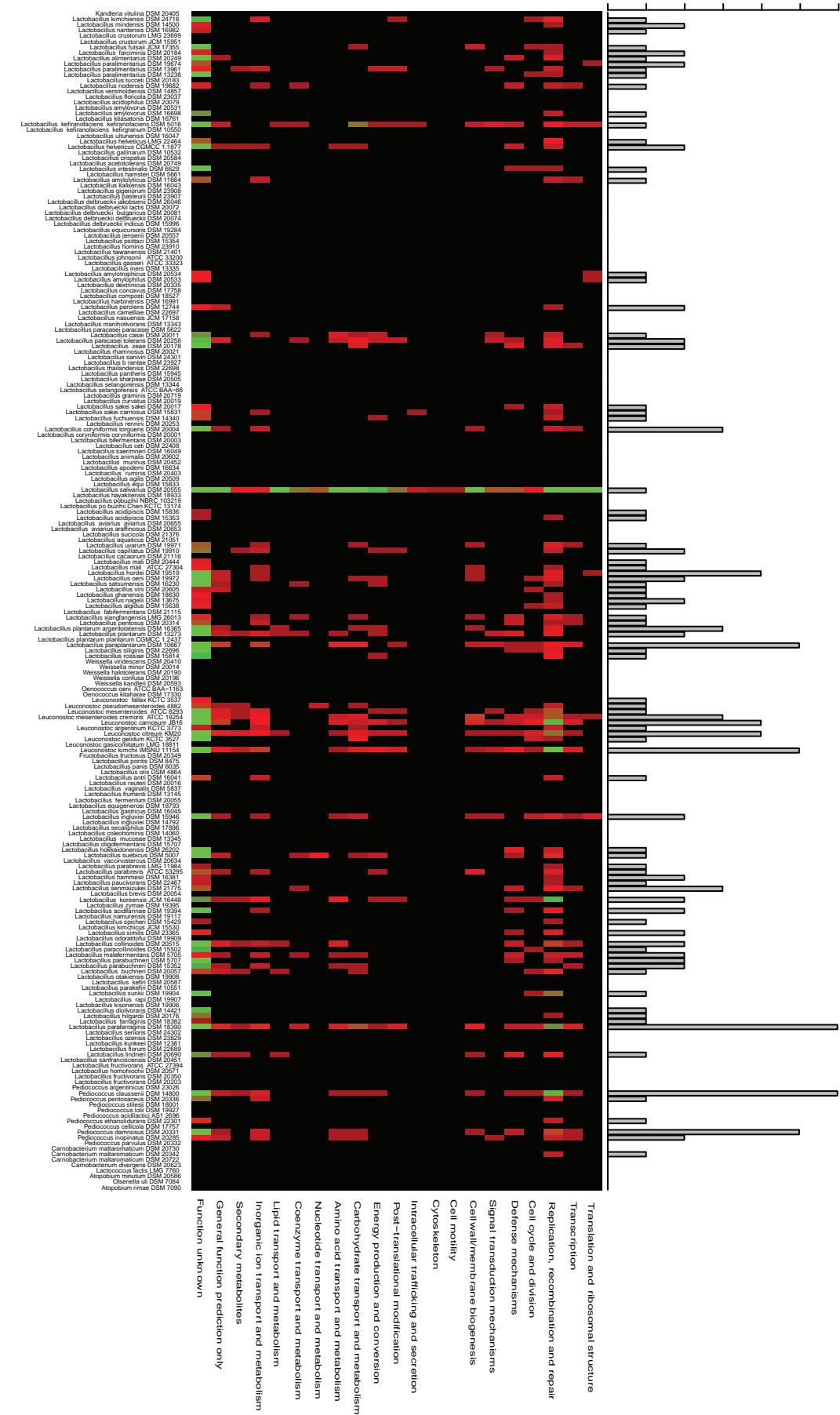
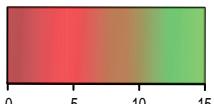
Supp. Figure 12. Comparative analysis of Type II CRISPR-Cas systems. The tree in panel A is derived from an alignment of the sequence of the Type II signature Cas protein, Cas9, to create a phylogenetic tree showing the relatedness of Cas9 proteins from Type II-A and II-C systems (see Fig. 5B). The tree in panel B is derived from an alignment of the predicted tracrRNA sequences for Type II-A systems.



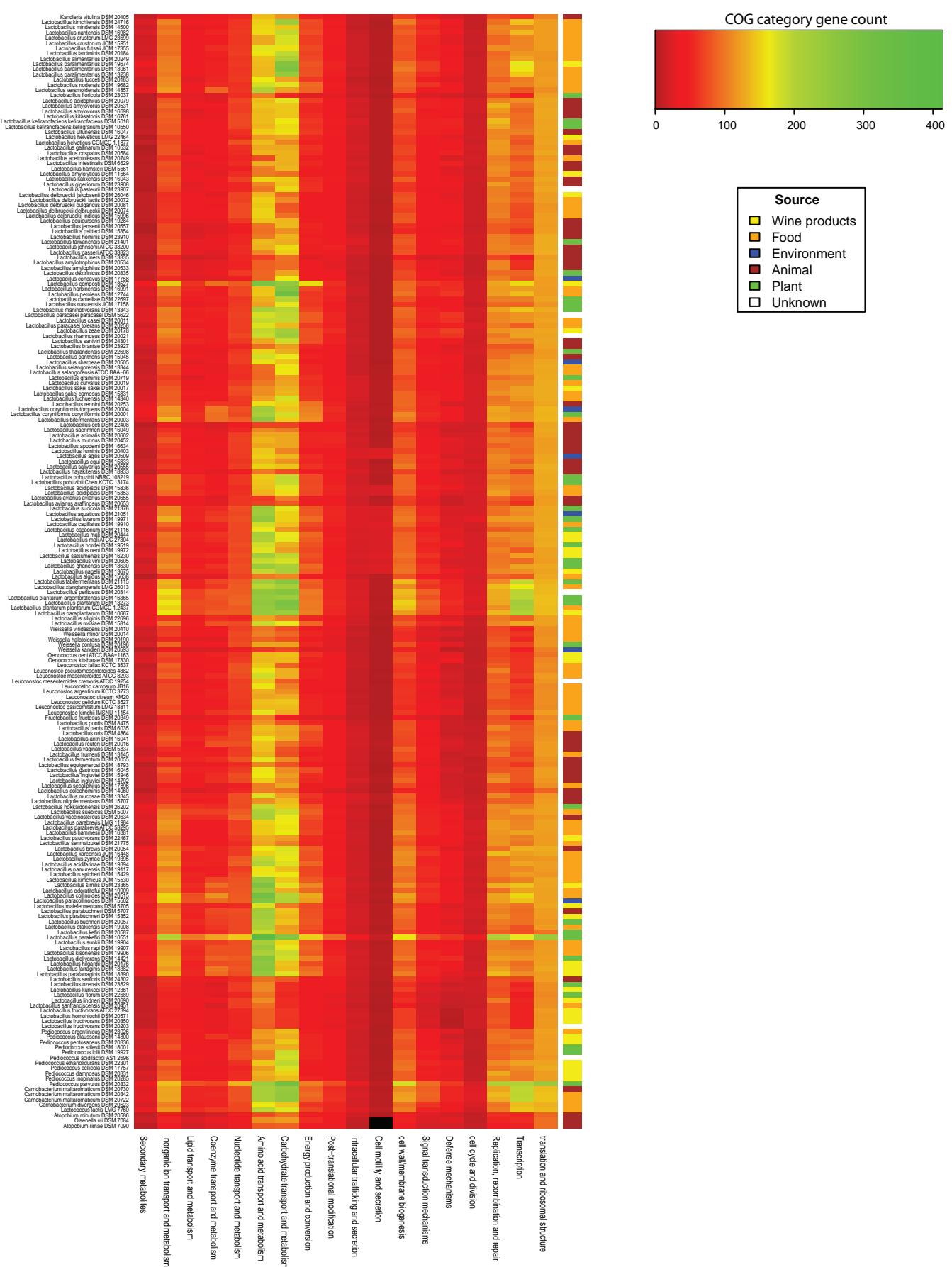
Supp. Figure 13. Heatmap of the distribution of 7 phage functional categories over the 213 genomes. The order of rows, from top to bottom, follows the order of genomes in the phylogenetic tree in Fig. 2 from top to bottom. The color key shows a gradation in color from black to red to yellow to green representing gene counts from 0 to 16.

plasmid count

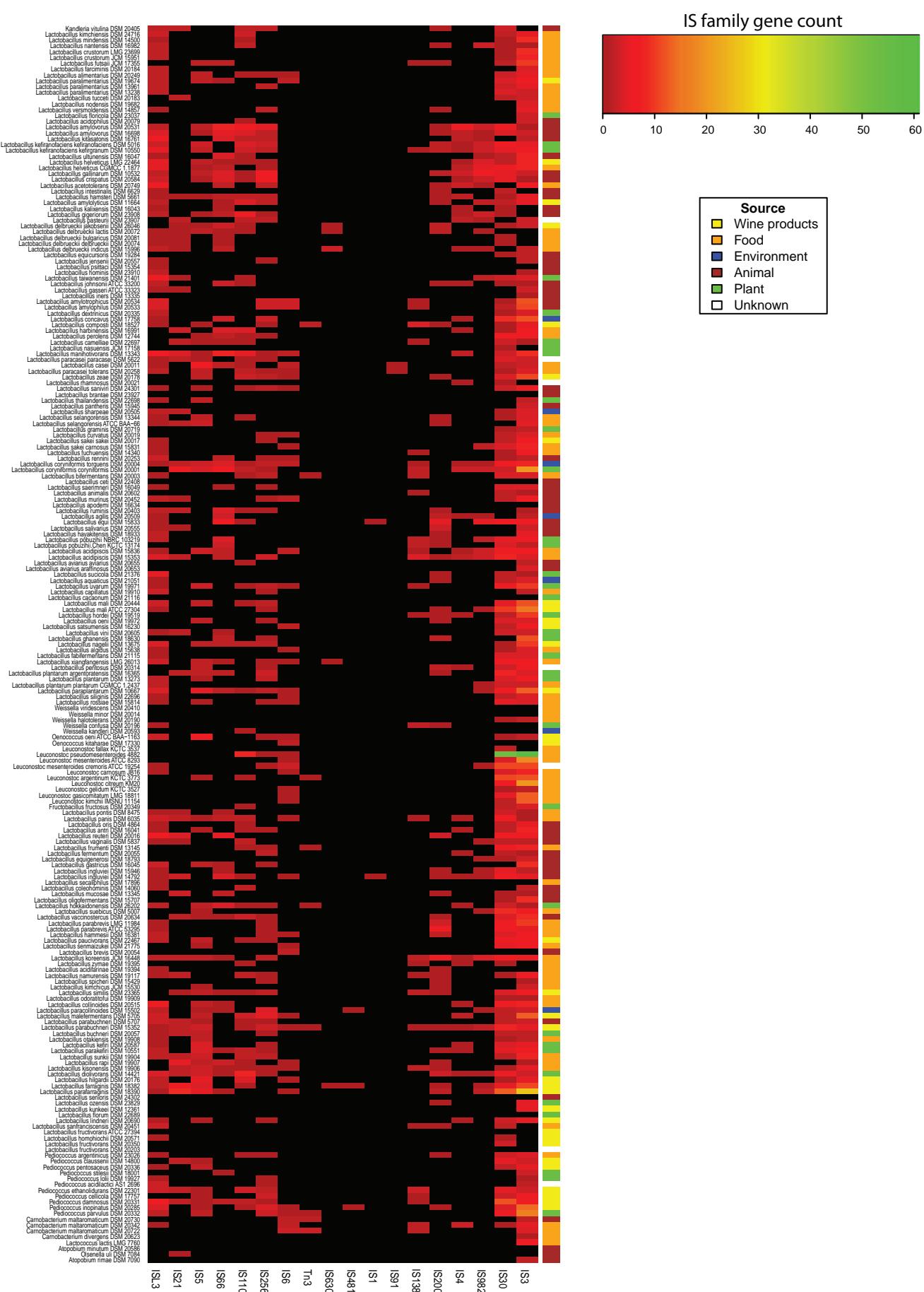
COG count



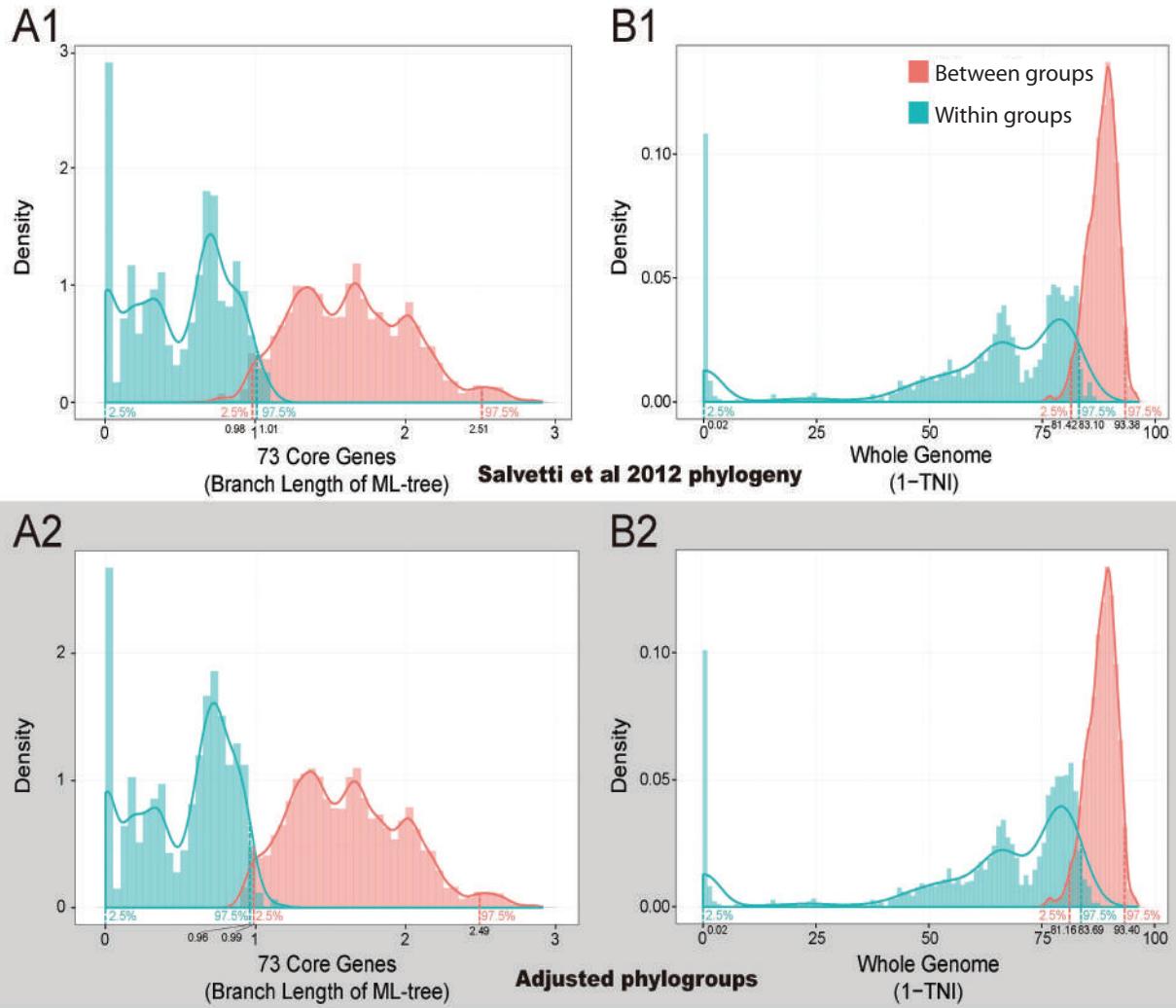
Supp. Figure 14. Heatmap and barplot showing the distribution of plasmid-associated COGs and the number of plasmids in the 213 genomes. The order of rows, from top to bottom, follows the order of genomes in the phylogenetic tree in Fig. 2 from top to bottom. The colour key shows a gradation in colour from black to red representing gene counts from 0 to 15.



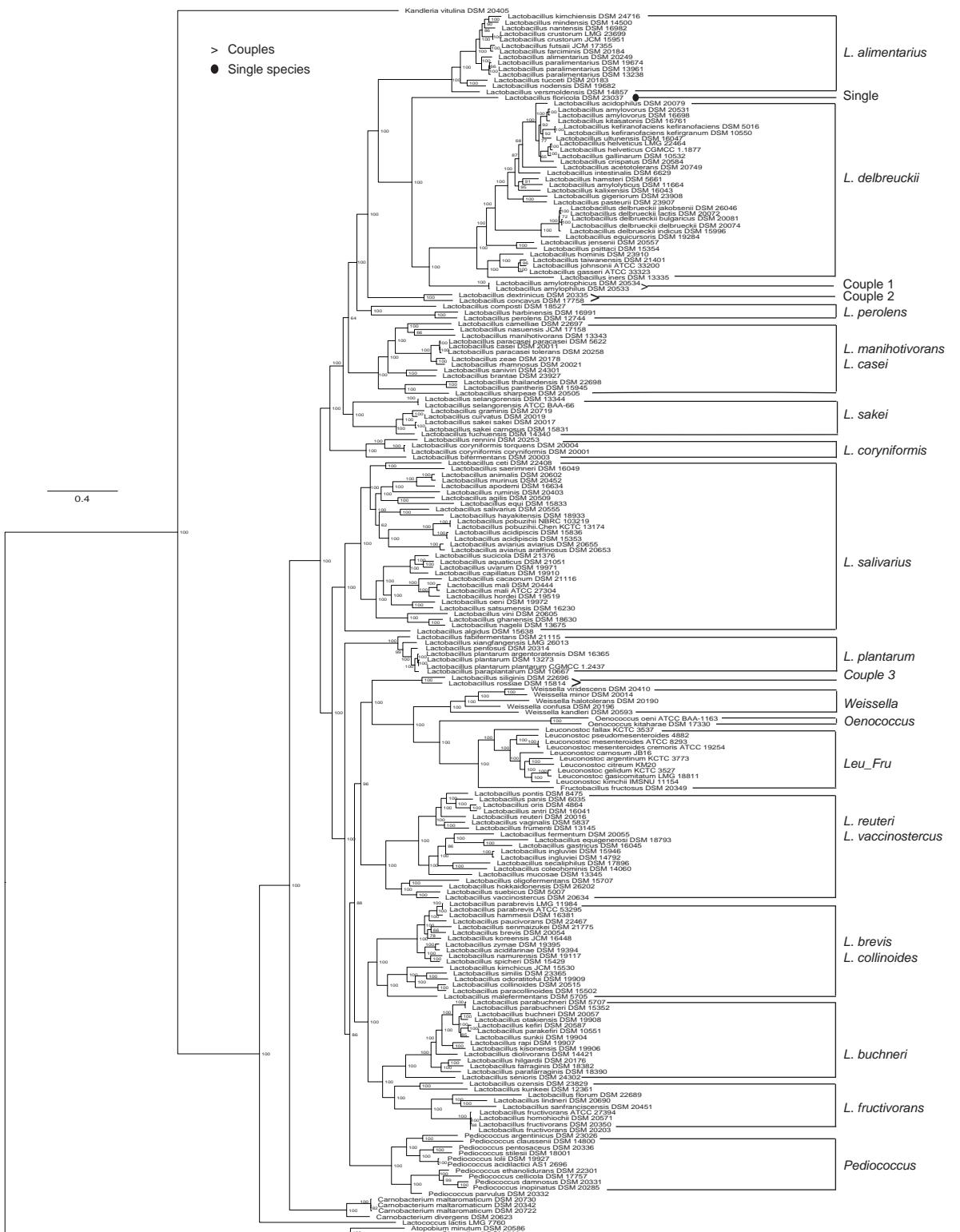
Supp. Figure 15. Heatmap of the distribution and abundance of 18 different COG categories across the 213 genomes. Number of genes assigned to each of the different COG categories is indicated by the color bar from black (absent) to green. Strains are ordered from top to bottom as they appear top-down in the phylogeny (Fig. 2) with source information indicated by the color bar along the top of the heatmap.



Supp. Figure 16. Heatmap of the distribution and abundance of 18 insertion sequence families across the 213 genomes. The number of genes assigned to each IS family is indicated by the color bar from black (absent) to green. The strains appear from top to bottom as they are featured top-down in the phylogeny (Fig. 2). Source information for each strain is indicated by a color bar along the top of the heatmap.



Supp. Figure 17. Branch length distribution and TNI value distribution (1-TNI). A current phylogrouping of the *Lactobacillus* Genus complex and a manually curated phylogrouping based on the maximum likelihood tree of 73 core genes (this study; see Supplementary Figure 18).



Supp. Figure 18. Manually curated phylogrouping of the *Lactobacillus* complex and associated genera based on 73 core genes maximum likelihood phylogeny.

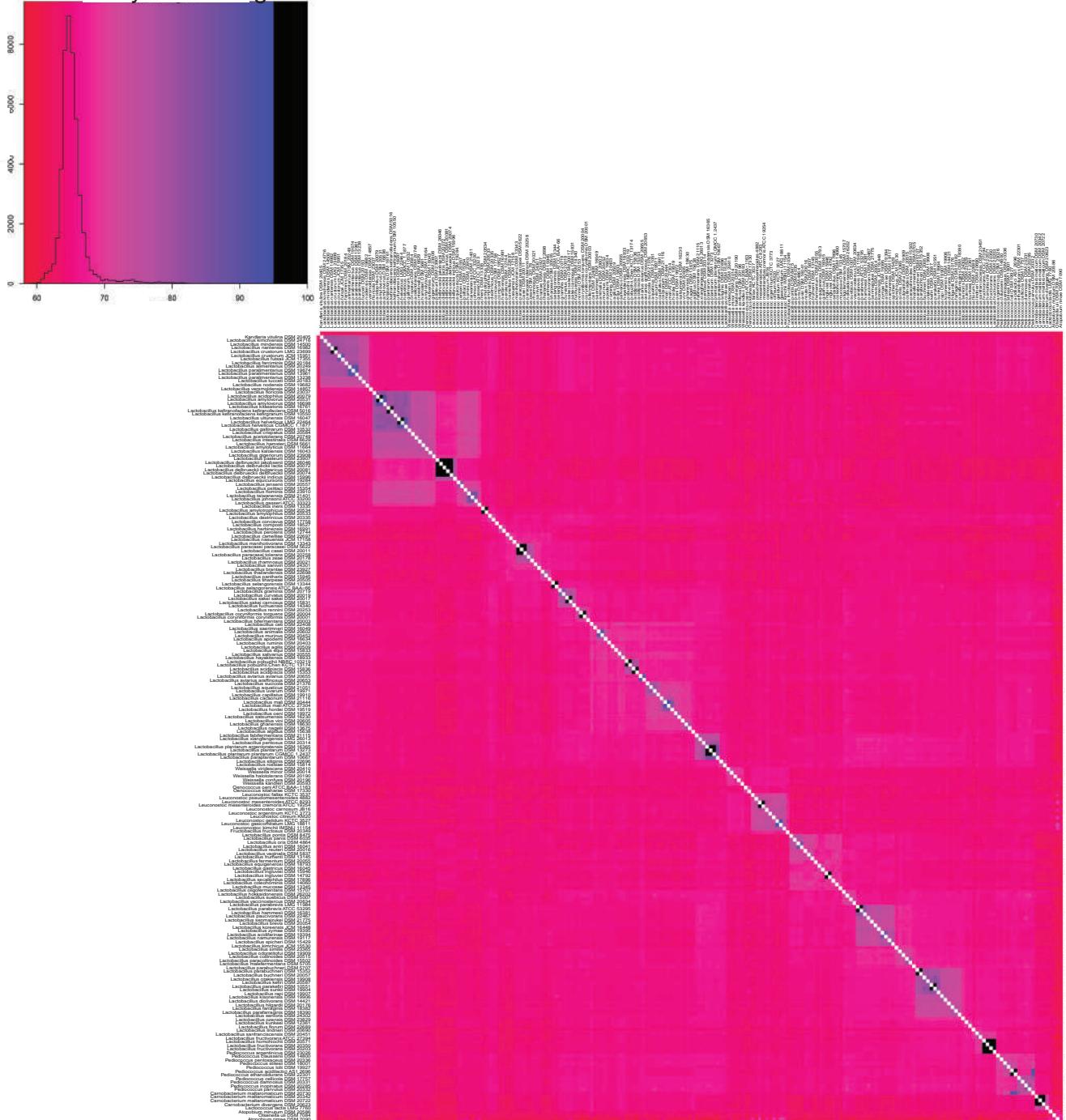
According to this revised phylogrouping, when the branch length between two strains is greater than 0.99, the probability is very high (>97.5%) that they belong to different phylogroups, and when the branch length is less than 0.96 between two strains, the probability that they belong to the same phylogroup is > 97.5%. Compared to the existing phylogrouping in Salvetti et al., 2012, the adjustments made here are:

1. Two species, *L. amylophilicus* and *L. amylophilus* that originally belonged to the *L. delbreuckii* group were excluded from *L. delbreuckii* and defined as a new Couple.
2. The single species *L. composti* was combined with the phylogroup *L. perolens*.
3. The phylogroup *L. casei* and *L. manihovorans* were combined together with the previously defined single species, *L. camelliae*, *L. sanitiviri*, *L. brantae*, *L. sharpeae*, and the Couple that contained *L. thailandensis* and *L. pantheris*, was defined as a single phylogroup.
4. The single species *L. algidus* was combined with the phylogroup *L. salivarius*.
5. *Leuconostoc* and *Fructobacillus* were defined as a single phylogroup.
6. The phylogroups *L. reuteri* and *L. vaccinostercus* were combined together.
7. The phylogroups *L. brevis* and *L. collinoides* and a single species, *L. malefermentans*, were combined as a single phylogroup.
8. *L. senioris* was combined with the phylogroup *L. buchneri*.
9. The couple that contained *L. ozensis* and *L. kunkeei* was combined into the phylogroup *L. fructivorans*.



Supp. Figure 19. Phylogeny inferred from a 100 core gene dataset (27 partial + 73 complete core genes).

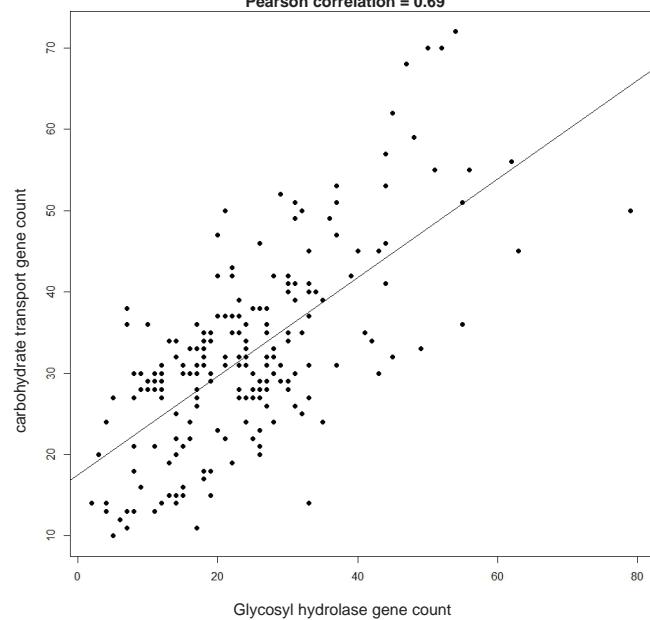
Color key and histogram



Supp. Figure 20. Heatmap of pairwise ANI values for 213 genomes. The order of the rows (top to bottom) and columns (left to right) is according to their position in the phylogenetic tree based on 73 core proteins (Fig. 2).

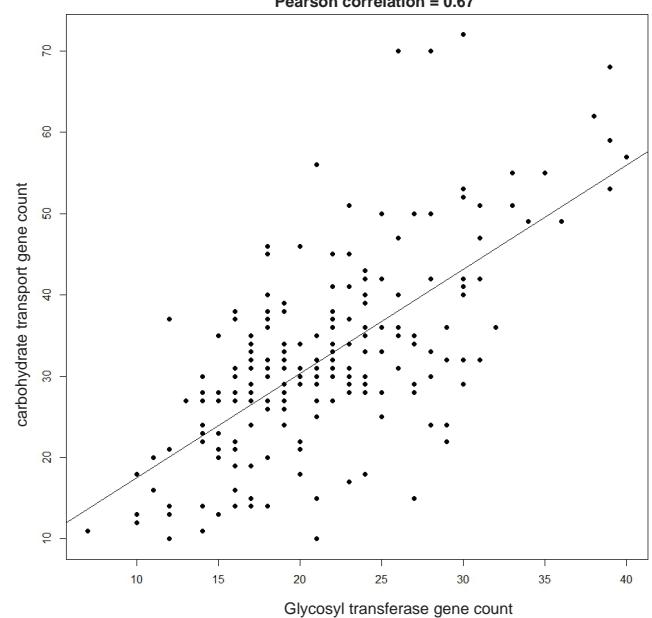
Carbohydrate transport against GHs

Pearson correlation = 0.69

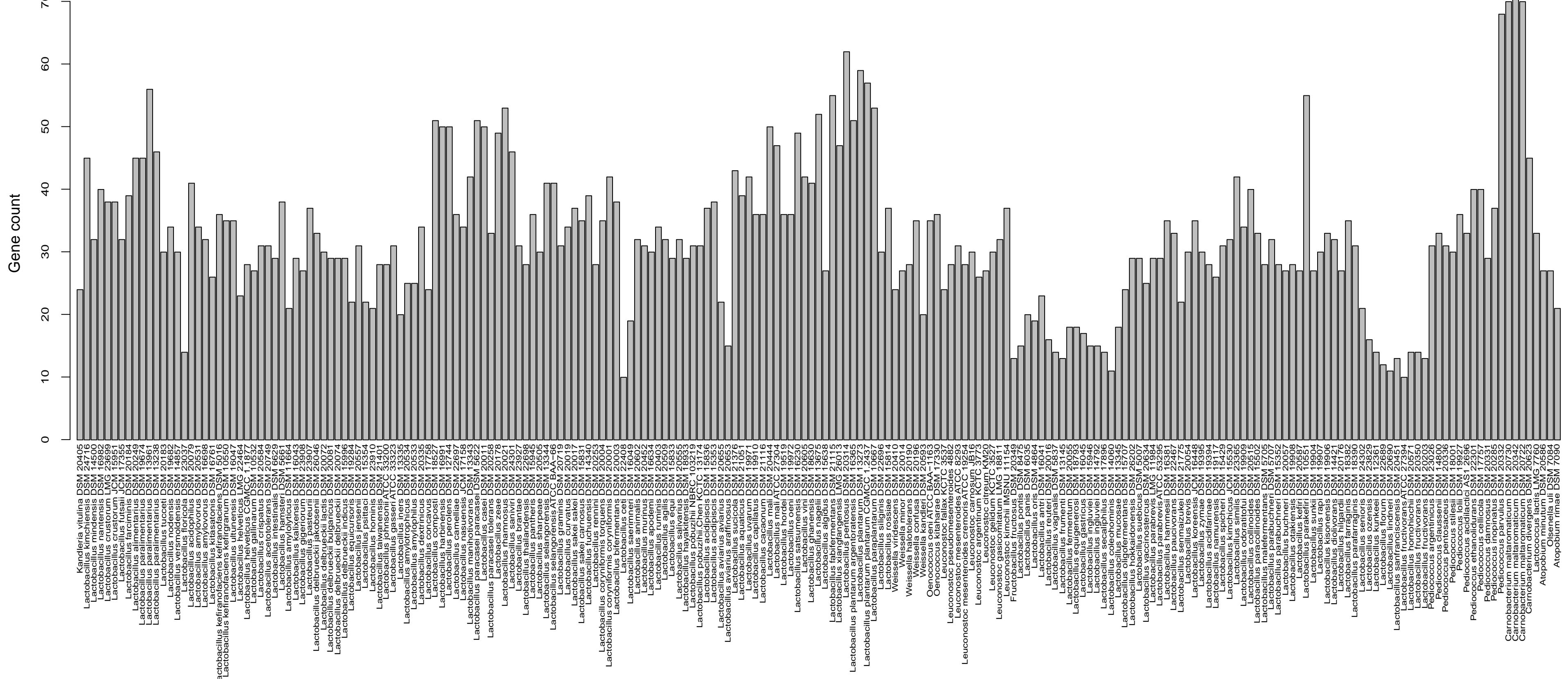


Carbohydrate transport against GTs

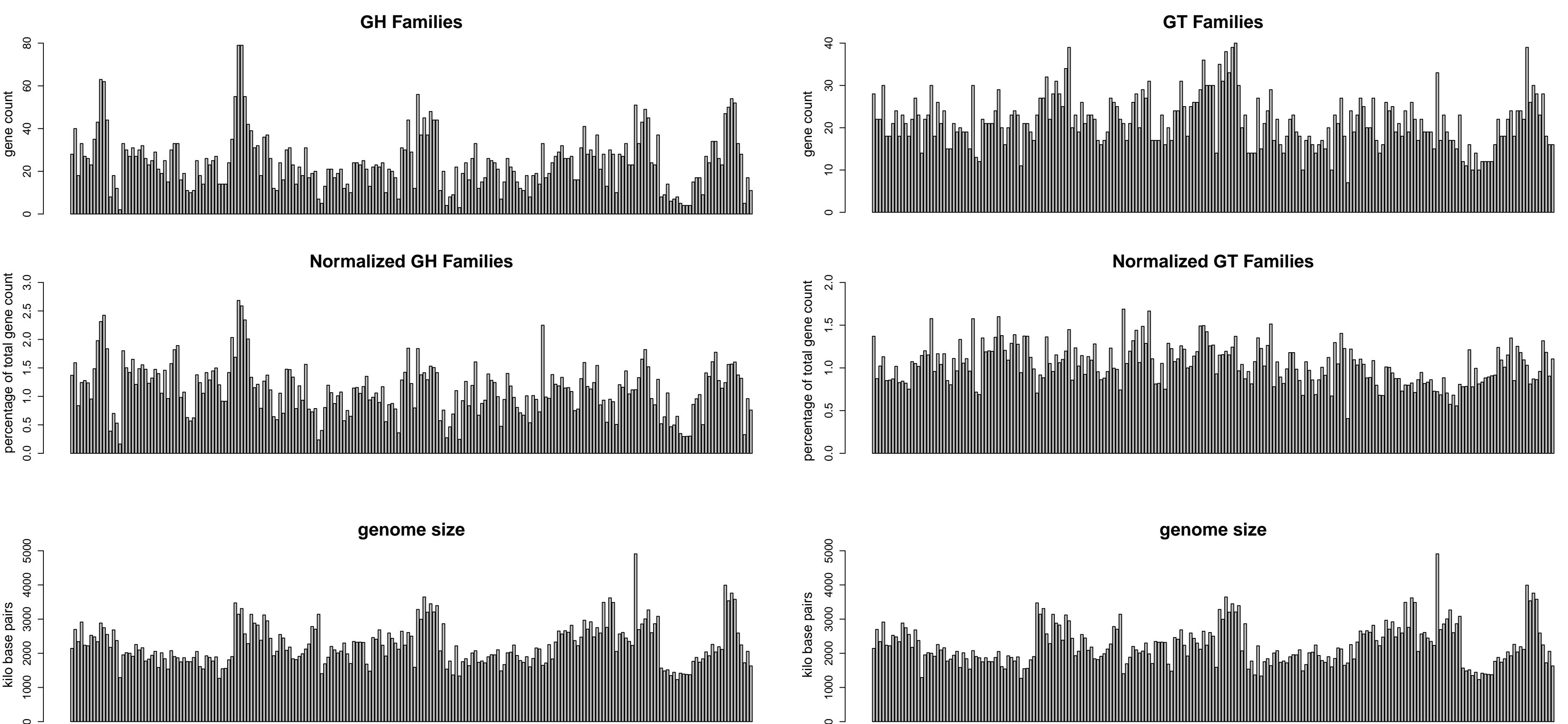
Pearson correlation = 0.67



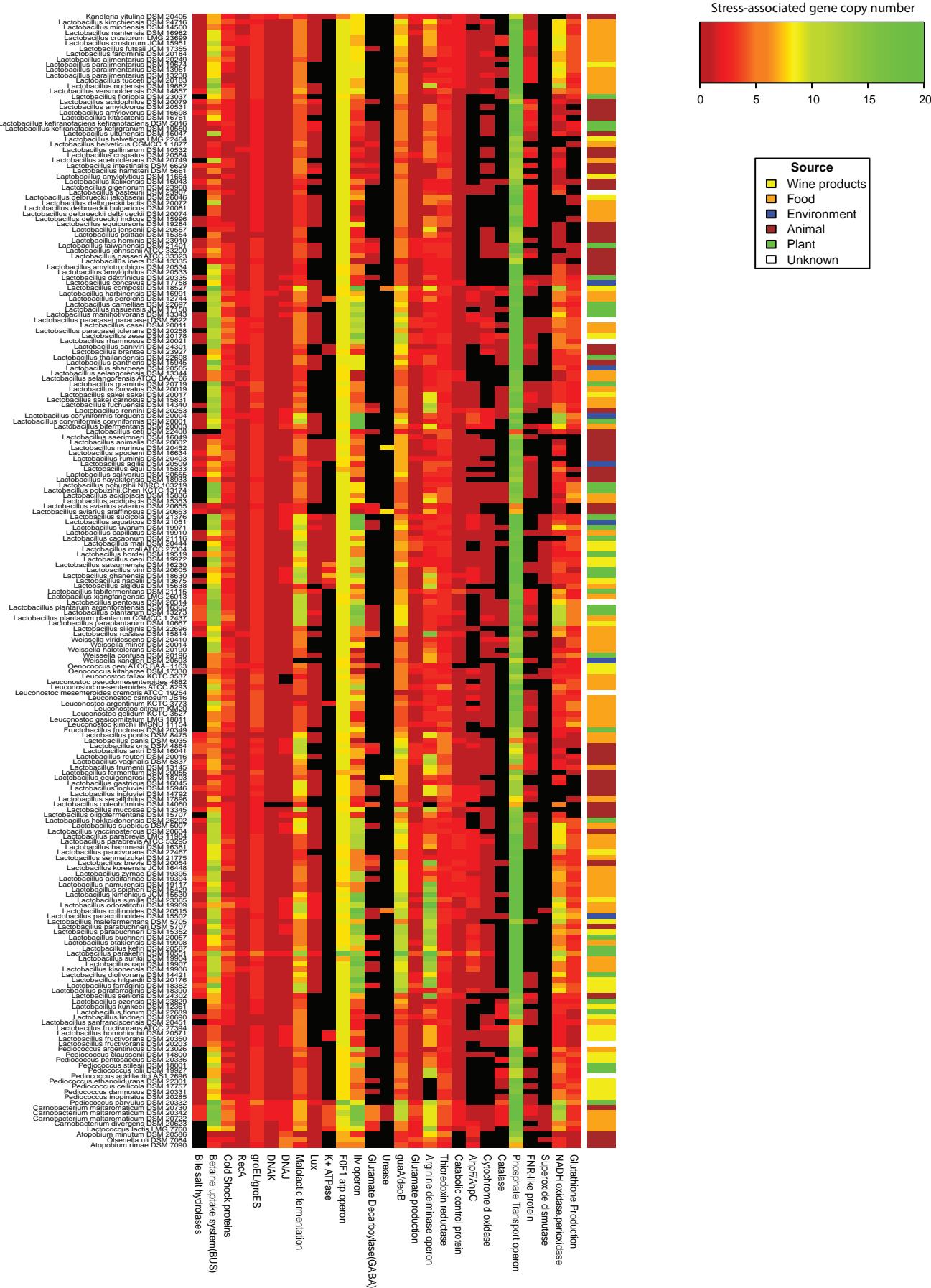
Supp. Figure 21. Scatterplots showing the correlation between the number of carbohydrate transport genes (y-axes) and the number of glycosyl hydrolase genes (x axis; left) and the number of glycosyl transferase genes (x-axis; right). The line of best fit for each plot was estimated using a least squares linear model.



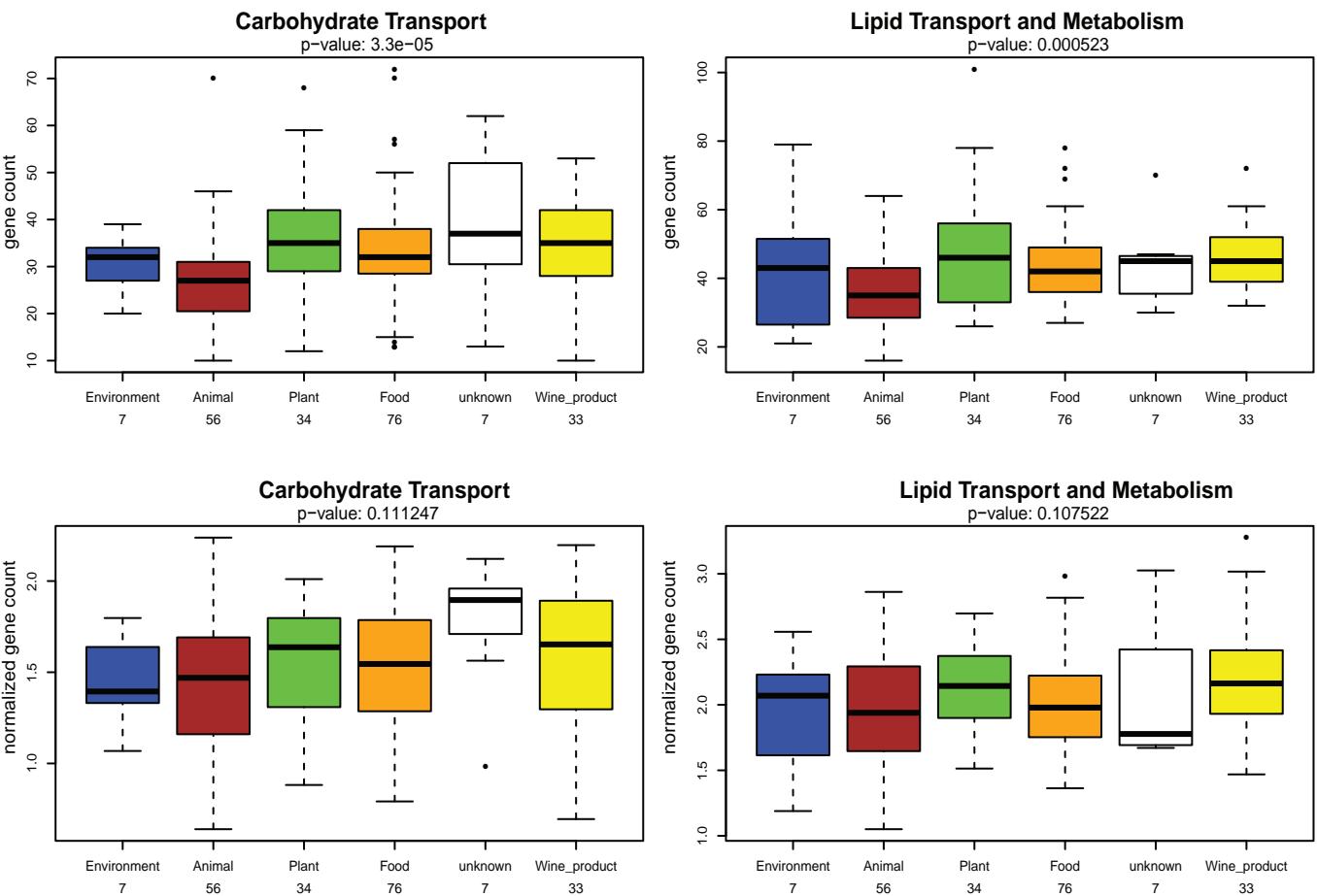
Supp. Figure 22. Barplot of the number of genes involved in carbohydrate transport for each strain. Strains are ordered according to the phylogenetic tree in Fig 2.



Supp. Figure 23. The effect of normalizing counts of GHs and GTs by genome size. The three bar-graphs on the left show, from top to bottom, GH gene counts, GH gene counts expressed as a percentage of the total gene count and genome size (in kbps). The equivalent for GTs is shown in the three bar-graphs on the right. Genome size is highly correlated with total number of genes per genome (Pearson; 0.99).



Supp. Figure 24. Heatmap of the distribution and abundance of stress response genes across the *Lactobacillus* Genus Complex and associated genera. Gene copy number for 27 stress-associated genes is indicated by the color key from black (absent) to green. Type of stress resistance is indicated by the names at the bottom of the figure. Strains are ordered from top to bottom as they appear top-down on the phylogeny (Fig. 2) with source information for each strain indicated by a color bar at the side of the heat-map.



Supp. Figure 25. Association of carbohydrate transport and lipid transport/metabolism with niche. Top panels display raw gene counts and bottom panels display gene counts normalized by total genes. Boxplots represent a five-point summary of the data in the following order (from bottom to top): minimum, first quartile, median, third quartile and maximum. Outliers are represented as individual points above or below the boxplot.

1 **Supplementary Tables**

2 **Supp. Table 1. Species analysed and their genomic features.**

3 **Supp. Table 2. Sequence information for the 73 core complete genes.** The core genome
4 was established based on genomes listed in Table 1. *Lactobacillus salivarius* genes are shown
5 as exemplars for sequence retrieval. Columns are protein ID numbers, gene designation,
6 locus tag, COG, annotation, co-ordinates in the *L. salivarius* UCC118 genome
7 (NC_007929.1), strand and gene length.

8 **Supp. Table 3. Genera used in building the tree of bacteria.**

9 **Supp. Table 4. Sixteen marker genes used to build the bacterial tree composed of 452**

10 genera and 213 *Lactobacillus* genomes and *Lactobacillus* associated genera.

11 **Supp. Table 5. Distribution of LPXTG-containing and sortase enzymes across the 213**

12 genomes.

13 **Supp. Table 6. Distribution and abundance of cell envelope proteases (CEPs) and**

14 associated anchoring domains and motifs.

15 **Supp. Table 7. CRISPR occurrence and diversity.** CRISPR-Cas system type designation
16 was determined by the presence of CRISPR repeats, spacers, the universal *cas1* gene, and
17 the signature for each type, namely *cas3*, *cas9*, and *cas10*, for Type I, II, and III, respectively.
18 A “Y” in the each of the gene columns designates that the gene was positioned next to the
19 CRISPR locus. In instances where only a partial gene was annotated, the symbol “Y*” is
20 shown. An N indicates that no *cas* gene was found. The type of the CRISPR-Cas system is
21 noted in the “Type” column. When multiple CRISPR loci of the same type were present in the
22 same genome, the type was then designated with a differential letter demarking their order in
23 identification. When *cas* genes could not be annotated, the CRISPR-Cas system type was
24 labeled “undefined.” Strains that contained neither CRISPR repeats nor *cas* genes were
25 labeled with the “N/A” designation. “DR length” corresponds to the number of nucleotides in
26 the CRISPR direct repeat. The number of spacers was also determined for each repeat-
27 spacer array.

28 **Supp. Table 8. Sequence information for the 27 core partial genes.**

29 **Supp. Table 9. Presence and absence of the complete pathways for production of the**

30 20 standard amino acids. A = auxotrophic; P = prototrophic.

31 **Supp. Table 10. Presence of sirtuin homologs in the 213 genomes analyzed.**

32 **Supp. Table 11. Genomic regions related to bacteriocin production identified in the**
33 **213 genomes.**

Lactobacillus selangorensis	DSM-13344	2000	Chili bo	2.09	46.45	2082	95.83	32	SRX690300	JQAZ00000000	Food	<i>L. sakei</i>
Lactobacillus senioris	DSM-24302	2012	Facess of a healthy 100-year-old Japanese fema	1.57	39.09	1567	153.08	16	ERX359768	JYZR00000000	Animal	<i>L. buchneri</i>
Lactobacillus snyderiae	DSM-21775	2008	Semiarakaze, a Japanese pickle	2.22	48.64	2149	107.96	68	SRX456331	JYZH00000000	Food	<i>L. brevis</i> , <i>L. collinoides</i>
Lactobacillus sakei	DSM-1035	1990	Mustard pickle	2.45	33.38	2371	90.46	79	ERX359770	JYVX00000000	Environment	<i>L. brevis</i> , <i>L. collinoides</i>
Lactobacillus siliginea	DSM-22696	2006	Wheat sourdough	2.07	44.08	2059	96.60	52	SRX359753	JQCD00000000	Food	Couple3
Lactobacillus similis	DSM-23365	2010	Fermented cane molasses at alcohol plants	3.49	46.99	3206	68.75	282	ERX359772	JYZM00000000	Wine product	<i>L. brevis</i> , <i>L. collinoides</i>
Lactobacillus sucirola	DSM-15429	2004	Rice sourdough	2.75	55.91	2494	87.23	51	ERX359773	JZF00000000	Food	<i>L. brevis</i> , <i>L. collinoides</i>
Lactobacillus sucirola	DSM-21376	2009	Sap of an Oak tree	2.46	38.48	2335	97.51	31	ERX450947	JYZF00000000	Plant	<i>L. salivarius</i>
Lactobacillus subieicus	DSM-5007	1989	Apple mash	2.65	38.98	2517	45.26	81	SRX456334	JZGF00000000	Food	<i>L. reuteri</i> , <i>L. vaccinostercus</i>
Lactobacillus sunmoriensis	DSM-10404	2006	Sunki, a Japanese traditional pickle	2.69	42.06	2574	89.12	79	SRX456335	JZEA00000000	Food	<i>L. buchneri</i>
Lactobacillus thailandensis	DSM-31401	2009	Village-style feed	1.88	33.97	1881	127.73	93	ERX359776	JYZD00000000	Plant	<i>L. delbrueckii</i>
Lactobacillus tucareti	DSM-22698	2007	Fermented tea leaves (<i>miang</i>)	2.06	53.5	1943	116.47	23	SRX456337	JZKZ00000000	Plant	<i>L. manshutovorans</i> , <i>L. casei</i>
Lactobacillus ultenensis	DSM-20183	2009	Sausage	2.17	34.07	2093	110.40	51	ERX359779	JZDG00000000	Food	<i>L. alimentarius</i>
Lactobacillus ultenensis	DSM-16047	2005	Gastric biopsies, Human stomach mucosa	2.16	35.95	2113	55.51	105	SRX456274	JZF00000000	Animal	<i>L. delbrueckii</i>
Lactobacillus uvarum	DSM-19971	2009	Mast of Bobal grape variety	2.69	36.88	2606	89.34	164	ERX359780	JZEG00000000	Plant	<i>L. salivarius</i>
Lactobacillus uvarinosterucus	DSM-20635	1983	Cow dung	2.57	43.48	2485	93.56	112	ERX359731	JYYY00000000	Animal	<i>L. reuteri</i> , <i>L. vaccinostercus</i>
Lactobacillus uvarinosterucus	DSM-1893	1990	Vinegar with	1.79	40.86	1735	67.13	19	SRX456338	JZGZ00000000	Animal	<i>L. reuteri</i> , <i>L. uvarinosterucus</i>
Lactobacillus uvarinosterucus	DSM-14857	2002	Poultry salami	2.37	38.27	2319	50.46	632	SRX456244	JZFJA00000000	Food	<i>L. alimentarius</i>
Lactobacillus vini	DSM-20605	2006	Must of grape	2.24	37.54	2191	53.62	269	SRX456339	JAXY00000000	Plant	<i>L. salivarius</i>
Lactobacillus viungangensis	LMG-26013	2012	Pickles	3.00	45.1	2801	80.11	145	ERX359765	JOCLO00000000	Food	<i>L. plantarum</i>
Lactobacillus zeae	DSM-20178	1959	Corn steep liquor	3.12	47.74	3043	38.45	55	SRX456269	JZCT00000000	Wine product	<i>L. manihotovorans</i> , <i>L. casei</i>
Lactobacillus zymae	DSM-19393	2005	Artisanal wheat sourdough	2.71	53.57	2460	88.66	75	ERX399772	JZDW00000000	Food	<i>#L. brevis</i> , <i>L. collinoides</i>
Leuconostoc lactis	LMG-17760	1873	Anchu mash	2.25	35.02	2237	89.07	45	ERX359766	JQAZ00000000	Food	Other
Leuconostoc lactis	KCTC-13973	1997	Baw moh	1.72	42.89	1821	NA	98	AEC00000000	Food	Leu_Fru	
Leuconostoc camousum	JB16	1989	Kimchi	1.77	37.13	1711	NA	5	NA	P003851- CP00385:	Food	Leu_Fru
Leuconostoc citrum	KM20	2008	Kimchi	1.90	38.87	1866	NA	5	NA	P0489736- DQ48974	Food	Leu_Fru
Leuconostoc fallax	KCTC-3537	1992	Sauerkraut	1.64	37.53	1916	NA	30	NA	AEI20000000	Food	Leu_Fru
Leuconostoc gascomitatum	LMG-18811	2001	Tomato-marinated broiler meat strips	1.95	36.66	1929	NA	1	NA	FN822744	Food	Leu_Fru
Leuconostoc gascomitatum	KCTC-3527	1996	Vacuum packaged beef	1.96	36.6	1951	NA	43	NA	AEM00000000	Food	Leu_Fru
Leuconostoc linhthai	DSM-1184	2000	Kimchi	2.10	37.91	2110	NA	6	NA	P001751- CP001751:	Food	Leu_Fru
Leuconostoc mesenteroides	ATCC-8293	1978	Fermenting olives	2.08	37.67	2086	NA	2	NA	C.008496..0085:	Food	Leu_Fru
Leuconostoc mesenteroides, clemoris	ATCC-19254	1929	Hansen's dried starter powder	1.74	37.9	1791	NA	29	NA	C2KK01	Unknown	Leu_Fru
Leuconostoc pseudomesenteroides	4882	N/A	N/A	2.01	39.06	2180	NA	106	NA	CAKV00000000	Food	Leu_Fru
Oenococcus kitharae	DSM-17330	2006	Distilled residue of shochu mushes	1.84	42.68	1900	NA	2	NA	AFV20000000	Wine product	Oenococcus
Oenococcus oeni	ATCC-BAA-1163	2002	Fermented beverages	1.75	37.94	2055	NA	62	NA	AAU00000000	Wine product	Oenococcus
Oenococcus oeni	AS1-2696	N/A	Human gingival crevicle	1.85	46.99	1888	97.25	13	ERX359771	JQAZ00000000	Animal	
Pediococcus acidilactici	DSM-23206	2008	Fermented wheat flour	1.76	36.67	1772	56.66	93	SRX456746	JQCB00000000	Unknown	Pediococcus
Pediococcus acidilactici	DSM-17757	2005	Distilled piri-fermenting cellar	2.05	39.04	1974	49.06	22	SRX456747	JQBRE00000000	Wine product	Pediococcus
Pediococcus clausenii	DSM-14802	2002	Spoiled beer	1.88	36.74	1807	53.21	44	SRX456748	JQBRE00000000	Wine product	Pediococcus
Pediococcus damnosus	DSM-20336	1903	Lager beer yeast	2.19	38.23	2085	45.63	201	SRX456749	JQBD00000000	Wine product	Pediococcus
Pediococcus halophilodurans	DSM-22300	2006	Walls of a distilled-spirit-fermenting cellar	2.26	37.18	2180	44.23	66	SRX456750	JQBY00000000	Wine product	Pediococcus
Pediococcus insipidus	DSM-19395	1988	Brewery yeast	2.11	38.61	2081	47.40	157	SRX456751	JQCC00000000	Wine product	Pediococcus
Pediococcus loli	DSM-19227	1887	Ryegrass silage	2.04	42.13	1971	49.00	31	SRX456752	JQCC00000000	Plant	Pediococcus
Pediococcus parvulus	DSM-20323	1961	Silage	3.99	40.38	3917	25.04	153	SRX456754	JQBEE00000000	Plant	Pediococcus
Pediococcus pentosaceus	DSM-20336	1934	Dried American beer yeast	1.74	37.25	1687	57.49	28	SRX456755	JQBPF00000000	Wine product	Pediococcus
Pediococcus stilesii	DSM-18001	2006	White maize grains	1.84	38.11	1834	54.41	47	SRX456756	JQBX00000000	Plant	Pediococcus
Weissella confusa	DSM-20106	1982	Sugar cane	2.22	44.73	2075	90.16	40	ERX359760	JQAY00000000	Plant	Weissella
Weissella confusa	DSM-20190	1985	Sap	1.37	45.06	1341	146.41	12	SRX359698	JQAZ00000000	Food	Weissella
Weissella kandleri	DSM-20593	1982	Desert spring	1.33	39.67	1281	149.88	21	ERX359708	JQBPH00000000	Environment	Weissella
Weissella minor	DSM-20014	1983	Milking machine slime	1.77	39.29	1777	112.85	59	ERX399773	JQCD00000000	Food	Weissella
Weissella viridescens	DSM-20410	1957	Cured meat products	1.54	41.09	1525	130.11	8	ERX359781	JQBM00000000	Food	Weissella

Table S2. Sequence information for the 73 core genes

PID*	Gene	locus tag*	COG	Annotation	Co-ordinates*	Strand*	Length*
90960992	<i>dnaN</i>	LSL_0002	COG0592L	DNA polymerase III subunit beta	1532..2671	+	1140
90960995	<i>gyrB</i>	LSL_0005	COG0187L	DNA gyrase subunit B	4451..6409	+	1959
90960996	<i>gyrA</i>	LSL_0006	COG0188L	DNA gyrase subunit A	6446..8998	+	2553
90960997	<i>rpsF</i>	LSL_0007	COG0360J	30S ribosomal protein S6	9217..9507	+	291
90960998	<i>ssb</i>	LSL_0008	COG0629L	Single-strand DNA binding protein	9548..10099	+	552
90960999	<i>rpsR</i>	LSL_0009	COG0238J	30S ribosomal protein S18	10121..10357	+	237
90961180	<i>rpoC</i>	LSL_0198	COG0086K	DNA-directed RNA polymerase subunit beta'	238653..242318	+	3666
90961182	<i>rpsL</i>	LSL_0200	COG0048J	30S ribosomal protein S12	243364..243777	+	414
90961184	<i>efg</i>	LSL_0202	COG0480J	elongation factor G	244403..246496	+	2094
90961200	<i>trpS</i>	LSL_0218	COG0180J	Tryptophanyl-tRNA synthetase II	271973..272992	+	1020
90961335	<i>murF</i>	LSL_0355	COG0770M	UDP-N-acetyl muramoyl-tripeptide-D-alanyl-D-alanine ligase	387857..389227	+	1371
90961453	-	LSL_0477	COG0537FGR	bis(5'-nucleosyl)-tetraphosphatase	525874..526302	-	429
90961460	-	LSL_0484	COG0073R	tRNA-binding domain-containing protein	530707..531357	+	651
90961464	<i>polA</i>	LSL_0488	COG0749L	DNA polymerase I	536545..539223	+	2679
90961470	<i>thrS</i>	LSL_0494	COG0441J	threonyl-tRNA synthetase	543759..545708	+	1950
90961471	<i>infC</i>	LSL_0495	COG0290J	translation initiation factor IF-3	545916..546440	+	525
90961480	-	LSL_0504	COG0799S	Iojap-related protein	552036..552389	+	354
90961487	<i>rpsB</i>	LSL_0511	COG0052J	30S ribosomal protein S2	557234..558031	+	798
90961488	<i>tsf</i>	LSL_0512	COG264J	elongation factor Ts	558125..559000	+	876
90961519	-	LSL_0543	COG0218R	GTP-binding protein	584473..585063	+	591
90961537	<i>pyrH</i>	LSL_0561	COG0528F	uridylate kinase	600230..600952	+	723
90961538	<i>frr</i>	LSL_0562	COG0233J	ribosome recycling factor	600955..601518	+	564
90961539	<i>uppS</i>	LSL_0563	COG0020I	undecaprenyl pyrophosphate synthetase	601653..602435	+	783
90961540	<i>cdsA</i>	LSL_0564	COG0575I	phosphatidate cytidylyltransferase	602438..603226	+	789
90961544	-	LSL_0568	COG0779S	hypothetical protein	610934..611407	+	474
90961545	<i>nusA</i>	LSL_0569	COG0195K	transcription elongation factor NusA	611433..612560	+	1128
90961567	-	LSL_0591	COG2890J	peptide release factor-glutamine N5-methyltransferase	634747..635586	+	840
90961569	<i>upp</i>	LSL_0593	COG0035F	uracil phosphoribosyltransferase	636708..637337	+	630
90961575	<i>atpG</i>	LSL_0599	COG0224C	F0F1 ATP synthase subunit gamma	641185..642114	+	930
90961576	<i>atpD</i>	LSL_0600	COG0055C	F0F1 ATP synthase subunit beta	642139..643545	+	1407
90961630	<i>typA</i>	LSL_0653	COG1217T	GTP-binding protein	697034..698875	+	1842
90961790	<i>pheS</i>	LSL_0813	COG0016J	phenylalanyl-tRNA synthetase subunit alpha	829184..830230	+	1047
90961836	<i>pfs</i>	LSL_0859	COG0775F	5'-methylthioadenosine nucleosidase	878731..879417	+	687
90962022	<i>ftsZ</i>	LSL_1047	COG0206D	cell division protein FtsZ	1070613..1071866	-	1254
90962026	<i>murD</i>	LSL_1051	COG0771M	UDP-N-acetyl muramoyl-L-alanyl-D-glutamate synthetase	1075412..1076788	-	1377
90962027	<i>mraY</i>	LSL_1052	COG0472M	phospho-N-acetyl muramoyl-pentapeptide-transferase	1076817..1077785	-	969
90962042	<i>mreC</i>	LSL_1067	COG1792M	rod shape-determining protein MreC	1089421..1090275	-	855
90962072	<i>obgE</i>	LSL_1097	COG0536R	GTPase ObgE	1125579..1126877	-	1299
90962084	-	LSL_1109	COG0816L	Holliday junction resolvase-like protein	1136102..1136533	-	432
90962100	<i>ruvB</i>	LSL_1125	COG2255L	Holliday junction DNA helicase RuvB	1153506..1154516	-	1011
90962101	<i>ruvA</i>	LSL_1126	COG0632L	Holliday junction DNA helicase RuvA	1154555..1155160	-	606
90962106	<i>pgsA</i>	LSL_1131	COG0558I	CDP-diacylglycerol-glycerol-3-phosphate 3-phosphatidyltransferase	1163042..1163629	-	588
90962110	<i>ftsK</i>	LSL_1135	COG1674D	cell division protein	1168139..1170418	-	2280
90962125	-	LSL_1150	COG0802R	ATP/GTP hydrolase	1183489..1183941	-	453
90962130	<i>smpB</i>	LSL_1155	COG0691O	SsrA-binding protein	1187273..1187740	-	468
90962160	<i>prfB</i>	LSL_1185	COG1186J	peptide chain release factor 2	1222388..1223455	-	1068
90962186	<i>groEL</i>	LSL_1211	COG0459O	molecular chaperone GroEL	1246385..1248007	-	1623
90962187	<i>groS</i>	LSL_1212	COG0234O	molecular chaperone GroES	1248037..1248321	-	285
90962191	<i>gcp</i>	LSL_1216	COG0533O	O-sialoglycoprotein endopeptidase	1252076..1253107	-	1032
90962198	<i>holB</i>	LSL_1223	COG2812L	DNA polymerase III subunit delta'	1257561..1258553	-	993
90962199	<i>tmk</i>	LSL_1224	COG125F	thymidylate kinase	1258588..1259214	-	627
90962204	-	LSL_1229	COG0590FJ	cytosine/adenosine deaminase	1262491..1262991	-	501
90962212	<i>rplJ</i>	LSL_1238	COG0244J	50S ribosomal protein L10	1271311..1271814	-	504
90962215	<i>nusG</i>	LSL_1241	COG0250K	transcription antitermination protein	1273402..1273962	-	561
90962219	<i>spoU</i>	LSL_1245	COG0566J	tRNA/rRNA methyltransferase	1275013..1275759	-	747
90962221	<i>cysS</i>	LSL_1247	COG0215J	cysteinyl-tRNA synthetase	1276159..1277571	-	1413
90962278	<i>ppnK</i>	LSL_1304	COG0061G	inorganic polyphosphate/ATP-NAD kinase	1351254..1352060	-	807
90962327	-	LSL_1355	COG0037D	tRNA(Ile)-lysidine synthase TisS	1418710..1420068	-	1359
90962332	<i>mfd</i>	LSL_1360	COG197LK	transcription-repair coupling factor	1422969..1426493	-	3525
90962333	<i>pth</i>	LSL_1361	COG0193J	peptidyl-tRNA hydrolase	1426515..1427072	-	558
90962374	<i>rplM</i>	LSL_1403	COG0102J	50S ribosomal protein L13	1478312..1478755	-	444
90962376	<i>cbiQ</i>	LSL_1405	COG0619P	cobalt transport permease	1479676..1480470	-	795
90962380	<i>rpoA</i>	LSL_1409	COG0202K	DNA-directed RNA polymerase subunit alpha	1482888..1483832	-	945
90962387	<i>rplO</i>	LSL_1416	COG0200J	50S ribosomal protein L15	1487364..1487798	-	435
90962389	<i>rpsE</i>	LSL_1418	COG0098J	30S ribosomal protein S5	1488030..1488530	-	501
90962391	<i>rplF</i>	LSL_1420	COG0097J	50S ribosomal protein L6	1488953..1489489	-	537
90962392	<i>rpsH</i>	LSL_1421	COG0096J	30S ribosomal protein S8	1489522..1489920	-	399
90962394	<i>rplE</i>	LSL_1423	COG0094J	50S ribosomal protein L5	1490153..1490695	-	543
90962400	<i>rpsC</i>	LSL_1429	COG0092J	30S ribosomal protein S3	1492402..1493058	-	657
90962404	<i>rplW</i>	LSL_1433	COG0089J	50S ribosomal protein L23	1494625..1494909	-	285
90962405	<i>rplD</i>	LSL_1434	COG0088J	50S ribosomal protein L4	1494909..1495532	-	624
90962406	<i>rplC</i>	LSL_1435	COG0087J	50S ribosomal protein L3	1495557..1496180	-	624
90962696	<i>rplI</i>	LSL_1727	COG0359J	50S ribosomal protein L9	1809124..1809573	-	450

*These columns are provided according to the reference genome *L. salivarius* UCC118

Table S3. Genera used in building the tree of bacteria

Accession No.	Phylum	Class	Order	Family	Genus	Species
NC_012483	Acidobacteria	Acidobacteria	Acidobacterales	Acidobacteriaceae	<i>Acidobacterium</i>	<i>Acidobacterium capsulatum</i>
NC_014963	Acidobacteria	Acidobacteria	Acidobacterales	Acidobacteriaceae	<i>Terriglobus</i>	<i>Terriglobus saanensis</i>
NC_015064	Acidobacteria	Acidobacteria	Acidobacterales	Acidobacteriaceae	<i>Granulicella</i>	<i>Granulicella tundricola</i>
NC_008536	Acidobacteria	Solibacteres	Solibacterales	Solibacteraceae	<i>Candidatus Solibacter</i>	<i>Candidatus Solibacter usitatus</i>
NC_008009	Acidobacteria	Unclassified	Unclassified	Unclassified	<i>Candidatus Koribacter</i>	<i>Candidatus Koribacter versatilis</i>
NC_009962	Actinobacteria	Actinobacteria	Actinomycetales	Mycobacteriaceae	<i>Mycobacterium</i>	<i>Mycobacterium tuberculosis</i>
NC_002935	Actinobacteria	Actinobacteria	Actinomycetales	Corynebacteriaceae	<i>Corynebacterium</i>	<i>Corynebacterium diphtheriae</i>
NC_003155	Actinobacteria	Actinobacteria	Actinomycetales	Streptomycetaceae	<i>Streptomyces</i>	<i>Streptomyces avermitilis</i>
NC_004307	Actinobacteria	Actinobacteria	Bifidobacterales	Bifidobacteriaceae	<i>Bifidobacterium</i>	<i>Bifidobacterium longum</i>
NC_004551	Actinobacteria	Actinobacteria	Actinomycetales	Unclassified	<i>Tropheryma</i>	<i>Tropheryma whipplei</i>
NC_006085	Actinobacteria	Actinobacteria	Actinomycetales	Propionibacteriaceae	<i>Propionibacterium</i>	<i>Propionibacterium acnes</i>
NC_006087	Actinobacteria	Actinobacteria	Actinomycetales	Microbacteriaceae	<i>Leifsonia</i>	<i>Leifsonia xyli</i>
NC_006361	Actinobacteria	Actinobacteria	Actinomycetales	Nocardiaceae	<i>Nocardia</i>	<i>Nocardia farcinica</i>
NC_007333	Actinobacteria	Actinobacteria	Actinomycetales	Nocardiopsaceae	<i>Thermobifida</i>	<i>Thermobifida fusca</i>
NC_007777	Actinobacteria	Actinobacteria	Actinomycetales	Frankiaceae	<i>Frankia</i>	<i>Unclassified</i>
NC_008148	Actinobacteria	Actinobacteria	Rubrobacterales	Rubrobacteraceae	<i>Rubrobacter</i>	<i>Rubrobacter xylanophilus</i>
NC_008268	Actinobacteria	Actinobacteria	Actinomycetales	Nocardiaceae	<i>Rhodococcus</i>	<i>Rhodococcus jostii</i>
NC_008541	Actinobacteria	Actinobacteria	Actinomycetales	Micrococcaceae	<i>Arthrobacter</i>	<i>Unclassified</i>
NC_008578	Actinobacteria	Actinobacteria	Actinomycetales	Acidothermaceae	<i>Acidothermus</i>	<i>Acidothermus cellulolyticus</i>
NC_008699	Actinobacteria	Actinobacteria	Actinomycetales	Noardioidaceae	<i>Noardioides</i>	<i>Noardioides</i>
NC_009142	Actinobacteria	Actinobacteria	Actinomycetales	Pseudonocardiaceae	<i>Saccharopolyspora</i>	<i>Saccharopolyspora erythraea</i>
NC_009380	Actinobacteria	Actinobacteria	Actinomycetales	Micromonosporaceae	<i>Salinispora</i>	<i>Salinispora tropica</i>
NC_009480	Actinobacteria	Actinobacteria	Actinomycetales	Microbacteriaceae	<i>Clavibacter</i>	<i>Clavibacter michiganensis</i>
NC_009664	Actinobacteria	Actinobacteria	Actinomycetales	Kineosporiaceae	<i>Kineococcus</i>	<i>Kineococcus radiotolerans</i>
NC_010168	Actinobacteria	Actinobacteria	Actinomycetales	Micrococcaceae	<i>Renibacterium</i>	<i>Renibacterium salmoninarum</i>
NC_010617	Actinobacteria	Actinobacteria	Actinomycetales	Micrococcaceae	<i>Kocuria</i>	<i>Kocuria rhizophila</i>
NC_012669	Actinobacteria	Actinobacteria	Actinomycetales	Beutenbergiaceae	<i>Beutenbergia</i>	<i>Beutenbergia cavernae</i>
NC_012803	Actinobacteria	Actinobacteria	Actinomycetales	Micrococcaceae	<i>Micrococcus</i>	<i>Micrococcus luteus</i>
NC_013093	Actinobacteria	Actinobacteria	Actinomycetales	Pseudonocardiaceae	<i>Actinosynema</i>	<i>Actinosynema mirum</i>
NC_013124	Actinobacteria	Actinobacteria	Acidimicrobiales	Acidimicrobiaceae	<i>Acidimicrobium</i>	<i>Acidimicrobium ferrooxidanis</i>
NC_013131	Actinobacteria	Actinobacteria	Actinomycetales	Catenulisporeae	<i>Catenulispora</i>	<i>Catenulispora acidiphila</i>
NC_013159	Actinobacteria	Actinobacteria	Actinomycetales	Pseudonocardiaceae	<i>Saccharomonospora</i>	<i>Saccharomonospora viridis</i>
NC_013165	Actinobacteria	Actinobacteria	Coriobacterales	Coriobacteriaceae	<i>Slackia</i>	<i>Slackia heliotrinireducens</i>
NC_013169	Actinobacteria	Actinobacteria	Actinomycetales	Dermacoccaceae	<i>Kytococcus</i>	<i>Kytococcus sedentarius</i>
NC_013170	Actinobacteria	Actinobacteria	Coriobacterales	Coriobacteriaceae	<i>Cryptobacterium</i>	<i>Cryptobacterium curtum</i>
NC_013172	Actinobacteria	Actinobacteria	Actinomycetales	Dermabacteraceae	<i>Brachybacterium</i>	<i>Brachybacterium faecium</i>
NC_013174	Actinobacteria	Actinobacteria	Actinomycetales	Jonesiaceae	<i>Jonesia</i>	<i>Jonesia denitrificans</i>
NC_013203	Actinobacteria	Actinobacteria	Coriobacterales	Coriobacteriaceae	<i>Atopobium</i>	<i>Atopobium parvulum</i>
NC_013204	Actinobacteria	Actinobacteria	Coriobacterales	Coriobacteriaceae	<i>Eggerthella</i>	<i>Eggerthella lenta</i>
NC_013235	Actinobacteria	Actinobacteria	Actinomycetales	Nakamurellaceae	<i>Nakamurella</i>	<i>Nakamurella multipartita</i>
NC_013510	Actinobacteria	Actinobacteria	Actinomycetales	Thermomonosporaceae	<i>Thermomonospora</i>	<i>Thermomonospora curvata</i>
NC_013521	Actinobacteria	Actinobacteria	Actinomycetales	Sanguibacteraceae	<i>Sanguibacter</i>	<i>Sanguibacter keddlei</i>
NC_013530	Actinobacteria	Actinobacteria	Actinomycetales	Promicromonosporaceae	<i>Xylanimonas</i>	<i>Xylanimonas cellulolysitica</i>
NC_013595	Actinobacteria	Actinobacteria	Actinomycetales	Streptosporangiaceae	<i>Streptosporangium</i>	<i>Streptosporangium roseum</i>
NC_013715	Actinobacteria	Actinobacteria	Actinomycetales	Micrococcaceae	<i>Rothia</i>	<i>Rothia mucilaginosa</i>
NC_013721	Actinobacteria	Actinobacteria	Bifidobacterales	Bifidobacteriaceae	<i>Gardnerella</i>	<i>Gardnerella vaginalis</i>
NC_013739	Actinobacteria	Actinobacteria	Solirubrobacterales	Conexibacteraceae	<i>Conexibacter</i>	<i>Conexibacter woesei</i>
NC_013947	Actinobacteria	Actinobacteria	Actinomycetales	Glycomycetaceae	<i>Stackebrandtia</i>	<i>Stackebrandtia nassauensis</i>
NC_014151	Actinobacteria	Actinobacteria	Actinomycetales	Cellulomonadaceae	<i>Cellulomonas</i>	<i>Cellulomonas flavigena</i>
NC_014158	Actinobacteria	Actinobacteria	Actinomycetales	Tsukamurellaceae	<i>Tsukamurella</i>	<i>Tsukamurella paurometabola</i>
NC_014165	Actinobacteria	Actinobacteria	Actinomycetales	Pseudonocardiaceae	<i>Thermobispora</i>	<i>Thermobispora bispora</i>
NC_014168	Actinobacteria	Actinobacteria	Actinomycetales	Segniliparaceae	<i>Segniliparus</i>	<i>Segniliparus rotundus</i>
NC_014218	Actinobacteria	Actinobacteria	Actinomycetales	Actinomycetaceae	<i>Arcanobacterium</i>	<i>Arcanobacterium haemolyticum</i>
NC_014246	Actinobacteria	Actinobacteria	Actinomycetales	Actinomycetaceae	<i>Mobiluncus</i>	<i>Mobiluncus curtisi</i>
NC_014318	Actinobacteria	Actinobacteria	Actinomycetales	Pseudonocardiaceae	<i>Amycolatopsis</i>	<i>Amycolatopsis mediterranei</i>
NC_014363	Actinobacteria	Actinobacteria	Coriobacterales	Coriobacteriaceae	<i>Olsenella</i>	<i>Olsenella uli</i>
NC_014830	Actinobacteria	Actinobacteria	Actinomycetales	Intrasporangiaceae	<i>Intrasporangium</i>	<i>Intrasporangium calvum</i>
NC_015125	Actinobacteria	Actinobacteria	Actinomycetales	Microbacteriaceae	<i>Microbacterium</i>	<i>Microbacterium testaceum</i>
NC_015312	Actinobacteria	Actinobacteria	Actinomycetales	Pseudonocardiaceae	<i>Pseudonocardia</i>	<i>Pseudonocardia diaxanivorans</i>
NC_015389	Actinobacteria	Actinobacteria	Coriobacterales	Coriobacteriaceae	<i>Coriobacterium</i>	<i>Coriobacterium glomerans</i>
NC_015564	Actinobacteria	Actinobacteria	Actinomycetales	Mycobacteriaceae	<i>Amycolicicoccus</i>	<i>Amycolicicoccus subflavus</i>
NC_015588	Actinobacteria	Actinobacteria	Actinomycetales	Promicromonosporaceae	<i>Isoptericola</i>	<i>Isoptericola variabilis</i>
NC_015635	Actinobacteria	Actinobacteria	Actinomycetales	Propionibacteriaceae	<i>Microlunatus</i>	<i>Microlunatus phosphovorus</i>
NC_000918	Aquificae	Aquificae	Aquificales	Aquificaceae	<i>Aquifex</i>	<i>Aquifex aeolicus</i>
NC_010730	Aquificae	Aquificae	Aquificales	Hydrogenothermaceae	<i>Sulfurhydrogenibium</i>	<i>Unclassified</i>
NC_011126	Aquificae	Aquificae	Aquificales	Aquificaceae	<i>Hydrogenobaculum</i>	<i>Unclassified</i>
NC_012440	Aquificae	Aquificae	Aquificales	Hydrogenothermaceae	<i>Persephonella</i>	<i>Persephonella marina</i>
NC_013799	Aquificae	Aquificae	Aquificales	Aquificaceae	<i>Hydrogenobacter</i>	<i>Hydrogenobacter thermophilus</i>
NC_013894	Aquificae	Aquificae	Aquificales	Aquificaceae	<i>Thermocrinis</i>	<i>Thermocrinis albus</i>
NC_014926	Aquificae	Aquificae	Aquificales	Desulfurobacteriaceae	<i>Thermovibrio</i>	<i>Thermovibrio ammonificans</i>
NC_015185	Aquificae	Aquificae	Aquificales	Desulfurobacteriaceae	<i>Desulfurobacterium</i>	<i>Desulfurobacterium thermolithotrophum</i>
NC_002950	Bacteroidetes	Bacteroidia	Bacteroidales	Porphyromonadaceae	<i>Porphyromonas</i>	<i>Porphyromonas gingivalis</i>
NC_003228	Bacteroidetes	Bacteroidia	Bacteroidales	Bacteroidaceae	<i>Bacteroides</i>	<i>Bacteroides fragilis</i>
NC_009615	Bacteroidetes	Bacteroidia	Bacteroidales	Porphyromonadaceae	<i>Parabacteroides</i>	<i>Parabacteroides distasonis</i>
NC_011565	Bacteroidetes	Bacteroidia	Bacteroidales	Unclassified	<i>Candidatus Azobacteroides</i>	<i>Candidatus Azobacteroides pseudotrichonymphae</i>
NC_014033	Bacteroidetes	Bacteroidia	Bacteroidales	Prevotellaceae	<i>Prevotella</i>	<i>Prevotella ruminicola</i>
NC_014734	Bacteroidetes	Bacteroidia	Bacteroidales	Porphyromonadaceae	<i>Paludibacter</i>	<i>Paludibacter propionicigenes</i>
NC_015160	Bacteroidetes	Bacteroidia	Bacteroidales	Porphyromonadaceae	<i>Odoribacter</i>	<i>Odoribacter splanchnicus</i>
NC_008255	Bacteroidetes	Cytophagia	Cytophagales	Cytophagaceae	<i>Cytophaga</i>	<i>Cytophaga hutchinsonii</i>
NC_013037	Bacteroidetes	Cytophagia	Cytophagales	Cytophagaceae	<i>Dyadobacter</i>	<i>Dyadobacter fermentans</i>
NC_013730	Bacteroidetes	Cytophagia	Cytophagales	Cytophagaceae	<i>Spirosoma</i>	<i>Spirosoma linguale</i>
NC_014655	Bacteroidetes	Cytophagia	Cytophagales	Cytophagaceae	<i>Leadbetterella</i>	<i>Leadbetterella byssophila</i>
NC_014759	Bacteroidetes	Cytophagia	Cytophagales	Flammeovirgaceae	<i>Marivirga</i>	<i>Marivirga tractiosa</i>
NC_015703	Bacteroidetes	Cytophagia	Cytophagales	Cytophagaceae	<i>Ranella</i>	<i>Ranella sithyformis</i>
NC_015914	Bacteroidetes	Cytophagia	Cytophagales	Cyclobacteriaceae	<i>Cyclobacterium</i>	<i>Cyclobacterium marinum</i>
NC_008571	Bacteroidetes	Flavobacteria	Flavobacterales	Flavobacteriaceae	<i>Gramella</i>	<i>Gramella forsetii</i>
NC_009441	Bacteroidetes	Flavobacteria	Flavobacterales	Flavobacteriaceae	<i>Flavobacterium</i>	<i>Flavobacterium johnsoniae</i>

NC_013525	Unclassified	Unclassified	Unclassified	Unclassified	<i>Thermobaculum</i>	<i>Thermobaculum terrenum</i>
NC_010571	Verrucomicrobia	Opitutae	Unclassified	Opitutaceae	<i>Opitutus</i>	<i>Opitutus terrae</i>
NC_014008	Verrucomicrobia	Opitutae	Puniceicoccales	Puniceicoccaceae	<i>Coraliomargarita</i>	<i>Coraliomargarita akajimensis</i>
NC_010794	Verrucomicrobia	Unclassified	Methylacidiphilales	Methylacidiphilaceae	<i>Methylacidiphilum</i>	<i>Methylacidiphilum infernorum</i>
NC_010655	Verrucomicrobia	Verrucomicrobiae	Verrucomicrobiales	Verrucomicrobiaceae	<i>Akkermansia</i>	<i>Akkermansia muciniphila</i>

Table S4. List of genes used in building the tree of bacteria

PID	Gene	Locus tag*	COG	Predicted product	Position*	Length (bp)*
90961179	<i>rpoB</i>	LSL_0197	COG0085K	DNA-directed RNA polymerase subunit beta	235025..238624	3600
90961471	<i>infC</i>	LSL_0495	COG0290J	translation initiation factor IF-3	545916..546440	525
90961487	<i>rpsB</i>	LSL_0511	COG0052J	30S ribosomal protein S2	557234..558031	798
90961488	<i>tsf</i>	LSL_0512	COG0264J	elongation factor Ts	558125..559000	876
90961538	<i>frr</i>	LSL_0562	COG0233J	ribosome recycling factor	600955..601518	564
90961545	<i>nusA</i>	LSL_0569	COG0195K	transcription elongation factor NusA	611433..612560	1128
90962130	<i>smpB</i>	LSL_1155	COG0691O	SsrA-binding protein	1187273..1187740	468
90962374	<i>rplM</i>	LSL_1403	COG0102J	50S ribosomal protein L13	1478312..1478755	444
90962389	<i>rpsE</i>	LSL_1418	COG0098J	30S ribosomal protein S5	1488030..1488530	501
90962391	<i>rplF</i>	LSL_1420	COG0097J	50S ribosomal protein L6	1488953..1489489	537
90962394	<i>rplE</i>	LSL_1423	COG0094J	50S ribosomal protein L5	1490153..1490695	543
90962399	<i>rplP</i>	LSL_1428	COG0197J	50S ribosomal protein L16	1491964..1492398	435
90962400	<i>rpsC</i>	LSL_1429	COG0092J	30S ribosomal protein S3	1492402..1493058	657
90962403	<i>rplB</i>	LSL_1432	COG0090J	50S ribosomal protein L2	1493766..1494599	834
90962405	<i>rplD</i>	LSL_1434	COG0088J	50S ribosomal protein L4	1494909..1495532	624
90962406	<i>rplC</i>	LSL_1435	COG0087J	50S ribosomal protein L3	1495557..1496180	624

*These columns are provided according to the reference genome *L. salivarius* UCC118

Lactobacillus koreensis	JCM_16448	9	JCM_16448GL000985, JCM_16448GL001438, JCM_16448GL001675, JCM_16448GL000224, JCM_16448GL000225	Yes	1	1	
Lactobacillus zymelactis	DSM-19394	14	DSM_19394GL000101, DSM_19394GL000381, DSM_19394GL000905	No	0	0	
Lactobacillus naumannii	DSM-19117	15	DSM_19117GL000223, DSM_19117GL000216, DSM_19117GL	Yes	0	0	
Lactobacillus spicheri	DSM-15429	15	DSM_15429GL000237, DSM_15429GL001364, DSM_15429GL	No	0	0	
Lactobacillus kimchicus	JCM_15530	7	JCM_15530GL0002468, JCM_15530GL000731, JCM_15530GL	Yes	0	0	
Lactobacillus similis	DSM-23365	21	DSM_23365GL000101, DSM_23365GL000121, DSM_23365GL	Yes	0	0	
Lactobacillus plantarum	DSM-19999	20	DSM_19999GL000101, DSM_19999GL000120, DSM_19999GL	Yes	0	0	
Lactobacillus collinensis	DSM-20515	10	DSM_20515GL000349, DSM_20515GL000267, DSM_20515GL	Yes	0	0	
Lactobacillus paracollinensis	DSM-15920	15	DSM_15920GL000136, DSM_15920GL000701, DSM_15920GL	Yes	0	0	
Lactobacillus malefermentans	DSM-5705	12	DSM_5705GL000172, DSM_5705GL000489, DSM_5705GL000	No	0	0	
Lactobacillus paradoxus	DSM-1507	9	DSM_1507GL000101, DSM_1507GL000100, DSM_1507GL000	No	0	0	
Lactobacillus pentosevorus	DSM-15352	11	DSM_15352GL000152, DSM_15352GL000258, DSM_15352GL	No	0	0	
Lactobacillus buchneri	DSM-20057	5	DSM_20057GL000250, DSM_20057GL000220, DSM_20057GL	Yes	1	1	DSM_15352GL001527-DSM_15352GL001531
Lactobacillus raffinolactis	DSM-19908	6	DSM_19908GL000070, DSM_19908GL000706, DSM_19908GL	Yes	0	0	
Lactobacillus kefir	DSM-18857	3	DSM_18857GL000101, DSM_18857GL000100, DSM_18857GL	No	0	0	
Lactobacillus sakei	DSM-10551	6	DSM_10551GL000197, DSM_10551GL000184, DSM_10551GL	Yes	0	0	
Lactobacillus sakeiakai	DSM-19994	4	DSM_19994GL000561, DSM_19994GL000217, DSM_19994GL	No	0	0	
Lactobacillus rapi	DSM-19907	6	DSM_19907GL000146, DSM_19907GL0002649, DSM_19907GL	Yes	0	0	
Lactobacillus diastaticus	DSM-19906	6	DSM_19906GL000249, DSM_19906GL000206, DSM_19906GL	Yes	0	0	
Lactobacillus faragaldii	DSM-18716	4	DSM_18716GL000142, DSM_18716GL000260, DSM_18716GL	Yes	0	0	
Lactobacillus parafarraginis	DSM-18717	2	DSM_18717GL000183, DSM_20716GL000973	Yes	0	0	
Lactobacillus sensitis	DSM-24392	3	DSM_24392GL0001476, DSM_24392GL0001193, DSM_24392GL	Yes	1	1	DSM_24302GL001193-DSM_24302GL001196
Lactobacillus kunkelii	DSM-12361	6	DSM_12361GL000115, DSM_12361GL0001310, DSM_12361GL	Yes	1	1	
Lactobacillus florum	DSM-22689	6	DSM_22689GL000154, DSM_22689GL000195, DSM_22689GL	No	0	0	
Lactobacillus lindneri	DSM-20699	6	DSM_20699GL000120, DSM_20699GL0001342, DSM_20699GL	No	0	0	
Lactobacillus acidilactici	DSM-15131	4	DSM_15131GL000101, DSM_15131GL000100, DSM_15131GL	No	0	0	
Lactobacillus fructivorans	ATCC-27394	9	ATCC_27394GL0008993, ATCC_27394GL000880, ATCC_27394	Yes	0	0	
Lactobacillus homothrochi	DSM-20571	9	DSM_20571GL000572, DSM_20571GL000664, DSM_20571GL	Yes	0	0	
Lactobacillus fructivora	DSM-20350	8	DSM_20350GL000774, DSM_20350GL000324, DSM_20350GL	Yes	0	0	
Lactobacillus plantarum	DSM-20303	8	DSM_20303GL0001943, DSM_20303GL0001944, DSM_20303GL	Yes	0	0	
Pediococcus argentinicus	DSM-20326	4	DSM_20326GL000175, DSM_20326GL000244, DSM_20326GL	No	0	0	
Pediococcus classenii	DSM-14800	3	DSM_14800GL000637, DSM_14800GL001642, DSM_14800GL	Yes	1	1	DSM_14800GL001113-DSM_14800GL001115
Pediococcus pentosaceus	DSM-20336	3	DSM_20336GL000909, DSM_20346GL000913, DSM_20336GL	No	0	0	
Pediococcus stilesii	DSM-18001	0		Yes	0	0	
Pediococcus acidilactici	DSM-19977	3	DSM_19977GL000147, DSM_19977GL0001600, DSM_19977GL	No	0	0	
Pediococcus acidilactici	AS1-26996	3	AS1-26996GL0001049, AS1-26996GL0001783, AS1-26996GL000415	Yes	0	0	
Pediococcus ethanolidans	DSM-22301	9	DSM_22301GL000128, DSM_22301GL000004, DSM_22301GL	Yes	1	1	DSM_22301GL000125-DSM_22301GL000128
Pediococcus cellicola	DSM-17757	7	DSM_17757GL000421, DSM_17757GL001524, DSM_17757GL	No	0	0	
Pediococcus cellicola	DSM-17758	10	DSM_17758GL000100, DSM_17758GL000101, DSM_17758GL	Yes	1	1	DSM_2031GL001009-DSM_2031GL001093
Pediococcus inopinatus	DSM-20285	10	DSM_20285GL000008, DSM_20285GL000010, DSM_20285GL	Yes	1	1	DSM_20285GL000846-DSM_20285GL000849
Pediococcus parvulus	DSM-20332	10	DSM_20332GL002774, DSM_20332GL003420, DSM_20332GL	Yes	1	2	DSM_20332GL003313-DSM_20332GL003316, DSM_20332GL003316
Carnobacterium malabaricum	DSM-20730	22	DSM_20730GL000263, DSM_20730GL000714, DSM_20730GL	Yes	0	0	
Carnobacterium malabaricum	DSM-20342	27	DSM_20342GL002038, DSM_20342GL002040, DSM_20342GL	Yes	1	1	DSM_20342GL002041-DSM_20342GL002944
Carnobacterium malabaricum	DSM-20722	22	DSM_20722GL000804, DSM_20722GL000805, DSM_20722GL	Yes	1	1	DSM_20722GL000754-DSM_20722GL000757
Carnobacterium divergens	DSM-20623	18	DSM_20623GL000163, DSM_20623GL000792, DSM_20623GL	Yes	1	1	
Lactococcus lactis	LMG-7760	7	LMG_7760GL001946, LMG_7760GL001608, LMG_7760GL00	Yes	1	1	LMG_7760GL001946-LMG_7760GL001949
Atopobium minutum	DSM-20586	9	DSM_20586GL001354, DSM_20586GL000632, DSM_20586GL	No	0	0	
Olsenella uili	DSM-7084	5	DSM_7084GL001434, DSM_7084GL000839, DSM_7084GL001	Yes	1	1	DSM_7084GL001746-DSM_7084GL001749
Aerococcus rimae	DSM-7090	0		No	0	0	
Total		1628		57	51	67	

Footnote: The gene sequences for all locus tags listed above are provided as supplementary datasets 3 and 4.

* Cells colored in light gray indicate strain harboring at least 1 pilus gene cluster with a similar gene order to *L. rhamnosus* strain GG.

Table S6. Distribution and abundance of cell envelope proteins and associated anchoring domains and motifs

Species name	StrainID	CEPs	Cell anchor type		
			LPxTG	SLAP	T
Kandleria vitulina	DSM-20405	0	0	0	0
Lactobacillus kimchiensis	DSM-24716	0	0	0	0
Lactobacillus mindensis	DSM-14500	0	0	0	0
Lactobacillus nantensis	DSM-16982	0	0	0	0
Lactobacillus crustorum	LMG-23699	0	0	0	0
Lactobacillus crustorum	JCM-15951	0	0	0	0
Lactobacillus futsaii	JCM-17355	0	0	0	0
Lactobacillus farciminis	DSM-20184	0	0	0	0
Lactobacillus alimentarius	DSM-20249	0	0	0	0
Lactobacillus paralimentarius	DSM-19674	0	0	0	0
Lactobacillus paralimentarius	DSM-13961	0	0	0	0
Lactobacillus paralimentarius	DSM-13238	0	0	0	0
Lactobacillus tucceti	DSM-20183	0	0	0	0
Lactobacillus nodensis	DSM-19682	0	0	0	0
Lactobacillus versmoldensis	DSM-14857	0	0	0	0
Lactobacillus floricola	DSM-23037	0	0	0	0
Lactobacillus acidophilus	DSM-20079	1	0	1	0
Lactobacillus amylovorus	DSM-20531	0	0	0	0
Lactobacillus amylovorus	DSM-16698	0	0	0	0
Lactobacillus kitasatonis	DSM-16761	0	0	0	0
Lactobacillus kefiranofaciens kefiranofaciens	DSM-5016	2	0	2	0
Lactobacillus kefiranofaciens kefirgranum	DSM-10550	0	0	0	0
Lactobacillus ultunensis	DSM-16047	1	0	1	0
Lactobacillus helveticus	LMG-22464	1	0	1	0
Lactobacillus helveticus	CGMCC-1.1877	1	0	1	0
Lactobacillus gallinarum	DSM-10532	1	0	1	0
Lactobacillus crispatus	DSM-20584	0	0	0	0
Lactobacillus acetotolerans	DSM-20749	0	0	0	0
Lactobacillus intestinalis	DSM-6629	1	0	1	0
Lactobacillus hamsteri	DSM-5661	0	0	0	0
Lactobacillus amyloyticus	DSM-11664	0	0	0	0
Lactobacillus kalixensis	DSM-16043	1	0	1	0
Lactobacillus gigerorum	DSM-23908	1	0	1	0
Lactobacillus pasteurii	DSM-23907	0	0	0	0
Lactobacillus delbrueckii jakobsenii	DSM-26046	1	0	1	0
Lactobacillus delbrueckii lacticis	DSM-20072	1	0	1	0
Lactobacillus delbrueckii bulgaricus	DSM-20081	1	0	1	0
Lactobacillus delbrueckii delbrueckii	DSM-20074	1	0	1	0
Lactobacillus delbrueckii indicus	DSM-15996	1	0	1	0
Lactobacillus equicursoris	DSM-19284	1	0	1	0
Lactobacillus jensenii	DSM-20557	0	0	0	0
Lactobacillus psittaci	DSM-15354	0	0	0	0
Lactobacillus hominis	DSM-23910	1	0	1	0
Lactobacillus taiwanensis	DSM-21401	1	1	0	0
Lactobacillus johnsonii	ATCC-33200	0	0	0	0
Lactobacillus gasseri	ATCC-33323	0	0	0	0
Lactobacillus iners	DSM-13335	0	0	0	0
Lactobacillus amylophilicus	DSM-20534	0	0	0	0
Lactobacillus amylophilus	DSM-20533	0	0	0	0
Lactobacillus dextrinicus	DSM-20335	0	0	0	0
Lactobacillus concavus	DSM-17758	1	1	0	0
Lactobacillus composti	DSM-18527	0	0	0	0
Lactobacillus harbinensis	DSM-16991	0	0	0	0
Lactobacillus perlens	DSM-12744	1	0	0	1
Lactobacillus camelliae	DSM-22697	2	2	0	0
Lactobacillus nasuensis	JCM-17158	1	1	0	0
Lactobacillus manihotivorans	DSM-13343	1	1	0	0
Lactobacillus paracasei paracasei	DSM-5622	2	2	0	0
Lactobacillus casei	DSM-20011	1	1	0	0
Lactobacillus paracasei tolerans	DSM-20258	0	0	0	0
Lactobacillus zea	DSM-20178	2	2	0	0
Lactobacillus rhamnosus	DSM-20021	2	2	0	0
Lactobacillus saniviri	DSM-24301	0	0	0	0
Lactobacillus brantae	DSM-23927	0	0	0	0
Lactobacillus thailandensis	DSM-22698	0	0	0	0
Lactobacillus pantheris	DSM-15945	1	1	0	0
Lactobacillus sharpeae	DSM-20505	0	0	0	0
Lactobacillus selangorensis	DSM-13344	0	0	0	0
Lactobacillus selangorensis	ATCC-BAA-66	0	0	0	0
Lactobacillus graminis	DSM-20719	0	0	0	0
Lactobacillus curvatus	DSM-20019	0	0	0	0
Lactobacillus sakei sakei	DSM-20017	0	0	0	0
Lactobacillus sakei carnosus	DSM-15831	0	0	0	0
Lactobacillus fuchuensis	DSM-14340	0	0	0	0
Lactobacillus rennini	DSM-20253	0	0	0	0
Lactobacillus coryniformis torquens	DSM-20004	0	0	0	0
Lactobacillus coryniformis coryniformis	DSM-20001	0	0	0	0
Lactobacillus bifermentans	DSM-20003	0	0	0	0
Lactobacillus ceti	DSM-22408	0	0	0	0
Lactobacillus saerimneri	DSM-16049	0	0	0	0
Lactobacillus animalis	DSM-20602	1	1	0	0
Lactobacillus murinus	DSM-20452	1	1	0	0
Lactobacillus apodemi	DSM-16634	1	1	0	0
Lactobacillus ruminis	DSM-20403	1	1	0	0
Lactobacillus agilis	DSM-20509	0	0	0	0
Lactobacillus equi	DSM-15833	1	1	0	0
Lactobacillus salivarus	DSM-20555	1	1	0	0
Lactobacillus hayakitensis	DSM-18933	0	0	0	0
Lactobacillus pobuzihii	NBRC-103219	0	0	0	0

Lactobacillus pobuzihii.Chen	KCTC-13174	0	0	0
Lactobacillus acidipiscis	DSM-15836	0	0	0
Lactobacillus acidipiscis	DSM-15353	0	0	0
Lactobacillus aviarius aviarius	DSM-20655	0	0	0
Lactobacillus aviarius araffinosis	DSM-20653	0	0	0
Lactobacillus sucicola	DSM-21376	0	0	0
Lactobacillus aquaticus	DSM-21051	0	0	0
Lactobacillus uvarum	DSM-19971	0	0	0
Lactobacillus capillatus	DSM-19910	0	0	0
Lactobacillus cacaonum	DSM-21116	0	0	0
Lactobacillus mali	DSM-20444	0	0	0
Lactobacillus mali	ATCC-27304	0	0	0
Lactobacillus hordei	DSM-19519	0	0	0
Lactobacillus oeni	DSM-19972	0	0	0
Lactobacillus satsumensis	DSM-16230	0	0	0
Lactobacillus vini	DSM-20605	0	0	0
Lactobacillus ghanensis	DSM-18630	0	0	0
Lactobacillus nagelii	DSM-13675	0	0	0
Lactobacillus algidus	DSM-15638	0	0	0
Lactobacillus fabifementans	DSM-21115	0	0	0
Lactobacillus xiangfangensis	LMG-26013	0	0	0
Lactobacillus pentosus	DSM-20314	0	0	0
Lactobacillus plantarum argentoratensis	DSM-16365	0	0	0
Lactobacillus plantarum	DSM-13273	0	0	0
Lactobacillus plantarum plantarum	CGMCC-1.2437	0	0	0
Lactobacillus paraplanтарum	DSM-10667	0	0	0
Lactobacillus siliginis	DSM-22696	0	0	0
Lactobacillus rossiae	DSM-15814	0	0	0
Weissella viridescens	DSM-20410	0	0	0
Weissella minor	DSM-20014	0	0	0
Weissella halotolerans	DSM-20190	0	0	0
Weissella confusa	DSM-20196	0	0	0
Weissella kandleri	DSM-20593	0	0	0
Oenococcus oeni	ATCC-BAA-1163	0	0	0
Oenococcus kitaharae	DSM-17330	0	0	0
Leuconostoc fallax	KCTC-3537	0	0	0
Leuconostoc pseudomesenteroides	4882-	1	0	1
Leuconostoc mesenteroides	ATCC-8293	0	0	0
Leuconostoc mesenteroides cremoris	ATCC-19254	0	0	0
Leuconostoc carnosum	JB16-	0	0	0
Leuconostoc argentinum	KCTC-3773	0	0	0
Leuconostoc citreum	KM20-	0	0	0
Leuconostoc gelidum	KCTC-3527	0	0	0
Leuconostoc gasicomitatum	LMG-18811	0	0	0
Leuconostoc kimchii	IMSNU-11154	0	0	0
Fructobacillus fructosus	DSM-20349	0	0	0
Lactobacillus pontis	DSM-8475	0	0	0
Lactobacillus panis	DSM-6035	0	0	0
Lactobacillus oris	DSM-4864	0	0	0
Lactobacillus antri	DSM-16041	0	0	0
Lactobacillus reuteri	DSM-20016	0	0	0
Lactobacillus vaginalis	DSM-5837	0	0	0
Lactobacillus frumenti	DSM-13145	0	0	0
Lactobacillus fermentum	DSM-20055	0	0	0
Lactobacillus equigenerosi	DSM-18793	0	0	0
Lactobacillus gastricus	DSM-16045	0	0	0
Lactobacillus ingluviei	DSM-15946	0	0	0
Lactobacillus ingluviei	DSM-14792	0	0	0
Lactobacillus secaliphilus	DSM-17896	0	0	0
Lactobacillus coleohominis	DSM-14060	0	0	0
Lactobacillus mucosae	DSM-13345	0	0	0
Lactobacillus oligofermentans	DSM-15707	0	0	0
Lactobacillus hokkaidonensis	DSM-26202	0	0	0
Lactobacillus suebicus	DSM-5007	0	0	0
Lactobacillus vaccinostercus	DSM-20634	0	0	0
Lactobacillus parabrevis	LMG-11984	0	0	0
Lactobacillus parabrevis	ATCC-53295	0	0	0
Lactobacillus hammesii	DSM-16381	0	0	0
Lactobacillus paucivorans	DSM-22467	0	0	0
Lactobacillus sennaizukei	DSM-21775	0	0	0
Lactobacillus brevis	DSM-20054	0	0	0
Lactobacillus koreensis	JCM-16448	0	0	0
Lactobacillus zymae	DSM-19395	0	0	0
Lactobacillus acidifarinae	DSM-19394	0	0	0
Lactobacillus namurensis	DSM-19117	0	0	0
Lactobacillus spicheri	DSM-15429	0	0	0
Lactobacillus kimchicus	JCM-15530	0	0	0
Lactobacillus similis	DSM-23365	0	0	0
Lactobacillus odoratitofui	DSM-19909	0	0	0
Lactobacillus collinoides	DSM-20515	1	1	0
Lactobacillus paracollinoides	DSM-15502	0	0	0
Lactobacillus malefermentans	DSM-5705	0	0	0
Lactobacillus parabuchneri	DSM-5707	1	0	0
Lactobacillus parabuchneri	DSM-15352	1	0	0
Lactobacillus buchneri	DSM-20057	1	0	0
Lactobacillus otakiensis	DSM-19908	1	0	0
Lactobacillus kefiri	DSM-20587	1	0	0
Lactobacillus parakefiri	DSM-10551	2	0	2
Lactobacillus sunkii	DSM-19904	1	0	1
Lactobacillus rapi	DSM-19907	0	0	0
Lactobacillus kisonensis	DSM-19906	0	0	0
Lactobacillus diolivovans	DSM-14421	2	1	1
Lactobacillus hilgardii	DSM-20176	0	0	0
Lactobacillus farraginis	DSM-18382	1	0	1

Lactobacillus parafarraginis	DSM-18390	1	0	0	1
Lactobacillus senioris	DSM-24302	0	0	0	0
Lactobacillus ozensis	DSM-23829	0	0	0	0
Lactobacillus kunkeei	DSM-12361	0	0	0	0
Lactobacillus florum	DSM-22689	0	0	0	0
Lactobacillus findneri	DSM-20690	0	0	0	0
Lactobacillus sanfranciscensis	DSM-20451	0	0	0	0
Lactobacillus fructivorans	ATCC-27394	0	0	0	0
Lactobacillus homohiochii	DSM-20571	0	0	0	0
Lactobacillus fructivorans	DSM-20350	0	0	0	0
Lactobacillus fructivorans	DSM-20203	0	0	0	0
Pediococcus argentinicus	DSM-23026	0	0	0	0
Pediococcus claussenii	DSM-14800	0	0	0	0
Pediococcus pentosaceus	DSM-20336	0	0	0	0
Pediococcus stilesii	DSM-18001	0	0	0	0
Pediococcus lolii	DSM-19927	0	0	0	0
Pediococcus acidilactici	AS1-2696	0	0	0	0
Pediococcus ethanolidurans	DSM-22301	1	1	0	0
Pediococcus cellicola	DSM-17757	1	1	0	0
Pediococcus damnosus	DSM-20331	0	0	0	0
Pediococcus inopinatus	DSM-20285	0	0	0	0
Pediococcus parvulus	DSM-20332	0	0	0	0
Carnobacterium maltaromaticum	DSM-20730	1	1	0	0
Carnobacterium maltaromaticum	DSM-20342	1	1	0	0
Carnobacterium maltaromaticum	DSM-20722	1	1	0	0
Carnobacterium divergens	DSM-20623	1	1	0	0
Lactococcus lactis	LMG-7760	0	0	0	0
Atopobium minutum	DSM-20586	2	2	0	0
Olsenella uli	DSM-7084	0	0	0	0
Atopobium rimae	DSM-7090	0	0	0	0
Total		60	30	17	13

Table S8. Sequence information for the 27 core partial genes

PID*	Gene	LSL locus tag*	COG	Annotation	Co-ordinates*	Strand*	Length*
90960991	<i>dnaA</i>	LSL_0001	COG0593L	chromosomal replication initiation protein	1..1365	+	454
90960994	<i>recF</i>	LSL_0004	COG1195L	recombination protein F	3309..4448	+	379
90961461	<i>murC</i>	LSL_0485	COG0773M	UDP-N-acetylmuramate-L-alanine ligase	533809..535140	+	443
90961551	<i>ribF</i>	LSL_0575	COG0196H	riboflavin kinase/FMN adenyllyltransferase	616816..617772	+	318
90961553	<i>grpE</i>	LSL_0577	COG0576O	GrpE protein HSP-70 cofactor	618942..619538	+	198
90961554	<i>dnaK</i>	LSL_0578	COG0443O	molecular chaperone DnaK	619577..621424	+	615
90961556	<i>lepA</i>	LSL_0580	COG0481M	GTP-binding protein LepA	622843..624672	+	609
90961566	<i>prfA</i>	LSL_0590	COG0216J	peptide chain release factor 1	633672..634754	+	360
90961572	<i>atpF</i>	LSL_0596	COG0711C	ATP synthase subunit B	638577..639104	+	175
90961615	<i>rpsO</i>	LSL_0638	COG0184J	30S ribosomal protein S15	681489..681758	+	89
90962017	<i>ileS</i>	LSL_1042	COG0060J	isoleucyl-tRNA synthetase	1065203..1067998	-	931
90962030	<i>mraW</i>	LSL_1055	COG0275M	S-adenosyl-methyltransferase MraW	1080326..1081270	-	314
90962140	<i>pgk</i>	LSL_1165	COG0126G	phosphoglycerate kinase	1199234..1200436	-	400
90962149	<i>uvrA</i>	LSL_1174	COG0178L	excinuclease ABC subunit A	1208147..1210981	-	944
90962150	<i>uvrB</i>	LSL_1175	COG0556L	excinuclease ABC subunit B	1211000..1213000	-	666
90962161	<i>secA</i>	LSL_1186	COG0653U	preprotein translocase subunit SecA	1223594..1225957	-	787
90962203	<i>dnaX</i>	LSL_1228	COG2812L	DNA polymerase III subunit gamma/tau	1260545..1262284	-	579
90962211	<i>rplL</i>	LSL_1237	COG0222J	50S ribosomal protein L7/L12	1270871..1271239	-	122
90962242	<i>galU</i>	LSL_1268	COG1210M	UTP--glucose-1-phosphate uridylyltransferase	1304213..1305085	-	290
90962313	<i>trmA</i>	LSL_1341	COG2265J	tRNA (Uracil-5-) -methyltransferase	1392529..1393908	-	459
90962321	<i>pcrA</i>	LSL_1349	COG0210L	ATP-dependent DNA helicase	1403083..1405317	-	744
90962375	<i>truA</i>	LSL_1404	COG0101J	tRNA pseudouridine synthase A	1478890..1479660	-	256
90962386	<i>secY</i>	LSL_1415	COG0201U	preprotein translocase subunit SecY	1486063..1487361	-	432
90962403	<i>rplB</i>	LSL_1432	COG0090J	50S ribosomal protein L2	1493766..1494599	-	277
90962565	<i>parB</i>	LSL_1596	COG1475K	chromosome partitioning protein, DNA-binding protein	1676145..1677020	-	291
90962568	<i>gidB</i>	LSL_1599	COG0357M	16S rRNA methyltransferase GidB	1678668..1679393	-	241
90962695	<i>dnaB</i>	LSL_1726	COG0305L	replicative DNA helicase	1807560..1808951	-	463

*These columns are provided according to the reference genome *L. salivarius* UCC118

Lactobacillus koreensis	JCM-16448	A	A	A	A	A	P	P	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	
Lactobacillus zymae	DSM-19395	A	A	A	A	A	P	P	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	
Lactobacillus acidifarinae	DSM-19394	A	A	A	A	A	A	P	P	A	A	A	A	A	A	A	A	A	A	A	A	A	A	
Lactobacillus kimchiae	DSM-15477	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	
Lactobacillus spicheri	DSM-15429	A	A	A	A	A	A	P	P	A	A	A	P	P	P	P	A	A	A	A	A	A	A	
Lactobacillus kimchiaeus	JCM-15520	P	P	P	P	P	P	P	A	A	P	P	P	P	P	P	P	A	A	A	A	P	P	P
Lactobacillus simulis	DSM-23365	P	P	P	P	P	P	P	A	A	P	P	P	P	P	P	P	A	A	A	P	P	P	P
Lactobacillus odoratofaci	DSM-19909	P	A	A	A	A	P	P	P	A	A	P	P	P	P	P	P	A	A	A	P	P	P	P
Lactobacillus collinooides	DSM-20515	P	P	P	P	P	P	P	P	A	A	P	P	P	P	P	P	A	A	A	P	P	P	P
Lactobacillus paracollinoides	DSM-15502	P	P	P	P	P	P	P	P	A	A	P	P	P	P	P	P	A	A	A	P	P	P	P
Lactobacillus magnamentans	DSM-5705	A	A	A	A	A	A	P	P	A	A	A	A	A	A	A	P	A	A	A	P	B	P	P
Lactobacillus parabuchneri	DSM-5379	P	P	P	P	P	P	P	P	A	A	A	A	A	A	P	P	A	A	A	P	A	P	P
Lactobacillus parabuchneri	DSM-15352	P	A	P	A	P	P	P	A	A	A	A	A	A	A	P	A	A	A	A	P	A	A	A
Lactobacillus buekneri	DSM-20057	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Lactobacillus otakiensis	DSM-19901	P	A	A	A	P	P	P	A	A	P	A	A	A	P	A	A	A	A	A	P	A	P	P
Lactobacillus kefiri	DSM-20587	A	A	A	A	A	P	P	A	A	A	A	A	A	A	A	P	A	A	A	P	A	P	P
Lactobacillus parakefiri	DSM-10551	P	A	A	A	A	P	P	P	A	A	A	P	P	P	P	P	A	A	A	P	P	P	P
Lactobacillus saekkii	DSM-12014	A	A	A	A	A	P	P	P	A	A	A	A	A	A	P	P	A	A	A	P	A	A	A
Lactobacillus spp.	DSM-19907	P	P	P	P	P	P	P	P	A	A	A	A	A	A	P	P	A	A	A	P	A	P	P
Lactobacillus koreanus	DSM-19906	A	A	A	A	A	P	P	P	A	A	A	A	A	A	P	P	P	A	A	A	A	A	A
Lactobacillus diolivans	DSM-14421	P	A	A	A	P	P	P	A	A	A	P	P	P	P	P	P	A	A	A	P	A	P	P
Lactobacillus hilgardi	DSM-20176	P	P	P	P	P	P	P	A	A	A	P	P	P	P	P	P	A	A	A	P	A	P	P
Lactobacillus faraginis	DSM-18382	A	A	A	A	A	A	P	P	P	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Lactobacillus parafaraginis	DSM-18390	P	P	P	P	P	P	P	A	A	A	P	P	P	P	P	P	A	A	A	P	A	P	P
Lactobacillus senioris	DSM-13024	A	A	A	A	A	P	P	A	A	A	A	A	A	A	P	A	A	A	A	A	A	A	A
Lactobacillus sanguinis	DSM-23893	A	A	A	A	A	A	P	P	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Lactobacillus tankevi	DSM-12361	A	A	A	A	A	P	P	P	A	A	A	A	A	A	A	A	A	A	A	P	A	A	A
Lactobacillus florum	DSM-22689	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Lactobacillus lindneri	DSM-20690	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Lactobacillus sanfranciscensis	DSM-20451	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Lactobacillus fructivorans	ATCC-27394	A	A	A	A	A	A	F	F	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Lactobacillus fructochoci	DSM-20171	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Lactobacillus fructivorans	DSM-20350	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Lactobacillus fructivorans	DSM-20203	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Pediococcus argentinicus	DSM-23026	A	A	A	A	A	P	P	A	A	A	A	A	A	A	P	A	A	A	A	A	A	A	A
Pediococcus clausenii	DSM-14800	A	A	A	A	A	P	P	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	P
Pediococcus pentaseucus	DSM-20336	A	A	A	A	A	A	P	P	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Pediococcus stilesii	DSM-18001	A	A	A	A	A	A	P	P	P	A	A	A	A	A	A	A	P	A	A	A	A	P	A
Pediococcus acidilactici	DSM-19927	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Pediococcus acidilactici	AS1-2695	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Pediococcus othmoliudurans	DSM-22301	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Pediococcus cellicola	DSM-17757	A	A	A	A	A	P	P	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	P
Pediococcus damnosus	DSM-20331	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Pediococcus inopinatus	DSM-20285	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Pediococcus parvulus	DSM-20720	A	A	A	A	A	A	A	P	P	A	A	A	A	A	A	A	A	A	A	P	A	A	P
Carnobacterium malonaticum	DSM-20342	A	P	P	P	P	P	P	A	A	P	A	A	A	A	P	P	P	A	A	P	A	A	P
Carnobacterium malonaticum	DSM-20722	A	P	P	P	P	P	P	A	A	A	A	A	A	A	P	P	P	A	A	P	A	A	P
Carnobacterium divergens	DSM-20623	A	P	P	P	P	P	P	P	P	A	A	A	P	P	P	P	A	A	A	P	P	A	A
Lactococcus lactis	LMG-7760	P	P	P	P	P	P	P	A	A	A	P	P	P	P	P	P	P	A	A	P	A	P	P
Atopobium minutum	DSM-20586	A	A	A	A	A	A	P	P	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Olsentella ili	DSM-7084	P	A	A	A	P	P	A	P	A	A	P	P	A	A	A	A	A	A	A	P	A	A	P
Atopobium rimae	DSM-17901	A	A	A	A	P	P	A	P	A	A	A	P	A	A	A	A	A	A	A	P	A	A	P

Table S10. Presence of sirtuin homologs in the 213 genomes analyzed

Species Name	StrainID	SIR2 family proteins (homolog of mammalian SIRT1)
		SIR2L1, Sir2α
<i>Kandleria vitulina</i>	DSM-20405	2
<i>Lactobacillus kimchiensis</i>	DSM-24716	1
<i>Lactobacillus mindensis</i>	DSM-14500	1
<i>Lactobacillus nantensis</i>	DSM-16982	1
<i>Lactobacillus crustorum</i>	LMG-23699	1
<i>Lactobacillus crustorum</i>	JCM-15951	1
<i>Lactobacillus futsaii</i>	JCM-17355	3
<i>Lactobacillus farciminis</i>	DSM-20184	2
<i>Lactobacillus alimentarius</i>	DSM-20249	2
<i>Lactobacillus paralimentarius</i>	DSM-19674	2
<i>Lactobacillus paralimentarius</i>	DSM-13961	2
<i>Lactobacillus paralimentarius</i>	DSM-13238	1
<i>Lactobacillus tucctei</i>	DSM-20183	2
<i>Lactobacillus nodensis</i>	DSM-19682	1
<i>Lactobacillus versmoldensis</i>	DSM-14857	3
<i>Lactobacillus floricola</i>	DSM-23037	0
<i>Lactobacillus acidophilus</i>	DSM-20079	1
<i>Lactobacillus amylovorus</i>	DSM-20531	1
<i>Lactobacillus amylovorus</i>	DSM-16698	1
<i>Lactobacillus kitasatonis</i>	DSM-16761	1
<i>Lactobacillus kefiranofaciens kefiranofaciens</i>	DSM-5016	3
<i>Lactobacillus kefiranofaciens kefirgranum</i>	DSM-10550	3
<i>Lactobacillus ultunensis</i>	DSM-16047	4
<i>Lactobacillus helveticus</i>	LMG-22464	3
<i>Lactobacillus helveticus</i>	CGMCC-1.1877	3
<i>Lactobacillus gallinarum</i>	DSM-10532	1
<i>Lactobacillus crispatus</i>	DSM-20584	1
<i>Lactobacillus acetotolerans</i>	DSM-20749	1
<i>Lactobacillus intestinalis</i>	DSM-6629	1
<i>Lactobacillus hamsteri</i>	DSM-5661	3
<i>Lactobacillus amylolyticus</i>	DSM-11664	1
<i>Lactobacillus kalixensis</i>	DSM-16043	1
<i>Lactobacillus gigerorum</i>	DSM-23908	1
<i>Lactobacillus pasteurii</i>	DSM-23907	1
<i>Lactobacillus delbrueckii jakobsenii</i>	DSM-26046	1
<i>Lactobacillus delbrueckii lactis</i>	DSM-20072	1
<i>Lactobacillus delbrueckii bulgaricus</i>	DSM-20081	1
<i>Lactobacillus delbrueckii delbrueckii</i>	DSM-20074	1
<i>Lactobacillus delbrueckii indicus</i>	DSM-15996	1
<i>Lactobacillus equicursoris</i>	DSM-19284	1
<i>Lactobacillus jensenii</i>	DSM-20557	1
<i>Lactobacillus psittaci</i>	DSM-15354	1
<i>Lactobacillus hominis</i>	DSM-23910	1
<i>Lactobacillus taiwanensis</i>	DSM-21401	1

Lactobacillus johnsonii	ATCC-33200	2
Lactobacillus gasseri	ATCC-33323	1
Lactobacillus iners	DSM-13335	0
Lactobacillus amylo trophicus	DSM-20534	1
Lactobacillus amylophilus	DSM-20533	1
Lactobacillus dextrinic us	DSM-20335	1
Lactobacillus concavus	DSM-17758	1
Lactobacillus composti	DSM-18527	1
Lactobacillus harbinensis	DSM-16991	1
Lactobacillus perolens	DSM-12744	1
Lactobacillus camelliae	DSM-22697	1
Lactobacillus nasuensis	JCM-17158	1
Lactobacillus manihotivorans	DSM-13343	1
Lactobacillus paracasei paracasei	DSM-5622	1
Lactobacillus casei	DSM-20011	1
Lactobacillus paracasei tolerans	DSM-20258	1
Lactobacillus zae	DSM-20178	2
Lactobacillus rhamnosus	DSM-20021	2
Lactobacillus saniviri	DSM-24301	1
Lactobacillus brantae	DSM-23927	1
Lactobacillus thailandensis	DSM-22698	1
Lactobacillus pantheris	DSM-15945	1
Lactobacillus sharpeae	DSM-20505	0
Lactobacillus selangorensis	DSM-13344	1
Lactobacillus selangorensis	ATCC-BAA-66	1
Lactobacillus graminis	DSM-20719	3
Lactobacillus curvatus	DSM-20019	3
Lactobacillus sakei sakei	DSM-20017	0
Lactobacillus sakei carnosus	DSM-15831	0
Lactobacillus fuchuensis	DSM-14340	0
Lactobacillus rennini	DSM-20253	1
Lactobacillus coryniformis torquens	DSM-20004	2
Lactobacillus coryniformis coryniformis	DSM-20001	1
Lactobacillus bif fermentans	DSM-20003	1
Lactobacillus ceti	DSM-22408	1
Lactobacillus saerimneri	DSM-16049	0
Lactobacillus animalis	DSM-20602	0
Lactobacillus murinus	DSM-20452	0
Lactobacillus apodemi	DSM-16634	1
Lactobacillus ruminis	DSM-20403	1
Lactobacillus agilis	DSM-20509	1
Lactobacillus equi	DSM-15833	2
Lactobacillus salivarius	DSM-20555	1
Lactobacillus hayakitensis	DSM-18933	0
Lactobacillus pobuzihii	NBRC-103219	1
Lactobacillus pobuzihii.Chen	KCTC-13174	1
Lactobacillus acidipiscis	DSM-15836	1

<i>Lactobacillus acidipiscis</i>	DSM-15353	1
<i>Lactobacillus aviarius aviarius</i>	DSM-20655	1
<i>Lactobacillus aviarius araffinosus</i>	DSM-20653	1
<i>Lactobacillus sucicola</i>	DSM-21376	1
<i>Lactobacillus aquaticus</i>	DSM-21051	1
<i>Lactobacillus uvarum</i>	DSM-19971	1
<i>Lactobacillus capillatus</i>	DSM-19910	1
<i>Lactobacillus cacaonum</i>	DSM-21116	0
<i>Lactobacillus mali</i>	DSM-20444	0
<i>Lactobacillus mali</i>	ATCC-27304	0
<i>Lactobacillus hordei</i>	DSM-19519	0
<i>Lactobacillus oeni</i>	DSM-19972	1
<i>Lactobacillus satsumensis</i>	DSM-16230	1
<i>Lactobacillus vini</i>	DSM-20605	1
<i>Lactobacillus ghanensis</i>	DSM-18630	1
<i>Lactobacillus nagelii</i>	DSM-13675	1
<i>Lactobacillus algidus</i>	DSM-15638	0
<i>Lactobacillus fabifementans</i>	DSM-21115	1
<i>Lactobacillus xiangfangensis</i>	LMG-26013	1
<i>Lactobacillus pentosus</i>	DSM-20314	1
<i>Lactobacillus plantarum argentoratensis</i>	DSM-16365	1
<i>Lactobacillus plantarum</i>	DSM-13273	1
<i>Lactobacillus plantarum plantarum</i>	CGMCC-1.2437	1
<i>Lactobacillus paraplantarum</i>	DSM-10667	1
<i>Lactobacillus siliginis</i>	DSM-22696	1
<i>Lactobacillus rossiae</i>	DSM-15814	1
<i>Weissella viridescens</i>	DSM-20410	0
<i>Weissella minor</i>	DSM-20014	0
<i>Weissella halotolerans</i>	DSM-20190	1
<i>Weissella confusa</i>	DSM-20196	1
<i>Weissella kandleri</i>	DSM-20593	0
<i>Oenococcus oeni</i>	ATCC-BAA-116	1
<i>Oenococcus kitaharae</i>	DSM-17330	1
<i>Leuconostoc fallax</i>	KCTC-3537	0
<i>Leuconostoc pseudomesenteroides</i>	4882	1
<i>Leuconostoc mesenteroides</i>	ATCC-8293	1
<i>Leuconostoc mesenteroides cremoris</i>	ATCC-19254	1
<i>Leuconostoc carnosum</i>	JB16	0
<i>Leuconostoc argentinum</i>	KCTC-3773	1
<i>Leuconostoc citreum</i>	KM20	1
<i>Leuconostoc gelidum</i>	KCTC-3527	0
<i>Leuconostoc gasicomitatum</i>	LMG-18811	0
<i>Leuconostoc kimchii</i>	IMSNU-11154	0
<i>Fructobacillus fructosus</i>	DSM-20349	1
<i>Lactobacillus pontis</i>	DSM-8475	1
<i>Lactobacillus panis</i>	DSM-6035	2
<i>Lactobacillus oris</i>	DSM-4864	1

<i>Lactobacillus antri</i>	DSM-16041	1
<i>Lactobacillus reuteri</i>	DSM-20016	1
<i>Lactobacillus vaginalis</i>	DSM-5837	1
<i>Lactobacillus frumenti</i>	DSM-13145	2
<i>Lactobacillus fermentum</i>	DSM-20055	1
<i>Lactobacillus equigenerosi</i>	DSM-18793	1
<i>Lactobacillus gastricus</i>	DSM-16045	1
<i>Lactobacillus ingluviei</i>	DSM-15946	1
<i>Lactobacillus ingluviei</i>	DSM-14792	1
<i>Lactobacillus secaliphilus</i>	DSM-17896	1
<i>Lactobacillus coleohominis</i>	DSM-14060	0
<i>Lactobacillus mucosae</i>	DSM-13345	1
<i>Lactobacillus oligofermentans</i>	DSM-15707	1
<i>Lactobacillus hokkaidonensis</i>	DSM-26202	1
<i>Lactobacillus suebicus</i>	DSM-5007	1
<i>Lactobacillus vaccinostercus</i>	DSM-20634	1
<i>Lactobacillus parabrevis</i>	LMG-11984	1
<i>Lactobacillus parabrevis</i>	ATCC-53295	1
<i>Lactobacillus hammesii</i>	DSM-16381	1
<i>Lactobacillus paucivorans</i>	DSM-22467	1
<i>Lactobacillus senmaizukei</i>	DSM-21775	1
<i>Lactobacillus brevis</i>	DSM-20054	1
<i>Lactobacillus koreensis</i>	JCM-16448	1
<i>Lactobacillus zymae</i>	DSM-19395	1
<i>Lactobacillus acidifarinae</i>	DSM-19394	1
<i>Lactobacillus namurensis</i>	DSM-19117	1
<i>Lactobacillus spicheri</i>	DSM-15429	1
<i>Lactobacillus kimchicus</i>	JCM-15530	1
<i>Lactobacillus similis</i>	DSM-23365	1
<i>Lactobacillus odoratitofui</i>	DSM-19909	1
<i>Lactobacillus collinoides</i>	DSM-20515	1
<i>Lactobacillus paracollinoides</i>	DSM-15502	1
<i>Lactobacillus malefermentans</i>	DSM-5705	1
<i>Lactobacillus parabuchneri</i>	DSM-5707	1
<i>Lactobacillus parabuchneri</i>	DSM-15352	1
<i>Lactobacillus buchneri</i>	DSM-20057	1
<i>Lactobacillus otakiensis</i>	DSM-19908	1
<i>Lactobacillus kefiri</i>	DSM-20587	1
<i>Lactobacillus parakefiri</i>	DSM-10551	2
<i>Lactobacillus sunkii</i>	DSM-19904	2
<i>Lactobacillus rapi</i>	DSM-19907	1
<i>Lactobacillus kisonensis</i>	DSM-19906	1
<i>Lactobacillus diolivorans</i>	DSM-14421	1
<i>Lactobacillus hilgardii</i>	DSM-20176	1
<i>Lactobacillus farraginis</i>	DSM-18382	1
<i>Lactobacillus parafarraginis</i>	DSM-18390	2
<i>Lactobacillus senioris</i>	DSM-24302	1

<i>Lactobacillus ozensis</i>	DSM-23829	1
<i>Lactobacillus kunkeei</i>	DSM-12361	1
<i>Lactobacillus florum</i>	DSM-22689	1
<i>Lactobacillus lindneri</i>	DSM-20690	1
<i>Lactobacillus sanfranciscensis</i>	DSM-20451	1
<i>Lactobacillus fructivorans</i>	ATCC-27394	1
<i>Lactobacillus homohiochii</i>	DSM-20571	1
<i>Lactobacillus fructivorans</i>	DSM-20350	1
<i>Lactobacillus fructivorans</i>	DSM-20203	1
<i>Pediococcus argentinicus</i>	DSM-23026	0
<i>Pediococcus claussenii</i>	DSM-14800	0
<i>Pediococcus pentosaceus</i>	DSM-20336	0
<i>Pediococcus stilesii</i>	DSM-18001	0
<i>Pediococcus lolii</i>	DSM-19927	0
<i>Pediococcus acidilactici</i>	AS1-2696	0
<i>Pediococcus ethanolidurans</i>	DSM-22301	0
<i>Pediococcus cellicola</i>	DSM-17757	0
<i>Pediococcus damnosus</i>	DSM-20331	0
<i>Pediococcus inopinatus</i>	DSM-20285	0
<i>Pediococcus parvulus</i>	DSM-20332	0
<i>Carnobacterium maltaromaticum</i>	DSM-20730	0
<i>Carnobacterium maltaromaticum</i>	DSM-20342	0
<i>Carnobacterium maltaromaticum</i>	DSM-20722	0
<i>Carnobacterium divergens</i>	DSM-20623	1
<i>Lactococcus lactis</i>	LMG-7760	0
<i>Atopobium minutum</i>	DSM-20586	0
<i>Olsenella uli</i>	DSM-7084	2
<i>Atopobium rimae</i>	DSM-7090	1

Lactobacillus agilis	DSM-20509	1	Plantacarin A	Unmodified	Yes							
Lactobacillus equi	DSM-15833	0										
Lactobacillus salivarius	DSM-20555	0										
Lactobacillus hayakienensis	DSM-18933	2	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon	Enterolysin	Bacteriocin >10kd	Not a bacteriocin operon				
Lactobacillus pobozhii	NBRC-103219	0										
Lactobacillus pobozhii.Chen	KCTC-13174	0										
Lactobacillus acidipiscis	DSM-15836	3	Plantaricin_K	/Cam	Unmodified	Potential but across multiple contigs	Pediocin/Colicin V	Unmodified	Potential but across multiple co	Pediocin/Colicin V	Unmodified	Potential but across multiple contigs
Lactobacillus acidipiscis	DSM-15353	0										
Lactobacillus aviarus	DSM-20655	0										
Lactobacillus aviarus araffinosis	DSM-20653	0										
Lactobacillus suciola	DSM-21376	0										
Lactobacillus aquaticus	DSM-21051	2	Mundificin_ATO6	Unmodified	Yes	Carnocin_CP52	Unmodified	Not a bacteriocin operon				
Lactobacillus uvarum	DSM-19971	0										
Lactobacillus capillatus	DSM-19910	0										
Lactobacillus cacaonum	DSM-21116	0										
Lactobacillus mali	DSM-20444	1	Carnocin_CP52	Unmodified	Yes							
Lactobacillus mali	ATCC-27304	0										
Lactobacillus hordei	DSM-19519	3	Carnocin_CP52	Unmodified	Potential	Plantacarin	Unmodified	yes	Coagulin	Bacteriocin >10kd	Not a bacteriocin operon	
Lactobacillus oeni	DSM-19972	0										
Lactobacillus satsumensis	DSM-16230	0										
Lactobacillus vini	DSM-20605	0										
Lactobacillus ghanensis	DSM-18630	0										
Lactobacillus nalgaei	DSM-13675	0										
Lactobacillus algidus	DSM-15638	0										
Lactobacillus fab fermentans	DSM-21115	1	Carnocin_CP52	Unmodified	Yes							
Lactobacillus xiangfanensis	LMG-26013	2	Carnocin_CP52	Unmodified	Not a bacteriocin operon	Lactococcin_B	Unmodified	Potential but across multiple contigs				
Lactobacillus pentosus	DSM-20314	2	Plantacarin_A	Unmodified	Yes	Pediocin	Unmodified	Not a bacteriocin operon				
Lactobacillus plantarum argentoratensis	DSM-16365	1	Enterocin		Potential but across multiple contigs							
Lactobacillus plantarum	DSM-13273	1	Plantaricin_K	Glycacin	Yes							
Lactobacillus plantarum plantarum	CGMCC-1,2437	1	Plantaricin_K	Glycycin	Yes							
Lactobacillus paraplantarum	DSM-10667	1	Plantaricin_K	Glycycin	Yes							
Lactobacillus oligensis	DSM-22696	0										
Lactobacillus rosiae	DSM-15814	2	Pediocin	Potential		Lactococcin	Unmodified	Potential but across multiple contigs				
Weissella viridescens	DSM-20410	0										
Weissella minor	DSM-20014	0										
Weissella halotolerans	DSM-20190	0										
Weissella confusa	DSM-20196	0										
Weissella kandleri	DSM-20693	0										
Oenococcus oeni	ATCC-BAA-1163	1	Unknown	Head_to_tail_cyclize	Potential							
Oenococcus kitaharae	DSM-17330	1	Streptolysin	LAPs	Yes							
Leuconostoc fallax	KCTC-3537	1	Lactobin_A	Unmodified	Yes							
Leuconostoc pseudomesenteroides	4882	0										
Leuconostoc mesenteroides	ATCC-8293	0										
Leuconostoc mesenteroides cremoris	ATCC-19254	0										
Leuconostoc carnosum	JB16	0										
Leuconostoc argentatum	KCTC-3773	1	Lactococcin_972	Unmodified	Potential but across multiple contigs							
Leuconostoc citreum	KM20	1	Unknown	Head_to_tail_cyclize	Yes							
Leuconostoc gelidum	KCTC-3527	2	Enterocin	Unmodified	Potential	Penocin_A	Unmodified	Yes				
Leuconostoc gasomitatum	LMG-18811	1	Plantaricin_NC8	Unmodified	Potential							
Leuconostoc kimchi	IMSNU-11154	1	Mesentericin_B105	Unmodified	Yes							
Fructobacillus fructosus	DSM-20349	0										
Lactobacillus pontis	DSM-8475	0										
Lactobacillus panis	DSM-6035	1	Enterolysin_A	Bacteriocin >10kd	Potential							
Lactobacillus oris	DSM-4864	0										
Lactobacillus antri	DSM-16041	2	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon	Enterolysin	Bacteriocin >10kd	Potential				
Lactobacillus reuteri	DSM-20016	2	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon	Bovicin	Bacteriocin >10kd	Not a bacteriocin operon				
Lactobacillus vaginalis	DSM-5837	2	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon				
Lactobacillus fermentum	DSM-13145	1	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon							
Lactobacillus fermentum	DSM-20055	1	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon							
Lactobacillus eutenerosi	DSM-18793	2	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon	Enterolysin	Bacteriocin >10kd	Potential				
Lactobacillus gastricus	DSM-16045	1	Cytolysin_A	Lamipeptide_class	Yes							
Lactobacillus ingluviei	DSM-15946	2	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon	Enterolysin	Bacteriocin >10kd	Potential				
Lactobacillus ingluviei	DSM-14792	1	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon							
Lactobacillus caseophilus	DSM-17896	0										
Lactobacillus caseomimeticus	DSM-14060	0										
Lactobacillus mucosae	DSM-13345	2	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon	Enterolysin	Bacteriocin >10kd	Not a bacteriocin operon				
Lactobacillus oligofermenans	DSM-15707	0										
Lactobacillus holkaidoneensis	DSM-26202	1	Leucocin_A like	Unmodified	Yes							
Lactobacillus subucus	DSM-5007	0										
Lactobacillus vaccinostercus	DSM-20634	0										
Lactobacillus parabrevis	LMG-11984	0										
Lactobacillus parabrevis	ATCC-53295	0										
Lactobacillus hamensis	DSM-16381	0										
Lactobacillus paucivorans	DSM-22467	0										
Lactobacillus semenzae	DSM-21775	0										
Lactobacillus brevis	DSM-20054	0										
Lactobacillus Korensis	JCM-16448	0										
Lactobacillus zymae	DSM-19395	0										
Lactobacillus acidifirinae	DSM-19394	0										
Lactobacillus namurensis	DSM-19117	0										
Lactobacillus spicheri	DSM-15429	0										
Lactobacillus kimchicus	JCM-15530	1	Pediocin	Unmodified	Yes							
Lactobacillus similis	DSM-23365	1	Unknown	Unknown	Potential but across multiple contigs							
Lactobacillus odoratofaciens	DSM-19909	0										
Lactobacillus collinoides	DSM-20515	0										
Lactobacillus paracollinoides	DSM-15502	0										
Lactobacillus mal fermentans	DSM-5705	0										
Lactobacillus parabuchneri	DSM-5707	1	Helveticin_J	Bacteriocin >10kd	Not a bacteriocin operon							

<i>Lactobacillus parabuchneri</i>	DSM-15352	0								
<i>Lactobacillus buchneri</i>	DSM-20057	1	Unknown	Head_to_tail_cyclize_Potential						
<i>Lactobacillus otakensis</i>	DSM-19908	1	Unknown	Head_to_tail_cyclize Not a bacteriocin operon						
<i>Lactobacillus kefiri</i>	DSM-20587	0								
<i>Lactobacillus parakefiri</i>	DSM-10551	0	Unknown	Unmodified	Potential					
<i>Lactobacillus sunkei</i>	DSM-19904	0								
<i>Lactobacillus rapi</i>	DSM-19907	0								
<i>Lactobacillus kisونensis</i>	DSM-19906	0								
<i>Lactobacillus diilverans</i>	DSM-14421	0								
<i>Lactobacillus hilgardi</i>	DSM-20176	0								
<i>Lactobacillus faraginis</i>	DSM-18382	1	Helveticin J	Bacteriocin >10kd	Not a bacteriocin operon					
<i>Lactobacillus parafaraginis</i>	DSM-18390	1	Helveticin J	Bacteriocin >10kd	Potential					
<i>Lactobacillus senioris</i>	DSM-24302	0								
<i>Lactobacillus ozensis</i>	DSM-23829	0								
<i>Lactobacillus kunkeei</i>	DSM-12361	0								
<i>Lactobacillus florum</i>	DSM-22689	0								
<i>Lactobacillus indiferi</i>	DSM-20690	0								
<i>Lactobacillus sanfranciscensis</i>	DSM-20451	0								
<i>Lactobacillus fructivorans</i>	ATCC-27394	0								
<i>Lactobacillus homohiochii</i>	DSM-20571	0								
<i>Lactobacillus fructivorans</i>	DSM-20350	0								
<i>Lactobacillus fructivorans</i>	DSM-20203	0								
<i>Pediococcus argentinicus</i>	DSM-23026	0								
<i>Pediococcus claussenii</i>	DSM-14800	1	SalivaricinA	Lactipeptide	Potential					
<i>Pediococcus pentosaceus</i>	DSM-20336	1	Pediocin	Unmodified	Potential					
<i>Pediococcus stilesii</i>	DSM-18001	2	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon	Carnocin_CPS2	Unmodified	Yes		
<i>Pediococcus lolii</i>	DSM-19927	1	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon					
<i>Pediococcus acidilactici</i>	AS1-2696	0								
<i>Pediococcus ethanolidurans</i>	DSM-22301	2	Pediocin like	Unmodified	Yes	Carnocin_CPS2	Unmodified	Not a bacteriocin operon		
<i>Pediococcus cellicola</i>	DSM-17757	1	Carnocin_CPS2	Unmodified	Potential					
<i>Pediococcus damnosus</i>	DSM-20331	2	Pediocin /Sacacin	Unmodified	Yes	SalivaricinA	Lanthipeptide_class	Not a bacteriocin operon		
<i>Pediococcus inopinatus</i>	DSM-20285	1	Enterolysin_A	Bacteriocin >10kd	Not a bacteriocin operon					
<i>Pediococcus parvulus</i>	DSM-20332	1	Enterolysin_A	Bacteriocin >10kd	Propaghe					
<i>Carmobacterium maltaromaticum</i>	DSM-20730	3	Piscicolin_126	Unmodified	Yes	Cytolysin_ClyLs	Lanthipeptide_class	Potential		
<i>Carmobacterium maltaromaticum</i>	DSM-20342	2	Carnocin_CPS2	Unmodified	Potential	Unknown	Sactipeptides	Not a bacteriocin operon		
<i>Carmobacterium maltaromaticum</i>	DSM-20722	4	Carmobacteriocin_B	Unmodified	Yes	Carmobacteriocin_BM1	Unmodified	Potential	Unknown	Sactipeptides
<i>Carmobacterium divergens</i>	DSM-20623	0								
<i>Lactococcus lactis</i>	LMG-7760	0	Plantacarin A	Unmodified	Not a bacteriocin operon					
<i>Atopobium minutum</i>	DSM-20586	0								
<i>Olsinella uii</i>	DSM-7084	0								
<i>Atopobium rimae</i>	DSM-7090	0								

34 **Supplementary Notes**

35 **Supplementary Note 1**

36 ***Lactobacillus* relatedness to other genera**

37 We also sequenced genomes of type strains from other genera associated with lactic
38 acid bacteria (LAB) including *Carnobacterium* and *Lactococcus* and other genera previously
39 misidentified as LAB, namely, *Atopobium*, *Kandleria* and *Olsenella*. *Carnobacterium* and
40 *Lactococcus* form distinct branches and are classified in the families *Carnobacteriaceae* and
41 *Streptococcaceae*, respectively, revealing closer genetic relatedness to genera other than
42 *Lactobacillus*. *Carnobacterium* is closely related to *Melissococcus* and *Lactococcus* to
43 *Streptococcus* (Fig. 1). This indicates that all currently known LAB descend from the same
44 ancestor as proposed previously¹. The un-relatedness of *Kandleria*, *Atopobium* and *Olsenella*
45 and other LAB is confirmed, with a closer relationship of *Kandleria* with *Erysipelothrix* in the
46 family *Erysipelotrichaceae* of the *Clostridium* subphylum cluster XVII of *Firmicutes*. *Atopobium*
47 and *Olsenella* revealed the greatest genetic distance from the genus *Lactobacillus*; they
48 belong to another phylum, *Actinobacteria*, which belongs to the high-GC, Gram-positive
49 bacteria, the most closely related genus being *Coriobacterium*.

50 **Supplementary Note 2**

51 **Validation of the core genome phylogeny**

52 We validated the branching order of the 73 core gene tree using alternative subsets of
53 gene datasets. We inferred a tree of the *Lactobacillus* genus complex, rooted on *Lactococcus*
54 *lactis*, comprised of 117 genes, and another tree of the *Lactobacillus* genus complex, rooted
55 on the *Carnobacteria*, comprised of 121 genes. We found there were only 4 minor branching
56 alterations. The robustness of the 73 core gene tree was also assessed using a range of
57 different models. Models tested were the WAG, LG, JTT, mtREV and Dayhoff models. The
58 resulting branching order of the strains was consistent across all the models with the exception
59 of mtREV. With this model the positions of *L. salivarius* DSM_20555, *L. hayakitensis*
60 DSM_18933 and *L. acetotolerans* DSM_20749 underwent minor node exchanges within the
61 same clade.

62 The impact of adding partial genes to the core gene dataset was assessed by inferring
63 a phylogeny (Supplementary Fig. 19) from the 73 complete core genes with an additional 27
64 partial core genes (listed in Supplementary Table 8) added to the dataset. The inferred
65 topology was highly congruent with the 73 complete core gene tree with only three very minor
66 branching alterations. Specifically, *L. koreensis* moved one branch within the *L. brevis/L.*
67 *collinoides* group. Minor displacements were also observed for *L. kefiranofaciens* DSM_5016
68 and DSM_10550 within the *L. delbrueckii* group and *L. salivarius* and *L. hayakitensis* within
69 the *L. salivarius* group.

70 **Supplementary Note 3**
71 **Relatedness of *Lactobacillus* species**

72 At the species level, the combination of ANI value and core genome phylogeny could
73 be proposed as the basis for optimal taxonomic classification², and its application could shed
74 new light on several issues including species widely used in the food and probiotic industry. A
75 long-debated case has been that of the *L. casei* group³. We propose that the species *L. casei*
76 and *L. paracasei* should be combined into a single species, *L. casei*, because the pair-wise
77 ANI values between the type strains of *L. casei* and two *L. paracasei* are 98~99%, larger than
78 the “un-official” species’ cut-off value of 95%⁴, and these three strains clustered together as a
79 monophyletic group (Fig. 2; Supplementary Fig. 20). The designation of *L. zeae* has been
80 controversial and there are reports suggesting its classification into the species *L. casei*^{3,5}.
81 However, the ANI value between *L. casei* and *L. zeae* is only 77%~78%, which is similar to
82 the value between *L. casei* and another well-defined species, *L. rhamnosus*. Therefore, our
83 genomic analysis supports retaining *L. zeae* as a single species. Remarkably, ANI values
84 seem not to be related to total DNA-DNA hybridization data reported previously⁵ for similarities
85 of *L. casei* type strain ATCC 393, which corresponds to the strain DSM 20011 included in this
86 study.

87 The genus *Lactobacillus* was recently defined as 16 phylogroups (incl. *Pediococcus*),
88 4 couples (groups containing only two species) and 10 single species⁶. Such delineations are
89 generally supported by the phylogenetic relationships constructed here based on the core
90 proteins, but two modifications are suggested (Fig. 2; Supplementary Fig. 18). Firstly, *L.*
91 *camelliae* could be included in the *L. manihotivorans* group and not in the *L. delbrueckii* group,
92 supported by a mean TNI value of <15% to the group, which is far lower than that within the
93 other *L. delbrueckii* group species (79.5%). Moreover, *L. amylo trophicus* and *L. amylophilus*
94 should also be classified as a single species, *L. amylophilus*, as the two type strains have an
95 ANI value of approximately 100%, although multilocus analysis and DNA-DNA hybridization
96 values suggested their separation.

97 **Supplementary Note 4**
98 **Phylogenomics of glycolysis and hexose fermentation**

99 In species characterized by the presence of Pfk, the distribution of the pyruvate
100 dehydrogenase operon (Pdh; composed of 4 genes) reflected carbohydrate metabolism since
101 it is absent in 57% of obligately homofermentative species while it is present in facultatively
102 heterofermentative members. Consistency was observed between Pdh distribution and
103 phylogenetic groupings: 90% of the species of the *L. delbrueckii* group lack *pdh*, while
104 members of groups like *L. salivarius*, *L. plantarum*, *L. casei* or *L. alimentarius* are
105 characterized by presence of both *pfk* and *pdh*, although they belong to different phenotypic
106 categorizations. Additionally, 80% of the species within the *L. delbrueckii* group lack

glucokinase, the first enzyme of the glycolytic pathway, except for the monophyletic subgroup
107 formed by the *L. delbrueckii* and *L. equicursoris* species. For growth on glucose, a PTS
108 transport system may obviate the need for this enzyme⁷ and most of these species (79%) are
109 classified as obligately homofermentative. Thus, comparative genomic analysis of glycolysis
110 reveals that species inside the historically defined groups have a coherent genotypic
111 background despite metabolic heterogeneity.

112 **Supplementary Note 5**

113 **Carbohydrate active enzymes**

114 Some GH families are present more uniformly across the dataset, indicating the
115 importance of the biotransformations associated with these families. Included in this are
116 enzymes involved in the hydrolysis of peptidoglycan (GH25 and GH73⁸), which play an
117 important role in cell division, growth and preserving cell wall integrity. Bacterial autolysis can
118 have a positive impact in the dairy fermentation process through the enhancement of cheese
119 flavour upon the release of enzymes and amino acids⁹. Starch degradation enzymes are also
120 present almost uniformly across the genome set. α-amylase enzymes are catalysts in the
121 hydrolysis of the α-1,4 glycosidic linkages of starch with GH13 being the main GH family acting
122 on substrates with α-glucoside linkages. α-glucan metabolism is important in the breakdown
123 of resistant starch. Another universal family of GHs is GH65; this family is mainly composed
124 of phosphorylases, including maltose and treholase phosphorylase, which are essential for
125 the survival of lactobacilli in sugar-rich environments such as sourdough, where maltose is the
126 predominant sugar¹⁰.

127 Six clades display unusually high GH abundance, namely, *L. (par)alimentarius*, *L.*
128 *perolens*, *L. plantarum*, *L. rapi*, *L. fructivorans* and *Carnobacterium* spp. The *Weissella* spp.
129 and *L. fructivorans* clade show an unusually low GH gene count. The most abundant GH
130 families are GH1, GH13, GH25 and GH73. GH73, an N-acetylmuramidase, is present in all
131 *Lactobacillus* species except *L. equi*. The four *Carnobacterium* genomes harbor several GH
132 families that are absent in all or most of the other genomes including GH18, GH24, GH84,
133 GH85 and GH119. GH18 enzymes are chitinases, GH24 are lysozymes while GH119
134 enzymes are involved in chitin binding.

135 Some rare GTs emerge in the dataset. *P. lolii* and *P. parvulus* are the only strains that
136 harbour genes for GT12, a ganglioside synthase. This activity is very interesting as it has been
137 reported in only 3 bacterial species and 28 eukaryotes. Production of GT11 may be a strain-
138 specific trait, because a single non-type strain of each of *L. johnsonii*, *L. amylovorus* and *L.*
139 *paraplatnarum* also appear to encode this activity (http://www.cazy.org/GT11_bacteria.html).

140 A number of GT families are present almost uniformly across the dataset. These
141 include GT51, which is involved in peptidoglycan synthesis⁸ and only *L. coleohominis* lacks

142 GT28, a galactosyltransferase involved in cell wall metabolism, suggesting alternative cell wall
143 structure in this vaginal isolate. Other ubiquitous families include more broad-spectrum GTs
144 that have been termed “polyspecific” because of their diverse functionality. Examples include
145 GT4 and GT2, which encompass at least 12 functions including cellulose synthase, chitin
146 synthase and mannosyltransferase¹¹.

147 The abundance of genes encoding carbohydrate transporters correlates strongly with
148 GH abundance and less strongly with GT abundance (Supplementary Fig. 21). The clade
149 distribution of carbohydrate transporter gene counts (Supplementary Fig. 22) mirrors this
150 correlation and highlights the relative abundance of carbohydrate management machinery in
151 the *L. alimentarius*, *L. casei*, *L. mali*, *L. plantarum* and *L. collinoides* clades, the pediococci
152 and carnobacteria. Normalization for genome size (Supplementary Fig. 23) reduces the
153 apparent overabundance of GT genes in some species, but not GH genes (Supplementary
154 Fig. 23), and it is debatable if such normalization is biologically relevant.

155 **Supplementary Note 6**
156 **Metabolic diversity of the lactobacilli**

157 A general representation of the genome content of the lactobacilli as Clusters of
158 Orthologous Groups (COGs; Supplementary Fig 15) reveals unexpected diversity in
159 categories including Transcription, Cell wall biogenesis, Energy production, Co-enzyme
160 transport, and Inorganic ion transport. The *L. plantarum*-related species from *L.*
161 *fabifermentans* through *L. paraplantum* (in Fig. 2) are particularly endowed with genes for
162 carbohydrate and amino acid transport and metabolism. Members of the *L. salivarius* clade
163 have the highest number of genes involved in cell motility and secretion, as expected from
164 previous studies in our group¹². Forty nine (23%) of the genomes screened had none of the
165 complete pathways for production of the 20 standard amino acids (AA), while no single
166 genome harboured the genes to produce all 20 AAs. The highest number of pathways
167 encoded by any one species was sixteen in *L. similis* (Supplementary Table 9). Some
168 phylogenetic clades harbored genes for the production of one or two amino acids; these
169 include *Weissella*, *L. brevis*, *Pediococcus* and *L. sakei*. The clade containing *L. collinoides*
170 and *L. kimchicus* as well as the *L. coryniformis* clade (with the exception of *L. rennini*) are
171 predicted to be prototrophic for at least 14 AAs. In contrast, ten of the twelve dairy isolates
172 have genes for five or fewer complete AA pathways, reflecting their evolution to the AA-rich
173 dairy environment.

174
175 **Supplementary Note 7**
176 **Sortase-anchored proteins**

177 The distribution of LPXTG proteins was not clearly correlated with the evolutionary
178 relationship of lactobacilli. Rather, it seems that the decoration of the cell wall with anchored

proteins is a common feature, indicating that most lactobacilli can establish interactions with the environment. In our dataset of 213 genomes, only twelve bacterial strains do not have LPXTG proteins (Supplementary Table 5). In addition, most lactobacilli genomes harbor at least one gene encoding sortase, presumably the housekeeping sortase. The copy number of sortase genes could be associated to some extent with the presence of pilus gene clusters (PGC) (Fig. 4). These PGCs were identified in 51 strains in total, mostly belonging to six clades. The bacterial species with the most pilus gene clusters was *Lactobacillus sharpeae*, a sewage isolate. We also observed a large diversity of PGCs in terms of gene organization and numbers, suggesting they may have distinct gene order and functions (Supplementary Fig. 10). One of the most common PGC types consists of three pilin genes and one sortase gene, which is similar to the PGC originally reported in *L. rhamnosus* GG¹³.

Supplementary Note 8

The *Lactobacillus* mobilome

The mobile component of the bacterial genome can expand coding capacity as in the case of plasmids and megaplasmids¹⁴ or may be associated with genome decay as in the case of Insertion Sequence (IS) elements¹⁵. Some IS elements exhibit a very limited distribution across the 213 genomes, for example, IS1 which is restricted to only *L. ingluviei* and *L. equi* (Supplementary Fig. 16). IS91 is present in the genomes of only two species, both of dairy-product origin (*L. casei* and *L. paracasei* subsp. *tolerans*), and IS481 is found in only 3 strains (*L. paracollinoides*, *L. farraginis* and *P. inopinatus*) all associated with brewing. IS3, on the other hand, exhibits a much greater distribution being present in almost all groups, with the exception of *Weissella/Leuconostoc*, *L. fructivorans*, some *L. delbrueckii* strains and a few singletons. Some genomes apparently harbor no IS elements, perhaps indicating a more rigid architecture and a selective pressure against the acquisition of such elements. These species include *Atopobium minutum*, *L. fructivorans*, *L. florum*, *L. senoris*, *Weissella viridescens*, *L. cacaonum*, *L. apodemi*, *L. ceti* and *L. brantae*. Of the 18 IS families in this database, the largest number of families found is 13, in the *L. parabuchneri* genome. It is clear that IS elements have played a general role in shaping the diversification and evolution of the lactobacilli.

Phages were detected in the genomes of 195 of the 213 genomes (Supplementary Fig. 13), and only the genomes of *L. floridana*, *L. ingluviei*, *L. psittaci*, *L. sakei* subsp. *sakei*, *L. sanfranciscensis*, *P. cellicola*, *P. clausenii* and *W. halotolerans* lacked homologues of phage proteins. Proteins corresponding to holins and endolysins, and fibers/fiber assembly proteins were apparently under-represented; this is likely due to the fact that holins are not well annotated, whereas fiber proteins are very divergent, and not always present in the phage. It is likely that this analysis under-reports phage genes and that fine tuning of homology cut-offs for individual genes will identify more temperate phage. We identified plasmids in 41% of the

215 213 genomes analysed (Supplementary Fig. 14) with numbers ranging from zero to six. 58%
216 of the plasmid-related genes were of unknown function, emphasizing the need for functional
217 genomics to elucidate this gene repertoire. Given the desirability of finding new plasmid
218 vectors for genetically manipulating lactobacilli in the laboratory and for food-grade strain
219 construction, these genomic data represent a valuable resource. Of the 87 strains predicted
220 to have plasmids, 75 had identifiable replication genes. The other 12 may harbor new
221 replication types that would be compatible with existing plasmid vectors.

222 **Supplementary Note 9**

223 **Stress resistance**

224 The broad range of niches that lactobacilli occupy is reflected in the multiple stress
225 resistance mechanisms their genomes encode (Supplementary Fig. 24). Knowledge of the
226 differential abundance of these systems can be exploited for identifying species and strains
227 that can withstand production stress, storage stress or intestinal survival¹⁶. Superoxide
228 dismutase and catalase show very limited distribution, as does the gene encoding glutamate
229 decarboxylase involved in proton scavenging and acid resistance (Supplementary Fig. 24),
230 the last of which is concentrated in *L. helveticus* and related species. Urease genes were
231 present in only 8 species including 3 of the 4 *Carnobacteria* species. Bile salt hydrolases
232 contribute to bile resistance in lactobacilli¹⁷; the pattern of their distribution in this genome data
233 resource, viewed in conjunction with other stress resistance genes, allows rational
234 identification of species likely to survive intestinal transit.

235 Protein acetyltransferases of prokaryotes like *Escherichia coli* confer resistance to heat
236 and oxidative stress¹⁸, and it was recently reported that homologs of the eukaryotic sirtuin
237 protein acetyltransferases contribute to stress resistance in *Lactobacillus paracasei*¹⁹. Forty of
238 the 213 genomes analyzed here lack homologs of any of SIRT1, SIR2L1, or Sir2a
239 (Supplementary Table 10). The remaining 173 genomes encode at least one homolog, with a
240 single species *L. ultunensis* harbouring 4 homologs, and 3 homologs being found in a large
241 number of food or intestinal lactobacilli. Since pre-treatment of *Lactobacillus paracasei* strains
242 with the sirtuin activator resveratrol (found in berries and red wine) alleviated growth inhibition
243 by cholate¹⁹, this presents the exciting prospect that some food ingredients might promote
244 shelf-life of lactobacilli in functional food products, or that certain prebiotics might rationally
245 promote survival and intestinal transit of lactobacilli administered in probiotics.

246 **Supplementary Note 10**

247 **Niche association and genome content**

248 A search for associations between niche and genome content for the 213 genomes
249 revealed moderate trends. The strongest trends were detected for species isolated from
250 animal sources (n=56), which, as noted in the main text, also had the lowest number of

predicted genes and the smallest average genome size (Supplementary Fig. 25). This is evident at a functional level where numerous functional gene groups displayed the lowest abundances in the animal niche. These groups include genes for the transport and metabolism of carbohydrates (Supplementary Fig. 25, top panel), amino acids, lipids (Supplementary Fig. 25, top panel), co-enzymes, nucleotides and inorganic acids. Normalization for genome size brought the difference above the significance threshold (Supplementary Fig. 25, bottom panels; p-values of 0.11 and 0.107 for carbohydrate transport and lipid metabolism, respectively). Genes involved in transcription, cell wall/membrane biogenesis and secondary metabolites also have the lowest average abundance in the animal niche. Gene decay is commonly associated with host-adapted microbes and this is a possible explanation for the trend observed in genomes that have been isolated from animal sources.

Across the broader range of niches/sources, the analyses failed to detect niche-specific genomic associations in our study, but rather a general pattern of gene decay in species from the animal niche. This study focuses largely on the comparison of type strains within the genus *Lactobacillus* and associated species so it is likely that the diversity among species in the dataset was too great to reveal niche-specific traits. Studies that focus on multiple strains of the same species are more suited to discovering niche-specific genes.

268

269 **Supplementary Note 11**

270 **Targeting competing microbes**

Bacteriocins are small, ribosomally synthesized antimicrobial peptides that can be exploited as antimicrobial-producing cultures in fermented foods, or the bacteriocins themselves added, e.g. pediocin and carnobacteriocin used as biopreservatives²⁰. Bacteriocins may also contribute to probiotic properties by limiting infection²¹ or signalling to the innate immune system²². Of the 213 genomes analysed here, 107 (50.2%; Supplementary Table 11) harbored at least one Area of Interest (AOI) relating to bacteriocin production by screening against the BAGEL database²³. Over half of these were larger proteins (>10 kDa) of the enterolysin/helveticin class, which are no longer considered classical bacteriocins²⁴. However, their widespread distribution suggests a central function for these currently cryptic antimicrobials. Manual inspection confirmed that 38 AOIs had the contiguous gene structure expected for typical bacteriocin operons. No dominant bacteriocin type was identified, although 58% of intact AOIs fell under the Unmodified Bacteriocin (Class II) category. Many of these included homologs of well-known bacteriocin operons including plantaricin, sakacin, salivaricin, subtilin, leucocin, carnobacteriocin and lacticin F. Surprisingly, AOIs were identified in many species not previously associated with bacteriocin production. For example, genes encoding pediocins were annotated in *L. kimchicus* and *L. taiwanensis* was found to harbour the machinery to produce subtilin. Predicted bacteriocin loci were found across all

288 clades of the phylogenetic tree with the exception of *Weissella* and the *L. brevis/L.parabrevis*
289 clade. Overall, the data do not support strict associations between species niche, the presence
290 of a bacteriocin or type of bacteriocin. Moreover, the overall prevalence of bacteriocins was
291 unexpectedly low compared to the literature describing this area, suggesting that many more
292 *Lactobacillus* species can produce bacteriocins than those represented by their type strains
293 examined in this study (for example *L. salivarius*²⁵).

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288

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