

Document S6. Oligonucleotide primers and plasmids utilized for the *Bartonella* and *Rickettsia* experiments.

Oligonucleotide primers used for the *Bartonella* experiments

Primers used for the generation of constructs utilized in the *Bartonella* bacterial two-hybrid assays are provided in **Table 1**. See manuscript (**Materials and Methods: Protein-Protein Interactions**) for further information regarding the design of each construct.

Plasmids used in the *Bartonella* experiments

All plasmids utilized in the *Bartonella* bacterial two-hybrid assays are provided in **Table 2**. See manuscript (**Materials and Methods: Protein-Protein Interactions**) for further information regarding the design of each construct.

Oligonucleotide primers utilized in the *Rickettsia typhi* RT-qPCR experiments

Primers used for the RT-qPCR assay during *R. typhi* infection of HeLa cells are provided in **Table 3**. Among the six reference genes utilized, *adr1* and *sca5* were selected for normalization of *rvhB8-I* and *rvhB8-II* gene expression.

Table 1 (Document S6)

Table 1 (Document S6). Oligonucleotide primers used for the *Bartonella* experiments.

Primer	Sequence (5'→3'), restriction sites underlined (restriction enzyme), mutagenic nucleotides underlined in bold
prAPV-36	TTTTTTGGATCCCATGAAAAACAGCAAGCTAAAC (BamHI)
prAPV-37	TTTTTTGGTACCTTAATTATTTAGCAGTATTTTC (KpnI)
prAPV-52	TTTGGATCCCATGAAAACAAAACAAGTAAAACC (BamHI)
prAPV-53	TTGGTACCTTAATCATTTAACAGTATTTCTGGGTC (KpnI)
prAPV-54	TTTGGATCCCATGAAGCACGATCTGATAAACTG (BamHI)
prAPV-55	TTGGTACCTCATTTTATCACCTCTGGATCAGATCT (KpnI)
prAPT0326	CATCTGTCCAACCTCCGCGACTC
prAPT0328	AGCGGACGTTCGAAGTTCTCGC
prAPV-38	CTTGGTTTCCAAGTGACAAGT
prAPV-39	TGC ATTAAGCAATCGATCAGATG
prAPV-40	GGCGGC GACAGATATTGGCTCAAC
prAPV-41	TTCGCCATAGCTAGTTTCATG

Table 2 (Document S6)**Table 2 (Document S6).** Plasmids used in the *Bartonella* experiments.

Plasmid	Relevant characteristics	Reference or source
pKT25	Bacterial two-hybrid plasmid, allows cloning to the 3' end of the T25 fragment of <i>Bordetella pertussis</i> adenylate cyclase.	Prof. Urs Jenal
pUT18c	Bacterial two-hybrid plasmid, allows cloning to the 3' end of the T18 fragment of <i>Bordetella pertussis</i> adenylate cyclase.	Prof. Urs Jenal
pKT25-<i>zip</i>	Bacterial two-hybrid plasmid, positive interaction control plasmid	Prof. Urs Jenal
pUT18c-<i>zip</i>	Bacterial two-hybrid plasmid, positive interaction control plasmid	Prof. Urs Jenal
pAPV001	Derivative of pKT25 encoding for T25-TrwG (<i>B. birtlesii</i>) phusion protein.	This work
pAPV002	Derivative of pUT18c encoding for T18-TrwG (<i>B. birtlesii</i>) phusion protein.	This work
pAPV003	Derivative of pKT25 encoding for T25-TrwG (<i>B. grahamii</i>) phusion protein.	This work
pAPV004	Derivative of pUT18c encoding for T18-TrwG (<i>B. grahamii</i>) phusion protein.	This work
pAPV005	Derivative of pKT25 encoding for T25-VirB8 (<i>B. grahamii</i>) phusion protein.	This work
pAPV006	Derivative of pUT18c encoding for T18-VirB8 (<i>B. grahamii</i>) phusion protein.	This work
pAPV007	Derivative of prAPV001 encoding for T25-TrwG-P214A (<i>B. birtlesii</i>) phusion protein.	This work
pAPV008	Derivative of prAPV001 encoding for T25-TrwG-V96G/V97A (<i>B. birtlesii</i>) phusion protein.	This work
pAPV009	Derivative of pUT18c encoding for T25-TrwG-P214A (<i>B. birtlesii</i>) phusion protein.	This work
pAPV010	Derivative of pUT18c encoding for T25-TrwG-V96G/V97A (<i>B. birtlesii</i>) phusion protein.	This work

Table 3 (Document S6)

Table 3 (Document S6). Oligonucleotide primers utilized in the *Rickettsia typhi* RT-qPCR experiments.

Gene	Forward Primer (5' to 3')	Reverse Primer (5' to 3')
<i>adr1</i>	CCACACCAGCACCAGCAAAA	GCACGGACAGTAGTTCAACC
<i>adr2</i>	CTTGCGCAGCAGTACCAAAA	CTGGTGTTCGGTCCTGCATTA
<i>gltA</i>	TACGAATTGCTGGCTCATCA	AAATGGATCATTTCCTTTAGCTTTAGC
<i>sca5</i>	TGGTATTACTGCTCAACAAGCT	CAGTAAAGTCTATTGATCCTACACC
<i>rpsL</i>	CTCCTGCCTTAGAATCCAACCC	TGAACGACCTTGCTTACGCC
<i>16S rRNA</i>	CGCAACCCTTATTCTTATTTG	CCTCTGTAAACACCATTGTAGCA
<i>rvhB8-I</i>	CACCTGTCATACGATATCAAAAGC	CAATTTTTGCTTCCCATAGCA
<i>rvhB8-II</i>	TCCAGTTGATTTTGCAACGA	TGGCTTCCTGATGTTTCATTT