

Probe name	5'-3' sequence
Probe b	CATGGCTTAATCTTTGAGAC
Probe c	GACTCTCCATCTCTTGCTTCTTG
Probe d	TGTTACCTCTGGGCC
Probe e	TTTCGCTGCGTTCTTCATC
Probe f	GGCCAGCAATTTCAAGTTA
Probe g	GAACATTGTTGCCTAGA
Probe h	CTCCGCTTATTGATATGC
Probe 5S	GGTCACCCACTACACTACTCGG

Supplementary Table S1. Oligonucleotides used as probes for RNA analysis.

	ChErb1 ₄₃₅₋₈₀₁ /ChYtm1	ChErb1 ₄₃₂₋₈₀₁ /ChYtm1	ChErb1[R486E] ₄₃₅₋₈₀₁ /ChYtm1
Data collection			
Space group	P 6 ₅ 2 2	P 2 ₁ 2 ₁ 2	P 6 ₅ 2 2
Cell dimensions			
<i>a</i> , <i>b</i> , <i>c</i> (Å)	169.11, 169.11, 154.04	86.443, 108.149, 108.618	170.75, 170.75, 155.66
α , β , γ (°)	90, 90, 120	90, 90, 90	90, 90, 120
Resolution (Å)	48.82 - 3.1 (3.31 - 3.1)*	48.53 - 2.1 (2.175 - 2.1)*	85.38-3.0 (3.18-3.0)*
<i>R</i> _{merge}	0.282 (2.838)	0.05838 (0.924)	0.176 (1.862)
<i>I</i> / σ <i>I</i>	17 (2.5)	18.69 (2.18)	13.5 (1.6)
Completeness (%)	100.00 (100.00)	99.90 (99.97)	100.00 (100.00)
Redundancy	39.1 (40.2)	6.5 (6.5)	12.2 (12.7)
Refinement			
Resolution (Å)	48.82 - 3.1 (3.31 - 3.1)	48.53 - 2.1 (2.175 - 2.1)	85.38-3.0 (3.18-3.0)
No. reflections	24180 (4313)	60041 (5923)	27367 (4333)
<i>R</i> _{work} / <i>R</i> _{free}	0.1936 (0.2940)/ 0.2390 (0.3557)	0.1639 (0.2386)/ 0.2132 (0.2703)	0.2016 (0.3153)/ 0.2621 (0.3914)
No. atoms	6170	6631	6182
Protein	6168	6279	6177
Ligand/ion	2 ^a	26 ^b	5 ^c
Water	0	326	0
<i>B</i> -factors	102.40	63.20	89.70
Protein	102.40	63.40	89.70
Ligand/ion	96.10	76.70	84.70
Water		57.80	
R.m.s. deviations			
Bond lengths (Å)	0.006	0.008	0.006
Bond angles (°)	1.26	1.18	1.35

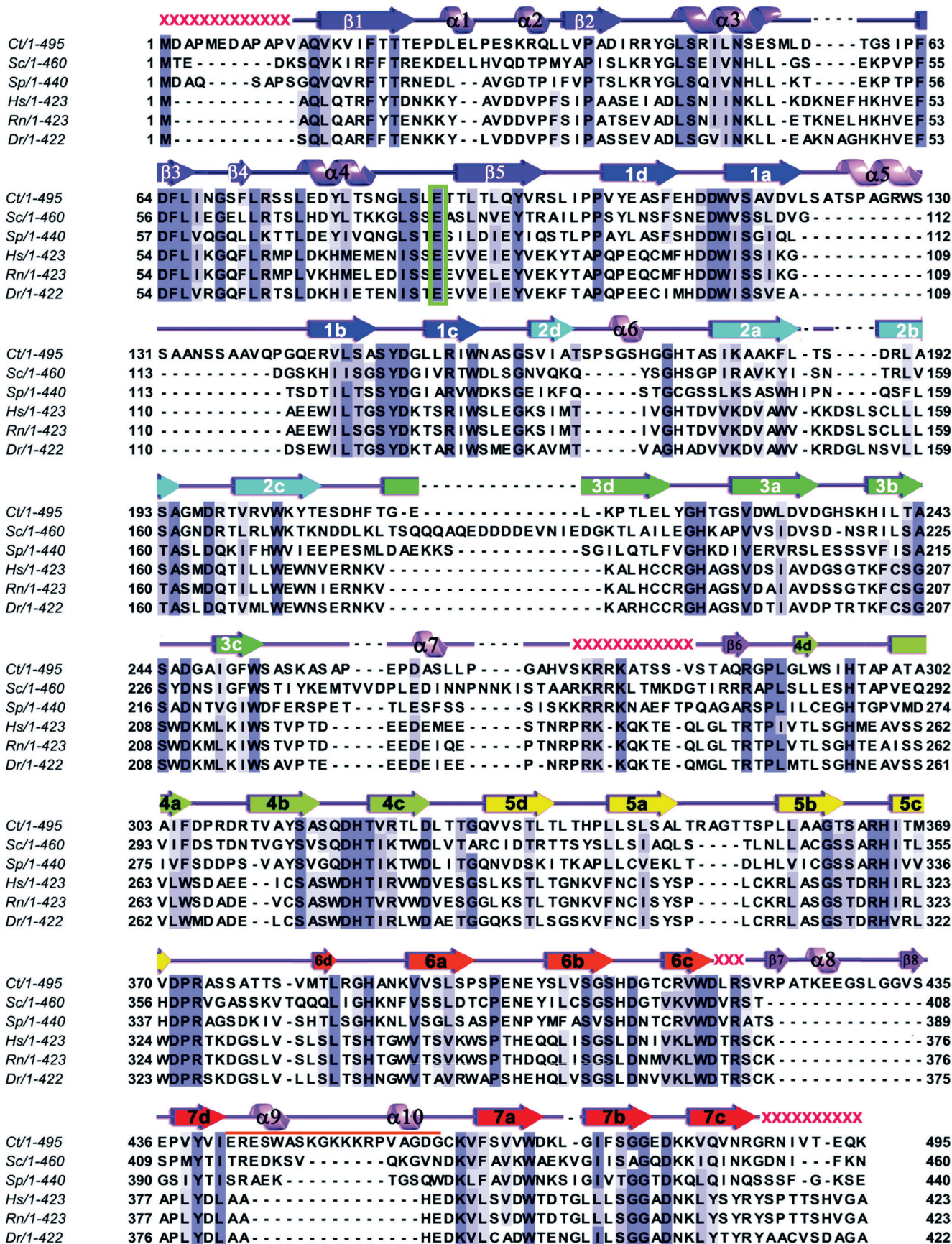
Supplementary table S2. Data collection and refinement statistics

*Values in parentheses are for highest-resolution shell.

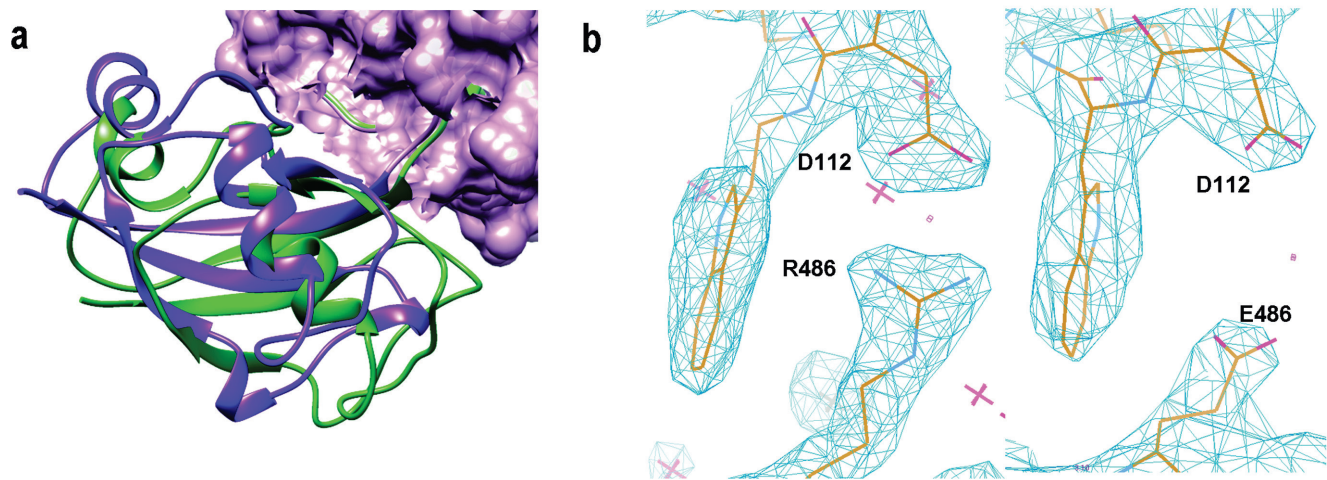
^a) Ligands: chloride ion

^b) Ligands: chloride ion, sodium ion, glycerol, ethylene glycol

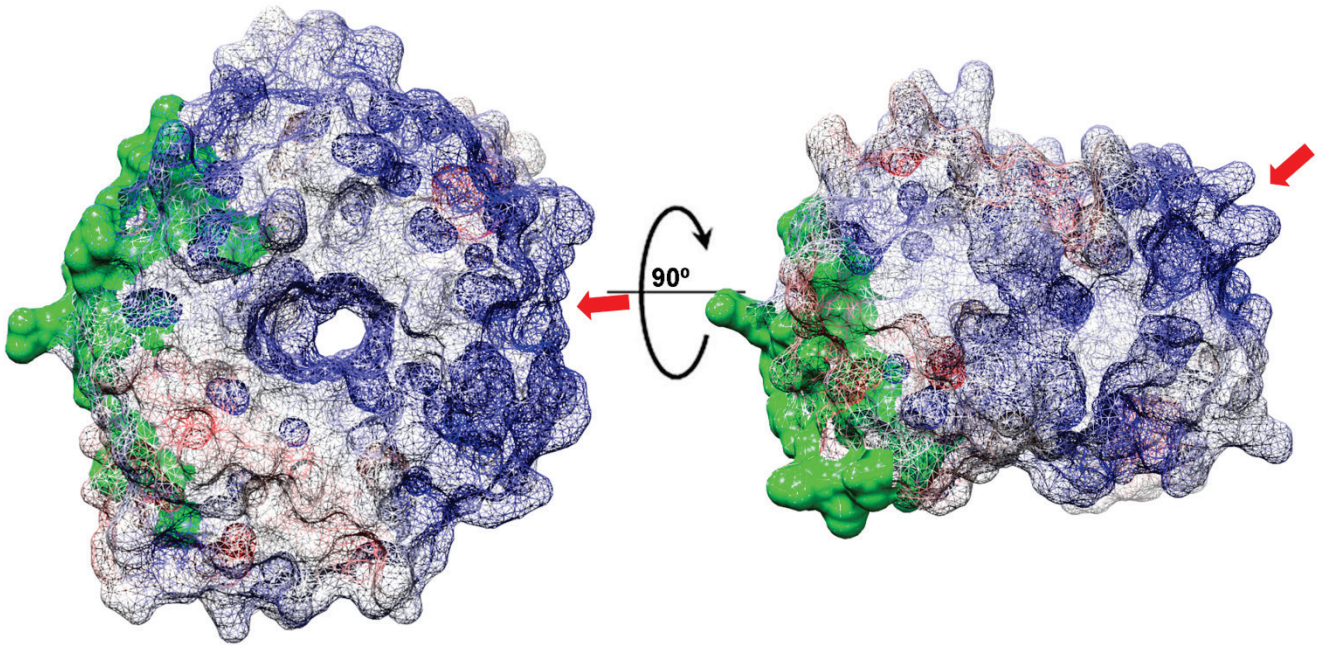
^c) Ligands: chloride ion. Data were collected from one crystal for each dataset.



Supplementary Figure S1. Multiple sequence alignment of Ytm1 orthologues. Sequences corresponding to Ytm1 (or WDR12) proteins from *Chaetomium thermophilum* (Ct), *Saccharomyces cerevisiae* (Sc), *Schizosaccharomyces pombe* (Sp), *Homo sapiens* (Hs), *Rattus norvegicus* (Rn) and *Danio rerio* (Dr) were aligned using JalView. Secondary structure elements as seen in the final model of ChYtm1 are represented above the alignment. β -strands are marked with arrows, α -helices are depicted with coils. Secondary structure elements corresponding to UBL are shown in purple. Coloring and numbering of the following β -strands correspond to the WD repeats. Conserved residues are marked with shadows. The conserved glutamic acid proposed to bind the MIDAS domain of Rea1 is marked with a green box. The sequence corresponding to the knob-like structure is shown with orange line



Supplementary Figure S2. (a) UBL of ChYtm1 presents certain degree of flexibility because it appears in two different orientations depending on the symmetry of the crystals. UBL in purple: P 21 21 2 and in green: P 65 2 2 space groups. (b) Upon successful crystallization of ChYtm1/ChErb1[R486E]₄₃₂₋₈₀₁ we proved that the β -propeller structure was intact and the only difference between both dimers was the lack of D112-R486 salt bridge (left) that is not formed in the mutant (right).



Supplementary Figure S3. Electrostatic surface potential of the β -propeller of Erb1 represented as mesh shows that highly electropositive area possibly involved in binding to RNA (red arrow) is located on the opposite side to Ytm1 binding site (in green)