

Fig. S1. Luciferase activity at different Cu concentrations. Total protein was extracted from 7 day-old *PLHY:LUC* seedlings grown under Cu deficiency ($\frac{1}{2}$ MS; -Cu). Kinetic measurements were performed after adding $10 \mu\text{M}$ CuSO_4 (Cu excess; +Cu) and a constant amount of D-luciferin (0.3 mg/mL). The activity was recorded every 10 min. Extract without substrate was used as a negative control (Ctrl). a.u.= arbitrary units.

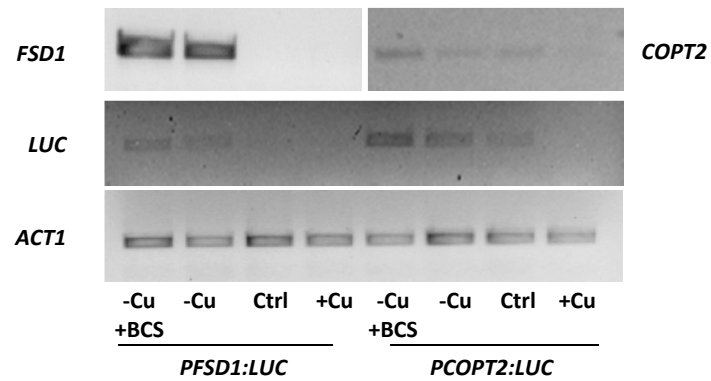


Fig. S2. *FSD1*, *COPT2* and *LUC* gene expression at different Cu contents. Six day-old *PFSD1:LUC* and *PCOPT2:LUC* seedlings were grown in severe Cu deficiency (½ MS with 50 µM BCS; -Cu + BCS), Cu deficiency (½ MS; -Cu), sufficiency (½ MS with 1 µM µM CuSO₄; Ctrl) or in Cu excess (½ MS with 10 µM µM CuSO₄; +Cu) in LDHC cycles. Samples were collected at 4 h after the onset of the light cycle and *LUC* and *FSD1* gene expression were analyzed by sqPCR. *ACT1* expression was used as loading control.

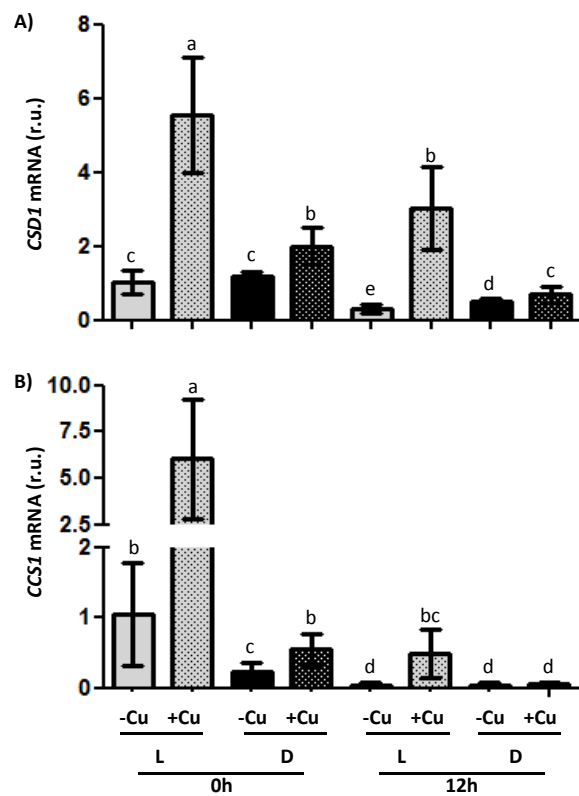


Fig. S3. Regulation of *CSD1* and *CCS1* gene expression by Cu and light conditions. The relative expression was determined in the same conditions shown in Fig. 4. Means with a different letter are significantly different ($P < 0.05$). r.u. = relative units.

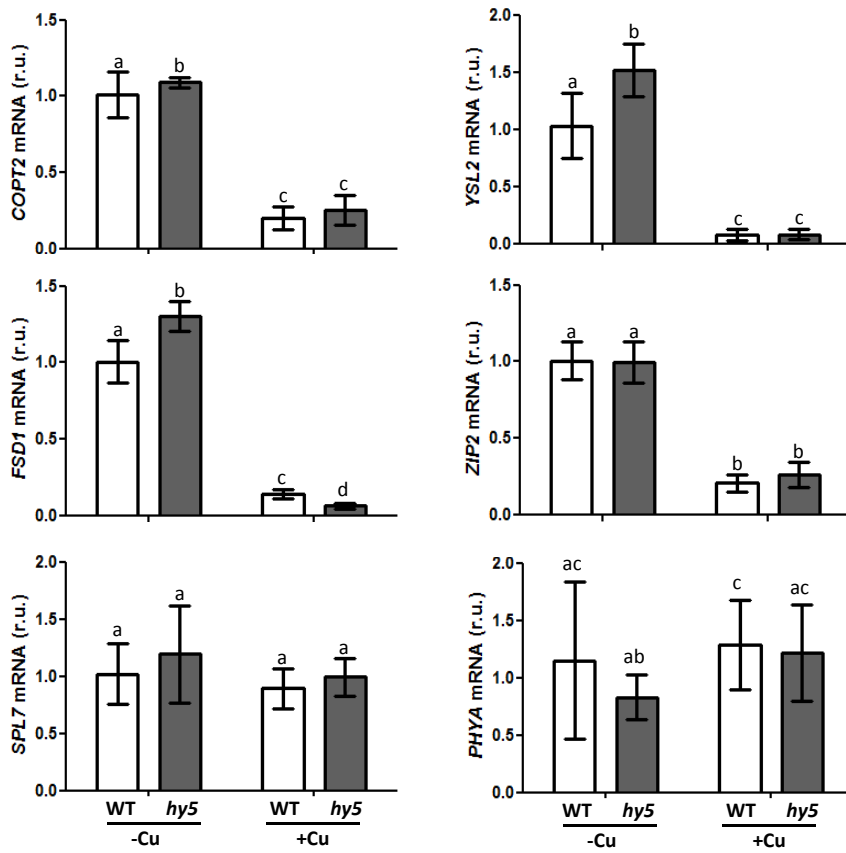


Fig. S4. Regulation of gene expression by Cu in the *hy5* mutant. The relative expression of the *COPT2*, *FSD1*, *SPL7*, *YSL2*, *ZIP2* and *PHYA* genes was determined by qPCR in 7 day-old WT seedlings grown under Cu deficiency (½ MS; -Cu, white bars) or Cu excess (½ MS with 10 μM CuSO₄; +Cu, grey bars). The *UBQ10* gene was used as reference. mRNA levels are expressed as relative units (r.u.). The bars represent the mean ± SD of 3 replicates. Means with a different letter are significantly different ($P < 0.05$).

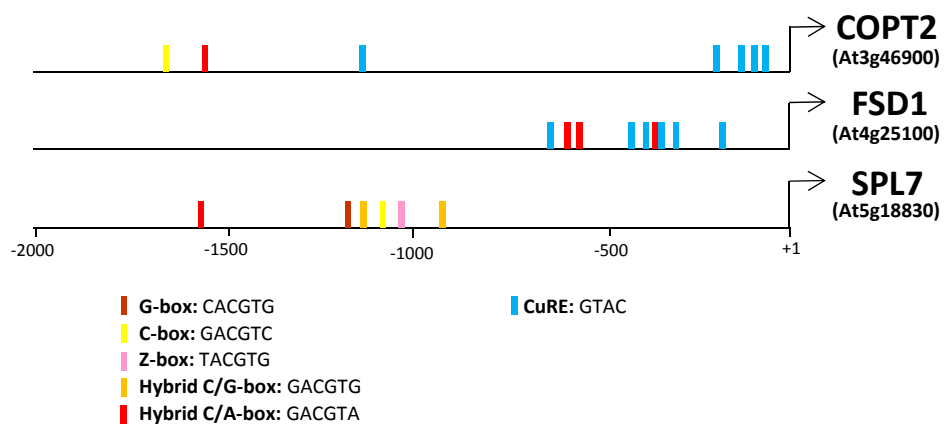


Fig. S5. Sequence analysis of the *COPT2*, *FSD1* and *SPL7* promoters for putative HY5 and SPL7 recognized sites. Positions of the *cis* regulatory elements are marked in different colors as indicated in figure legend. Putative CuRE elements bound by SPL7 (Yamasaki *et al.*, 2009) and different HY5 bound elements (Song *et al.*, 2008) are shown.

Table S1. Analysis of the peak time values under different circadian and diurnal oscillatory conditions for the genes regulated under Cu deficiency conditions. The locus MIPS code (<http://www.arabidopsis.org>) and the phases under different oscillatory conditions (<http://diurnal.mocklerlab.org>) are shown. D: dark, L: light, H: hot (22°C), C: cold (12°C). Letters between brackets mean entrainment conditions before constant conditions.

		Circadian					Diurnal						
Gene	Locus	DD(DDHC)	LL23(LDHH)	LL12(LDHH)	LL(LDHC)	LL(LLHC)	LDHH_SM	LDHH_ST	longday	shortday	LDHC	LLHC	Peak
<i>COPT2</i>	AT3G46900		20	22				21	0	22			21.8
<i>FSD1</i>	AT4G25100		6					1					3.5
<i>SPL7</i>	AT5G18830						19			22	23	22	21.5
<i>ZIP2</i>	AT5G59520									19			19
<i>CSD1</i>	AT1G08830	20		6		23							0.3
<i>CCS</i>	AT1G12520	23					22	21	23	18			21.4
<i>LHY</i>	AT1G01060	0	22	1	1	2	0	23	1	23	23	2	0.2
<i>CAS</i>	AT5G23060		21	23		3	2	23	2	1	2	2	0.7
<i>YSL2</i>	AT5G24380		19	21						4		21	22.3
<i>SMC6B</i>	AT5G61460								1	10		18	18
<i>IRT1</i>	AT4G19690			1	4				23				1.0
<i>ACA1</i>	AT3G52720		19		22	23	1	0	21	21	1		22.5
<i>TIR-NBS-like</i>	AT3G04210							16	21		7		22.7
<i>CYP83A1</i>	AT4G13770			7		22	8	8	9	6	8	23	5.4
<i>GSTF11</i>	AT3G03190						8	10		6		0	6
<i>BCAT4</i>	AT3G19710						7	8	7	6		3	6.2
<i>UGT78D2</i>	AT5G17050	21	19	22	23	23		19		5	22	21	22.1
<i>EXPA8</i>	AT2G40610		4	6	6			0		23	5	8	4.0
<i>Aux resp</i>	AT1G29440		1	5	3	3	3	1	3	23	3	6	2.6
<i>COPT1</i>	AT5G59030						21	21	22	22			21.5
<i>CCH</i>	AT3G56240							0		20			22
<i>CSD2</i>	AT2G28190		15	7		23						0	17.3

Table S2. Primers used for cloning the *FSD1* and *COPT2* promoters. The restriction site is indicated *in italics* in the sequence.

Name	Sequence (5'-)
<i>COPT2-1200F-HindIII</i>	GTGGTGTTAAAGCTTTTGCAACAA
<i>COPT2-ATG-R-HindIII</i>	CATAAGCTTGATACTAATGTTAATAGGGTTATGAT
<i>FSD1 HindIII F</i>	TTTAAGCTTGGAGTGAAGCTATATATA
<i>FSD1 HindIII R</i>	TTTAAGCTTAGCAGCCATTCTTTGTAATTGA

Table S3. Oligonucleotides used in sqPCR.

Gene Name	Sequence (5'-)
<i>ACTINARA1A</i>	GGCGATGAAGCTCAATCCAAC
<i>ACTINARA1B</i>	GGTCACGACCAGCAAGATCAAGACG
<i>FSD1 rt F1</i>	GCTTCAAGTGCTGTCACC
<i>FSD1 rt R1</i>	CAAGCCAGGCCAGCCAG
<i>LUC-F</i>	GGAAGACGCCAAAAACAT
<i>LUC-R</i>	GTTTTGTCACGATCAAAGGA
<i>COPT2-F</i>	ACGTGTCAGTGGCTCAACC
<i>COPT2-R</i>	GACGGCGGAAGAAGCTCGGCGG

Table S4. Oligonucleotides used in qPCR.

Gene Name	Sequence (5'-)	
	forward	reverse
<i>COPT2</i>	CCTTTCGTATTTGGTGATGCT	AAACACCTGCGTTAAAGGAC
<i>CCS</i>	TCTCCACGTCTCTGGGACTTT	AGCTGAGGCATGGCTCGAT
<i>CSD1</i>	CATCATTGGTCTCCAGGGCT	GACCTCCTTATTACATCAAT
<i>FSD1</i>	ACCGAAGACCAGATTACATA	TGGCACTTACAGTTCCCAA
<i>SPL7</i>	CAGGCAGACTGTTACCAGA	AGTTTGACGGGACCTGAATG
<i>UBQ10</i>	TAATCCCTGATGAATAAGTGTCTAC	AAAACGAAGCGATGATAAAGAAG
<i>ZIP2</i>	CGCTTGAGAAAACCTATGGA	CGACACCTATGGGACTCGAT
<i>HY5</i>	GTTTGGAGGAGAAGCTGTCTG	TCTTGCTTGCTGAGCTGAAA
<i>PHYA</i>	GTTACAAAACCTGGGCTGGA	ATGTTGGCCATGACTGCAA
<i>YSL2</i>	TCTTATAAATGGATTCATACTA	AATGCCCAAAAGAACTCAAA
<i>LHY</i>	TCGGCCTCTTCTCACAGTT	ACACCCGAGCAATTCTCATC
<i>CCA1</i>	GTTGCAGCTGCTAGTGCTTG	GAAGATCGAGCCTTTGATGC
<i>TOC1</i>	TGATCTCCAATGGCTAAGG	ACTTCGTCTTGCTCGACAT
<i>GI</i>	GCATCTAGTTGCTGGCCTTC	AGCTCGAAGGAGTCCACAA