

## Supplementary Information

# Iron oxide nanoparticle-mediated hyperthermia stimulates dispersal in bacterial biofilms and enhances antibiotic efficacy

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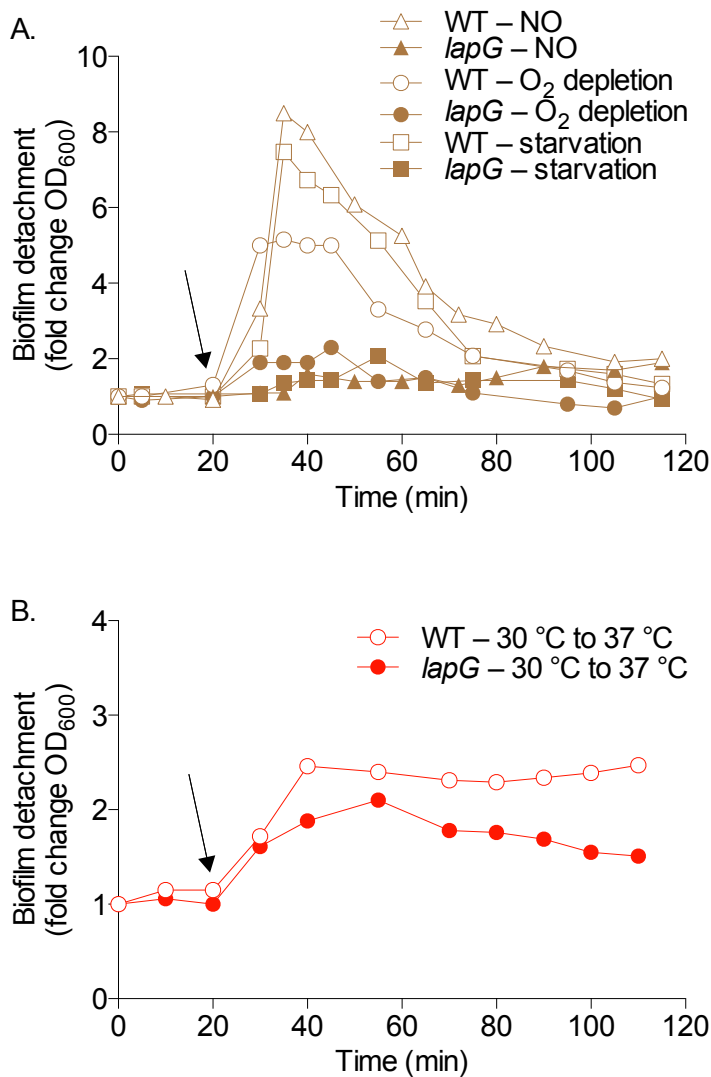
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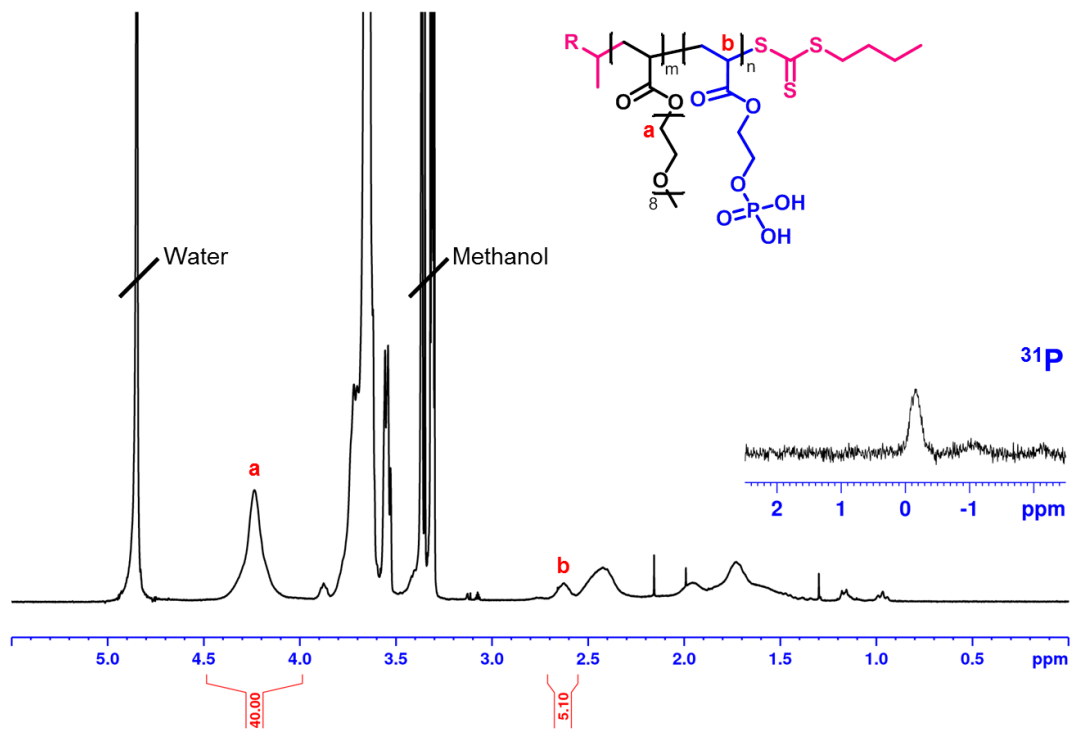
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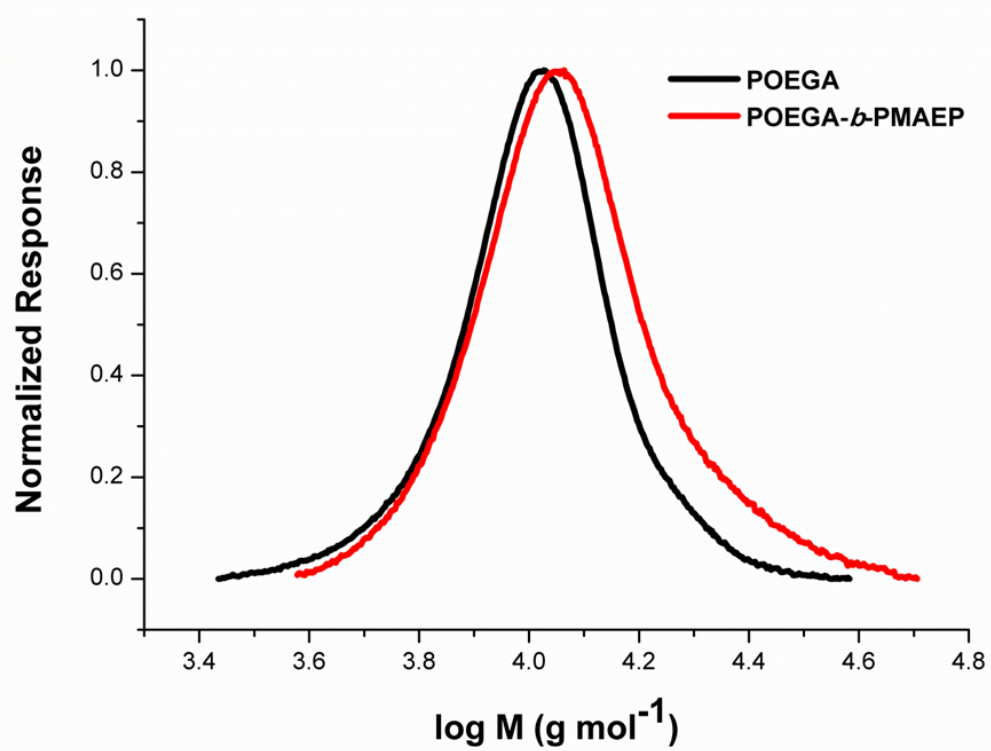
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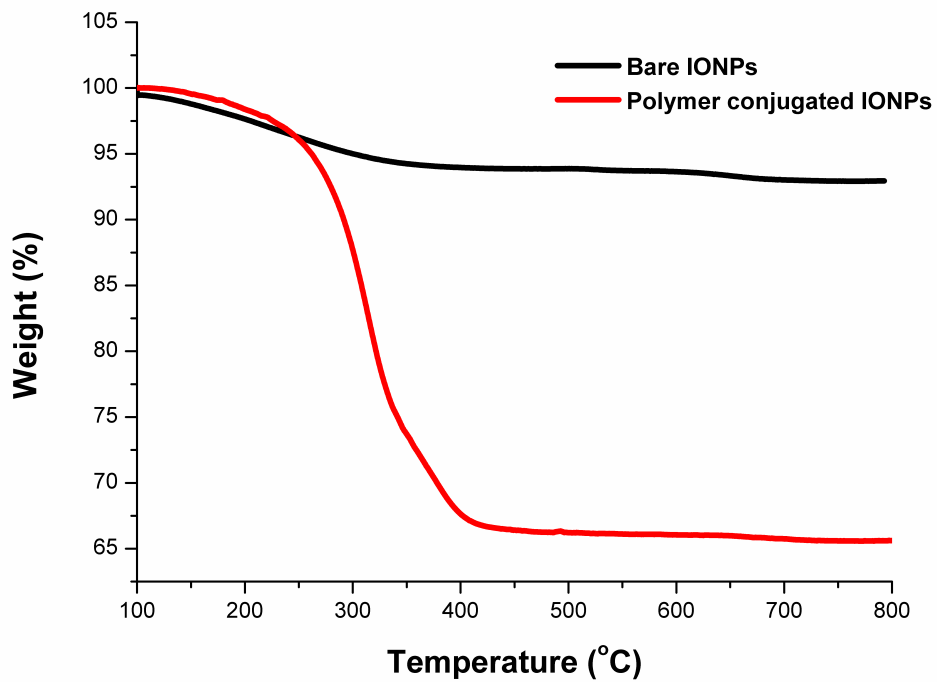
**Fig. S1.** A *P. aeruginosa* *lapG* mutant strain, which does not disperse in response to a range of signals and cues, is not strongly affected in temperature-mediated dispersal. (A) *P. aeruginosa* wild type (WT) and *lapG* mutant biofilms were grown in continuous flow microfermentor cultures at 37 °C for 24 h. At this time, biofilm dispersal was induced by: (i) adding the nitric oxide (NO) donor sodium nitroprusside (1 mM) to the biofilm medium; (ii) switching the microfermentor aeration from air to 100% nitrogen gas to induce oxygen (O<sub>2</sub>) depletion; or (iii) switching the biofilm medium to M9 medium without carbon source to induce starvation. The biofilm effluent was collected at regular intervals and quantified at OD<sub>600</sub>. Solid arrows indicate induction of dispersal. (B) *P. aeruginosa* wild type (WT) and *lapG* mutant biofilms were grown in continuous flow microfermentor cultures for 24 h at 30 °C before suddenly increasing the temperature to 37 °C. Solid arrows indicate temperature upshift. The data shown are representatives from at least 3 independent replicate experiments.



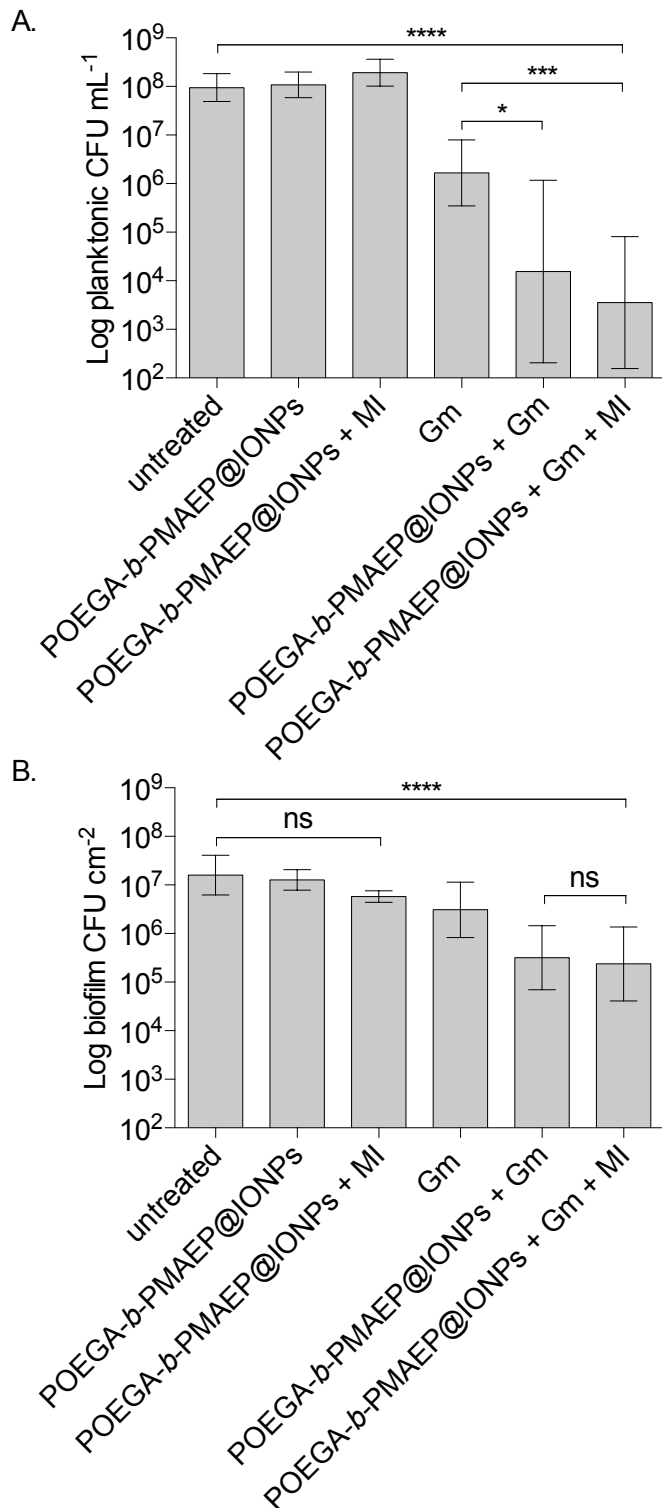
**Fig. S2.**  $^1\text{H}$  and  $^{31}\text{P}$  spectra of purified POEGA-*b*-PMAEP (recorded in  $\text{CD}_3\text{OD}$ ).



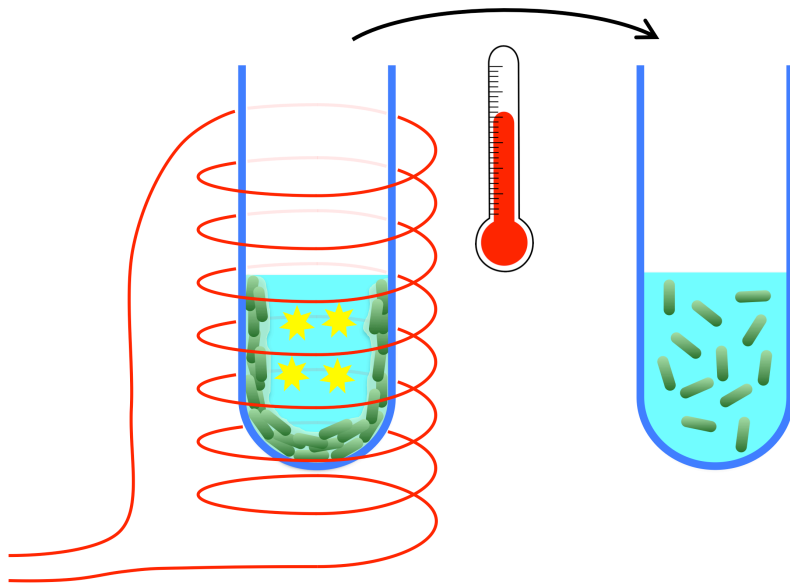
**Fig. S3.** SEC chromatograms of POEGA and POEGA-*b*-PMAEP



**Fig. S4.** TGA of bare IONPs and POEGA-*b*-PMAEP@IONPs synthesized using “grafting onto” approach.



**Fig. S5.** IONP-induced hyperthermia increases the efficacy of the antibiotic gentamicin against both biofilm and planktonic *P. aeruginosa*. Biofilms were grown as described before for 24 h in test tubes before adding POEGA-*b*-PMAEP@IONPs (1 mg mL<sup>-1</sup>), gentamicin (Gm, 2 mg mL<sup>-1</sup>), a combination of both or no treatment to the cultures, and exposing (+ MI) or not the cultures to an alternating magnetic field (6.5 T, 196 kHz) for a further 2 h. After treatment, colony-forming units (CFU) analyses were performed for both the planktonic (A) and biofilm (B) phases. MI, magnetic induction. Error bars represent standard errors ( $n \geq 4$ ). Asterisks indicate statistically significant difference of treatment versus untreated culture (\*,  $P < 0.1$ ; \*\*\*,  $P < 0.001$ ; \*\*\*\*,  $P < 0.0001$ ; ns, not significant).



Schematic representation of IONP-mediated hyperthermia inducing biofilm dispersal.