

Figure S1, related to Figure 1. IL-6 production and bacterial burden after infection of CD1 mice with WT or $\Delta hy IB$ GBS.

(A) Overnight bacterial culture supernatants from WT and $\Delta hy/B$ GBS were added to a hyaluronan agar plate. The plate was visualized after 24 h for a zone of clearance as an indication of the presence of hyaluronidase activity. (B-C) CD1 mice (n = 12 per group) were infected i.p. with WT, $\Delta hy/B$, or the complemented $\Delta hy/B$ GBS strain. (B) IL-6 in the spleen and (C) CFU in the blood and spleen were determined after 24 h. (D) CD1 mice were infected i.p. with either WT or $\Delta hy/B$ and CFU in the blood determined

after 48 hours. Data analysis was performed using ANOVA for (B) and (C) and unpaired two-tailed *t*-test for (D). *P < 0.05 and **P < 0.01.



Figure S2, related to Figure 1. Immunopathologic findings in C57BL/6 mice 48 h after infection with WT or $\Delta hyIB$ GBS.

Mice were injected i.p. with either WT or $\Delta hylB$ GBS and spleens were harvested after 48 h. Homogenized spleens were assayed for (A) KC (n=5 per group) or (B) fibrinogen by ELISA (*P* =0.06; n=10 per group). Each data point represents an individual mouse. Data analysis was performed using Mann-Whitney U for (A) and unpaired two-tailed *t*-test for (B). **P* < 0.05.



Figure S3, related to Figure 1. GBS hyaluronidase reduces inflammation in GAS and GBS murine models.

(A-B) C57BL/6 mice were injected i.p. with either WT GAS (no hyaluronidase) or GAS expressing hyaluronidase from GBS. (A) CFU and (B) TNF- α from the spleen were

determined after 48 h. (C-D) Infection of neonatal mice. Four day old C57BL/6 pups were injected i.p. with WT or $\Delta hy/B$ GBS. After 48 h, kidneys were homogenized and (C) bacterial loads and (D) TNF- α levels were enumerated. (E) Mice were administered WT or $\Delta hy/B$ GBS into the vaginal lumen as described in Figure 1H. The vaginal cavities were swabbed every other day and percentage of mice remaining colonized analyzed. Each data point represents an individual mouse. Data analysis was performed by Mann-Whitney U for (A) and (B), by unpaired two-tailed *t*-test for (C) and (D) and Fisher exact test for (E). **P* < 0.05.



Figure S4, related to Figure 2. Bacterial burden, cytokine production, and host survival in WT C57BL/6 and TLR2^{-/-} after infection with WT or $\Delta hyIB$ GBS.

(A) WT C57BL/6 and *TLR2^{-/-}* mice were injected i.p. with either WT or $\Delta hy/B$ GBS. IL-6 from the spleen was determined at 24 h. Analysis was performed using Mann-Whitney U. *P*-value: ***P* < 0.01.



Figure S5, related to Figure 3. Toxicity of HA disaccharides and effect of HA disaccharides on LPS-, PAM₃CSK₄-, and CpG- induced TNF- α release.

(A) Toxicity of HA disaccharides. BMDM were incubated for 6 h with HA disaccharides (10 µg/ml), GBS hyaluronidase (0.5mg/ml), or Triton X-100. An LDH assay was performed to quantify cell death. (B-C) Murine BMDM (WT or *MyD88^{-/-}*) were stimulated with LPS, Pam₃CSK₄ or GBS in the presence or absence of HA disaccharides. Supernatants were collected after 4 h for TNF- α or IL-6 determination. (B) Effect of varying LPS dose with HA disaccharides maintained at 10 µg/ml. (C) Effect of HA disaccharides (10 µg/ml) on stimulation of WT and *MyD88-/-* macrophages by various agonists. Data are shown as mean ± SD, where **P* < 0.05. ****P*<0.001, and results from (A) to (C) each are representative of three experiments. Data analysis was performed using ANOVA for (A) and (C), and unpaired two-tailed *t*-test for (B).

Strain, plasmid, or primer	Genotype or description	Reference or source
Bacterial Strains		
E. coli DH5α	φ80 Δ(lacZ)M15 Δ(argF-lac)U169 endA1 recA1hsdR17 (r−K m−K) deoRthi-1 supE44 gyrA96 relA1	Lab stocks
E. coli MC1061	F [−] araD139 Δ(ara-leu)7696 galE15 galK16 Δ(lac)X74 rpsL (Str ^R) hsdR2 (rK [−] mK ⁺) mcrA mcrB1	Lab stocks
GAS M49	Wild-type clinical isolate	Lab stocks
GBS A909	Wild-type clinical isolate	Lab stocks
S. aureus Newman	Wild-type lab strain	Lab stocks
$\Delta ny B$	GBS A909 <i>nyIB</i> mutant	This study
$\Delta hy B + pny B$	GBS A909 <i>nyIB</i> mutant complement with <i>hyiB</i>	This study
ΔnyiB + S. aureus hyl	GBS A909 <i>nyiB</i> mutant complement with <i>nysA</i> nyi gene from <i>S. aureus</i> Newman	i nis study
$\Delta hy B + S.$	GBS A909 <i>hylB</i> mutant complement with hyl gene from <i>S.</i> coelicolor	This study
GAS M49 + phylB	GAS M49 complement with <i>phyIB</i>	This study
Plasmids		
pDCerm	Shuttle vector encoding erythromycin resistance cassette	Jeng et al. 2003
pKODestErm	Created for Gateway cloning	Locke et al.
pUC19	Amp shuttle vector	Invitrogen
Primers	5'-3' orientation	
<u>GBS ΔhylB</u>		
hylB upst fwd	ATC CCGCGG CGAGCTGTTCTCTAACCCTA	This study
<i>hylB</i> dwnst rev	ATC GGATCC ACATACCTTATTGGTGCCGACGGT	This study
hylB upstr/cat rev	ATATCCAGTGATTTTTTTTCTCCAT	This study
hylB dwnstr/cat fwd	GIGIGCIACITIGAIAGIAAAIIGCC GCGATGAGTGGCAGGGCGGGGGCGTAAAACTTGGGCTGTTA	This study
	TCAAACACGA	
cat fwd	ATGGAGAAAAAAATCACTGGATATACCACC	This study
cat rev	ATGGAGAAAAAAATCACTGGATATACCACC	This study
Complement		
hylB compl fwd	AGGCAATTTACTATCAAAGTAGCAC	This study
hylB compl rev	TGCTGTAGAAGCCAAGTATG	This study
$\Delta hy B + S. aureus$		
<u>nyai (nysA)</u> hua A fud		
nysA two	GTTIGGTCGTCTAAAGTAATGTIGC	i nis study
<i>hysA</i> rev	CAAAGGGGATGTGGTCTGCG	This study

Table S1, related Figures 1 and 5. Strains, plasmids, and primers used in this study