

Molecular spectrum of *BRAF*, *NRAS* and *KRAS* gene mutations in plasma cell dyscrasias: implication for MEK-ERK pathway activation

Supplementary Material

Supplementary Materials and Methods.....	2
Supplementary Tables.....	7
Supplementary Figures.....	36

Supplementary Materials and Methods

Mutation analyses

Genomic DNA was amplified using FastStart High Fidelity Polymerase (Roche) and fusion primers (Roche) containing M13 adapter sequences and the sequence-specific primers spanning *BRAF* exons 11 and 15 (RefSeq NM_004333.4) (exon 11 forward 5'-TCCCTCTCAGGCATAAGGTAA-3'; exon 11 reverse 5'-CGAACAGTGAATATTTCTTTGAT-3'; exon 15 forward 5'-TCATAATGCTTGCTCTGATAGGA-3'; exon 15 reverse 5'-GGCCAAAATTTAATCAGTGGA-3') [1], *NRAS* exons 2 and 3 (exon 2 forward 5'-ATGGAAGGTCACACTAGGGTT-3'; exon 2 reverse 5'-TCCTTTAATACAGAATATGGG-3'; exon 3 forward 5'-ACCCCAGGATTCTTACAGA-3'; exon 3 reverse 5'-AGAGTTAATATCCGCAAATG-3') [2], and *KRAS* exons 2, 3 and 4 (exon 2 forward 5'-TTAACCTTATGTGTGACATGTTCTAA-3'; exon 2 reverse 5'-GAAAGTAAAGTTCCCATATTAATGGT-3'; exon 3 forward 5'-GCCTGTAATAATCCAGACT-3'; exon 3 reverse 5'-CATGGCATTAGCAAAGACTC-3' [3]; exon 4 forward 5'-AGTTGTGGACAGGTTTTGAAAGA-3'; exon 4 reverse: 5'-ATTTCAAGTGTACTTACCTGTCTTG-3' (Primer-BLAST at http://www.ncbi.nlm.nih.gov/tools/primer-blast/index.cgi?LINK_LOC=BlastHome)). Amplicon library A and B sequencing adapters and multiplex identifier (MID) tags were then added to both tails of amplicons by a second amplification step. PCR conditions were as follows: in the first amplification step, denaturing step at 94°C for 5 min followed by 25 cycles at 94°C (30 sec. per cycle), annealing step at 57°C (30 sec. per cycle), and extension at 72°C (45 sec. per cycle), followed by a final 7 min. extension at 72°C; in the second amplification step, denaturing step at 94°C for 5 min followed by 25 cycles at 94°C (20 sec. per cycle), annealing step at 55°C (20 sec. per cycle), and extension at 72°C (45 sec. per cycle), followed by a final 10 min. extension at 72°C. All PCR samples were purified using an AMPure XP (Agencourt) bead clean-up; samples were then quantitated using picogreen dye (Life Technologies) and the Victor X2 (Perkin Elmer) fluorometer. Samples were pooled together at equimolar ratios to prepare for Roche/454 pyrosequencing. The obtained amplicon library was added to the emulsion PCR at a ratio of 0.8 molecules per bead and subjected to deep sequencing on the Genome Sequencer Junior instrument (Roche-454 Life Sciences). The obtained sequencing reads were mapped to the reference sequence (RefSeq NG_007873.2 for *BRAF*, RefSeq NG_007572.1 for *NRAS*, RefSeq NG_007524.1 for *KRAS*) and analyzed by the Amplicon Variant Analyzer software (Roche) to establish the mutant allele frequency.

To verify the occurrence of *BRAF* variants at transcriptional level, total RNA of mutated samples was converted to cDNA using M-MLV reverse transcriptase (Invitrogen) and random hexamers, and subjected to deep sequencing of the exon harboring the variant/s detected on genomic DNA. Sequence-specific exonic PCR primers were designed in the Primer 3 program (<http://frodo.wi.mit.edu/primer3/>) and are as follows: exon 11 forward 5'-CTCAGCGAGAAAGGAAGTCA-3'; exon 11 reverse 5'-GGAATAGCCCATGAAGAGTAGGA-3'; exon 15 forward 5'-AGATATTGCACGACAGACTGC-3'; exon 15 reverse 5'-TTCTGATGACTTCTGGTGCC-3'.

Gene expression profiling and data analysis

For gene expression analysis, samples were profiled on the GeneChip Human Gene 1.0 ST array (Affymetrix, Santa Clara, CA, USA) as previously described [4]. The raw intensity expression values were processed by Robust Multi-array Average procedure (RMA) [5], with the re-annotated Chip Definition Files from BrainArray libraries version 18.0.0 [6], available at <http://brainarray.mbni.med.umich.edu>. Sno/scaRNA expression profiles were excluded in the following supervised analyses by Significant Analysis of Microarrays (SAM). In the patients' dataset, supervised analyses on gene expression profiles were carried out using the SAM software version 4.00 (excel front-end publicly available at <http://www-stat.stanford.edu/Btib/SAM/index.html>) [7]. The cutoff point for statistical significance (at a q -value = 0) was determined by tuning the Δ parameter on the false discovery rate (FDR) and controlling the q -value of the selected gene lists. A higher stringency level (90th percentile FDR = 0) was also applied to the differentially expressed gene lists at q -value 0. The list of differentially expressed genes was submitted to ToppGene Suite portal (<http://toppgene.cchmc.org>) for functional enrichment analysis by using ToppFun application [8]. A P -value cut-off of 0.05 and FDR Benjamini and Hochberg correction method were applied to all the annotation terms in the default parameter set, and those with a q -value < 0.05 were defined as significantly enriched. For gene expression analysis of U266 cells treated with BRAF-inhibitor or untreated (two independent replicas for each condition), microarray data were globally analyzed by Gene Set Enrichment Analysis (GSEA) [9] by means of gene sets of the MSigDB database (<http://www.broadinstitute.org/gsea/msigdb/index.jsp>) consisting of 5-1 000 genes (C1 Positional and C4 Computational gene sets were not considered). Gene set permutations (1 000) were set and default parameters were applied for statistical analyses. Gene sets showing a false discovery rate lower than 5% were selected as significantly enriched in U266 treated and control phenotypes, respectively.

The principal component analysis of the samples was performed by singular value decomposition of the considered data expression matrix using *prcomp* function in *stats* package and visualized with *plot3d* function in *rgl* package for R software.

Cell lines

Details regarding the source of all cell lines have been previously reported by us [10, 11], except for delta-47, UTM-2, MM.1S and MM1-144, that were kindly provided by Dr. G.Tonon - San Raffaele Scientific Institute, Milan. All cell lines have been recently authenticated by STR profiling and/or gene mutational analyses, and tested for mycoplasma contamination.

Cell-based assays

U266 cell line was maintained in RPMI 1640 Medium supplemented with 10% fetal calf serum. Vemurafenib (Sigma-Aldrich) was dissolved in dimethyl sulfoxide (DMSO) stock solutions and stored at -20°C. The drug was added to log phase cells and used at 30 µM, based on previous experiments testing cell proliferation and survival in a dose-dependent manner (data not shown) that confirmed the very weak sensitivity of U266 to the drug [12].

For proliferation assays, U266 cells were plated in 6 or 96-well plates, at 0.5×10^6 or 0.1×10^6 cells/well respectively and were treated with Vemurafenib or vehicle (DMSO) as control for 6, 12, 24, 48, 72 hrs. Cell number and viability were assessed by Trypan Blue exclusion and cell proliferation was evaluated by Thiazolyl Blue Tetrazolium Bromide (MTT) assay (Sigma Aldrich, USA), following the manufacturer's instructions.

For cell cycle analysis, cells were then collected after 6, 12, 24, 48, 72 h, fixed in 70% ethanol, and stored at 4°C until analysis. Before cytofluorimetric analysis, 10^6 cells were washed twice in PBS and incubated in staining solution (10 mg/ml propidium iodide, 100 mg/ml RNase) at room temperature in the dark for at least 2 h. Cell cycle profiles were assessed using a FACSCalibur device (Becton Dickinson, San Jose, CA, USA) and analyzed using Multicycle AV™ software (version 4; De Novo Software). For apoptosis analysis, cells were harvested after 6, 12, 24, 48, 72 h and treated with Annexin V/7-aminoactinomycin D (7-AAD) solution (BD Pharmingen) according to the manufacturer's instructions. The fluorescent intensity of at least 20 000 cells from each sample was measured. Data were acquired using CellQuest software (BD, San Jose, CA) and analyzed using FCS Express software (version 4, De Novo Software).

Western Blot Analysis

Standard SDS-PAGE and Western blotting procedures were applied to total cell extracts. Anti-BRAF (F7) antibody (catalog no.sc-5284) was purchased from Santa Cruz

Biotechnology; anti-phospho-ERK1/2 (product no. M8159), anti-ERK1/2 (product no.M5670) and anti-B actin (product no. A2066) antibodies were purchased from Sigma-Aldrich.

References

1. Davies H, Bignell GR, Cox C, Stephens P, Edkins S, Clegg S, Teague J, Woffendin H, Garnett MJ, Bottomley W, Davis N, Dicks E, Ewing R, *et al.* Mutations of the BRAF gene in human cancer. *Nature*. 2002; 417:949-954.
2. Chng WJ, Gonzalez-Paz N, Price-Troska T, Jacobus S, Rajkumar SV, Oken MM, Kyle RA, Henderson KJ, Van Wier S, Greipp P, Van Ness B, Fonseca R. Clinical and biological significance of RAS mutations in multiple myeloma. *Leukemia*. 2008; 22:2280-2284.
3. Chang YS, Yeh KT, Hsu NC, Lin SH, Chang TJ, Chang JG. Detection of N-, H-, and KRAS codons 12, 13, and 61 mutations with universal RAS primer multiplex PCR and N-, H-, and KRAS-specific primer extension. *Clinical biochemistry*. 2010; 43:296-301.
4. Todoerti K, Agnelli L, Fabris S, Lionetti M, Tuana G, Mosca L, Lombardi L, Grieco V, Bianchino G, D'Auria F, Statuto T, Mazzoccoli C, De Luca L, *et al.* Transcriptional characterization of a prospective series of primary plasma cell leukemia revealed signatures associated with tumor progression and poorer outcome. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2013; 19:3247-3258.
5. Irizarry RA, Hobbs B, Collin F, Beazer-Barclay YD, Antonellis KJ, Scherf U, Speed TP. Exploration, normalization, and summaries of high density oligonucleotide array probe level data. *Biostatistics*. 2003; 4:249-264.
6. Dai MH, Wang PL, Boyd AD, Kostov G, Athey B, Jones EG, Bunney WE, Myers RM, Speed TP, Akil H, Watson SJ, Meng F. Evolving gene/transcript definitions significantly alter the interpretation of GeneChip data. *Nucleic acids research*. 2005; 33:e175.
7. Tusher VG, Tibshirani R, Chu G. Significance analysis of microarrays applied to the ionizing radiation response. *Proceedings of the National Academy of Sciences of the United States of America*. 2001; 98:5116-5121.
8. Chen J, Bardes EE, Aronow BJ, Jegga AG. ToppGene Suite for gene list enrichment analysis and candidate gene prioritization. *Nucleic acids research*. 2009; 37:W305-311.

9. Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, Paulovich A, Pomeroy SL, Golub TR, Lander ES, Mesirov JP. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proceedings of the National Academy of Sciences of the United States of America*. 2005; 102:15545-15550.
10. Lombardi L, Poretti G, Mattioli M, Fabris S, Agnelli L, Bicciato S, Kwee I, Rinaldi A, Ronchetti D, Verdelli D, Lambertenghi-Deliliers G, Bertoni F, Neri A. Molecular characterization of human multiple myeloma cell lines by integrative genomics: Insights into the biology of the disease. *Gene Chromosome Canc*. 2007; 46:226-238.
11. Verdelli D, Nobili L, Todoerti K, Mosca L, Fabris S, D'Anca M, Pellegrino E, Piva R, Inghirami G, Capelli C, Introna M, Baldini L, Chiaramonte R, *et al*. Molecular events underlying interleukin-6 independence in a subclone of the CMA-03 multiple myeloma cell line. *Genes, chromosomes & cancer*. 2014; 53:154-167.
12. Lode L, Moreau P, Menard A, Godon C, Touzeau C, Amiot M, Le Gouill S, Bene MC, Pellat-Deceunynck C. Lack of BRAF V600E mutation in human myeloma cell lines established from myeloma patients with extramedullary disease. *Blood Cancer J*. 2013; 3:e163.

Supplementary Table 1A: Clinical and molecular characteristics of 20 MM/PCL patients carrying non-synonymous *BRAF* mutations

Sample name	Disease	Variant (GRCh38)	% of mutated sequencing reads	AA change	Sex	Age	PP [†]	del(13)*	del(17p)*	1q gain*	1p loss*	t(4;14)	t(11;14)	t(14;16)	t(14;20)	HD [°]
MM-036	MM	140753355C>T	5.61%	D594N	M	65	Gκ	+	-	+	nd	-	-	-	-	-
MM-039	MM	140753333T>G	28.53%	K601T	M	50	Gλ	-	-	-	nd	-	-	-	-	+
MM-140	MM	140753355C>T	1.68%	D594N	M	62	na	-	-	-	nd	-	+	-	-	-
MM-177	MM	140753336A>T	27.18%	V600E	M	73	Gκ	+	-	+	-	-	-	-	-	-
MM-219	MM	140753354T>C	7.78%	D594G	M	73	Aλ	+	-	-	-	-	-	-	-	+
MM-224	MM	140753349C>G	35.44%	G596R	F	52	Gκ	-	-	+	-	-	-	+	-	-
MM-268	MM	140753336A>T	29.57%	V600E	M	77	Gκ	-	-	-	-	-	-	-	-	-
MM-295	MM	140753355C>T	50.00%	D594N	F	74	λ	+	-	nd	nd	-	-	-	-	nd
MM-313	MM	140753336A>T	44.59%	V600E	M	66	Gλ	-	-	-	-	-	+	-	-	-
MM-335	MM	140781602C>G	50.52%	G469A	F	68	Aκ	+	-	-	-	-	-	+	-	-
MM-411	MM	140753354T>C	0.86%	D594G	M	67	Gλ	-	-	-	-	-	-	-	-	+
MM-435	MM	140753354T>C	22.22%	D594G	na	42	Gλ	-	-	-	-	-	-	-	-	+
MM-441	MM	140753336A>T	35.74%	V600E	na	na	Gλ	-	-	-	-	-	-	-	-	-
MM-446	MM	140753336A>T	26.48%	V600E	na	52	Gλ	-	-	+	-	-	-	-	-	+
PCL-015	pPCL	140753336A>T	41.18%	V600E	M	78	k	+	-	+	-	-	-	+	-	-
PCL-023	pPCL	140781602C>G	70.70%	G469A	M	60	Gκ	+	+	+	+	-	-	+	-	-
PCL-026	pPCL	140753355C>T	43.05%	D594N	F	59	Gκ	+	-	+	-	-	-	+	-	-
		140753379C>T	42.15%	E586K												
PCL-028	pPCL	140753353A>C	4.65%	D594E	F	57	κ	+	-	-	-	-	+	-	-	-
PCL-042	sPCL	140781602C>G	51.26%	G469A	F	69	Gλ	-	-	-	-	-	+	-	-	-
PCL-043	pPCL	140753336A>T	44.59%	V600E	F	68	Gλ	-	-	-	-	-	+	-	-	-

[†]Paraprotein; *del(13), del(17), 1p loss and 1q gain were determined by FISH. [°]HD = presence of the hyperdiploid status on the basis of FISH evaluation criteria.

Supplementary Table 1B: Clinical and molecular characteristics of the 167 MM/PCL patients analyzed for *BRAF* mutations

Characteristic	All patients (n=167)		<i>BRAF</i> wild type (n=147)		<i>BRAF</i> mutated (n=20)		P value ^a
	N	%	n	%	n	%	
MM	132	79	118	80.3	14	70	
pPCL	24	14.4	19	12.9	5	25	n.s.
sPCL	11	6.6	10	6.8	1	5	
del(13q)	79	47.6	70	47.9	9	45	
chr 13 disomic patients	87	52.4	76	52.1	11	55	n.s.
del(17p)	18	10.9	17	11.7	1	5	
17p disomic patients	147	89.1	128	88.3	19	95	n.s.
1q gain	68	43	61	43.9	7	36.8	
1q disomic patients	90	57	78	56.1	12	63.2	n.s.
1p loss	18	12.7	17	13.5	1	6.25	
1p disomic patients	124	87.3	109	86.5	15	93.75	n.s.
<i>IGH</i> trx	82	49.7	72	49.6	10	50	
no <i>IGH</i> trx	83	50.3	73	50.4	10	50	n.s.
hyperdiploid	50	32.9	45	33.8	5	26.3	
non-hyperdiploid	102	67.1	88	66.2	14	73.7	n.s.

^a Significance was assessed by Freeman-Halton extension of Fisher's exact test for disease type, and by Fisher's exact test for all other variables.

Supplementary Table 2A: Non-synonymous *NRAS* mutations identified by NGS in MM/PCL patients

Variant (GRCh38)	AA change	dbSNP ID	COSMIC ID (v71)	MM literature*	Mutated patients (percentage of mutated sequencing reads)													
					MM-424 (95.95%)	PCL-041 (90.50%)	MM-174 (80.77%)	PCL-039 (63.37%)	MM-212 (46.70%)	MM-034 (46.37%)	MM-049 (34.44%)	MM-435 (29.87%)	MM-241 (28.98%)	MM-381 (21.99%)	MM-261 (21.63%)	MM-229 (11.68%)	MM-387 (4.56%)	
114713908T>C	Q61R	rs11554290	COSM584	1, 2, 3	MM-424 (95.95%)	PCL-041 (90.50%)	MM-174 (80.77%)	PCL-039 (63.37%)	MM-212 (46.70%)	MM-034 (46.37%)	MM-049 (34.44%)	MM-435 (29.87%)	MM-241 (28.98%)	MM-381 (21.99%)	MM-261 (21.63%)	MM-229 (11.68%)	MM-387 (4.56%)	
114713909G>T	Q61K	rs121913254	COSM580	1, 2, 3, 4	MM-434 (88.27%)	MM-449 (84.36%)	PCL-005 (45.05%)	MM-246 (40.98%)	MM-330 (33.91%)	MM-414 (31.87%)	MM-087 (31.74%)	MM-448 (26.60%)	MM-037 (12.22%)	PCL-039 (8.26%)	MM-327 (4.28%)	MM-284 (5.86%)		
114713907T>G	Q61H	/	COSM586	1, 3, 4	MM-398 (47.45%)	MM-405 (43.42%)	MM-341 (31.48%)	MM-387 (18.46%)	PCL-027 (65.62%)	MM-302 (15.73%)								
114716124C>G	G13R	rs121434595	COSM569	1, 3, 4	MM-372 (52.60%)	MM-066 (30.19%)	MM-302 (17.44%)	MM-284 (4.61%)										
114716126C>T	G12D	rs121913237	COSM564	3	MM-334 (96.49%)	MM-422 (11.95%)	MM-295 (8.89%)											
114713907delT	E62Kfs*6	/	/	/	MM-405 (2.67%)	MM-212 (1.28%)	MM-229 (1.27%)											
114713907T>A	Q61H	rs121913255	COSM585	1, 3, 4	MM-317 (48.82%)	MM-146 (6.11%)												
114713908T>A	Q61L	rs11554290	COSM583	3	MM-276 (45.41%)	MM-448 (10.37%)												
114716123C>T	G13D	rs121434596	COSM573	3	PCL-011 (93.06%)													
114713908_114713909TG>GT	Q61T	/	/	/	MM-314 (82%)													
114713878T>C	Y71C	/	/	/	MM-276 (45.95%)													
114713831T>A	S87C	/	/	/	MM-411 (45.45%)													
114713831delT	S87Afs*17	/	/	/	MM-411 (2.24%)													

*1: Chapman *et al.*, Nature 2011; 2: Walker *et al.*, Blood 2012; 3: Lohr *et al.*, Cancer Cell 2014; 4: Bolli *et al.*, Nature Communications 2014.

Supplementary Table 2B: Clinical and molecular characteristics of 40 MM/PCL patients carrying non-synonymous NRAS mutations

Sample name	Disease	Variant (GRCh38)	% of mutated sequencing reads	AA change	Sex	Age	PP [†]	del(13)*	del(17p)*	1q gain*	1p loss*	t(4;14)	t(11;14)	t(14;16)	t(14;20)	HD [°]
MM-405	MM	114713907T>G/g.114713907delT	43.42%/2.67%	Q61H/E62Kfs*6	M	69	Gk	-	-	-	-	-	+	-	-	-
MM-034	MM	114713908T>C	46.37%	Q61R	M	71	Gk	-	-	-	-	-	-	-	-	+
MM-037	MM	114713909G>T	12.22%	Q61K	F	50	Gk,Ak	+	-	-	-	-	+	-	-	-
MM-049	MM	114713908T>C	34.44%	Q61R	M	62	k	-	-	-	-	-	-	-	-	+
MM-066	MM	114716124C>G	30.19%	G13R	F	77	Ak	-	-	-	na	+	-	-	-	-
MM-087	MM	114713909G>T	31.74%	Q61K	F	84	Gλ	+	-	+	na	+	-	-	-	-
MM-146	MM	114713907T>A	6.11%	Q61H	M	68	Gλ	-	-	+	na	-	-	-	-	+
MM-174	MM	114713908T>C	80.77%	Q61R	M	85	Ak	-	-	-	-	-	-	-	-	+
MM-212	MM	114713908T>C/g.114713907delT	46.70%/1.28%	Q61R/E62Kfs*6	F	55	Gk	-	-	-	-	-	+	-	-	-
MM-229	MM	114713908T>C/g.114713907delT	11.68%/1.27%	Q61R/E62Kfs*6	M	75	Gk	-	-	-	-	-	-	-	-	-
MM-241	MM	114713908T>C	28.98%	Q61R	M	54	Gk	-	-	-	-	-	-	-	-	+
MM-246	MM	114713909G>T	40.98%	Q61K	M	71	Aλ	-	-	-	-	-	+	-	-	-
MM-261	MM	114713908T>C	21.63%	Q61R	F	66	Gk	+	-	+	-	-	-	-	-	+
MM-276	MM	114713878T>C/g.114713908T>A	45.95%/45.41%	Y71C/Q61L	F	70	Gk	+	-	-	-	+	-	-	-	-
MM-284	MM	114716124C>G/g.114713909G>T	4.61%/5.86%	G13R/Q61K	M	49	k	na	-	-	-	-	+	-	-	-
MM-295	MM	114716126C>T	8.89%	G12D	F	74	λ	+	-	na	na	-	-	-	-	na
MM-302	MM	114716124C>G/g.114713907T>G	17.44%/15.73%	G13R/Q61H	M	65	Ak	-	-	+	-	-	-	-	-	+
MM-314	MM	114713908_114713909TG>GT	82.04%	Q61T	F	70	Aλ	-	-	+	-	-	+	-	-	-
MM-317	MM	114713907T>A	48.82%	Q61H	M	56	Gλ	-	-	+	-	-	+	-	-	-
MM-327	MM	114713909G>T	4.28%	Q61K	F	62	absent	+	-	+	-	-	-	-	-	+
MM-330	MM	114713909G>T	33.91%	Q61K	F	61	Ak	+	-	+	-	+	-	-	-	-
MM-334	MM	114716126C>T	96.49%	G12D	F	45	Gλ	+	+	+	+	-	-	-	-	+
MM-341	MM	114713907T>G	31.48%	Q61H	M	65	Gk	-	-	+	-	-	-	-	-	-
MM-372	MM	114716124C>G	52.60%	G13R	M	54	Gk	-	-	+	-	-	+	-	-	-
MM-381	MM	114713908T>C	21.99%	Q61R	F	69	Gk	-	-	-	-	-	-	-	na	na
MM-387	MM	114713907T>G/g.114713908T>C	18.46%/4.56%	Q61H/Q61R	F	44	Gk	-	-	+	-	-	-	-	-	+
MM-398	MM	114713907T>G	47.45%	Q61H	F	65	Gk	+	-	+	-	-	+	-	-	-
MM-411	MM	114713831T>A/g.114713831delT	45.45%/2.24%	S87C/ S87Afs*17	M	67	Gλ	-	-	-	-	-	-	-	-	+
MM-414	MM	114713909G>T	31.87%	Q61K	F	67	Gλ	-	-	-	-	-	+	-	-	-
MM-422	MM	114716126C>T	11.95%	G12D	F	74	k	+	-	-	-	-	-	-	-	-
MM-424	MM	114713908T>C	95.95%	Q61R	F	84	Gk	+	-	-	+	-	-	-	-	+
MM-434	MM	114713909G>T	88.27%	Q61K	F	53	k	+	-	-	+	-	-	-	-	-
MM-435	MM	114713908T>C	29.87%	Q61R	na	42	Gλ	-	-	-	-	-	-	-	-	+
MM-448	MM	114713909G>T/g.114713908T>A	26.60%/10.37%	Q61K/Q61L	na	59	Ak	+	-	+	-	-	-	-	-	-
MM-449	MM	114713909G>T	84.36%	Q61K	na	54	Gk	+	-	-	+	-	-	-	-	-
PCL-005	sPCL	114713909G>T	45.05%	Q61K	M	76	Ak	-	-	-	-	-	+	-	-	-
PCL-011	sPCL	114716123C>T	93.06%	G13D	M	76	Gk	+	+	+	-	-	-	+	-	-
PCL-027	pPCL	114713907T>G	65.62%	Q61H	M	65	λ	-	+	-	-	-	+	-	-	-
PCL-039	sPCL	114713908T>C/g.114713909G>T	63.37%/8.26%	Q61R/Q61K	M	54	Ak	-	+	-	+	-	-	-	-	-
PCL-041	sPCL	114713908T>C	90.50%	Q61R	M	na	Gk	-	-	na	na	-	na	na	na	na

[†]Paraprotein; *del(13), del(17), 1p loss and 1q gain were determined by FISH. [°]HD = presence of the hyperdiploid status on the basis of FISH evaluation criteria.

Supplementary Table 2C: Clinical and molecular characteristics of the 167 MM/PCL patients analyzed for *NRAS* mutations

Characteristic	All patients (n=167)		NRAS wild type (n=127)		NRAS mutated (n=40)		P value ^a
	N	%	n	%	n	%	
MM	132	79	97	76.4	35	87.5	0.008
pPCL	24	14.4	23	18.1	1	2.5	
sPCL	11	6.6	7	5.5	4	10	
del(13q)	79	47.6	64	50.4	15	38.5	n.s.
chr 13 disomic patients	87	52.4	63	49.6	24	61.5	
del(17p)	18	10.9	14	11.2	4	10	n.s.
17p disomic patients	147	89.1	111	88.8	36	90	
1q gain	68	43	53	44.2	15	39.5	n.s.
1q disomic patients	90	57	67	55.8	23	60.5	
1p loss	18	12.7	13	12.1	5	14.3	n.s.
1p disomic patients	124	87.3	94	87.9	30	85.7	
<i>IGH</i> trx	82	49.7	64	49.8	18	46.1	n.s.
no <i>IGH</i> trx	83	50.3	62	50.2	21	53.8	
hyperdiploid	50	32.9	37	32.2	13	35.1	n.s.
non-hyperdiploid	102	67.1	78	67.8	24	64.9	

^a Significance was assessed by Freeman-Halton extension of Fisher's exact test for disease type, and by Fisher's exact test for all other variables.

Supplementary Table 3A: Non-synonymous *KRAS* mutations identified by NGS in MM/PCL patients

Variant (GRCh38)	AA change	dbSNP ID	COSMIC ID (v71)	MM literature*	Mutated patients (percentage of mutated sequencing reads)									
25227341T>G	Q61H	rs17851045	COSM554	1, 2, 3, 4	MM-055 (52.53%)	MM-442 (45.93%)	MM-430 (39.77%)	MM-440 (38%)	MM-327 (37.34%)	MM-202 (35.54%)	MM-151 (35.29%)	MM-402 (34.62%)	MM-200 (23.74%)	
					MM-229 (12.07%)	MM-381 (9.51%)	MM-441 (8.45%)	PCL-046 (7.73%)	MM-300 (7.61%)					
25245350C>T	G12D	rs121913529	COSM521	3	MM-154 (45.28%)	MM-030 (45%)	MM-152 (31.69%)	MM-433 (31.78%)	MM-026 (30.59%)	MM-429 (21.47%)	PCL-046 (9.01%)			
25245351C>G	G12R	rs121913530	COSM518	2, 3, 4	MM-039 (46.94%)	PCL-038 (42.42%)	MM-445 (28.85%)	MM-159 (27.78%)						
25245347C>T	G13D	rs112445441	COSM532	1, 2, 3	MM-385 (40.68%)	MM-308 (39.42%)	MM-151 (9.35%)	MM-219 (5.88%)						
25227342T>C	Q61R	rs121913240	COSM552	2, 3, 4	MM-038 (40.82%)	MM-351 (28.66%)	MM-159 (3.75%)							
25245321G>T	Q22K	rs121913236	COSM543	4	MM-280 (50%)	MM-026 (32.94%)								
25225628C>T	A146T	rs121913527	COSM19404	1	MM-413 (51.24%)	MM-131 (40.52%)								
25245348C>A	G13C	rs121913535	COSM527	/	PCL-001 (48.64%)	MM-149 (29.13%)								
25225713T>G	K117N	/	COSM19940	1, 3	MM-310 (47.09%)	MM-269 (3.07%)								
25245328C>G	L19F	/	COSM12703	1	MM-386 (42.73%)	MM-335 (18.6%)								
25245350C>A	G12V	rs121913529	COSM520	3, 4	MM-146 (24.58%)	MM-150 (8.93%)								
25227334A>T	Y64N	/	/	3	MM-115 (54.2%)									
25227266A>T	N86K	/	/	/	MM-115 (44.27%)									
25245332G>T	A18D	/	COSM542	3	PCL-021 (43.8%)									
25227334A>C	Y64D	/	/	1	MM-410 (41.96%)									
25245351C>A	G12C	rs121913530	COSM516	4	MM-243 (40.5%)									
25227348G>C	A59G	/	COSM28518	3	MM-079 (23.13%)									
25227330C>A	S65I	/	/	/	PCL-009 (15.84%)									
25245350C>G	G12A	rs121913529	COSM522	3, 4	MM-140 (14.81%)									
25227342T>A	Q61L	rs121913240	COSM553	3, 4	MM-284 (9.54%)									
25245351C>T	G12S	rs121913530	COSM517	/	MM-004 (9.18%)									
25225657C>T	S136N	/	/	/	MM-202 (4.86%)									
25225627G>A	A146V	/	COSM19900	4	MM-445 (2.53%)									

*1: Chapman *et al.*, Nature 2011; 2: Walker *et al.*, Blood 2012; 3: Lohr *et al.*, Cancer Cell 2014; 4: Bolli *et al.*, Nature Communications 2014.

Supplementary Table 3B: Clinical and molecular characteristics of 49 MM/PCL patients carrying non-synonymous KRAS mutations

Sample name	Disease	Variant (GRCh38)	% of mutated sequencing reads	AA change	Sex	Age	PP [†]	del(13)*	del(17p)*	1q gain*	1p loss*	t(4;14)	t(11;14)	t(14;16)	t(14;20)	HD [°]
MM-004	MM	25245351C>T	9.18%	G12S	F	58	Gκ	-	na	na	na	-	-	+	-	na
MM-026	MM	25245321G>T/g.25245350C>T	32.94%/30.59%	Q22K/G12D	F	72	κ	-	-	-	na	-	+	-	-	na
MM-030	MM	25245350C>T	45.00%	G12D	M	69	Gλ	-	-	-	+	-	-	-	-	+
MM-038	MM	25227342T>C	40.82%	Q61R	F	67	κ	+	-	-	-	-	-	-	-	+
MM-039	MM	25245351C>G	46.94%	G12R	M	50	Gλ	-	-	-	na	-	-	-	-	+
MM-055	MM	25227341T>G	52.53%	Q61H	F	69	Gκ	-	-	-	na	-	+	-	-	-
MM-079	MM	25227348G>C	23.13%	A59G	F	74	Gκ+Gλ	-	-	-	-	-	-	-	-	+
MM-115	MM	25227334A>T/g.25227266A>T	54.20%/44.27%	Y64N/N86K	F	53	Gλ	+	-	na	na	-	+	-	-	-
MM-131	MM	25225628C>T	40.52%	A146T	M	73	Gκ	-	-	-	-	-	-	-	-	-
MM-140	MM	25245350C>G	14.81%	G12A	M	62	Gκ	-	-	-	na	-	+	-	-	-
MM-146	MM	25245350C>A	24.58%	G12V	M	68	Gλ	-	-	+	na	-	-	-	-	+
MM-149	MM	25245348C>A	29.13%	G13C	F	52	Gλ	-	-	-	-	-	-	-	-	+
MM-150	MM	25245350C>A	8.93%	G12V	F	68	Gλ	-	-	+	-	-	-	-	-	+
MM-151	MM	25227341T>G/g.25245347C>T	35.29%/9.35%	Q61H/G13D	F	71	Gλ	-	-	-	-	-	-	-	-	+
MM-152	MM	25245350C>T	31.69%	G12D	M	66	Gκ	-	-	-	na	-	-	-	-	+
MM-154	MM	25245350C>T	45.28%	G12D	F	71	Gκ	+	-	+	-	-	-	+	-	-
MM-159	MM	25245351C>G/g.25227342T>C	27.78%/3.75%	G12R/Q61R	M	56	κ	+	-	-	-	-	+	-	-	-
MM-200	MM	25227341T>G	23.74%	Q61H	F	63	Aλ	-	-	-	-	-	-	-	-	+
MM-202	MM	25227341T>G/g.25225657C>T	35.54%/4.86%	Q61H/S136N	M	64	Gκ	-	-	+	na	-	-	-	-	+
MM-219	MM	25245347C>T	5.88%	G13D	M	73	Aλ	+	-	-	-	-	-	-	-	+
MM-229	MM	25227341T>G	12.07%	Q61H	M	75	Gκ	-	-	-	-	-	-	-	-	-
MM-243	MM	25245351C>A	40.50%	G12C	F	68	Gκ	-	-	+	-	-	-	-	-	+
MM-269	MM	25225713T>G	3.07%	K117N	F	67	Gκ	+	-	-	-	-	-	-	-	+
MM-280	MM	25245321G>T	50.00%	Q22K	M	62	Gλ	-	-	-	-	-	+	-	-	-
MM-284	MM	25227342T>A	9.54%	Q61L	M	49	κ	na	-	-	-	-	+	-	-	-
MM-300	MM	25227341T>G	7.61%	Q61H	M	65	Gκ	-	-	na	na	-	-	-	-	na
MM-308	MM	25245347C>T	39.42%	G13D	M	58	Aκ	-	-	-	-	-	-	-	-	-
MM-310	MM	25225713T>G	47.09%	K117N	M	67	Gι	-	-	-	-	-	+	-	-	-
MM-327	MM	25227341T>G	37.34%	Q61H	F	62	absent	+	-	+	-	-	-	-	-	+
MM-335	MM	25245328C>G	18.60%	L19F	F	68	Aκ	+	-	-	-	-	-	+	-	-
MM-351	MM	25227342T>C	28.66%	Q61R	M	na	Aκ	-	-	-	-	-	-	-	-	-
MM-381	MM	25227341T>G	9.51%	Q61H	F	69	Gκ	-	-	-	-	-	-	-	na	na
MM-385	MM	25245347C>T	40.68%	G13D	F	76	Gλ	-	-	-	-	-	-	-	-	+
MM-386	MM	25245328C>G	42.73%	L19F	F	76	Aλ	-	-	na	na	-	na	na	na	na
MM-402	MM	25227341T>G	34.62%	Q61H	M	67	Gκ	+	-	-	-	-	+	-	-	-
MM-410	MM	25227334A>C	41.96%	Y64D	F	79	λ	-	-	+	-	-	-	-	-	-

Sample name	Disease	Variant (GRCh38)	% of mutated sequencing reads	AA change	Sex	Age	PP [†]	del(13)*	del(17p)*	1q gain*	1p loss*	t(4;14)	t(11;14)	t(14;16)	t(14;20)	HD [°]
MM-413	MM	25225628C>T	51.24%	A146T	F	51	Gκ	+	-	+	-	-	-	-	-	+
MM-429	MM	25245350C>T	21.47%	G12D	F	63	Gκ	-	-	+	-	-	-	-	-	+
MM-430	MM	25227341T>G	39.77%	Q61H	na	62	Gλ	-	-	-	-	-	-	-	-	-
MM-433	MM	25245350C>T	31.78%	G12D	M	72	κ	-	-	-	-	-	+	-	-	-
MM-440	MM	25227341T>G	38.00%	Q61H	na	61	Aκ	-	-	-	+	-	-	-	-	+
MM-441	MM	25227341T>G	8.45%	Q61H	na	na	Gλ	-	-	-	-	-	-	-	-	-
MM-442	sPCL	25227341T>G	45.93%	Q61H	F	65	Gκ	+	-	+	+	-	-	-	-	-
MM-445	MM	25245351C>G/g.25225627G>A	28.85%/2.53%	G12R/A146V	na	65	κ	+	-	+	-	-	+	-	-	-
PCL-001	pPCL	25245348C>A	48.64%	G13C	F	51	κ	+	-	na	na	-	-	-	+	na
PCL-009	sPCL	25227330C>A	15.84%	S65I	F	77	Gκ	+	-	+	-	+	-	-	-	-
PCL-021	pPCL	25245332G>T	43.80%	A18D	M	48	Gλ	+	-	+	-	+	-	-	-	-
PCL-038	pPCL	25245351C>G	42.42%	G12R	M	57	Gκ	+	-	+	+	-	-	-	-	+
PCL-046	pPCL	25227341T>G/g.25245350C>T	7.73%/9.01%	Q61H/G12D	F	50	κ	+	-	+	-	-	-	-	-	+

[†]Paraprotein; *del(13), del(17), 1p loss and 1q gain were determined by FISH. [°]HD = presence of the hyperdiploid status on the basis of FISH evaluation criteria.

Supplementary Table 3C: Clinical and molecular characteristics of the 167 MM/PCL patients analyzed for *KRAS* mutations.

Characteristic	All patients (n=167)		<i>KRAS</i> wild type (n=118)		<i>KRAS</i> mutated (n=49)		P value ^a
	N	%	n	%	n	%	
MM	132	79	89	75.4	43	87.7	
pPCL	24	14.4	20	17	4	8.2	n.s.
sPCL	11	6.6	9	7.6	2	4.1	
del(13q)	79	47.6	62	52.5	17	35.4	
chr 13 disomic patients	87	52.4	56	47.5	31	64.6	n.s.
del(17p)	18	10.9	18	15.4	0	0	
17p disomic patients	147	89.1	99	84.6	48	100	0.0019
1q gain	68	43	53	46.5	15	34.1	
1q disomic patients	90	57	61	53.5	29	55.9	n.s.
1p loss	18	12.7	14	13.3	4	10.8	
1p disomic patients	124	87.3	91	86.7	33	89.2	n.s.
<i>IGH</i> trx	82	49.7	64	54.7	18	37.5	
no <i>IGH</i> trx	83	50.3	53	45.3	30	62.5	n.s.
hyperdiploid	50	32.9	29	26.6	21	48.8	
non-hyperdiploid	102	67.1	80	73.4	22	51.2	0.0124

^a Significance was assessed by Freeman-Halton extension of Fisher's exact test for disease type, and by Fisher's exact test for all other variables.

Supplementary Table 4A: Modulated genes between *BRAF/NRAS/KRAS* mutated and wild type patients

<i>Gene</i>	<i>SAM score</i> [‡]	<i>Gene</i>	<i>SAM score</i>
<i>PLEKHA1*</i>	-5.163765	<i>ETV5*</i>	6.24113
<i>ARL6IP5*</i>	-4.342202	<i>CD300A*</i>	5.77917
<i>ZHX1*</i>	-4.311072	<i>SPRED2*</i>	5.37656
<i>SMAD3*</i>	-4.287826	<i>DUSP6*</i>	4.36551
<i>AHNAK*</i>	-4.268764	<i>NRXN1*</i>	4.23105
<i>TBC1D1*</i>	-4.237025	<i>MORC1*</i>	4.20022
<i>MTMR4*</i>	-4.167526	<i>F12*</i>	4.15845
<i>COPZ2*</i>	-4.103426	<i>RRBP1*</i>	4.07138
<i>RASGRP1*</i>	-4.07288	<i>TDO2*</i>	4.03133
<i>FLNB</i>	-3.961337	<i>LGALS3BP</i>	3.97907
<i>BMP2K</i>	-3.918772	<i>PRKD2</i>	3.94335
<i>RIOK1</i>	-3.907891	<i>ACVR1B</i>	3.94093
<i>TSHZ1</i>	-3.904952	<i>ASPHD2</i>	3.92401
<i>NOTCH2</i>	-3.883492	<i>GPR56</i>	3.89699
<i>ZNF681</i>	-3.859508	<i>ACOXL</i>	3.85393
<i>DHCR24</i>	-3.82483	<i>SPRED1</i>	3.8092
<i>LIFR</i>	-3.819499	<i>LY9</i>	3.76526
<i>ATL3</i>	-3.818015	<i>ABI3BP</i>	3.74618
<i>TGIF2</i>	-3.794277	<i>DPEP1</i>	3.70693
<i>GLO1</i>	-3.776065	<i>TMEM184B</i>	3.70255
<i>C6orf89</i>	-3.76636	<i>PDE4D</i>	3.69971
<i>ZNF570</i>	-3.738105	<i>P2RY6</i>	3.68381
<i>CYB5R1</i>	-3.727057	<i>CASP4</i>	3.66862
<i>PHTF2</i>	-3.721193	<i>MIR221</i>	3.65834
<i>DENND5A</i>	-3.687141	<i>SEMA4D</i>	3.65632
<i>CD52</i>	-3.67123	<i>MAPKAPK2</i>	3.64609
<i>ZADH2</i>	-3.659897	<i>SIX4</i>	3.64407
<i>RRP8</i>	-3.635111		
<i>BCL2</i>	-3.627357		
<i>JAZF1</i>	-3.621626		
<i>CD86</i>	-3.612113		
<i>LAMTOR3</i>	-3.611917		
<i>SAMSN1</i>	-3.587501		
<i>NTAN1</i>	-3.586428		
<i>PLCL1</i>	-3.549426		
<i>LRIG3</i>	-3.547287		
<i>PPM1L</i>	-3.545335		
<i>GRB2</i>	-3.537068		
<i>NECAP2</i>	-3.522351		
<i>KDM1B</i>	-3.513606		
<i>KAT6A</i>	-3.51165		
<i>BCL11A</i>	-3.499956		
<i>NEU1</i>	-3.491002		
<i>KIAA1147</i>	-3.487048		
<i>NUDT7</i>	-3.455732		
<i>SLC23A2</i>	-3.446583		
<i>LSS</i>	-3.442877		
<i>LYPD6B</i>	-3.440096		
<i>ARHGEF3</i>	-3.436137		
<i>CLN3</i>	-3.423259		
<i>INSIG1</i>	-3.421565		
<i>LYN</i>	-3.382145		
<i>APBB1IP</i>	-3.378885		
<i>MYL2</i>	-3.372364		
<i>TOP3A</i>	-3.359096		
<i>AKAP5</i>	-3.358922		
<i>RPSAP58</i>	-3.357769		
<i>EDN1</i>	-3.355883		
<i>SQLE</i>	-3.353543		

*Differentially expressed gene at the highest stringency level. [‡]Negative scores indicate downregulated genes in mutated cases.

Supplementary Table 4B: Enriched biological processes in the MAPK mutation-associated 86 gene signature (q value<0.005)

<i>ID</i>	<i>Name</i>	<i>q value</i>	<i>Hit in Query List</i>
GO:0010563	negative regulation of phosphorus metabolic process	1.29E03	SEMA4D, AKAP5, PDE4D, SPRED2, CD300A, DUSP6, SPRED1, SAMS1N1, LYN, EDN1, SMAD3
GO:0045936	negative regulation of phosphate metabolic process	1.29E03	SEMA4D, AKAP5, PDE4D, SPRED2, CD300A, DUSP6, SPRED1, SAMS1N1, LYN, EDN1, SMAD3
GO:0042325	regulation of phosphorylation	1.29E03	PRKD2, SEMA4D, AKAP5, PDE4D, SPRED2, MAPKAPK2, CD300A, ARL6IP5, DUSP6, GRB2, LAMTOR3, SPRED1, BCL2, PLCL1, ACVR1B, SAMS1N1, LYN, EDN1, SMAD3
GO:0001932	regulation of protein phosphorylation	1.29E03	PRKD2, SEMA4D, AKAP5, PDE4D, SPRED2, MAPKAPK2, CD300A, DUSP6, LAMTOR3, SPRED1, BCL2, PLCL1, ACVR1B, SAMS1N1, LYN, EDN1, SMAD3
GO:0016126	sterol biosynthetic process	1.29E03	DHCR24, INSIG1, SQLE, LSS, CYB5R1
GO:1902531	regulation of intracellular signal transduction	1.29E03	PRKD2, SEMA4D, RASGRP1, AKAP5, SPRED2, MAPKAPK2, TBC1D1, CD300A, ARL6IP5, DUSP6, GRB2, LAMTOR3, GPR56, PLEKHA1, SPRED1, BCL2, DENND5A, ARHGFE3, LYN, EDN1, NOTCH2
GO:0023056	positive regulation of signaling	1.39E03	PRKD2, SEMA4D, AKAP5, SPRED2, NRXN1, MAPKAPK2, CD300A, ARL6IP5, GRB2, LAMTOR3, GPR56, SPRED1, BCL2, ACVR1B, LYN, EDN1, NOTCH2, SMAD3
GO:0010647	positive regulation of cell communication	1.39E03	PRKD2, SEMA4D, AKAP5, SPRED2, NRXN1, MAPKAPK2, CD300A, ARL6IP5, GRB2, LAMTOR3, GPR56, SPRED1, BCL2, ACVR1B, LYN, EDN1, NOTCH2, SMAD3
GO:1902533	positive regulation of intracellular signal transduction	1.39E03	PRKD2, SEMA4D, AKAP5, SPRED2, MAPKAPK2, CD300A, ARL6IP5, LAMTOR3, GPR56, SPRED1, BCL2, LYN, EDN1, NOTCH2
GO:0048584	positive regulation of response to stimulus	1.39E03	PRKD2, SEMA4D, AKAP5, PDE4D, SPRED2, MAPKAPK2, CD86, CD300A, ARL6IP5, DUSP6, GRB2, LAMTOR3, GPR56, PLEKHA1, SPRED1, BCL2, ACVR1B, LYN, EDN1, NOTCH2, SMAD3
GO:0001933	negative regulation of protein phosphorylation	1.39E03	SEMA4D, PDE4D, SPRED2, CD300A, DUSP6, SPRED1, SAMS1N1, LYN, SMAD3
GO:0019220	regulation of phosphate metabolic process	1.63E03	PRKD2, SEMA4D, RASGRP1, AKAP5, PDE4D, SPRED2, MAPKAPK2, TBC1D1, CD300A, ARL6IP5, DUSP6, GRB2, LAMTOR3, SPRED1, BCL2, PLCL1, DENND5A, ARHGFE3, ACVR1B, SAMS1N1, LYN, EDN1, SMAD3
GO:0002757	immune response activating signal transduction	1.63E03	PRKD2, PDE4D, MAPKAPK2, CD86, CD300A, DUSP6, GRB2, PLEKHA1, BCL2, LYN
GO:0051174	regulation of phosphorus metabolic process	1.63E03	PRKD2, SEMA4D, RASGRP1, AKAP5, PDE4D, SPRED2, MAPKAPK2, TBC1D1, CD300A, ARL6IP5, DUSP6, GRB2, LAMTOR3, SPRED1, BCL2, PLCL1, DENND5A, ARHGFE3, ACVR1B, SAMS1N1, LYN, EDN1, SMAD3
GO:0009967	positive regulation of signal transduction	1.63E03	PRKD2, SEMA4D, AKAP5, SPRED2, MAPKAPK2, CD300A, ARL6IP5, GRB2, LAMTOR3, GPR56, SPRED1, BCL2, ACVR1B, LYN, EDN1, NOTCH2, SMAD3
GO:0002764	immune response regulating signaling pathway	2.03E03	PRKD2, RASGRP1, PDE4D, MAPKAPK2, CD86, CD300A, DUSP6, GRB2, PLEKHA1, BCL2, LYN
GO:0051094	positive regulation of developmental process	2.03E03	PRKD2, SEMA4D, TGIF2, AKAP5, NRXN1, CD86, ETV5, BCL2, BCL11A, ACVR1B, SIX4, LYN, EDN1, NOTCH2, SMAD3
GO:0023014	signal transduction by phosphorylation	2.03E03	PRKD2, SPRED2, MAPKAPK2, CD300A, ARL6IP5, DUSP6, PPM1L, GRB2, LAMTOR3, SPRED1, ACVR1B, LYN, EDN1
GO:0031324	negative regulation of cellular metabolic process	2.74E03	SEMA4D, TGIF2, AKAP5, PDE4D, SPRED2, CLN3, CD300A, INSIG1, DUSP6, KAT6A, JAZF1, SPRED1, RRP8, BCL2, BCL11A, SAMS1N1, LYN, ZHX1, EDN1, NOTCH2, SMAD3
GO:0043407	negative regulation of MAP kinase activity	2.74E03	SPRED2, CD300A, DUSP6, SPRED1, LYN
GO:0042326	negative regulation of phosphorylation	2.74E03	SEMA4D, PDE4D, SPRED2, CD300A, DUSP6, SPRED1, SAMS1N1, LYN, SMAD3
GO:0009892	negative regulation of metabolic process	2.74E03	SEMA4D, TGIF2, AKAP5, PDE4D, SPRED2, CLN3, CD300A, INSIG1, DUSP6, KAT6A, JAZF1, SPRED1, RRP8, BCL2, BCL11A, ACVR1B, SAMS1N1, LYN, ZHX1, EDN1, NOTCH2, SMAD3
GO:0006468	protein phosphorylation	2.74E03	PRKD2, SEMA4D, RIOK1, AKAP5, PDE4D, SPRED2, BMP2K, MAPKAPK2, CD300A, DUSP6, LAMTOR3, SPRED1, BCL2, PLCL1, ACVR1B, SAMS1N1, LYN, EDN1, SMAD3
GO:2000026	regulation of multicellular organismal development	2.76E03	PRKD2, SEMA4D, TGIF2, NEU1, AKAP5, NRXN1, BMP2K, CD86, DUSP6, ETV5, GPR56, BCL2, BCL11A, ACVR1B, SIX4, LYN, EDN1, NOTCH2, SMAD3

<i>ID</i>	<i>Name</i>	<i>q value</i>	<i>Hit in Query List</i>
GO:0002253	activation of immune response	2.77E03	<i>PRKD2, PDE4D, MAPKAPK2, CD86, CD300A, DUSP6, GRB2, PLEKHA1, BCL2, LYN</i>
GO:0050853	B cell receptor signaling pathway	3.23E03	<i>CD300A, PLEKHA1, BCL2, LYN</i>
GO:0002768	immune response regulating cell surface receptor signaling pathway	3.29E03	<i>PRKD2, RASGRP1, PDE4D, CD86, CD300A, GRB2, PLEKHA1, BCL2, LYN</i>
GO:0030334	regulation of cell migration	3.29E03	<i>DPEP1, PRKD2, SEMA4D, P2RY6, CD300A, GPR56, BCL2, ACVR1B, LYN, EDN1, SMAD3</i>
GO:0051270	regulation of cellular component movement	3.29E03	<i>DPEP1, PRKD2, SEMA4D, PDE4D, P2RY6, CD300A, GPR56, BCL2, ACVR1B, LYN, EDN1, SMAD3</i>
GO:0000165	MAPK cascade	3.29E03	<i>PRKD2, SPRED2, MAPKAPK2, CD300A, ARL6IP5, DUSP6, PPM1L, GRB2, LAMTOR3, SPRED1, LYN, EDN1</i>
GO:0032269	negative regulation of cellular protein metabolic process	3.62E03	<i>SEMA4D, PDE4D, SPRED2, CLN3, CD300A, DUSP6, SPRED1, SAMS1N1, LYN, EDN1, SMAD3</i>
GO:0016310	phosphorylation	3.73E03	<i>PRKD2, SEMA4D, RIOK1, AKAP5, PDE4D, SPRED2, BMP2K, MAPKAPK2, CD300A, ARL6IP5, DUSP6, PPM1L, GRB2, LAMTOR3, SPRED1, BCL2, PLCL1, ACVR1B, SAMS1N1, LYN, EDN1, SMAD3</i>
GO:0006695	cholesterol biosynthetic process	3.73E03	<i>DHCR24, INSIG1, SQLE, LSS</i>
GO:0007167	enzyme linked receptor protein signaling pathway	4.06E03	<i>LIFR, PRKD2, TGIF2, MTMR4, MAPKAPK2, CD86, DUSP6, PPM1L, GRB2, PLEKHA1, ARHGEF3, ACVR1B, LYN, NOTCH2, SMAD3</i>
GO:0043583	ear development	4.06E03	<i>LRIG3, MAPKAPK2, INSIG1, TSHZ1, BCL2, SIX4, EDN1</i>
GO:0050851	antigen receptor mediated signaling pathway	4.06E03	<i>PRKD2, PDE4D, CD300A, PLEKHA1, BCL2, LYN</i>
GO:0031399	regulation of protein modification process	4.06E03	<i>PRKD2, SEMA4D, AKAP5, PDE4D, SPRED2, MAPKAPK2, CD300A, DUSP6, LAMTOR3, SPRED1, BCL2, PLCL1, ACVR1B, SAMS1N1, LYN, EDN1, SMAD3</i>
GO:0043408	regulation of MAPK cascade	4.06E03	<i>PRKD2, SPRED2, MAPKAPK2, CD300A, ARL6IP5, DUSP6, GRB2, LAMTOR3, SPRED1, LYN, EDN1</i>
GO:2000145	regulation of cell motility	4.08E03	<i>DPEP1, PRKD2, SEMA4D, P2RY6, CD300A, GPR56, BCL2, ACVR1B, LYN, EDN1, SMAD3</i>
GO:0050776	regulation of immune response	4.11E03	<i>PRKD2, RASGRP1, PDE4D, MAPKAPK2, CD86, CD300A, DUSP6, GRB2, PLEKHA1, BCL2, SAMS1N1, LYN, SMAD3</i>
GO:0018193	peptidyl amino acid modification	4.90E03	<i>PRKD2, SEMA4D, PDE4D, SPRED2, MAPKAPK2, CD300A, KAT6A, SPRED1, ASPHD2, BCL2, PLCL1, ACVR1B, SAMS1N1, LYN</i>

Supplementary Table 5A: Description of the 150 genes at the top of the ranked list in GSEA analysis of U266 treated versus control cells

<i>GENE_SYMBOL</i>	<i>GENE_TITLE</i>	<i>SCORE</i>
<i>INHBE</i>	inhibin, beta E	0.676317
<i>CTH</i>	cystathionase (cystathionine gamma-lyase)	0.634
<i>STC2</i>	stanniocalcin 2	0.600332
<i>FBXW10</i>	F-box and WD-40 domain protein 10	0.574939
<i>TCP11L2</i>	t-complex 11 (mouse) like 2	0.532132
<i>ESRP1</i>	epithelial splicing regulatory protein 1	0.509604
<i>DDIT4</i>	DNA-damage-inducible transcript 4	0.498076
<i>TRIB3</i>	tribbles homolog 3 (Drosophila)	0.474272
<i>GRB10</i>	growth factor receptor-bound protein 10	0.471464
<i>ASS1</i>	argininosuccinate synthetase 1	0.458725
<i>TMEM154</i>	transmembrane protein 154	0.457523
<i>GDF15</i>	growth differentiation factor 15	0.452052
<i>FAM106CP</i>	family with sequence similarity 106, member C, pseudogene	0.448272
<i>SESN2</i>	sestrin 2	0.437719
<i>BEX2</i>	brain expressed X-linked 2	0.422433
<i>PCK2</i>	phosphoenolpyruvate carboxykinase 2 (mitochondrial)	0.41365
<i>SLC35F1</i>	solute carrier family 35, member F1	0.405951
<i>CREB5</i>	cAMP responsive element binding protein 5	0.403822
<i>SLC10A5</i>	solute carrier family 10 (sodium/bile acid cotransporter family), member 5	0.401765
<i>HMOX1</i>	heme oxygenase (decycling) 1	0.39905
<i>SNORD116-18</i>	small nucleolar RNA, C/D box 116-18	0.397014
<i>FDPSP2</i>	farnesyl diphosphate synthase pseudogene 2	0.385425
<i>VSNL1</i>	visinin-like 1	0.381415
<i>SNORD116-25</i>	small nucleolar RNA, C/D box 116-25	0.38046
<i>SLC7A11</i>	solute carrier family 7, (cationic amino acid transporter, y+ system) member 11	0.379498
<i>LOC101928082</i>	uncharacterized LOC101928082	0.371708
<i>EIF4EBP1</i>	eukaryotic translation initiation factor 4E binding protein 1	0.36092
<i>TUBE1</i>	tubulin, epsilon 1	0.358196
<i>TRAV27</i>	T cell receptor alpha variable 27	0.356685
<i>CBS</i>	cystathionine-beta-synthase	0.354561
<i>ISY1</i>	ISY1 splicing factor homolog (S. cerevisiae)	0.354496
<i>SNORD115-24</i>	small nucleolar RNA, C/D box 115-24	0.353983
<i>ANK2</i>	ankyrin 2, neuronal	0.351251
<i>LOC101928574</i>	uncharacterized LOC101928574	0.351248
<i>VLDLR</i>	very low density lipoprotein receptor	0.34859
<i>SLC43A1</i>	solute carrier family 43, member 1	0.342847
<i>CXorf61</i>	chromosome X open reading frame 61	0.342714
<i>STON1</i>	stonin 1	0.341989
<i>CTAGE5</i>	CTAGE family, member 5	0.341841
<i>PRSS8</i>	protease, serine, 8 (prostasin)	0.340513
<i>TRAV10</i>	T cell receptor alpha variable 10	0.340494
<i>DDIT3</i>	DNA-damage-inducible transcript 3	0.336389
<i>RPS10P7</i>	ribosomal protein S10 pseudogene 7	0.334938
<i>CHAC1</i>	ChaC, cation transport regulator homolog 1 (E. coli)	0.332567
<i>FAM227B</i>	family with sequence similarity 227, member B	0.332222
<i>ALDH2</i>	aldehyde dehydrogenase 2 family (mitochondrial)	0.331961
<i>SLC6A9</i>	solute carrier family 6 (neurotransmitter transporter, glycine), member 9	0.330724
<i>MIR186</i>	microRNA 186	0.328755
<i>ZNF763</i>	zinc finger protein 763	0.328666
<i>HIST1H2AJ</i>	histone cluster 1, H2aj	0.327899
<i>RNU5D-1</i>	RNA, U5D small nuclear 1	0.323913
<i>ALOX5AP</i>	arachidonate 5-lipoxygenase-activating protein	0.32156
<i>FBXO32</i>	F-box protein 32	0.32125
<i>ZC3H6</i>	zinc finger CCCH-type containing 6	0.316641
<i>GPT2</i>	glutamic pyruvate transaminase (alanine aminotransferase) 2	0.314711

<i>GENE_SYMBOL</i>	<i>GENE_TITLE</i>	<i>SCORE</i>
<i>COLGALT2</i>	collagen beta(1-O)galactosyltransferase 2	0.314565
<i>WEE2-AS1</i>	WEE2 antisense RNA 1	0.314304
<i>PPM1M</i>	protein phosphatase 1M (PP2C domain containing)	0.313921
<i>SNORA31</i>	small nucleolar RNA, H/ACA box 31	0.313363
<i>C11orf74</i>	chromosome 11 open reading frame 74	0.313307
<i>ZMAT1</i>	zinc finger, matrin type 1	0.313217
<i>RND3</i>	Rho family GTPase 3	0.309211
<i>CLGN</i>	calmegin	0.305442
<i>UGT1A10</i>	UDP glucuronosyltransferase 1 family, polypeptide A10	0.30265
<i>CIR1</i>	corepressor interacting with RBPJ	0.301674
<i>CXorf22</i>	chromosome X open reading frame 22	0.299363
<i>ALDH1L2</i>	aldehyde dehydrogenase 1 family, member L2	0.298943
<i>RBM11</i>	RNA binding motif protein 11	0.298281
<i>SNORD116-13</i>	small nucleolar RNA, C/D box 116-13	0.298082
<i>TTL1</i>	tubulin tyrosine ligase-like family, member 1	0.297329
<i>CEBPB</i>	CCAAT/enhancer binding protein (C/EBP), beta	0.296077
<i>C2orf27A</i>	chromosome 2 open reading frame 27A	0.295393
<i>FAM129A</i>	family with sequence similarity 129, member A	0.286598
<i>KIAA1257</i>	KIAA1257	0.285531
<i>LOC644717</i>	sarcoma antigen 2, pseudogene	0.285277
<i>AGMO</i>	alkylglycerol monooxygenase	0.280998
<i>PSAT1</i>	phosphoserine aminotransferase 1	0.278228
<i>THAP9-AS1</i>	THAP9 antisense RNA 1	0.277759
<i>SMPX</i>	small muscle protein, X-linked	0.277239
<i>COBLL1</i>	COBL-like 1	0.272505
<i>WNT5A</i>	wingless-type MMTV integration site family, member 5A	0.271617
<i>MIR29A</i>	microRNA 29a	0.271519
<i>CBX4</i>	chromobox homolog 4 (Pc class homolog, Drosophila)	0.270723
<i>MT2A</i>	metallothionein 2A	0.267114
<i>ATF3</i>	activating transcription factor 3	0.26646
<i>PDLIM3</i>	PDZ and LIM domain 3	0.265137
<i>LYPLA1</i>	lysophospholipase I	0.263954
<i>PRH2</i>	proline-rich protein HaeIII subfamily 2	0.2622
<i>SMIM14</i>	small integral membrane protein 14	0.261547
<i>SCN4A</i>	sodium channel, voltage-gated, type IV, alpha	0.260615
<i>PEX5L</i>	peroxisomal biogenesis factor 5-like	0.260427
<i>RAB39B</i>	RAB39B, member RAS oncogene family	0.259919
<i>BEST1</i>	bestrophin 1	0.259136
<i>MIR2682</i>	microRNA 2682	0.259071
<i>LOC339803</i>	hypothetical protein LOC339803	0.25466
<i>MAP1LC3B2</i>	microtubule-associated protein 1 light chain 3 beta 2	0.254636
<i>FAAH2</i>	fatty acid amide hydrolase 2	0.254589
<i>RNU5B-1</i>	RNA, U5B small nuclear 1	0.25144
<i>ZNF117</i>	zinc finger protein 117	0.251039
<i>SCARNA9</i>	small Cajal body-specific RNA 9	0.250541
<i>NR5A2</i>	nuclear receptor subfamily 5, group A, member 2	0.249632
<i>CHDC2</i>	calponin homology domain containing 2	0.249293
<i>CCPG1</i>	cell cycle progression 1	0.248701
<i>ZNF674</i>	zinc finger protein 674	0.248269
<i>PIP5KL1</i>	phosphatidylinositol-4-phosphate 5-kinase-like 1	0.248112
<i>INPP1</i>	inositol polyphosphate-1-phosphatase	0.247267
<i>IFRD1</i>	interferon-related developmental regulator 1	0.245181
<i>LYPLAL1</i>	lysophospholipase-like 1	0.245028
<i>KIF21B</i>	kinesin family member 21B	0.244496
<i>LRRC37A3</i>	leucine rich repeat containing 37, member A3	0.244306
<i>TBC1D3P2</i>	TBC1 domain family, member 3 pseudogene 2	0.243629
<i>OPN1LW</i>	opsin 1 (cone pigments), long-wave-sensitive (color blindness, protan)	0.243267

<i>GENE_SYMBOL</i>	<i>GENE_TITLE</i>	<i>SCORE</i>
<i>PCDHA1</i>	protocadherin alpha 1	0.243149
<i>IGHV1-45</i>	immunoglobulin heavy variable 1-45	0.241021
<i>ANKRD31</i>	ankyrin repeat domain 31	0.240202
<i>KIF27</i>	kinesin family member 27	0.239263
<i>OR2M3</i>	olfactory receptor, family 2, subfamily M, member 3	0.238097
<i>MT1X</i>	metallothionein 1X	0.237609
<i>COX7B</i>	cytochrome c oxidase subunit VIIb	0.23747
<i>ZNF429</i>	zinc finger protein 429	0.237151
<i>MBNL2</i>	muscleblind-like 2 (Drosophila)	0.235401
<i>ACSM3</i>	acyl-CoA synthetase medium-chain family member 3	0.2354
<i>CYP19A1</i>	cytochrome P450, family 19, subfamily A, polypeptide 1	0.234959
<i>SLC1A4</i>	solute carrier family 1 (glutamate/neutral amino acid transporter), member 4	0.233497
<i>TP53</i>	tumor protein p53 (Li-Fraumeni syndrome)	0.233082
<i>RGMB</i>	RGM domain family, member B	0.229311
<i>SNORD116-6</i>	small nucleolar RNA, C/D box 116-6	0.226827
<i>ARHGAP10</i>	Rho GTPase activating protein 10	0.226407
<i>MST1</i>	macrophage stimulating 1 (hepatocyte growth factor-like)	0.226056
<i>RFPL4AL1</i>	ret finger protein-like 4A-like 1	0.225059
<i>FAM45B</i>	family with sequence similarity 45, member B	0.224498
<i>ERO1LB</i>	ERO1-like beta (S. cerevisiae)	0.223822
<i>FBXO48</i>	F-box protein 48	0.223482
<i>C5orf28</i>	chromosome 5 open reading frame 28	0.22115
<i>DDR2</i>	discoidin domain receptor family, member 2	0.22074
<i>TNFRSF10B</i>	tumor necrosis factor receptor superfamily, member 10b	0.220335
<i>AMY2B</i>	amylase, alpha 2B (pancreatic)	0.219811
<i>SNORD116-26</i>	small nucleolar RNA, C/D box 116-26	0.217503
<i>TRNASUP1</i>	transfer RNA suppressor 1 (anticodon UUA)	0.217355
<i>PBLD</i>	phenazine biosynthesis-like protein domain containing	0.217007
<i>IGHV1-69</i>	immunoglobulin heavy variable 1-69	0.215382
<i>IRAK1BP1</i>	interleukin-1 receptor-associated kinase 1 binding protein 1	0.214788
<i>ARHGAP9</i>	Rho GTPase activating protein 9	0.214722
<i>PHGDH</i>	phosphoglycerate dehydrogenase	0.213742
<i>PYCR1</i>	pyrroline-5-carboxylate reductase 1	0.213522
<i>SCARNA9L</i>	small Cajal body-specific RNA 9-like	0.213237
<i>DEPTOR</i>	DEP domain containing MTOR-interacting protein	0.212935
<i>BCAT1</i>	branched chain aminotransferase 1, cytosolic	0.212422
<i>SHMT2</i>	serine hydroxymethyltransferase 2 (mitochondrial)	0.211524
<i>ALPK2</i>	alpha-kinase 2	0.211106

Supplementary Table 5B: Description of the 150 genes at the bottom of the ranked list in GSEA analysis of U266 treated versus control cells

<i>GENE_SYMBOL</i>	<i>GENE_TITLE</i>	<i>SCORE</i>
<i>OR4M1</i>	olfactory receptor, family 4, subfamily M, member 1	-0.18006
<i>PRLR</i>	prolactin receptor	-0.18045
<i>FAM19A1</i>	family with sequence similarity 19 (chemokine (C-C motif)-like), member A1	-0.18057
<i>TNFSF13</i>	tumor necrosis factor (ligand) superfamily, member 13	-0.18059
<i>TUBA3E</i>	tubulin, alpha 3e	-0.18091
<i>FOS</i>	v-fos FBJ murine osteosarcoma viral oncogene homolog	-0.181
<i>MIR146A</i>	microRNA 146a	-0.18117
<i>EPGN</i>	epithelial mitogen homolog (mouse)	-0.18137
<i>SNORD27</i>	small nucleolar RNA, C/D box 27	-0.18153
<i>LOC339524</i>	pothetical LOC339524	-0.18153
<i>OR7D4</i>	olfactory receptor, family 7, subfamily D, member 4	-0.18161
<i>IL5RA</i>	interleukin 5 receptor, alpha	-0.18194
<i>SERPINB3</i>	serpin peptidase inhibitor, clade B (ovalbumin), member 3	-0.18204
<i>OR7E5P</i>	olfactory receptor, family 7, subfamily E, member 5 pseudogene	-0.1823
<i>WBP4</i>	WW domain binding protein 4 (formin binding protein 21)	-0.18311
<i>STARD4</i>	START domain containing 4, sterol regulated	-0.18332
<i>VAV3</i>	vav 3 oncogene	-0.18398
<i>S100A10</i>	S100 calcium binding protein A10	-0.18437
<i>SNORD115-32</i>	small nucleolar RNA, C/D box 115-32	-0.18475
<i>MIR101-2</i>	microRNA 101-2	-0.1848
<i>C4BPB</i>	complement component 4 binding protein, beta	-0.18487
<i>CNN2</i>	calponin 2	-0.18517
<i>OR2J3</i>	olfactory receptor, family 2, subfamily J, member 3	-0.18566
<i>MPV17L</i>	MPV17 mitochondrial membrane protein-like	-0.18629
<i>SMPDL3A</i>	sphingomyelin phosphodiesterase, acid-like 3A	-0.18643
<i>ZNF492</i>	zinc finger protein 492	-0.18661
<i>IGHV3-35</i>	immunoglobulin heavy variable 3-35	-0.18676
<i>OR3A1</i>	olfactory receptor, family 3, subfamily A, member 1	-0.18715
<i>CYP2B7P1</i>	cytochrome P450, family 2, subfamily B, polypeptide 7 pseudogene 1	-0.18722
<i>SQLE</i>	squalene epoxidase	-0.1877
<i>CYP4A22</i>	cytochrome P450, family 4, subfamily A, polypeptide 22	-0.18797
<i>PDIA3</i>	protein disulfide isomerase family A, member 3	-0.18818
<i>OR13J1</i>	olfactory receptor, family 13, subfamily J, member 1	-0.18828
<i>SAA2</i>	serum amyloid A2	-0.18866
<i>OR56A3</i>	olfactory receptor, family 56, subfamily A, member 3	-0.18884
<i>GAS6</i>	growth arrest-specific 6	-0.18925
<i>PLN</i>	phospholamban	-0.18931
<i>C1QTNF9B-AS1</i>	C1QTNF9B antisense RNA 1	-0.18973
<i>LAX1</i>	lymphocyte transmembrane adaptor 1	-0.19029
<i>EGFL6</i>	EGF-like-domain, multiple 6	-0.19069
<i>RBM46</i>	RNA binding motif protein 46	-0.19214
<i>RFPL1</i>	ret finger protein-like 1	-0.19246
<i>SLCO4A1</i>	solute carrier organic anion transporter family, member 4A1	-0.19335
<i>BTN2A3P</i>	butyrophilin, subfamily 2, member A3, pseudogene	-0.19382
<i>EFCAB2</i>	EF-hand calcium binding domain 2	-0.19434
<i>GYPB</i>	glycophorin B (MNS blood group)	-0.19494
<i>KRTAP19-8</i>	keratin associated protein 19-8	-0.19593
<i>PBK</i>	PDZ binding kinase	-0.19827
<i>IGHV3-33</i>	immunoglobulin heavy variable 3-33	-0.19842
<i>HSPH1</i>	heat shock 105kDa/110kDa protein 1	-0.19858
<i>MIR377</i>	microRNA 377	-0.19874
<i>PRICKLE1</i>	prickle homolog 1 (Drosophila)	-0.19891
<i>ZNF300P1</i>	zinc finger protein 300 pseudogene 1 (functional)	-0.1992
<i>CDC25A</i>	cell division cycle 25A	-0.20011
<i>POU2F2</i>	POU domain, class 2, transcription factor 2	-0.20045

<i>GENE_SYMBOL</i>	<i>GENE_TITLE</i>	<i>SCORE</i>
<i>SHCBP1</i>	SHC SH2-domain binding protein 1	-0.20052
<i>TRAV8-3</i>	T cell receptor alpha variable 8-3	-0.20052
<i>PSG10P</i>	pregnancy specific beta-1-glycoprotein 10, pseudogene	-0.2013
<i>BCL3</i>	B-cell CLL/lymphoma 3	-0.20196
<i>RPS27A</i>	ribosomal protein S27a	-0.20228
<i>SNORA2A</i>	small nucleolar RNA, H/ACA box 2A	-0.20261
<i>OR1N1</i>	olfactory receptor, family 1, subfamily N, member 1	-0.20264
<i>OR10G3</i>	olfactory receptor, family 10, subfamily G, member 3	-0.20269
<i>PKP2</i>	plakophilin 2	-0.20317
<i>RNU6-76P</i>	RNA, U6 small nuclear 76, pseudogene	-0.20332
<i>PGM2L1</i>	phosphoglucomutase 2-like 1	-0.20358
<i>DTX3L</i>	deltex 3-like (Drosophila)	-0.20393
<i>IFIT1</i>	interferon-induced protein with tetratricopeptide repeats 1	-0.20488
<i>GOLGA8DP</i>	golgin A8 family, member D, pseudogene	-0.20604
<i>NETO1</i>	neuropilin (NRP) and tolloid (TLL)-like 1	-0.20608
<i>ADAM21</i>	ADAM metallopeptidase domain 21	-0.2074
<i>STON1-GTF2A1L</i>	STON1-GTF2A1L readthrough	-0.2077
<i>PTP4A3</i>	protein tyrosine phosphatase type IVA, member 3	-0.20809
<i>MIR17HG</i>	microRNA host gene 1 (non-protein coding)	-0.20824
<i>PCDH11Y</i>	protocadherin 11 Y-linked	-0.2088
<i>FAM172BP</i>	family with sequence similarity 172, member B, pseudogene	-0.21178
<i>HMGCR</i>	3-hydroxy-3-methylglutaryl-Coenzyme A reductase	-0.212
<i>MIR224</i>	microRNA 224	-0.2121
<i>LOC101929115</i>	uncharacterized LOC101929115	-0.21248
<i>TMEM75</i>	transmembrane protein 75	-0.21334
<i>PPT2-EGFL8</i>	PPT2-EGFL8 readthrough (NMD candidate)	-0.21366
<i>SPRED2</i>	sprouty-related, EVH1 domain containing 2	-0.21369
<i>KLRC2</i>	killer cell lectin-like receptor subfamily C, member 2	-0.21392
<i>MIR103A2</i>	microRNA 103a2	-0.21466
<i>LOC730102</i>	hypothetical protein LOC730102	-0.21508
<i>TLR7</i>	toll-like receptor 7	-0.21603
<i>FASN</i>	fatty acid synthase	-0.21608
<i>MSMO1</i>	methylsterol monooxygenase 1	-0.21786
<i>OR2T29</i>	olfactory receptor, family 2, subfamily T, member 29	-0.2184
<i>TMPRSS11B</i>	transmembrane protease, serine 11B	-0.22077
<i>RPL13A</i>	ribosomal protein L13a	-0.22087
<i>EGR1</i>	early growth response 1	-0.22169
<i>C3orf14</i>	chromosome 3 open reading frame 14	-0.22297
<i>SLA</i>	Src-like-adaptor	-0.2233
<i>UGP2</i>	UDP-glucose pyrophosphorylase 2	-0.22551
<i>BCL6</i>	B-cell CLL/lymphoma 6 (zinc finger protein 51)	-0.22575
<i>IGKV4-1</i>	immunoglobulin kappa variable 4-1	-0.22598
<i>E2F2</i>	E2F transcription factor 2	-0.23065
<i>LOC100288637</i>	OTU deubiquitinase 7A pseudogene	-0.23075
<i>KRTAP21-1</i>	keratin associated protein 21-1	-0.23221
<i>SPANXN1</i>	SPANX family, member N1	-0.23226
<i>UGT2B28</i>	UDP glucuronosyltransferase 2 family, polypeptide B28	-0.23258
<i>KRTAP5-4</i>	keratin associated protein 5-4	-0.23337
<i>ACOXL</i>	acyl-Coenzyme A oxidase-like	-0.23394
<i>PLA2G10</i>	phospholipase A2, group X	-0.23428
<i>LDLR</i>	low density lipoprotein receptor (familial hypercholesterolemia)	-0.23852
<i>C9orf153</i>	chromosome 9 open reading frame 153	-0.24111
<i>TPRX1</i>	tetra-peptide repeat homeobox 1	-0.24176
<i>NAIP</i>	NLR family, apoptosis inhibitory protein	-0.24236
<i>AKAP5</i>	A kinase (PRKA) anchor protein 5	-0.24382
<i>HMGCS1</i>	3-hydroxy-3-methylglutaryl-Coenzyme A synthase 1 (soluble)	-0.25189
<i>MYH8</i>	myosin, heavy chain 8, skeletal muscle, perinatal	-0.2531

<i>GENE_SYMBOL</i>	<i>GENE_TITLE</i>	<i>SCORE</i>
<i>PSG1</i>	pregnancy specific beta-1-glycoprotein 1	-0.25377
<i>ZNF718</i>	zinc finger protein 718	-0.26188
<i>MROH7</i>	maestro heat-like repeat family member 7	-0.26352
<i>LBH</i>	limb bud and heart development homolog (mouse)	-0.26527
<i>ID3</i>	inhibitor of DNA binding 3, dominant negative helix-loop-helix protein	-0.26606
<i>PRDM9</i>	PR domain containing 9	-0.2697
<i>DUSP6</i>	dual specificity phosphatase 6	-0.27505
<i>ZFP36</i>	zinc finger protein 36, C3H type, homolog (mouse)	-0.27713
<i>IL6</i>	interleukin 6 (interferon, beta 2)	-0.28043
<i>RNU6-23P</i>	RNA, U6 small nuclear 23, pseudogene	-0.28877
<i>TBC1D3B</i>	TBC1 domain family, member 3B	-0.29116
<i>ETV4</i>	ets variant gene 4 (E1A enhancer binding protein, E1AF)	-0.29659
<i>MYOT</i>	myotilin	-0.29938
<i>OCM2</i>	oncomodulin 2	-0.31006
<i>IDI1</i>	isopentenyl-diphosphate delta isomerase 1	-0.31097
<i>SPRED1</i>	sprouty-related, EVH1 domain containing 1	-0.31603
<i>IFNA7</i>	interferon, alpha 7	-0.31626
<i>MIR142</i>	microRNA 142	-0.31715
<i>DHCR24</i>	24-dehydrocholesterol reductase	-0.31717
<i>PRR23A</i>	proline rich 23A	-0.33145
<i>TDGF1</i>	teratocarcinoma-derived growth factor 1	-0.33705
<i>INSIG1</i>	insulin induced gene 1	-0.34427
<i>GAGE1</i>	G antigen 1	-0.34578
<i>LOC286359</i>	hypothetical LOC286359	-0.35466
<i>ETV5</i>	ets variant gene 5 (ets-related molecule)	-0.38229
<i>IFNA10</i>	interferon, alpha 10	-0.38231
<i>ARL2-SNX15</i>	ARL2-SNX15 readthrough (NMD candidate)	-0.38978
<i>PARP9</i>	poly (ADP-ribose) polymerase family, member 9	-0.39203
<i>ZNF578</i>	zinc finger protein 578	-0.40315
<i>TRIM51</i>	tripartite motif-containing 51	-0.40741
<i>CCR1</i>	chemokine (C-C motif) receptor 1	-0.40874
<i>TRAV5</i>	T cell receptor alpha variable 5	-0.41448
<i>OR13C2</i>	olfactory receptor, family 13, subfamily C, member 2	-0.42899
<i>CD180</i>	CD180 molecule	-0.43024
<i>MIR148A</i>	microRNA 148a	-0.43208
<i>TRAV17</i>	T cell receptor alpha variable 17	-0.46508
<i>VTRNA1-1</i>	vault RNA 1-1	-0.47056
<i>GPRC5D</i>	G protein-coupled receptor, family C, group 5, member D	-0.5071

Supplementary Table 6A: Gene sets enriched in U266 control cells

<i>NAME</i>	<i>NES</i>	<i>FDR q-value</i>
ROSTY_CERVICAL_CANCER_PROLIFERATION_CLUSTER	-2.50567	0
KOBAYASHI_EGFR_SIGNALING_24HR_DN	-2.47026	0
DUTERTRE ESTRADIOL_RESPONSE_24HR_UP	-2.45821	0
CROONQUIST_NRAS_SIGNALING_DN	-2.43549	0
CROONQUIST_IL6_DEPRIVATION_DN	-2.42036	0
GRAHAM_NORMAL QUIESCENT_VS_NORMAL_DIVIDING_DN	-2.41937	0
ZHANG_TLX_TARGETS_UP	-2.41358	0
LEE_EARLY_T_LYMPHOCYTE_UP	-2.39146	0
ZHAN_MULTIPLE_MYELOMA_PR_UP	-2.37026	0
CHANG_CYCLING_GENES	-2.3691	0
SCHMIDT_POR_TARGETS_IN_LIMB_BUD_UP	-2.36034	0
SOTIRIOU_BREAST_CANCER_GRADE_1_VS_3_UP	-2.35916	0
KANG_DOXORUBICIN_RESISTANCE_UP	-2.35159	0
KONG_E2F3_TARGETS	-2.23723	4.46E-05
GOBERT_OLIGODENDROCYTE_DIFFERENTIATION_UP	-2.24137	4.67E-05
MANALO_HYPOXIA_DN	-2.24976	4.89E-05
BENPORATH_ES_2	-2.25888	5.13E-05
REACTOME_CHOLESTEROL_BIOSYNTHESIS	-2.26001	5.40E-05
GSE15750_DAY6_VS_DAY10_EFF_CD8_TCELL_UP	-2.26123	5.70E-05
ISHIDA_E2F_TARGETS	-2.26645	6.04E-05
FURUKAWA_DUSP6_TARGETS_PCI35_DN	-2.31819	6.41E-05
VERNELL_RETINOBLASTOMA_PATHWAY_UP	-2.33317	6.84E-05
ZHOU_CELL_CYCLE_GENES_IN_IR_RESPONSE_24HR	-2.33854	7.33E-05
WINNEPENINCKX_MELANOMA_METASTASIS_UP	-2.22891	8.12E-05
BLUM_RESPONSE_TO_SALIRASIB_DN	-2.23351	8.46E-05
ZHOU_CELL_CYCLE_GENES_IN_IR_RESPONSE_6HR	-2.20068	9.85E-05
HORTON_SREBF_TARGETS	-2.2023	1.02E-04
GOLDRATH_EFF_VS_MEMORY_CD8_TCELL_UP	-2.20359	1.05E-04
ODONNELL_TFRC_TARGETS_DN	-2.21489	1.09E-04
SARRIO_EPITHELIAL_MESENCHYMAL_TRANSITION_UP	-2.21638	1.13E-04
ZHANG_TLX_TARGETS_60HR_DN	-2.22135	1.17E-04
ODONNELL_TARGETS_OF_MYC_AND_TFRC_DN	-2.18752	1.20E-04
WANG_RESPONSE_TO_GSK3_INHIBITOR_SB216763_DN	-2.18812	1.23E-04
GRAHAM_CML_DIVIDING_VS_NORMAL QUIESCENT_UP	-2.18997	1.27E-04
MITSIADES_RESPONSE_TO_APLIDIN_DN	-2.17531	1.75E-04
CHIANG_LIVER_CANCER_SUBCLASS_PROLIFERATION_UP	-2.16301	2.27E-04
KEGG_STEROID_BIOSYNTHESIS	-2.12567	2.63E-04
HOFFMANN_LARGE_TO_SMALL_PRE_BII_LYMPHOCYTE_UP	-2.12669	2.69E-04
BURTON_ADIPOGENESIS_3	-2.14326	2.77E-04
AMUNDSON_GAMMA_RADIATION_RESPONSE	-2.11155	3.49E-04
GSE15750_DAY6_VS_DAY10_TRAF6KO_EFF_CD8_TCELL_UP	-2.11248	3.58E-04
PODAR_RESPONSE_TO_ADAPHOSTIN_DN	-2.10595	4.15E-04
ZHANG_TLX_TARGETS_36HR_DN	-2.10456	4.29E-04
CAFFAREL_RESPONSE_TO_THC_DN	-2.08356	9.54E-04
BENPORATH_PROLIFERATION	-2.08173	9.56E-04
FUJII_YBX1_TARGETS_DN	-2.07129	0.001047
M_PHASE_OF_MITOTIC_CELL_CYCLE	-2.06083	0.001171
GSE30962_PRIMARY_VS_SECONDARY_ACUTE_LCMV_INF_CD8_TCELL_UP	-2.06335	0.001177

<i>NAME</i>	<i>NES</i>	<i>FDR q-value</i>
LY_AGING_OLD_DN	-2.05885	0.001186
KEGG_TERPENOID_BACKBONE_BIOSYNTHESIS	-2.05962	0.001189
YU_MYC_TARGETS_UP	-2.06141	0.001195
TIEN_INTESTINE_PROBIOTICS_24HR_UP	-2.05638	0.001202
MARKEY_RB1_CHRONIC_LOF_UP	-2.0486	0.001329
MITOSIS	-2.04949	0.001335
MARKEY_RB1_ACUTE_LOF_DN	-2.0437	0.001337
EGUCHI_CELL_CYCLE_RB1_TARGETS	-2.0454	0.001342
REACTOME_DNA_REPLICATION	-2.04318	0.001349
PENG_LEUCINE_DEPRIVATION_DN	-2.04199	0.001396
GARGALOVIC_RESPONSE_TO_OXIDIZED_PHOSPHOLIPIDS_TURQUOISE_DN	-2.03984	0.001425
LY_AGING_PREMATURE_DN	-2.03722	0.001453
M_PHASE	-2.0264	0.001719
KAMMINGA_EZH2_TARGETS	-2.02676	0.001731
PENG_Glutamine_DEPRIVATION_DN	-2.0262	0.00174
FINETTI_BREAST_CANCER_KINOME_RED	-2.01831	0.001986
REACTOME_G1_S_SPECIFIC_TRANSCRIPTION	-2.01672	0.002018
GARY_CD5_TARGETS_DN	-2.00731	0.002346
CROONQUIST_NRAS_VS_STROMAL_STIMULATION_DN	-2.00023	0.002594
PID_FOXM1PATHWAY	-2.00028	0.002632
WILCOX_PRESPONSE_TO_ROGESTERONE_UP	-1.99054	0.00315
LIU_IL13_PRIMING_MODEL	-1.98765	0.00334
SMIRNOV_RESPONSE_TO_IR_6HR_DN	-1.98641	0.003409
MISSIAGLIA_REGULATED_BY_METHYLATION_DN	-1.98407	0.003505
MOLENAAR_TARGETS_OF_CCND1_AND_CDK4_DN	-1.98123	0.003668
HORIUCHI_WTAP_TARGETS_DN	-1.9723	0.0042
REACTOME_MITOTIC_M_M_G1_PHASES	-1.97297	0.004201
WHITEFORD_PEDIATRIC_CANCER_MARKERS	-1.9688	0.004428
REGULATION_OF_MITOSIS	-1.96696	0.004504
LI_WILMS_TUMOR_ANAPLASTIC_UP	-1.96585	0.004538
INACTIVATION_OF_MAPK_ACTIVITY	-1.96465	0.004572
AFFAR_YY1_TARGETS_DN	-1.96296	0.004656
GSE10239_NAIVE_VS_DAY4.5_EFF_CD8_TCELL_DN	-1.95866	0.004966
IKEDA_MIR133_TARGETS_UP	-1.9544	0.005281
MORI_LARGE_PRE_BII_LYMPHOCYTE_UP	-1.95038	0.005527
REACTOME_CELL_CYCLE_MITOTIC	-1.94704	0.005724
PUJANA_XPRSS_INT_NETWORK	-1.94754	0.005756
SHEDDEN_LUNG_CANCER_POOR_SURVIVAL_A6	-1.94109	0.006255
REACTOME_MITOTIC_PROMETAPHASE	-1.93893	0.006489
MYELOID_LEUKOCYTE_DIFFERENTIATION	-1.93734	0.006574
PUJANA_BRCA_CENTERED_NETWORK	-1.93737	0.006637
FOURNIER_ACINAR_DEVELOPMENT_LATE_2	-1.9316	0.007106
MUELLER_PLURINET	-1.92509	0.007672
KAUFFMANN_MELANOMA_RELAPSE_UP	-1.92513	0.007756
REICHERT_MITOSIS_LIN9_TARGETS	-1.92548	0.007796
PID_PLK1_PATHWAY	-1.91885	0.008409
TARTE_PLASMA_CELL_VS_PLASMABLAST_DN	-1.91183	0.009402
GSE24634_TEFF_VS_TCONV_DAY7_IN_CULTURE_UP	-1.90901	0.009785
VECCHI_GASTRIC_CANCER_EARLY_UP	-1.90755	0.009959

<i>NAME</i>	<i>NES</i>	<i>FDR q-value</i>
AIYAR_COBRA1_TARGETS_DN	-1.90127	0.010795
CHIARADONNA_NEOPLASTIC_TRANSFORMATION_KRAS_UP	-1.90134	0.010894
REACTOME_GLUCURONIDATION	-1.89377	0.011913
WHITFIELD_CELL_CYCLE_LITERATURE	-1.89411	0.011991
FERREIRA_EWINGS_SARCOMA_UNSTABLE_VS_STABLE_UP	-1.88504	0.013656
CHROMOSOME	-1.87943	0.01473
REACTOME_E2F_MEDIATED_REGULATION_OF_DNA_REPLICATION	-1.87878	0.014736
V\$E2F_Q6_01	-1.87675	0.014958
WU_APOPTOSIS_BY_CDKN1A_VIA_TP53	-1.86938	0.016366
V\$E2F_Q4	-1.86613	0.017122
GSE36476_CTRL_VS_TSST_ACT_72H_MEMORY_CD4_TCELL_YOUNG_DN	-1.86473	0.017343
V\$E2F_Q6	-1.86231	0.017795
JACKSON_DNMT1_TARGETS_UP	-1.86236	0.017958
V\$E2F1_Q3	-1.86101	0.018107
OXFORD_RALA_OR_RALB_TARGETS_UP	-1.85882	0.018468
GAL_LEUKEMIC_STEM_CELL_DN	-1.85771	0.01855
REACTOME_OLFACTORY_SIGNALING_PATHWAY	-1.85671	0.018677
CELL_CYCLE_PHASE	-1.85472	0.019103
GARGALOVIC_RESPONSE_TO_OXIDIZED_PHOSPHOLIPIDS_GREEN_UP	-1.85159	0.019629
MARSON_BOUND_BY_E2F4_UNSTIMULATED	-1.84778	0.020681
PUJANA_BRCA2_PCC_NETWORK	-1.84141	0.022741
MIKKELSEN_PLURIPOTENT_STATE_UP	-1.83779	0.023408
GSE3982_MAC_VS_TH1_DN	-1.83602	0.023412
REN_BOUND_BY_E2F	-1.83629	0.023444
WEST_ADRENOCORTICAL_TUMOR_MARKERS_UP	-1.83803	0.023501
BERENJENO_TRANSFORMED_BY_RHOA_UP	-1.83856	0.023603
ONDER_CDH1_TARGETS_1_DN	-1.83643	0.023619
SGCGSSAAA_V\$E2F1DP2_01	-1.83459	0.023783
SHEPARD_BMYB_TARGETS	-1.83406	0.023806
TURASHVILI_BREAST_DUCTAL_CARCINOMA_VS_LOBULAR_NORMAL_UP	-1.83344	0.023828
GSE24634_TREG_VS_TCONV_POST_DAY7_IL4_CONVERSION_UP	-1.8322	0.024116
SENGUPTA_NASOPHARYNGEAL_CARCINOMA_UP	-1.83052	0.024494
FRASOR_RESPONSE_TO_SERM_OR_FULVESTRANT_DN	-1.82939	0.024668
DAUER_STAT3_TARGETS_DN	-1.82871	0.024769
NADERI_BREAST_CANCER_PROGNOSIS_UP	-1.82703	0.025243
V\$E2F1_Q6	-1.82417	0.025902
CHROMOSOMAL_PART	-1.82366	0.025946
PUJANA_BREAST_CANCER_WITH_BRCA1_MUTATED_UP	-1.82271	0.026028
BURTON_ADIPOGENESIS_PEAK_AT_24HR	-1.82124	0.026395
LU_TUMOR_VASCULATURE_UP	-1.81981	0.026758
ZHAN_MULTIPLE_MYELOMA_CD2_DN	-1.81788	0.027279
V\$E2F1DP1RB_01	-1.81724	0.027341
GSE29614_CTRL_VS_DAY7_TIV_FLU_VACCINE_PBMCD_DN	-1.8156	0.027703
MORI_IMMATURE_B_LYMPHOCYTE_DN	-1.80745	0.030029
NEGATIVE_REGULATION_OF_CYTOKINE_BIOSYNTHETIC_PROCESS	-1.80788	0.030096
BIOCARTA_NKT_PATHWAY	-1.80845	0.030105
MCDOWELL_ACUTE_LUNG_INJURY_UP	-1.80845	0.030319
SISTER_CHROMATID_SEGREGATION	-1.806	0.030501
LOPEZ_MESOTELIOMA_SURVIVAL_TIME_UP	-1.80246	0.031804

<i>NAME</i>	<i>NES</i>	<i>FDR q-value</i>
GSE29614_DAY3_VS_DAY7_TIV_FLU_VACCINE_PBMC_DN	-1.80131	0.031832
GSE22886_UNSTIM_VS_IL2_STIM_NKCELL_DN	-1.80164	0.031902
TANG_SENESCENCE_TP53_TARGETS_DN	-1.79979	0.032238
LY_AGING_MIDDLE_DN	-1.79848	0.03228
V\$E2F4DP1_01	-1.79876	0.032331
GAVIN_FOXP3_TARGETS_CLUSTER_P6	-1.7971	0.032743
RUIZ_TNC_TARGETS_DN	-1.79579	0.033173
NEGATIVE_REGULATION_OF_MAP_KINASE_ACTIVITY	-1.79467	0.033194
BARIS_THYROID_CANCER_DN	-1.79476	0.033356
GSE13485_DAY3_VS_DAY7_YF17D_VACCINE_PBMC_DN	-1.79296	0.033658
WEI_MYCN_TARGETS_WITH_E_BOX	-1.79224	0.033784
CELLULAR_DEFENSE_RESPONSE	-1.79139	0.034031
CASORELLI_ACUTE_PROMYELOCYTIC_LEUKEMIA_DN	-1.78855	0.035011
CELL_CYCLE_CHECKPOINT_GO_0000075	-1.78712	0.03553
CHASSOT_SKIN_WOUND	-1.78383	0.036878
XU_HGF_SIGNALING_NOT_VIA_AKT1_48HR_DN	-1.78179	0.037432
SIMBULAN_UV_RESPONSE_NORMAL_DN	-1.7818	0.037657
BASAKI_YBX1_TARGETS_UP	-1.7801	0.037975
CERIBELLI_PROMOTERS_INACTIVE_AND_BOUND_BY_NFY	-1.77865	0.038379
GSE36476_CTRL_VS_TSST_ACT_40H_MEMORY_CD4_TCELL_YOUNG_DN	-1.77775	0.038655
GSE3982_EFF_MEMORY_CD4_TCELL_VS_TH1_DN	-1.77369	0.039578
PID_AURORA_B_PATHWAY	-1.77544	0.039641
BENPORATH_CYCLING_GENES	-1.77397	0.03966
ROVERSI_GLIOMA_LOH_REGIONS	-1.77476	0.039747
GSE10239_NAIVE_VS_KLRG1INT_EFF_CD8_TCELL_DN	-1.77415	0.039803
MITOTIC_CELL_CYCLE	-1.77165	0.040045
CHROMOSOMEPERICENTRIC_REGION	-1.77209	0.040051
KORKOLA_TERATOMA	-1.77079	0.040263
KEGG_OLFACTORY_TRANSDUCTION	-1.77005	0.04039
SONG_TARGETS_OF_IE86_CMV_PROTEIN	-1.76934	0.040568
GSE30962_ACUTE_VS_CHRONIC_LCMV_SECONDARY_INF_CD8_TCELL_DN	-1.76059	0.044475
GSE1460_INTRATHYMIC_T_PROGENITOR_VS_NAIVE_CD4_TCELL_ADULT_BLOOD_UP	-1.7614	0.044513
V\$E2F_02	-1.7609	0.044558
WONG_ENDMETRIUM_CANCER_UP	-1.75917	0.044947
MITOTIC_SISTER_CHROMATID_SEGREGATION	-1.75507	0.04706
TURASHVILI_BREAST_DUCTAL_CARCINOMA_VS_DUCTAL_NORMAL_UP	-1.75334	0.047908
LU_TUMOR_ENDOTHELIAL_MARKERS_UP	-1.75275	0.048044
KUROZUMI_RESPONSE_TO_ONCOCYTIC_VIRUS_AND_CYCLIC_RGD	-1.75083	0.048362
GCNP_SHH_UP_LATE.V1_UP	-1.75104	0.048485
MALIK_REPRESSED_BY_ESTROGEN	-1.75139	0.048525
REACTOME_PROCESSING_OF_CAPPED_INTRON_CONTAINING_PRE_MRNA	-1.74938	0.048894
V\$E2F1DP1_01	-1.74752	0.049639

Gene sets are ordered according to FDR q -value. NES (normalized enrichment score) is also reported.

Supplementary Table 6B: Gene sets enriched in U266 treated cells

<i>NAME</i>	<i>NES</i>	<i>FDR q-value</i>
GARGALOVIC_RESPONSE_TO_OXIDIZED_PHOSPHOLIPIDS_RED_UP	2.351516	0
REACTOME_AMINO_ACID_SYNTHESIS_AND_INTERCONVERSION_TRANSAMINATION	2.37104	0
KRIGE_RESPONSE_TO_TOSEDOSTAT_6HR_UP	2.459397	0
MTOR_UP.N4.V1_UP	2.516844	0
PENG_LEUCINE_DEPRIVATION_UP	2.638843	0
KRIGE_RESPONSE_TO_TOSEDOSTAT_24HR_UP	2.659443	0
BHATI_G2M_ARREST_BY_2METHOXYESTRADIOL_DN	2.666824	0
TIEN_INTESTINE_PROBIOTICS_24HR_DN	2.67889	0
PODAR_RESPONSE_TO_ADAPHOSTIN_UP	2.694749	0
PACHER_TARGETS_OF_IGF1_AND_IGF2_UP	2.754331	0
BLUM_RESPONSE_TO_SALIRASIB_UP	2.762486	0
ZHAN_MULTIPLE_MYELOMA_CD1_UP	2.7971	0
HELLER_SILENCED_BY_METHYLATION_DN	2.81202	0
ZHAN_MULTIPLE_MYELOMA_CD1_VS_CD2_UP	2.872037	0
KRIGE_AMINO_ACID_DEPRIVATION	2.896582	0
KEGG_GLYCINE_SERINE_AND_THREONINE_METABOLISM	2.273052	3.01E-04
AMUNDSON_RESPONSE_TO_ARSENITE	2.241046	4.81E-04
TERAMOTO_OPN_TARGETS_CLUSTER_7	2.241159	5.09E-04
ONDER_CDH1_TARGETS_1_UP	2.22951	5.58E-04
WANG_HCP_PROSTATE_CANCER	2.19506	0.0012
FORTSCHEGGER_PHF8_TARGETS_UP	2.175743	0.00169
ALK_DN.V1_UP	2.171733	0.001789
KASLER_HDAC7_TARGETS_2_DN	2.157696	0.002089
PRAMOONJAGO_SOX4_TARGETS_UP	2.144733	0.002684
KAN_RESPONSE_TO_ARSENIC_TRIOXIDE	2.126259	0.003574
BMI1_DN_MEL18_DN.V1_UP	2.108819	0.004551
REACTOME_AMINO_ACID_TRANSPORT_ACROSS_THE_PLASMA_MEMBRANE	2.089989	0.006055
NEUTRAL_AMINO_ACID_TRANSMEMBRANE_TRANSPORTER_ACTIVITY	2.079356	0.006766
SCIBETTA_KDM5B_TARGETS_UP	2.075699	0.006995
XU_HGF_SIGNALING_NOT_VIA_AKT1_48HR_UP	2.064639	0.007916
IGARASHI_ATF4_TARGETS_DN	2.061486	0.008095
CONCANNON_APOPTOSIS_BY_EPOXOMICIN_UP	2.047539	0.009946
CHO_NR4A1_TARGETS	2.041657	0.010372
PARK_OSTEOBLAST_DIFFERENTIATION_BY_PHENYLAMIL_UP	2.019906	0.013799
NOJIMA_SFRP2_TARGETS_UP	2.008785	0.015458
HORIUCHI_WTAP_TARGETS_UP	2.009873	0.015707
SMITH_TERT_TARGETS_UP	2.005849	0.015714
FERRARI_RESPONSE_TO_FENRETINIDE_UP	1.992631	0.018693
MAHADEVAN_RESPONSE_TO_MP470_DN	1.964147	0.027304
HANN_RESISTANCE_TO_BCL2_INHIBITOR_DN	1.954325	0.030298
HUANG_FOXA2_TARGETS_UP	1.947281	0.032153
BOYALT_LIVER_CANCER_SUBCLASS_G123_DN	1.948409	0.032375
KEGG_SELENOAMINO_ACID_METABOLISM	1.941641	0.033749
WANG_RESPONSE_TO_GSK3_INHIBITOR_SB216763_UP	1.939409	0.033834
GSE24634_IL4_VS_CTRL_TREATED_NAIVE_CD4_TCELL_DAY7_DN	1.932102	0.036692
HELLER_HDAC_TARGETS_SILENCED_BY_METHYLATION_DN	1.92893	0.037313
SASSON_FSH_RESPONSE	1.904675	0.04931

Gene sets are ordered according to FDR q -value. NES (normalized enrichment score) is also reported.

Supplementary Table 7: Clinical details of the 167 patients analyzed by NGS

Sample	Sex	Age	Disease	Stage ^Δ	Phase [‡]	PP [‡]	t(4;14)	t(11;14)	t(14;16)	t(14;20)	del(13)*	del(17p)*	1q gain*	1p loss*	HD [°]
MM-004	F	58	MM	IA	D	Gκ	-	-	+	-	-	na	na	na	na
MM-015	M	71	MM	IIA	D	Gκ	-	+	-	-	-	-	-	-	-
MM-016	M	66	MM	IIIB	D	Gκ	-	-	-	-	-	-	+	-	+
MM-026	F	72	MM	IIIB	D	κ	-	+	-	-	-	-	-	na	na
MM-027	M	60	MM	IA	D	Gκ	-	-	-	-	+	-	na	na	na
MM-030	M	69	MM	IIIA	D	Gλ	-	-	-	-	-	-	-	+	+
MM-031	M	58	MM	IIIA	D	Aκ	-	+	-	-	-	-	+	+	-
MM-034	M	71	MM	IA	D	Gκ	-	-	-	-	-	-	-	-	+
MM-036	M	65	MM	IIA	D	Gκ	-	-	-	-	+	-	+	na	-
MM-037	F	50	MM	IIA	D	Gκ+Aκ	-	+	-	-	+	-	-	-	-
MM-038	F	67	MM	IIA	D	κ	-	-	-	-	+	-	-	-	+
MM-039	M	50	MM	IIA	D	Gλ	-	-	-	-	-	-	-	na	+
MM-042	M	54	MM	IIIA	D	Aλ	+	-	-	-	+	-	+	-	+
MM-043	F	73	MM	IA	D	Gκ+Gλ	-	-	-	-	-	-	+	-	-
MM-049	M	62	MM	IIIB	D	κ	-	-	-	-	-	-	-	-	+
MM-055	F	69	MM	IIIA	D	Gκ	-	+	-	-	-	-	-	na	-
MM-066	F	77	MM	IIIA	D	Aκ	+	-	-	-	-	-	-	na	-
MM-069	M	63	MM	IIA	D	Gκ	-	-	-	+	+	-	+	na	-
MM-078	F	59	MM	IIIA	D	κ	-	-	-	-	+	-	-	na	+
MM-079	F	74	MM	IIA	D	Gκ+Gλ	-	-	-	-	-	-	-	-	+
MM-087	F	84	MM	IIIA	D	Gλ	+	-	-	-	+	-	+	na	-
MM-115	F	53	MM	IIIA	D	Gλ	-	+	-	-	+	-	na	na	-
MM-123	M	55	MM	IIIA	D	Gκ	+	-	-	-	+	-	+	-	-
MM-131	M	73	MM	IA	D	Gκ	-	-	-	-	-	-	-	-	-
MM-140	M	62	MM	IIIB	D	Gκ	-	+	-	-	-	-	-	na	-
MM-143	M	61	MM	IIA	D	Gκ	-	-	-	-	-	-	+	-	+
MM-146	M	68	MM	IIA	D	Gλ	-	-	-	-	-	-	+	na	+
MM-148	F	55	MM	IA	D	Aκ	-	-	-	-	-	+	+	na	+
MM-149	F	52	MM	IA	D	Gλ	-	-	-	-	-	-	-	-	+
MM-150	F	68	MM	IIA	D	Gλ	-	-	-	-	-	-	+	-	+
MM-151	F	71	MM	IA	D	Gλ	-	-	-	-	-	-	-	-	+
MM-152	M	66	MM	IA	D	Gκ	-	-	-	-	-	-	-	na	+

Sample	Sex	Age	Disease	Stage ^Δ	Phase [‡]	PP [‡]	t(4;14)	t(11;14)	t(14;16)	t(14;20)	del(13)*	del(17p)*	1q gain*	1p loss*	HD [°]
MM-154	F	71	MM	IIA	D	Gκ	-	-	+	-	+	-	+	-	-
MM-159	M	56	MM	IIA	D	κ	-	+	-	-	+	-	-	-	-
MM-174	M	85	MM	IIA	D	Aκ	-	-	-	-	-	-	-	-	+
MM-177	M	73	MM	IIIA	D	Gκ	-	-	-	-	+	-	+	-	-
MM-179	M	50	MM	IIIA	D	Gλ	-	+	-	-	-	+	-	-	+
MM-195	M	62	MM	IIA	D	Gκ	+	-	-	-	+	-	-	-	-
MM-200	F	63	MM	IA	D	Aλ	-	-	-	-	-	-	-	-	+
MM-202	M	64	MM	IIIA	D	Gκ	-	-	-	-	-	-	+	na	+
MM-206	F	73	MM	IIA	D	Gκ	+	-	-	-	+	-	-	+	-
MM-207	F	68	MM	IA	D	Aκ	-	-	-	-	+	-	+	-	-
MM-208	M	74	MM	IIA	D	Aλ	+	-	-	-	-	-	-	-	-
MM-209	F	65	MM	IIA	D	Gλ	-	-	-	-	-	-	-	-	+
MM-210	M	65	MM	IIB	D	Aλ	-	-	-	-	-	-	-	-	-
MM-212	F	55	MM	IIIA	D	Gκ	-	+	-	-	-	-	-	-	-
MM-213	M	66	MM	IIIA	D	λ	-	+	-	-	+	-	-	-	-
MM-219	M	73	MM	IIIA	D	Aλ	-	-	-	-	+	-	-	-	+
MM-224	F	52	MM	IIIA	D	Gκ	-	-	+	-	-	-	+	-	-
MM-229	M	75	MM	IIA	D	Gκ	-	-	-	-	-	-	-	-	-
MM-238	M	58	MM	IIB	D	Gκ	-	-	-	-	+	-	+	-	-
MM-239	F	72	MM	IIA	D	Aκ	-	-	-	-	+	-	-	-	+
MM-240	M	70	MM	IA	D	Gλ	-	-	-	-	-	-	-	-	+
MM-241	M	54	MM	IIA	D	Gκ	-	-	-	-	-	-	-	-	+
MM-242	M	69	MM	IIIA	D	Aκ	-	-	-	-	+	-	-	-	+
MM-243	F	68	MM	IIA	D	Gκ	-	-	-	-	-	-	+	-	+
MM-246	M	71	MM	IIIA	D	Aλ	-	+	-	-	-	-	-	-	-
MM-252	F	77	MM	IIA	D	Aλ	-	+	-	-	-	-	+	-	-
MM-253	F	45	MM	IIA	D	λ	-	-	-	-	+	-	+	-	+
MM-256	M	58	MM	IIIA	D	Aκ	-	-	+	-	+	-	-	-	-
MM-261	F	66	MM	IIB	D	Gκ	-	-	-	-	+	-	+	-	+
MM-262	M	74	MM	IA	D	λ	-	-	-	-	-	-	+	-	-
MM-263	F	65	MM	IIB	D	κ	+	-	-	-	+	-	+	-	-
MM-267	M	74	MM	IIIA	D	Gλ	-	-	-	-	+	-	-	-	+
MM-268	M	77	MM	IIIB	D	Gκ	-	-	-	-	-	-	-	-	-

Sample	Sex	Age	Disease	Stage ^Δ	Phase [‡]	PP [‡]	t(4;14)	t(11;14)	t(14;16)	t(14;20)	del(13)*	del(17p)*	1q gain*	1p loss*	HD [°]
MM-269	F	67	MM	IIA	D	Gκ	-	-	-	-	+	-	-	-	+
MM-271	M	76	MM	IA	D	Gκ	-	-	-	-	+	-	-	-	-
MM-274	M	59	MM	IIA	D	Gκ	+	-	-	-	-	-	-	-	-
MM-276	F	70	MM	IIB	D	Gκ	+	-	-	-	+	-	-	-	-
MM-278	M	73	MM	IIA	D	Gλ	-	-	-	-	-	-	-	+	+
MM-279	F	71	MM	IIIA	D	Gλ	-	-	-	na	+	-	+	-	na
MM-280	M	62	MM	IIIA	D	Gλ	-	+	-	-	-	-	-	-	-
MM-281	F	77	MM	IIA	D	Gκ	-	-	-	-	-	-	-	-	-
MM-282	M	66	MM	IIIA	D	Gκ	-	-	-	-	-	-	+	-	+
MM-284	M	49	MM	IIIA	D	κ	-	+	-	-	na	-	-	-	-
MM-286	F	71	MM	IIA	D	Gκ	-	-	-	-	+	-	-	-	-
MM-295	F	74	MM	IIIA	D	λ	-	-	-	-	+	-	na	na	na
MM-300	M	65	MM	IIB	D	Gκ	-	-	-	-	-	-	na	na	na
MM-301	F	72	MM	IIA	D	Aλ	-	-	-	-	-	-	+	-	+
MM-302	M	65	MM	IA	D	Aκ	-	-	-	-	-	-	+	-	+
MM-308	M	58	MM	IA	D	Aκ	-	-	-	-	-	-	-	-	-
MM-313	M	66	MM	IIIA	D	Gλ	-	+	-	-	-	-	-	-	-
MM-310	M	67	MM	IIIA	D	Gλ	-	+	-	-	-	-	-	-	-
MM-314	F	70	MM	IIIA	D	Aλ	-	+	-	-	-	-	+	-	-
MM-317	M	56	MM	IIA	D	Gλ	-	+	-	-	-	-	+	-	-
MM-321	M	63	MM	IIIA	D	Gκ	-	-	-	-	+	-	-	-	+
MM-327	F	62	MM	IIA	D	absent	-	-	-	-	+	-	+	-	+
MM-330	F	61	MM	IIIA	D	Aκ	+	-	-	-	+	-	+	-	-
MM-334	F	45	MM	IA	D	Gλ	-	-	-	-	+	+	+	+	+
MM-335	F	68	MM	IIA	D	Aκ	-	-	+	-	+	-	-	-	-
MM-340	M	46	MM	IIIA	D	Gλ	-	+	-	-	+	-	-	-	-
MM-341	M	65	MM	IIIA	D	Gκ	-	-	-	-	-	-	+	-	-
MM-343	M	74	MM	IIIA	D	Aλ	-	+	-	-	-	-	-	-	-
MM-351	M	na	MM	IIA	D	Aκ	-	-	-	-	-	-	-	-	-
MM-362	F	80	MM	IIA	D	Gλ	-	-	-	-	+	+	+	-	-
MM-372	M	54	MM	IA	D	Gκ	-	+	-	-	-	-	+	-	-
MM-375	F	78	MM	IA	D	Gκ	+	-	-	-	-	+	-	-	-
MM-381	F	69	MM	IA	D	Gκ	-	-	-	na	-	-	-	-	na

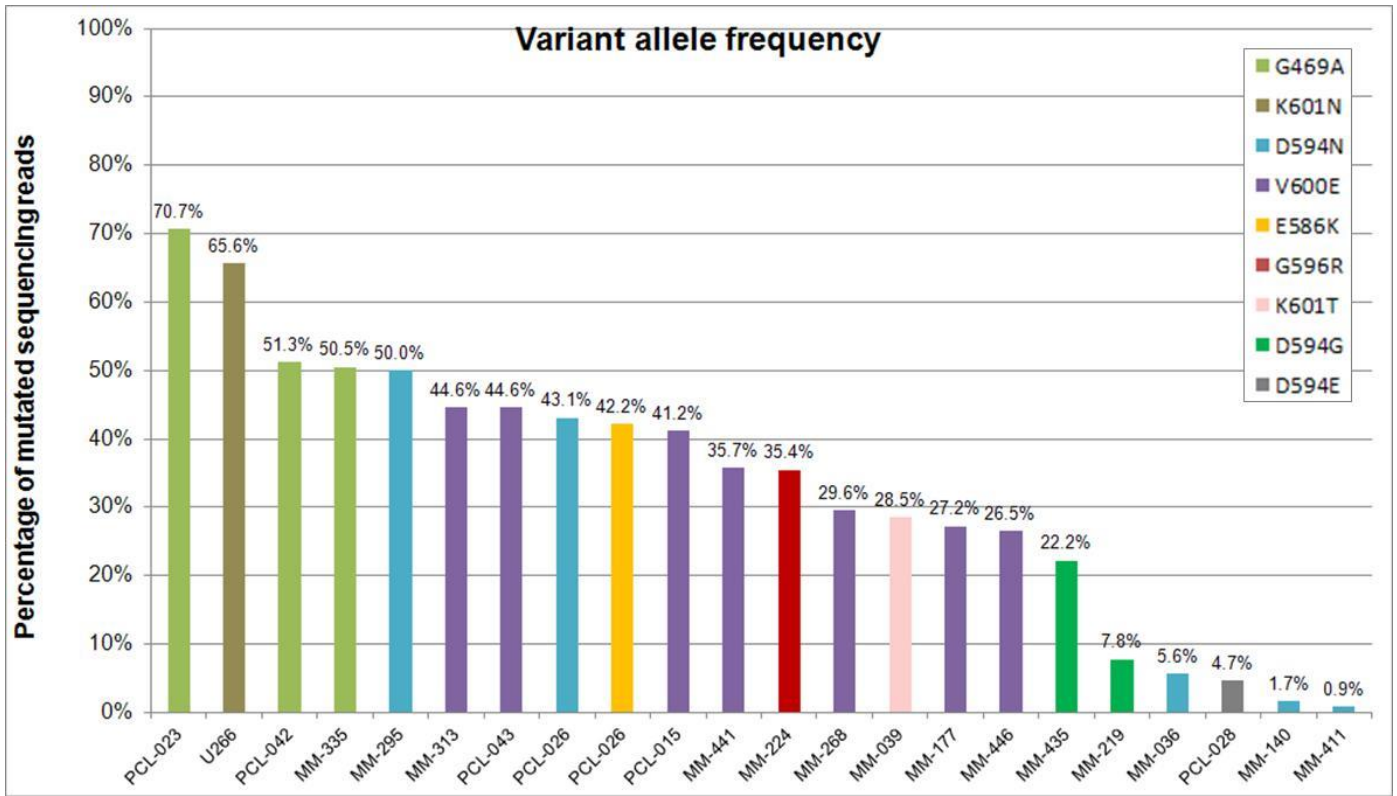
Sample	Sex	Age	Disease	Stage ^Δ	Phase [‡]	PP [‡]	t(4;14)	t(11;14)	t(14;16)	t(14;20)	del(13)*	del(17p)*	1q gain*	1p loss*	HD [°]
MM-382	M	85	MM	IIA	D	Gκ	-	-	-	-	+	+	na	na	na
MM-385	F	76	MM	IIIA	D	Gλ	-	-	-	-	-	-	-	-	+
MM-386	F	76	MM	IIA	D	Aλ	-	na	na	na	-	-	na	na	na
MM-387	F	44	MM	IIIA	D	Gκ	-	-	-	-	-	-	+	-	+
MM-392	F	53	MM	IA	D	Gλ	-	-	-	-	+	-	+	-	+
MM-398	F	65	MM	IIA	D	Gκ	-	+	-	-	+	-	+	-	-
MM-402	M	67	MM	IIA	D	Gκ	-	+	-	-	+	-	-	-	-
MM-405	M	69	MM	IA	D	Gκ	-	+	-	-	-	-	-	-	-
MM-406	M	62	MM	IIB	D	Aκ	-	+	-	-	-	-	+	-	na
MM-407	F	79	MM	IIIA	D	λ	-	-	-	-	+	-	-	-	-
MM-410	F	79	MM	IIIA	D	λ	-	-	-	-	-	-	+	-	-
MM-411	M	67	MM	IIIA	D	Gλ	-	-	-	-	-	-	-	-	+
MM-413	F	51	MM	IIA	D	Gκ	-	-	-	-	+	-	+	-	+
MM-414	F	67	MM	IIIA	D	Gλ	-	+	-	-	-	-	-	-	-
MM-422	F	74	MM	IIB	D	κ	-	-	-	-	+	-	-	-	-
MM-423	F	53	MM	IIA	D	λ	+	-	-	-	+	-	-	-	-
MM-424	F	84	MM	IA	D	Gκ	-	-	-	-	+	-	-	+	+
MM-425	F	65	MM	IIA	D	Aλ	-	-	-	-	+	-	+	-	-
MM-428	F	65	MM	IIA	D	Aλ	-	-	-	-	-	-	+	+	-
MM-429	F	63	MM	IIIA	D	Gκ	-	-	-	-	-	-	+	-	+
MM-430	na	62	MM	IIIA	D	Gλ	-	-	-	-	-	-	-	-	-
MM-431	F	70	MM	IA	D	Gλ	+	-	-	-	+	-	+	-	-
MM-433	M	72	MM	IIB	D	κ	-	+	-	-	-	-	-	-	-
MM-434	F	53	MM	IIA	D	κ	-	-	-	-	+	-	-	+	-
MM-435	na	42	MM	IIIA	D	Gλ	-	-	-	-	-	-	-	-	+
MM-437	F	72	MM	IIB	D	λ	-	-	-	-	+	-	+	-	-
MM-440	na	61	MM	IIA	D	Aκ	-	-	-	-	-	-	-	+	+
MM-441	na	na	MM	IA	D	Gλ	-	-	-	-	-	-	-	-	-
MM-442	F	65	sPCL	/	D	Gκ	-	-	-	-	+	-	+	+	-
MM-445	na	65	MM	IIA	D	κ	-	+	-	-	+	-	+	-	-
MM-446	na	52	MM	IIIA	D	Gλ	-	-	-	-	-	-	+	-	+
MM-447	na	57	MM	IIA	D	Gκ	-	+	-	-	-	-	+	-	-
MM-448	na	59	MM	IIA	D	Aκ	-	-	-	-	+	-	+	-	-

Sample	Sex	Age	Disease	Stage ^Δ	Phase [‡]	PP [‡]	t(4;14)	t(11;14)	t(14;16)	t(14;20)	del(13)*	del(17p)*	1q gain*	1p loss*	HD [°]
MM-449	na	54	MM	IIA	D	Gκ	-	-	-	-	+	-	-	+	-
MM-464	M	75	MM	IIIA	D	Gλ	-	-	-	-	+	-	+	-	-
PCL-001	F	51	pPCL	/	D	κ	-	-	-	+	+	-	na	na	na
PCL-002	F	69	sPCL	/	R	λ	-	+	-	-	+	na	-	na	na
PCL-004	M	72	pPCL	/	D	Gκ	-	-	+	-	-	-	-	-	-
PCL-005	M	76	sPCL	/	R	Aκ	-	+	-	-	-	-	-	-	-
PCL-007	M	65	sPCL	/	R	κ	-	-	-	-	+	+	+	na	na
PCL-008	M	57	pPCL	/	D	Gλ	-	-	+	-	+	-	-	na	na
PCL-009	F	77	sPCL	/	R	Gκ	+	-	-	-	+	-	+	-	-
PCL-011	M	76	sPCL	/	R	Gκ	-	-	+	-	+	+	+	-	-
PCL-012	F	62	sPCL	/	R	Aκ	-	-	+	-	-	+	-	-	-
PCL-014	M	72	pPCL	/	D	λ	-	+	-	-	+	+	-	+	-
PCL-015	M	78	pPCL	/	D	κ	-	-	+	-	+	-	+	-	-
PCL-016	F	57	pPCL	/	D	Gκ	-	-	+	-	+	-	+	+	-
PCL-017	F	68	pPCL	/	D	Gκ	-	-	+	-	+	+	+	+	-
PCL-018	F	59	pPCL	/	D	κ	-	+	-	-	+	+	-	-	-
PCL-019	F	67	pPCL	/	D	Mκ	-	-	+	-	+	+	+	+	-
PCL-020	F	79	pPCL	/	D	Gλ	-	-	-	-	-	-	+	-	-
PCL-021	M	48	pPCL	/	D	Gλ	+	-	-	-	+	-	+	-	-
PCL-023	M	60	pPCL	/	D	Gκ	-	-	+	-	+	+	+	+	-
PCL-026	F	59	pPCL	/	D	Gκ	-	-	+	-	+	-	+	-	-
PCL-027	M	65	pPCL	/	D	λ	-	+	-	-	-	+	-	-	-
PCL-028	F	57	pPCL	/	D	κ	-	+	-	-	+	-	-	-	-
PCL-029	M	51	pPCL	/	D	Aλ	-	-	+	-	-	-	-	-	-
PCL-030	F	52	pPCL	/	D	κ	-	-	-	+	+	+	-	-	-
PCL-031	F	59	sPCL	/	D	Gλ	-	-	+	-	-	+	+	-	-
PCL-032	M	51	pPCL	/	D	Gκ	+	-	-	-	+	-	+	-	-
PCL-035	F	76	pPCL	/	D	κ	-	+	-	-	-	-	-	-	-
PCL-036	M	72	pPCL	/	D	Gκ	-	+	-	-	+	-	-	-	-
PCL-037	M	72	pPCL	/	D	Aλ	-	+	-	-	+	-	-	-	-
PCL-038	M	57	pPCL	/	D	Gκ	-	-	-	-	+	-	+	+	+
PCL-039	M	54	sPCL	/	D	Aκ	-	-	-	-	-	+	-	+	-
PCL-041	M	na	sPCL	/	D	Gκ	-	na	na	na	-	-	na	na	na

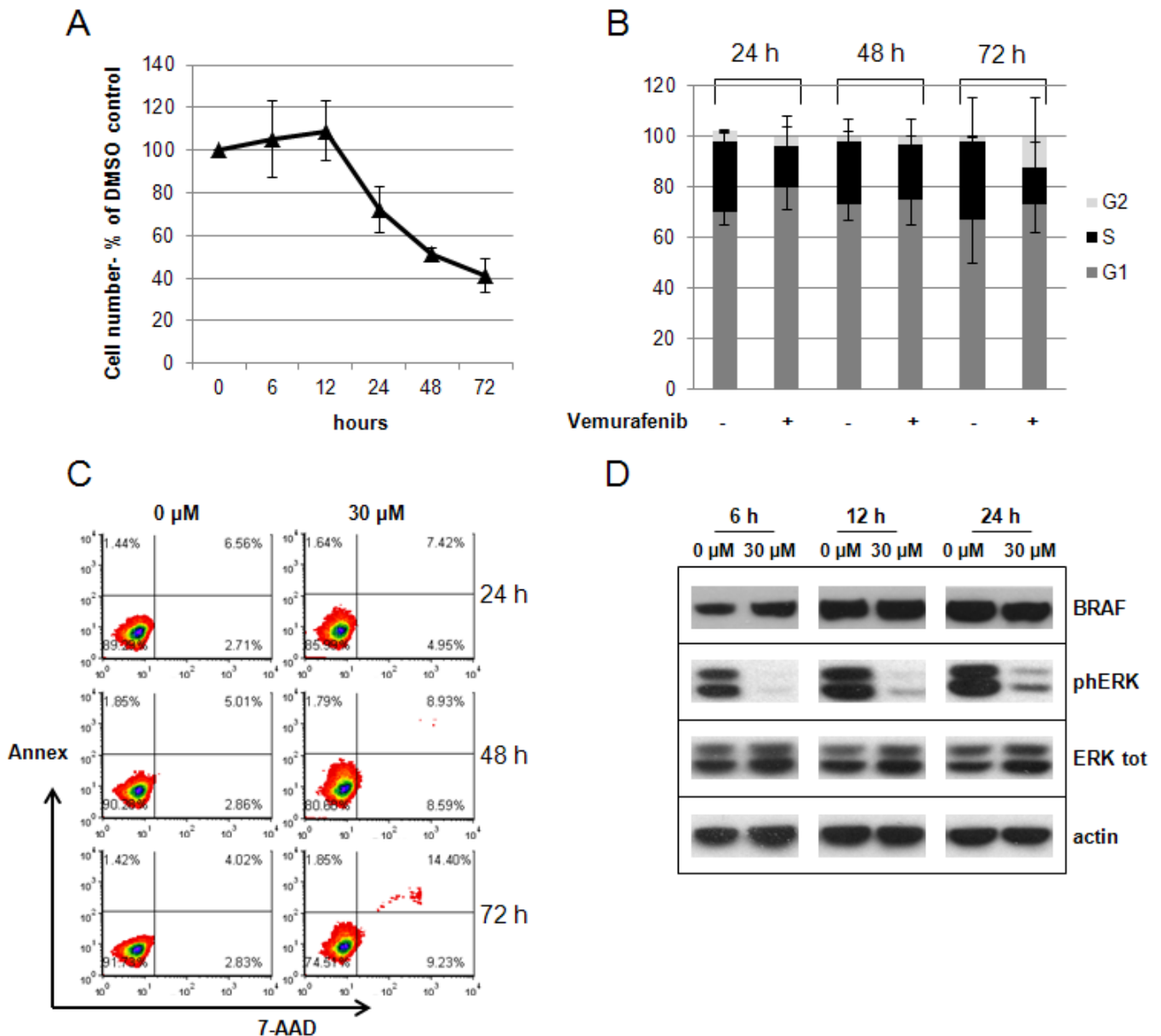
<i>Sample</i>	<i>Sex</i>	<i>Age</i>	<i>Disease</i>	<i>Stage</i> ^Δ	<i>Phase</i> [‡]	<i>PP</i> [†]	<i>t(4;14)</i>	<i>t(11;14)</i>	<i>t(14;16)</i>	<i>t(14;20)</i>	<i>del(13)*</i>	<i>del(17p)*</i>	<i>1q gain*</i>	<i>1p loss*</i>	<i>HD</i> [°]
PCL-042	F	69	sPCL	/	D	Gλ	-	+	-	-	-	-	-	-	-
PCL-043	F	68	pPCL	/	D	Gλ	-	+	-	-	-	-	-	-	-
PCL-046	F	50	pPCL	/	D	K	-	-	-	-	+	-	+	-	+

^ΔThe Durie clinical staging system was adopted; [‡]D: Diagnosis; R: relapse; [†]Paraprotein; *del(13), del(17), 1p loss and 1q gain were determined by FISH.

[°]HD = presence of the hyperdiploid status on the basis of FISH evaluation criteria.



Supplementary Figure 1: Percentage of variant *BRAF* sequencing reads on total sequencing reads for the nine non-synonymous somatic mutations identified by NGS analysis. Horizontal axis: patients are ordered according to decreasing mutation load. Each color represents a different mutation (reported in the legend).

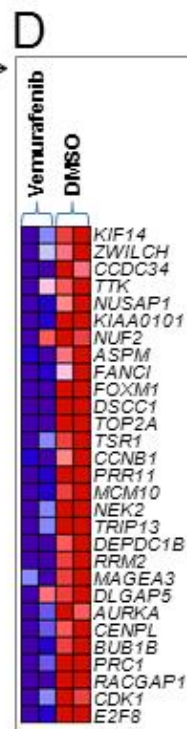
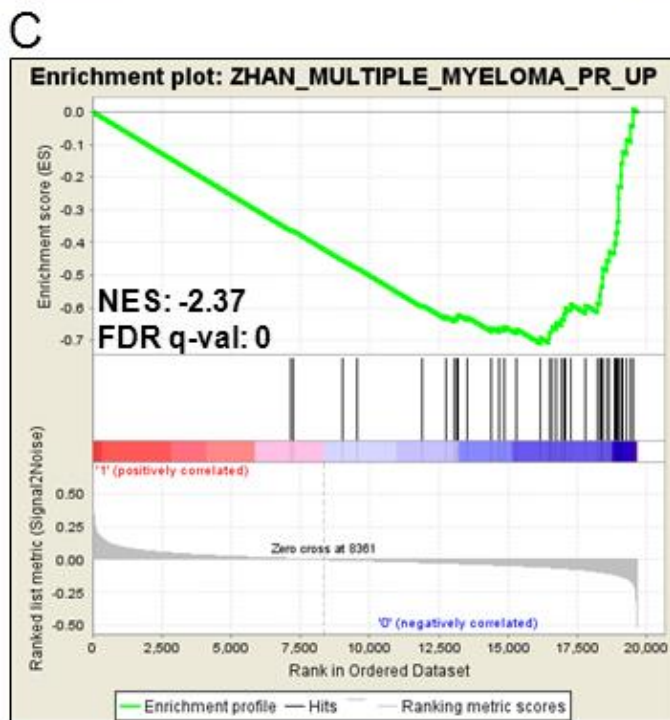
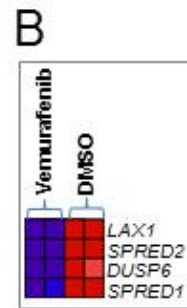
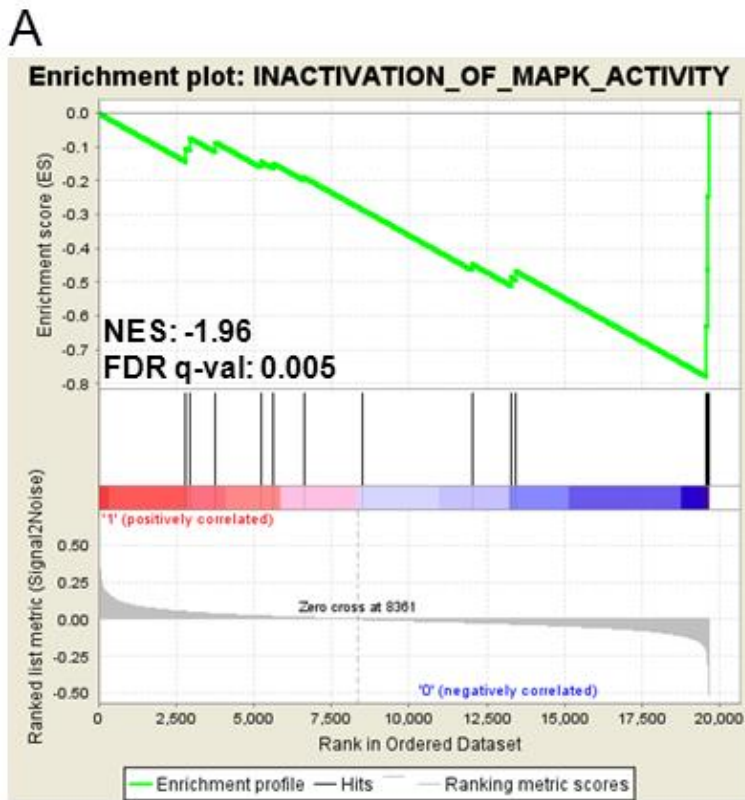


Supplementary Figure 2: (A) Effect of Vemurafenib on the growth of U266 cells assayed by the trypan blue dye exclusion test. Cells at a density of 0.5×10^6 /ml were cultured for 72 h with Vemurafenib at the concentration of 30 μ M. The data are presented as the mean \pm s.e. of three independent experiments.

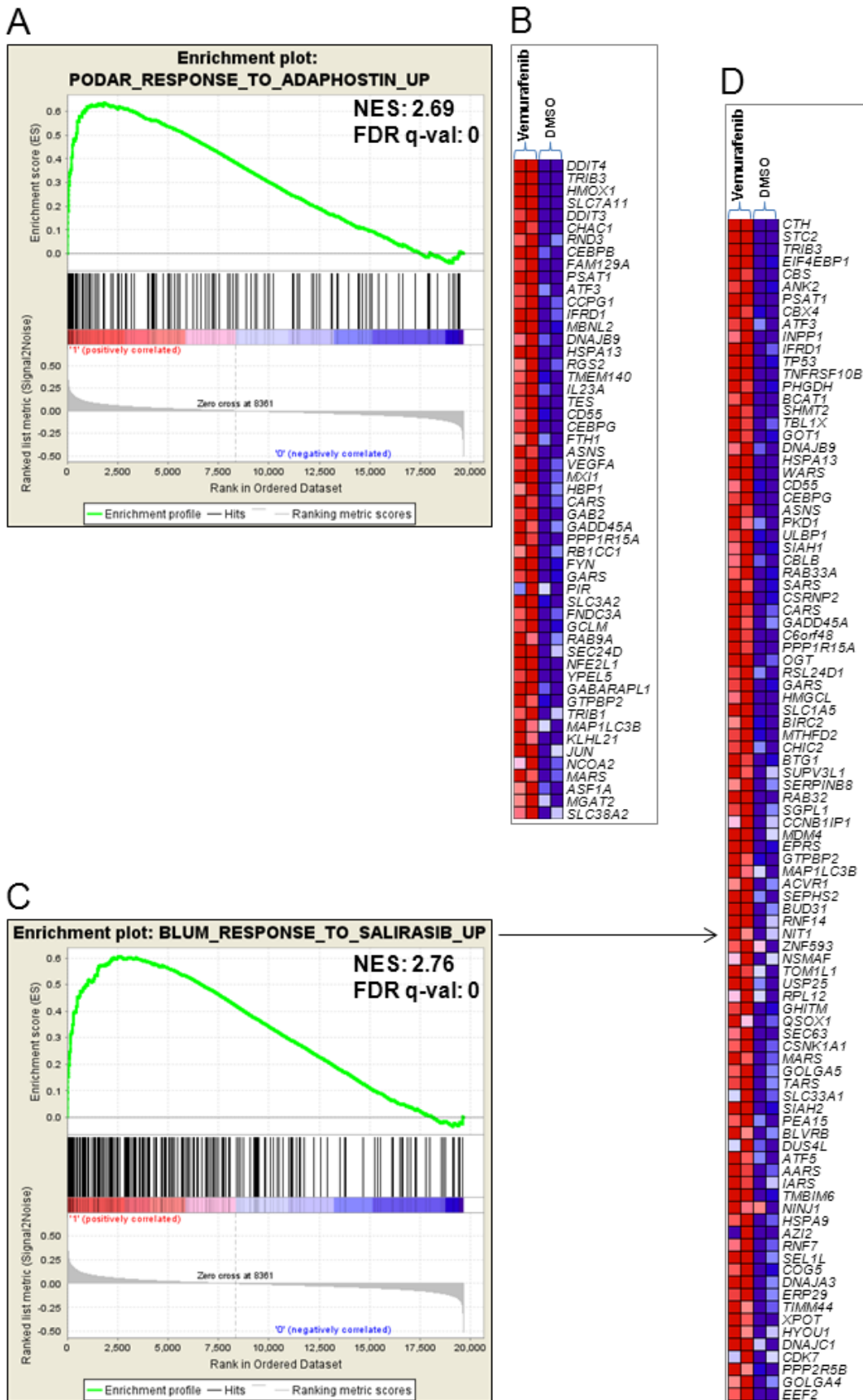
(B) Bar graph representation of cell cycle of U266 cells treated with DMSO or with 30 μ M Vemurafenib for 24, 48, and 72 hours, including standard deviations. Dark grey are cells in G1 phase, black are cells in S phase, light grey cells in G2 phase. The data are presented as the mean \pm s.e. of three independent experiments.

(C) Effect of Vemurafenib on the induction of apoptosis in U266 cells. Cells were cultured with Vemurafenib at the concentration of 30 μ M. Cells were stained with FITC-annexin V (AN) and 7-amino-actinomycin D (7AAD) at each time point, and analyzed by flow cytometry. Represented data are from one experiment representative of two independent experiments.

(D) Western blot analysis of U266 after 6-24 h of treatment with/without 30 μ M Vemurafenib. Cell lysates were analyzed for the expression of BRAF, phospho-ERK1/2 (Thr 202/Tyr 204), and total ERK1/2.



Supplementary Figure 3: (A, C) GSEA enrichment plots of selected gene sets down-regulated in U266 cells treated with Vemurafenib with respect to DMSO controls. The green curves show the enrichment score and reflect the degree to which each gene (black vertical lines) is represented at the bottom of the ranked gene list. Gene sets were considered enriched with an FDR <0.05. Normalized enrichment score (NES) and FDR are shown for both represented gene sets. (B, D) Heat-maps in U266 treated and control cells of the genes constituting the leading edge subsets within the gene sets depicted in panels (A) and (C), respectively.



Supplementary Figure 4: (A, C) GSEA enrichment plots of selected gene sets up-regulated in U266 cells treated with Vemurafenib with respect to DMSO controls. The green curves show the enrichment score and reflect the degree to which each gene (black vertical lines) is represented at the top of the ranked gene list. Gene sets were considered enriched with an FDR <0.05. Normalized enrichment score (NES) and FDR are shown for both

represented gene sets. **(B, D) Heat-maps in U266 treated and control cells of the genes constituting the leading edge subsets within the gene sets depicted in panels (a) and (c), respectively.**