

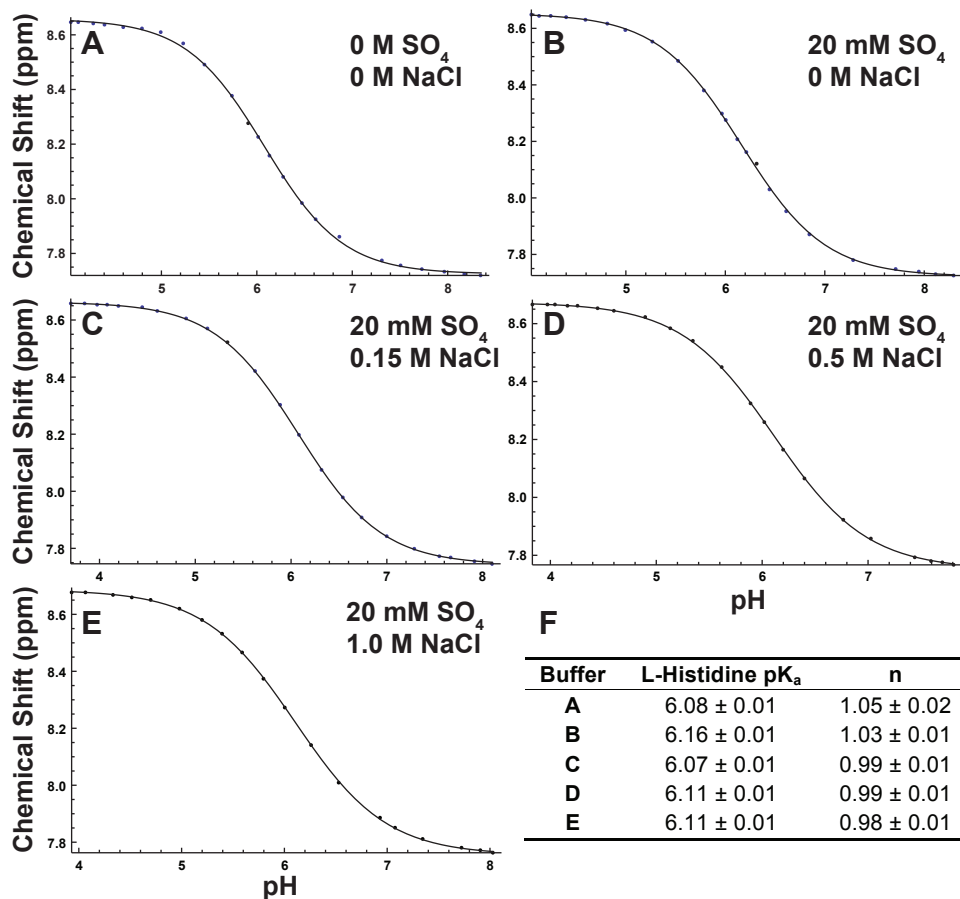
**Electrostatic Energetics of *Bacillus subtilis* Ribonuclease P Protein Determined by Nuclear Magnetic Resonance-Based Histidine pK<sub>a</sub> Measurements**

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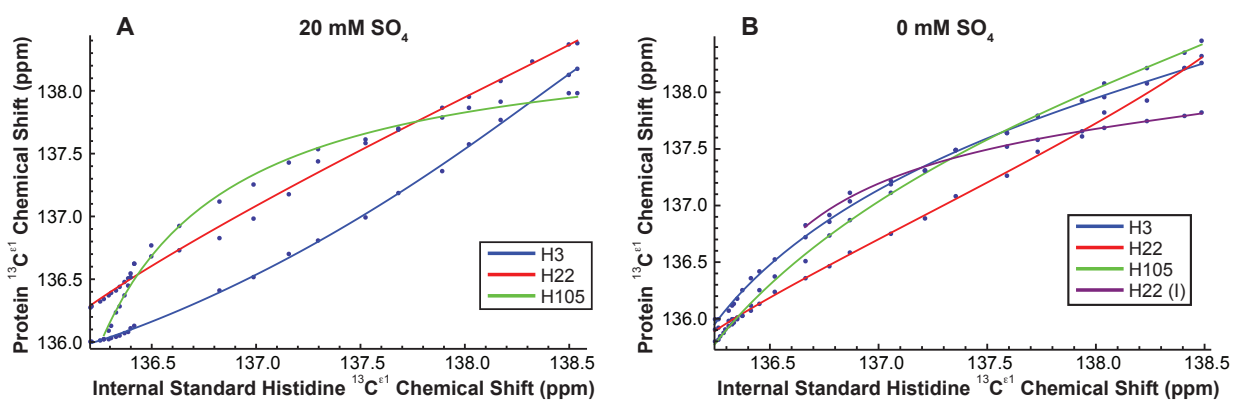
**Supporting Information**



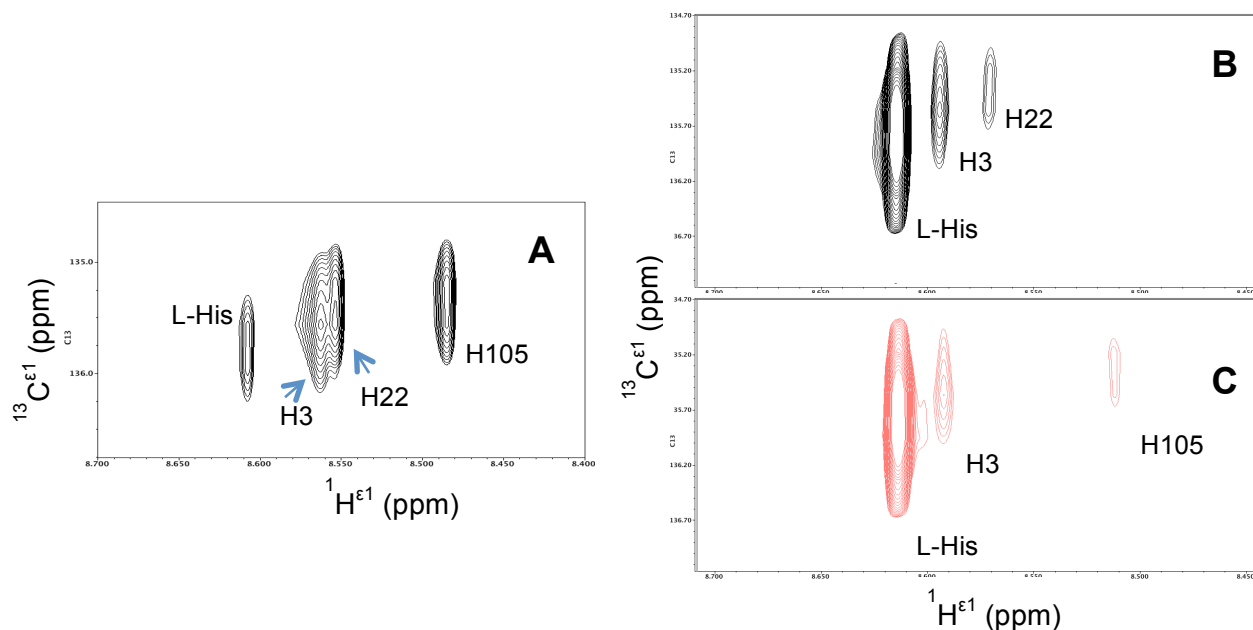
**Figure S1:** Titration of NMR sample with a 0.5–0.4  $\mu\text{l}$  aliquot of NaOH using a 5.0  $\mu\text{l}$  positive displacement syringe extended with polyethylene tubing.



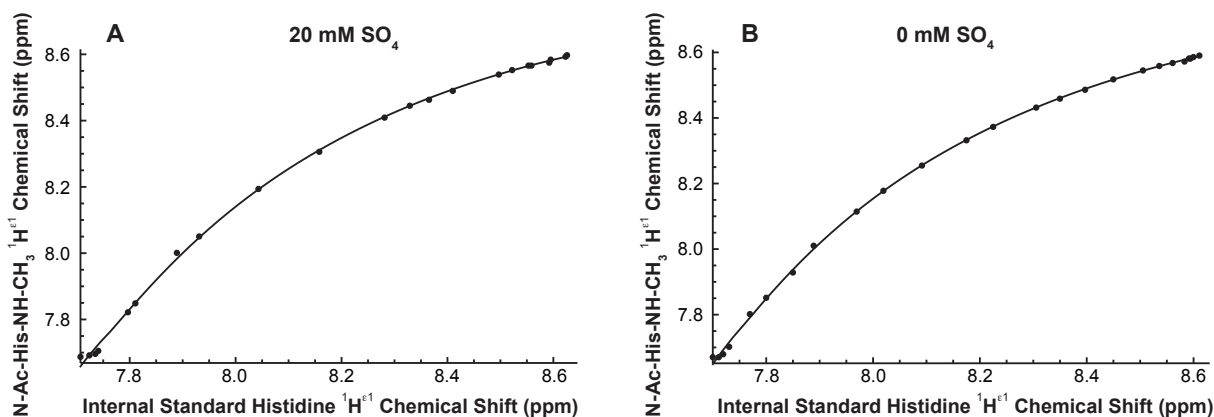
**Figure S2:** L-Histidine (internal standard) titration curves under different buffer conditions. In addition to the components indicated, each sample had 10 mM pyridine, 90% H<sub>2</sub>O, and 10% D<sub>2</sub>O. The <sup>13</sup>C<sup>ε1</sup> protons were used to determine the pK<sub>a</sub> values and Hill coefficients for L-Histidine, listed in panel F. Uncertainties are based on the standard errors of the fitted parameters.



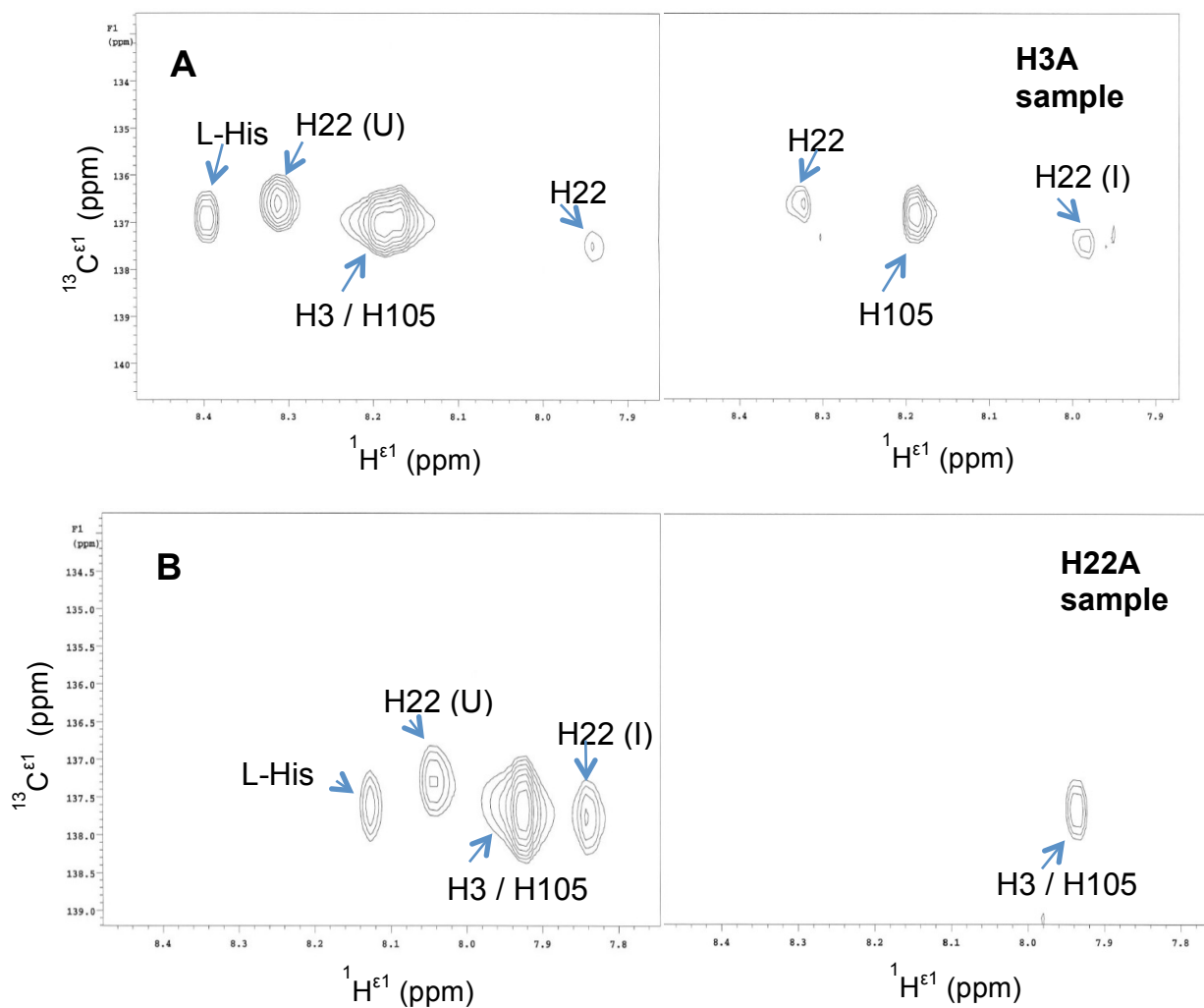
**Figure S3:** <sup>13</sup>C<sup>ε1</sup> P protein histidine chemical shifts versus <sup>13</sup>C<sup>ε1</sup> L-Histidine chemical shifts titration curves for (A) sulfate-bound folded and (B) unfolded P protein. H3, H22, and H105 are represented as the blue, red, and green curves, respectively. The purple curve in (B) represents the H22 intermediate peak.



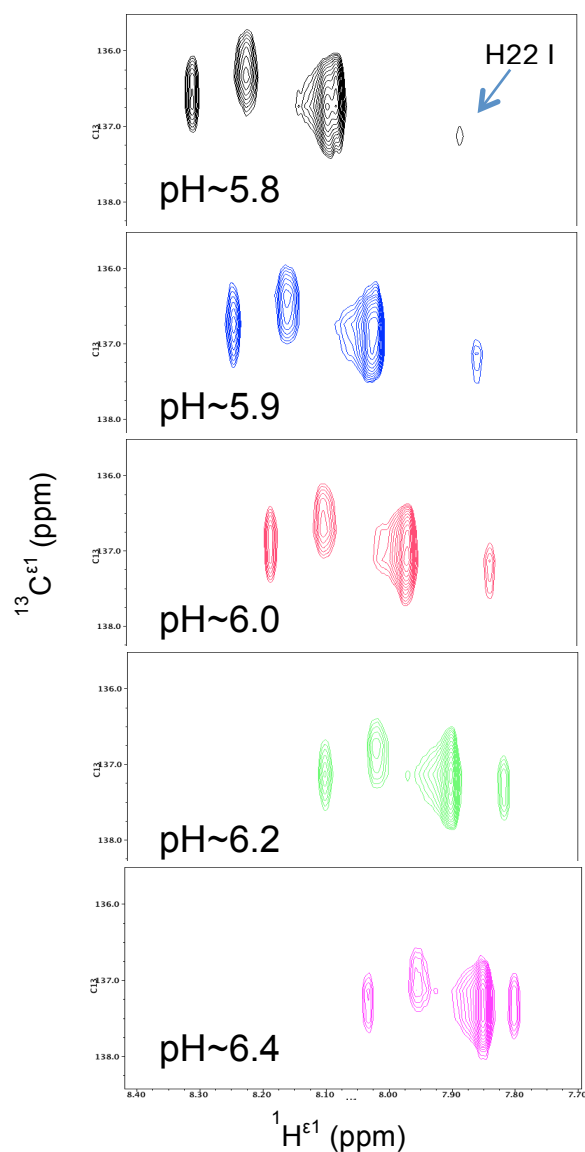
**Figure S4:** NMR resonances from all three histidine residues in the unfolded state of P protein were assigned by comparing spectra of (A) F107W P protein to spectra of two variants, (B) H105A and (C) H22A, that each lack a different histidine residues. All spectra were collected at a sample pH of 4. Assignment spectra for the sulfate-bound folded state are not shown.



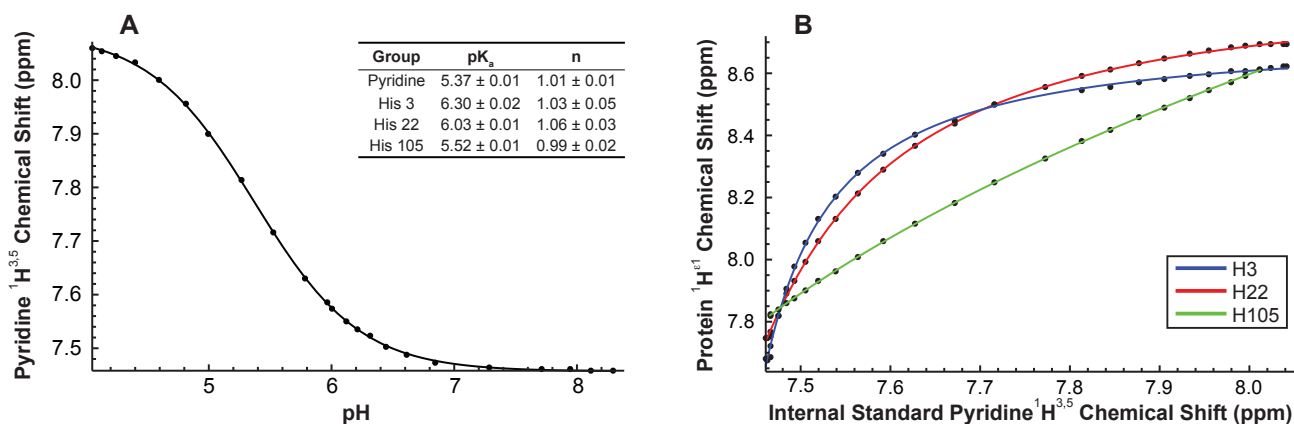
**Figure S5:**  $^1\text{H}^{\epsilon 1}$  N-acetyl-L-Histidine methylamide chemical shifts versus  $^1\text{H}^{\epsilon 1}$  L-Histidine chemical shifts titration curves in the (A) presence and (B) absence of 20 mM sodium sulfate. The estimated  $\text{pK}_a$  of N-acetyl-L-Histidine methylamide was (A)  $6.52 \pm 0.03$  and (B)  $6.44 \pm 0.02$ .



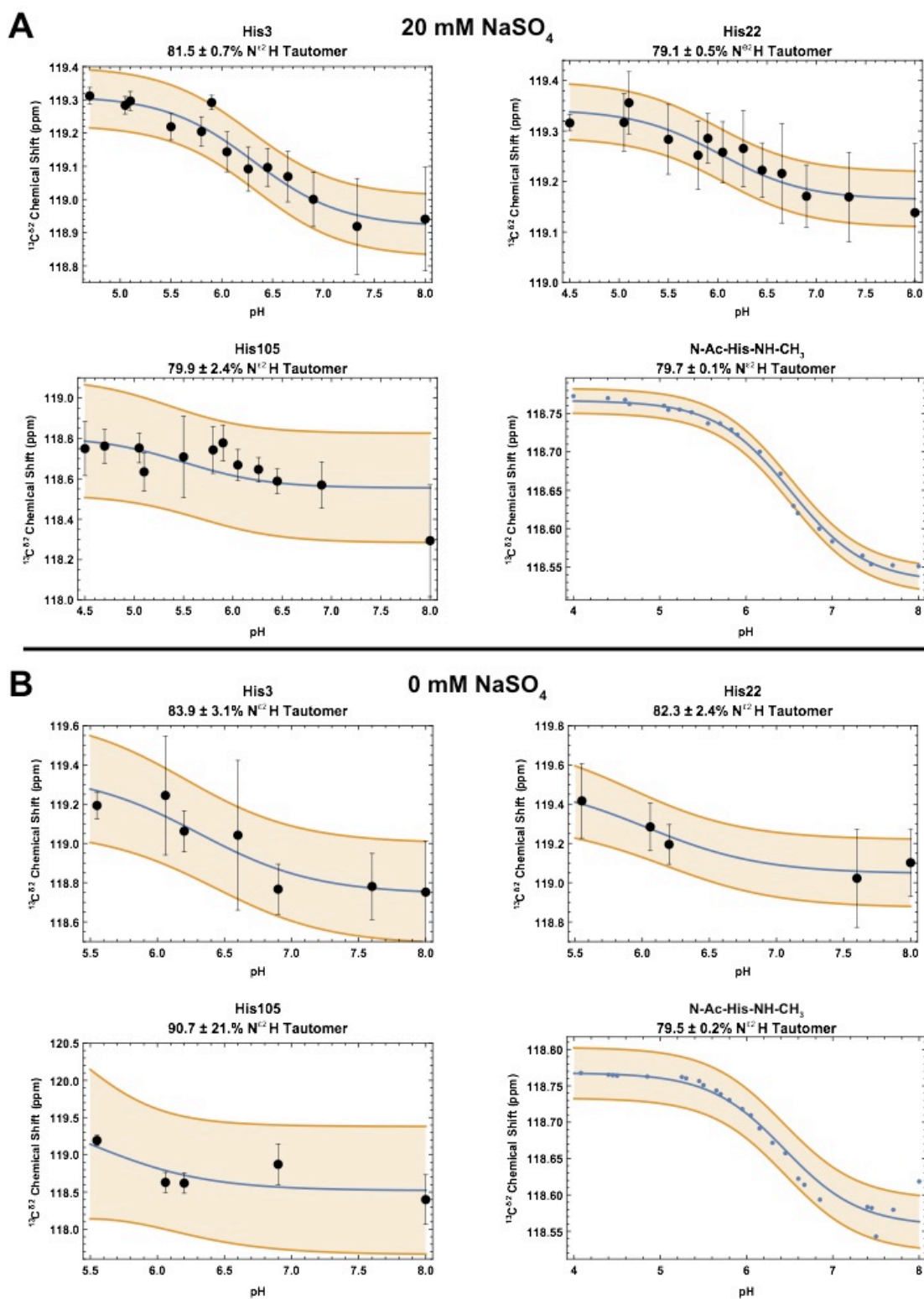
**Figure S6:** NMR resonance from intermediate peak in the unfolded state of P protein was assigned by comparing spectra of F107W P protein to spectra of two variants, **(A)** H3A and **(B)** H22A, that each lack a different histidine residue. The left sides of panels **(A)** and **(B)** correspond to spectra that were collected at a sample pH of 5.85 and 6.10, respectively.



**Figure S7:**  $^1\text{H}$ - $^{13}\text{C}$  HSQC unfolded P protein spectra in various pH buffers. The intensity of the H22 intermediate peak (H22 I) increases as the pH is increased.



**Figure S8:** (A) Pyridine (internal standard) and (B) P protein histidine residue titration curves in the presence of 20 mM sulfate using pyridine as the internal standard. Inset table lists the best-fit  $\text{pK}_a$  values, using pyridine as an internal standard instead of L-histidine.



**Figure S9:** Determination of the tautomeric state of the neutral form of the protein and model compound histidines in (A) 20 mM NaSO<sub>4</sub> (folded P protein) and (B) 0 mM NaSO<sub>4</sub> (unfolded P protein). All P protein histidines in both folding and unfolding conditions show very similar tautomeric state populations to the model compound N-acetyl-L-Histidine-methylamide, indicating that the protein histidines are highly solvated and form water hydrogen bonds under all conditions. The error bars represent 95% confidence levels obtained from peak fitting. The beige bands represent 95% confidence levels in the best-fit chemical shift. The fitting function is described in Materials and Methods.

**Table S1:** Histidine  $pK_a$  values and Hill coefficients in unfolded, intermediate, and sulfate-bound folded P protein based on fitting  $^{13}\text{C}^{\epsilon 1}$  data. <sup>a</sup>

Residue	$pK_a^F$	$n^F$	$pK_a^I$	$n^I$	$pK_a^U$	$n^U$
His 3	$6.32 \pm 0.02$	$1.10 \pm 0.06$	$5.73 \pm 0.04$	$0.94 \pm 0.09$	$5.73 \pm 0.04$	$0.94 \pm 0.09$
His 22	$6.10 \pm 0.05$	$0.99 \pm 0.12$	$5.40 \pm 0.55$	$0.91 \pm 0.41$	$6.11 \pm 0.04$	$0.95 \pm 0.09$
His 105	$5.55 \pm 0.05$	$0.90 \pm 0.03$	$5.80 \pm 0.03$	$0.96 \pm 0.08$	$5.80 \pm 0.03$	$0.96 \pm 0.08$

<sup>a</sup> Solutions contained 10 mM L-Histidine, 10 mM pyridine in 90% H<sub>2</sub>O, 10% D<sub>2</sub>O at 25°C with either 0 mM or 20 mM sodium sulfate. The  $pK_a$  values were obtained by fitting the data to Eq 5. The  $^{13}\text{C}^{\epsilon 1}$  shifts were used to determine the  $pK_a$  values. Uncertainties represent the 95% confidence intervals and were obtained as described in Materials and Methods.

**Table S2:** Solvent exposure of the histidine sidechains in P protein, relative to Gly-His-Gly, calculated as described in Materials and Methods.

Residue	Relative accessible surface area (%)
His 3	70
His 22	61
His 105	57

**Table S3:**  $^1\text{H}$  and  $^{13}\text{C}$  chemical shifts of internal standard L-Histidine, Pyridine, and P protein histidine residues when the imidazole ring is fully protonated (pH~4) or deprotonated (pH~8). Uncertainties represent the 95% confidence intervals and were obtained as described in Materials and Methods.

$\delta$ (ppm)	0 mM Na <sub>2</sub> SO <sub>4</sub>		20 mM Na <sub>2</sub> SO <sub>4</sub>	
	$^1\text{H}$	$^{13}\text{C}$	$^1\text{H}$	$^{13}\text{C}$
L-Histidine (pH~4)	$8.66 \pm 0.01$	$136.25 \pm 0.03$	$8.65 \pm 0.01$	$136.26 \pm 0.02$
L-Histidine (pH~8)	$7.72 \pm 0.01$	$138.59 \pm 0.03$	$7.72 \pm 0.01$	$138.59 \pm 0.02$
Pyridine (pH~4)	$8.08 \pm 0.01$	$129.47 \pm 0.05$	$8.09 \pm 0.01$	$129.49 \pm 0.03$
Pyridine (pH~8)	$7.45 \pm 0.01$	$126.58 \pm 0.02$	$7.46 \pm 0.01$	$126.57 \pm 0.01$
His 3 (pH~4)	$8.61 \pm 0.01$	$135.81 \pm 0.06$	$8.62 \pm 0.01$	$135.99 \pm 0.02$
His 3 (pH~8)	$7.69 \pm 0.01$	$138.33 \pm 0.07$	$7.67 \pm 0.01$	$137.98 \pm 0.04$
His 22 (pH~4)	$8.60 \pm 0.01$	$135.82 \pm 0.05$	$8.70 \pm 0.01$	$136.28 \pm 0.05$
His 22 (pH~8)	$7.67 \pm 0.01$	$138.48 \pm 0.08$	$7.74 \pm 0.01$	$138.46 \pm 0.09$
His 105 (pH~4)	$8.54 \pm 0.01$	$135.65 \pm 0.06$	$8.65 \pm 0.01$	$136.14 \pm 0.07$
His 105 (pH~8)	$7.69 \pm 0.01$	$138.52 \pm 0.06$	$7.81 \pm 0.01$	$137.99 \pm 0.05$