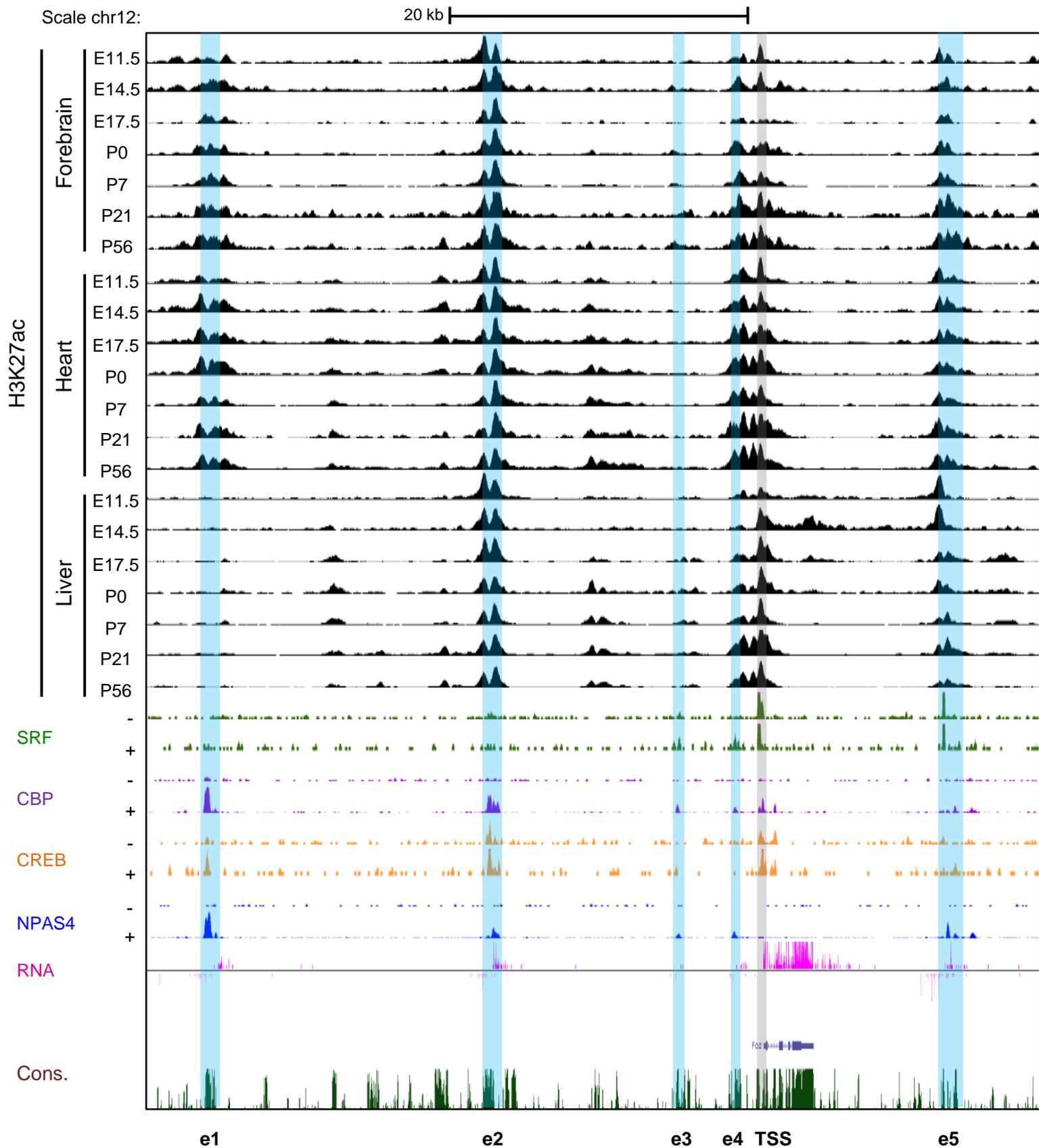


# Supplementary Figure 1



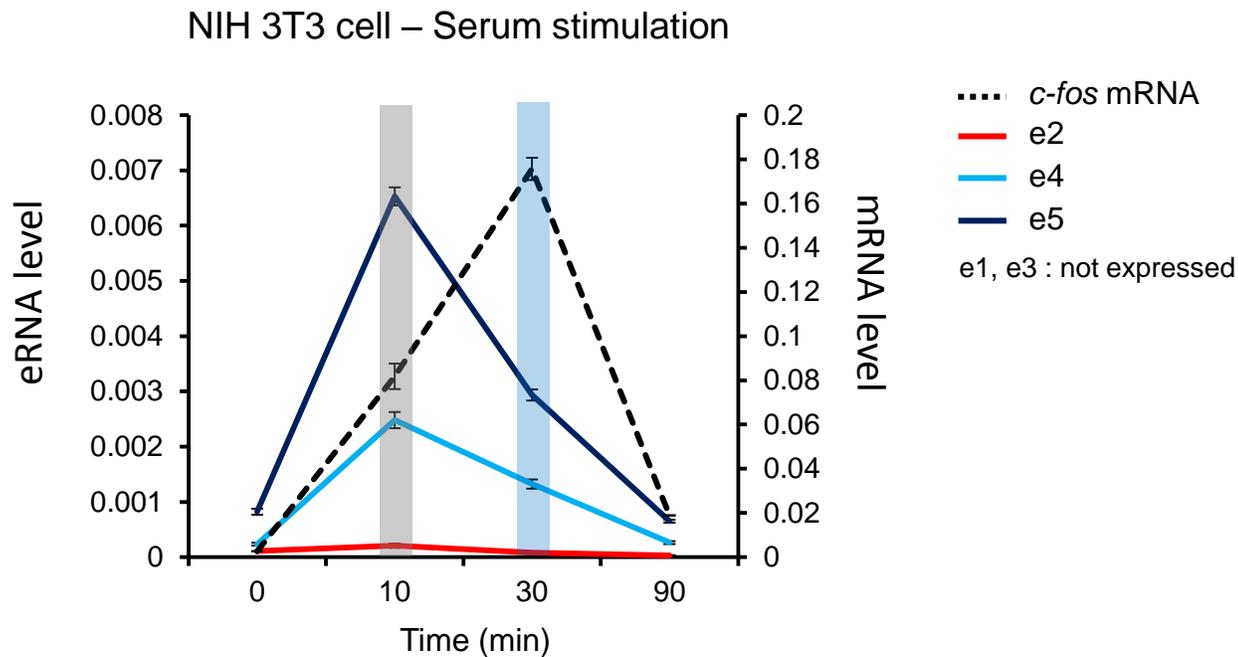
## Supplementary Figure 1

### Binding profiles of the H3K27ac mark at the *c-fos* enhancers and the promoter in various tissues throughout development

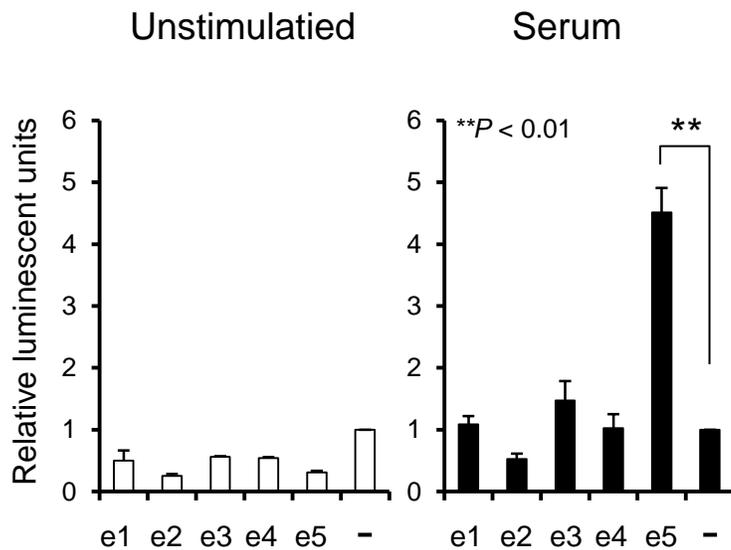
H3K27ac binding profiles were adapted from the data generated by Nord *et al.* 2013. Blue and gray vertical bars indicate the locations of the *c-fos* enhancers and the promoter, respectively.

# Supplementary Figure 2

a



b



## Supplementary Figure 2

### The *c-fos* enhancer activity in NIH 3T3 cells during serum stimulation.

(a) Time course analysis of *c-fos* mRNA and eRNA in NIH 3T3 cells following serum stimulation. Expression levels of *c-fos* mRNA and five eRNAs were measured at indicated time points after serum stimulation and normalized to *Gapdh* mRNA (n = 3 biological replicates). (b) Individual enhancer activities measured by luciferase reporter assay. The luciferase activity in NIH 3T3 cells transfected with each enhancer reporter construct was measured after 6 h serum stimulation ( $P = 0.0075$ ,  $t(2) = 11.513$ , n = 3 biological replicates, unpaired *t*-test with Welch correction). Error bars indicate SEM; p values are from a two-tailed *t* test.



e3 GCACCCGAGCTGTCCCGTTTCCTCAGCTCCAGATGGATCTGCTTCTGGCATTACACAGCCTTGTTTAAAGTGCTTGCAATTAG CAGCAGGATGGGGTTCCTACTGAAGCATCATCTCTTGTGAGAAAGCCAAGAGTTGCTCTGAATGAAAACAGAATTCTTCCC AAAGAGGC**TAAATCA**CTTGCCTAATTAGTCGCGGTGGTGGCGGGTGCCTGGGGTGGTGTCTAGTGG**TAAATCT**GTGCCT GTGCTCTTCTAAATAAGGACAACCTGGGGCTCTGTTTACAAGCAGAGGCATACGATGATGAGCCTTGATCCCACTAAGACCC TGTGTTGGGCTCACAATCGTGTGAGTCTCGCAGAGCATGTGAGATACTTCTTGTCAAATCCAGACATTTAGAACGGACACA TTTAGAATACTTTCAACAGTTACTGTCATTGTAGACTATGCGCTGTCTGTGTGTGGGCACTGTGGCCCTTCTCGTGGGGTA GCATAGCAGCCGGAGGCTGGTCTCTGATCTCCACCCACCATGAATACTGATGTTACACCACCTAGACTGCAACAGC TGAAGGATAGCTCAAGCGTGTACG**TATTTA**TCTTTCCAGTTTTTACTATTTCATACCCAAGAACGTTAAGTCTTGTCC

e4 TGAGGTGTTGGCCAGAGCTGCTTCAAACCTGAAAAGGAAA**TAAATCA**AATCTCTGCAG**TTTTCT**CTAAGAGTCCCTGAGA CGCAGTTATGAGTGAAGGCAAGGCCAGACAGACCCAGGAACCTCAAAGGTTGACCTACTTTGTGCAAATTAACAAGAGGG AAGACGTCTGCA**TATTTT**TCTGTCTCTCCCTTTCACAGTAAAGAACGGTGGGAGCACATGACAGACGAGCTGCAGGCCAC GAAATCCCTTCAAGGATTGAGCAGCTACACGAAGCCTGTGTTTCTATATCGTGTGTTCTCTTTCCCTCCCCCTTCCGAG TGAGAAAAAAGGGGGCCATAAATCCACCAACATAAACTAATGACATACAATGATGAAATTCTGTTTTACCTCTGCCT GTGACAGGGAATGCAAAAATAGCAAGTGGCCTATTTCCACGAATCCCCGCTCCCTGCCCTCCCCGCTCCCGCTGGGTTT GGATCTTAAGAATGGAGGCTAACGCAGAGGCAGGAGCCAGCCGGGATCAGCCCCCGAACCCGAGGGTGTGTCAGTC GCGGGAGCGGACGCAGCCTGGCCAGGGAGGCAGAGGTTCTAGGGAGCAGGGAGGGCGGGTGGCCTGGATGGGCG GACCGTGCGCCCGGCCGCTCCGTCGCCCTGGAGACGGCCCTGGCGGGCTGTGTTGCTGTAACAGCCGCTTCGCTGTTA CTATCTATAGAAAGATCGCCTAGCTCCCGGGTGCGGCGGCCGAGGCAGGTTTGTGGTCTGCACCC

TSS CTGACCCCCC**TTTTCT**TCTCTGCACTGATTTGGGATGGGGGGCTGATGTGGGCAAGCTTTCCTTTAGGAACAGAGGCTT CGAGCCTTAAAGGCTGCGTACTTGTCTTCTCCTAATACCAGAGACTC**AAAAAAAAAAAAA**AGTTCCAGATTGCTGGACAAT GACCCGGGTCTCATCCCTTGACCTGGGAACCGGGTCCACATTGAATCAGGTGCGAATGTTTCGCTCGCCTTCTCTGCCTT TCCCGCTCCCTCCCCCGGCCGCGGCCCGGTTCCCCCTGCGCTGCACCCTCAGAGTTGGCTGCAGCCGGCGGAGC TGTTCC**CGTCA**ATCCCTCCCTCCTTACACAGGATG**TCCATATTAG**ACATCTG**CGTCA**GCAGGTTCCACGGCCGGTCC CTGTTGTTCTGGGGGGGGACCATCTCCGAAATCTACACGCGGAAGGTCTAGGAGACCCCTAAGATCCCAAATGTGAA CACTCATAGGTGAAAGATGTATGCCAAGACGGGGTTGAAAGCCTGGGGCGTAGAGT**TGACG**ACAGAGCGCCCGCAGAG GGCCTTGGGGCGCGCTTCCCCCCTTCCAGTTCGCCAG**TGACG**TAGGAAGTCCATTCACAGCGCTTCTATAAA GCGCCAGCTG

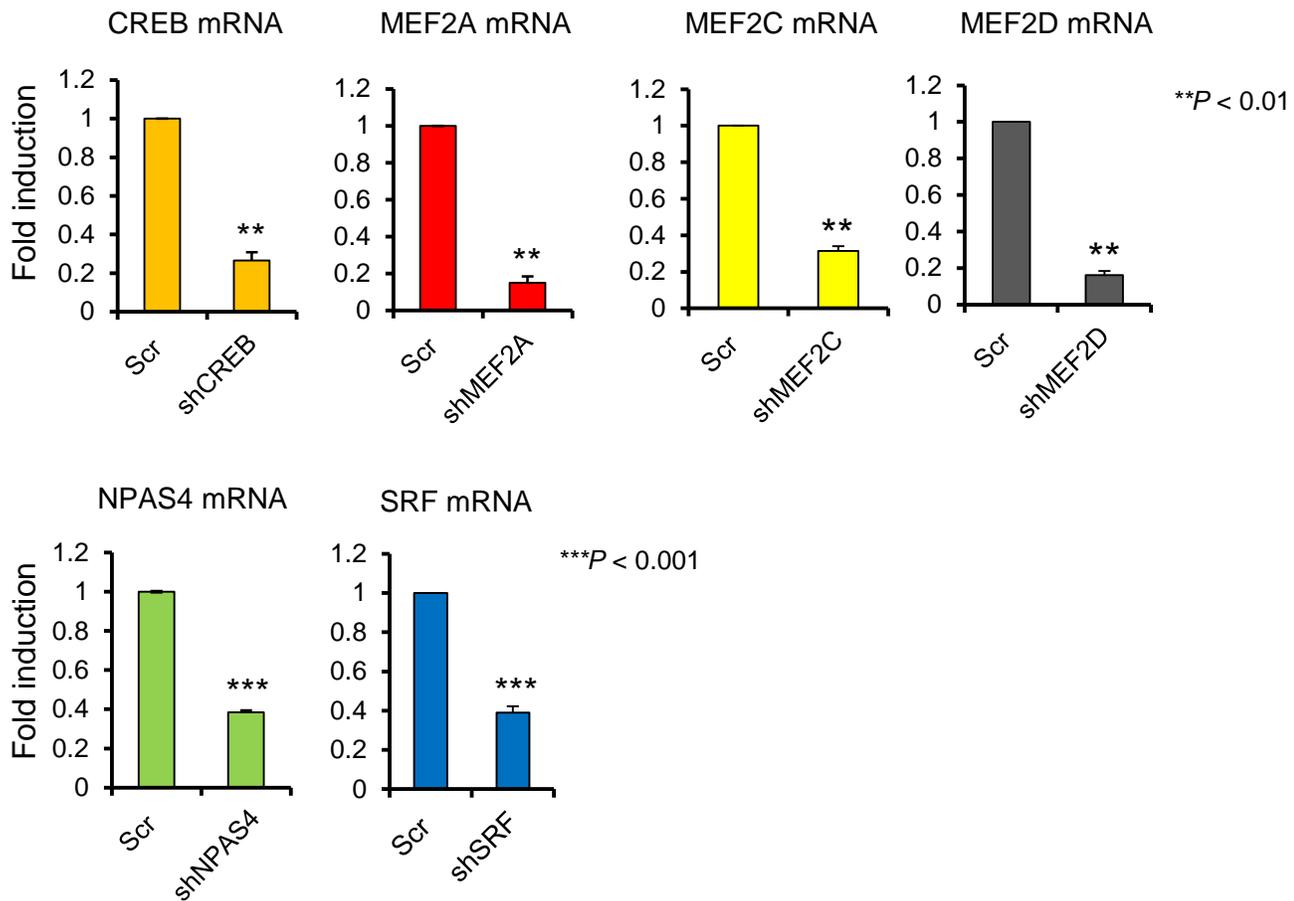
e5 GGAGCCGGCCAGTCTCTATTTACTGTTTGGTTTCCACGGTGACACGTAGGCGC**CGTCA**AGCGCCTCC**CGTCA**AGCGGCC CCACCCGGCGCCG**TGACG**TCTGCCCGCGCTTCCGTTCTGC**TCATATTGG**AAAGGCGCCGCGGGCAGGGAGGGGTTG GGAGCCGACTCCCGCGGCGCCCGCCCCCTGCCGGTCCGCCCGCACCCGAGGAGCCATGGGCTGTGCA CCCCTCCTCTACCCCGACCCTGCGGAGGCGGGCGGGGGCTCGGGTCCGACGATCAGATTTTCGCTTCTGTGCCGAAATCCT TCAAAGCTTAGGGCCTGGCTCCTCTGTTCCCGGTGGACGATCCGGGATCAGATTTTCGCTTCTGTGCCGAAATCCT GAAACAATCCCGGTCTAACCTCCCTCTCTTTGATACACTTGATTTTCAAGATTCTCGTCTTTGGCGCAGTGAATAGCTTTAA GGACACAAGTTCGGGGGTGGGTGGTCTGCGTTTTCCCTGGGACATAAGTGACCTTGGGAAGGCCAC**TTTTTTTTTTTTC** TGCTGGGGCC**TTTTAATTTTTT**CAACCCTGTCATCTATTTAGCGAGGTCTTCCATCACCTCTAAGTACCCCTCTT TGGTACATAAATGCAAAGTAGGTGAAGTCTTGAAGACCCCGCAGTCTTATGCCTGCACTACTTGTATCTGTGCGTCTC CTGGGCTTGTCAATTTAGACGC**TTAATTA**ACTTTAGTTAACTGCTGAATGAATGATCCGAGACGGGTTTACCTTCATA AGTCCGATCCCGCAATTCC**TTTTACA**GGCGGAGGGAATATTGCGATGA**TTAATAA**TCGCGCGGCAGGGTTGCTGATGTAA AGCGATTTGCGGATGTAAAGGGCAATAAACATGCGGATGTAAAGGAATCTATCGAGATTCCAGAAGTCTCTAAGGAGCCGT GGGT

### Supplementary Figure 3

#### Transcription factor binding motifs present at the *c-fos* enhancers and the promoter.

(a) Transcription factor binding motifs present at each *c-fos* enhancer. (b) Transcription factor binding motifs present in each of the *c-fos* enhancers and the promoter. Binding motifs for CREB, MEF2, and SRF are indicated by blue, red, and orange letters, respectively.

# Supplementary Figure 4



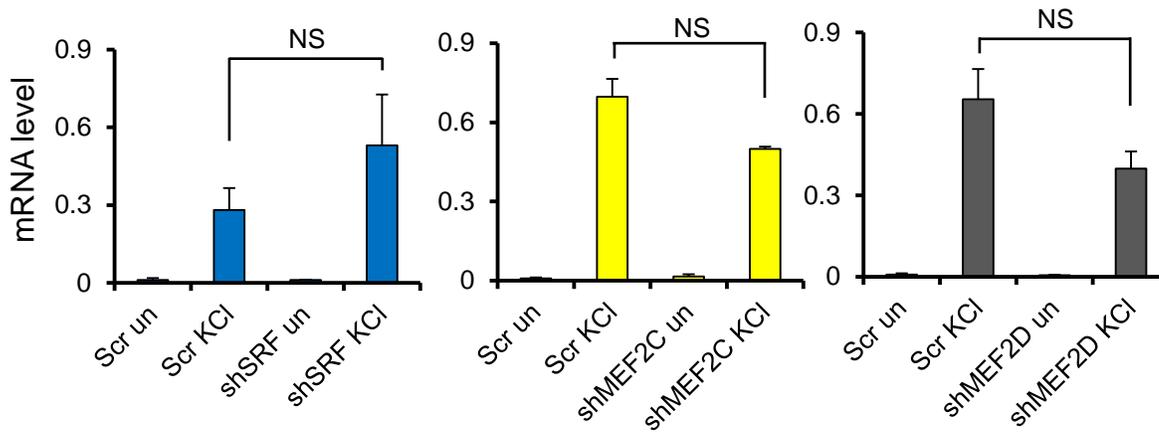
## Supplementary Figure 4

### Demonstration of the efficiency of knockdown.

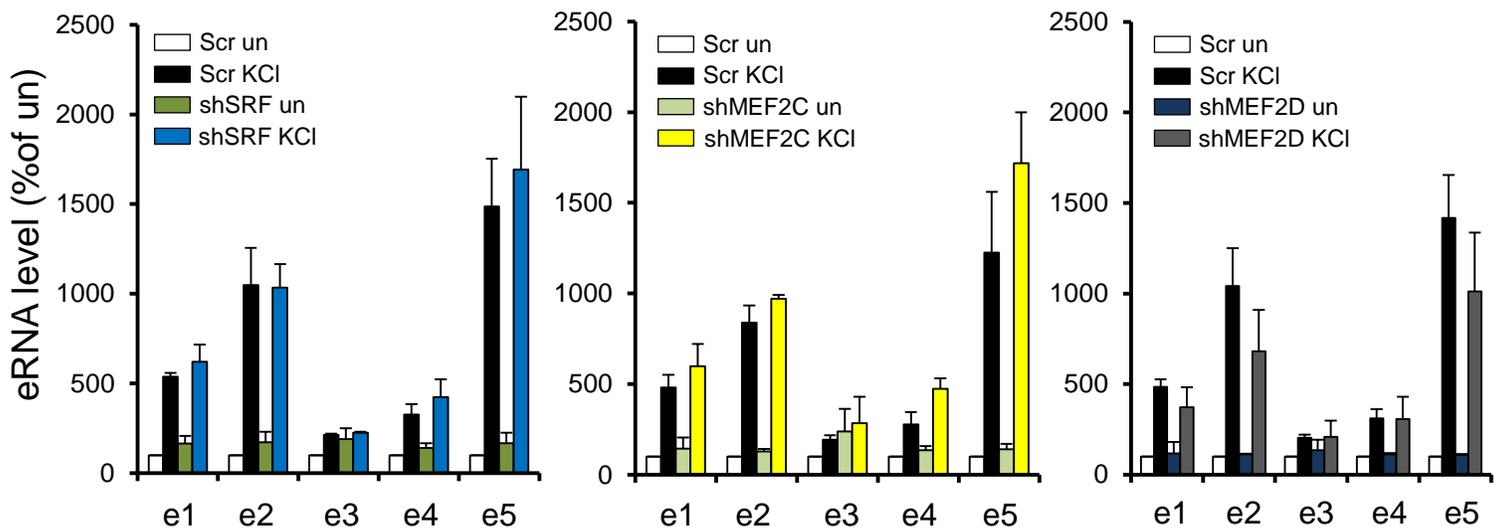
RT-qPCR analysis of *Creb*, *Mef2a*, *Mef2c*, *Mef2d*, *Npas4*, and *Srf* mRNA expression after knockdown in cortical neurons, related to **Fig. 4** and **Supplementary Fig. 5** (**CREB mRNA**:  $P = 0.0031$ ,  $t(2) = 18.040$ ,  $n = 3$ ; **MEF2A mRNA**:  $P = 0.0029$ ,  $t(2) = 18.616$ ,  $n = 3$ ; **MEF2C mRNA**:  $P = 0.0019$ ,  $t(2) = 22.975$ ,  $n = 3$ ; **MEF2D mRNA**:  $P = 0.0012$ ,  $t(2) = 28.633$ ,  $n = 3$ ; **NPAS4 mRNA**:  $P = 0.0019$ ,  $t(2) = 22.709$ ,  $n = 3$ ; **SRF mRNA**:  $P = 0.0014$ ,  $t(2) = 26.942$ ,  $n = 3$  biological replicates). All unpaired  $t$ -test with Welch correction. Error bars indicate SEM; p values are from a two-tailed  $t$  test.

# Supplementary Figure 5

## *c-fos* mRNA



## *c-fos* eRNA

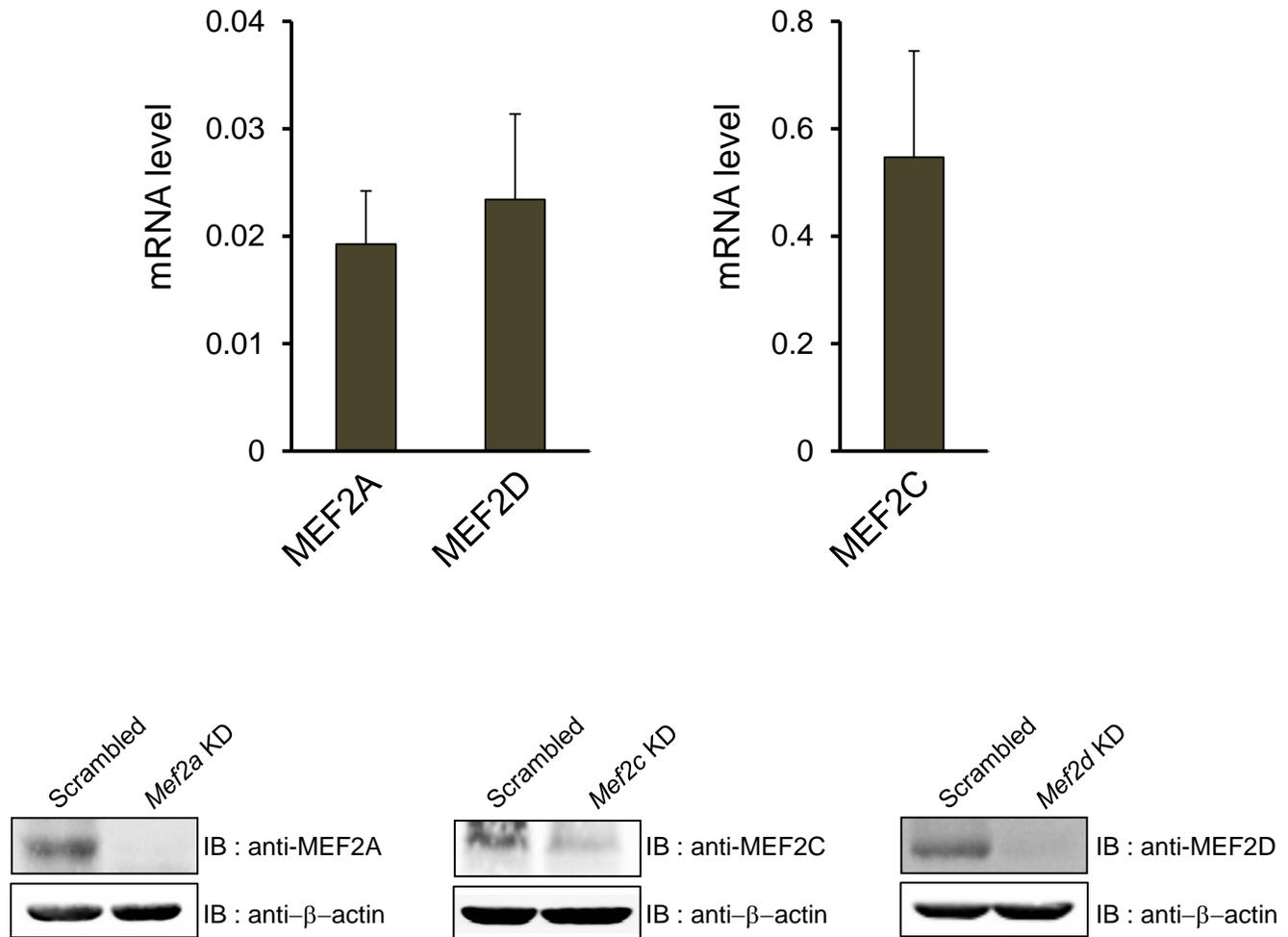


### Supplementary Figure 5

#### Effect of SRF, MEF2C, and MEF2D knockdown on KCl-mediated induction of *c-fos* mRNA and eRNAs.

Effect of transcription factor knockdown in KCl-mediated *c-fos* mRNA and eRNA induction (n = 3 biological replicates). Error bars indicate SEM; p values are from a two-tailed *t* test. NS, not significant.

## Supplementary Figure 6



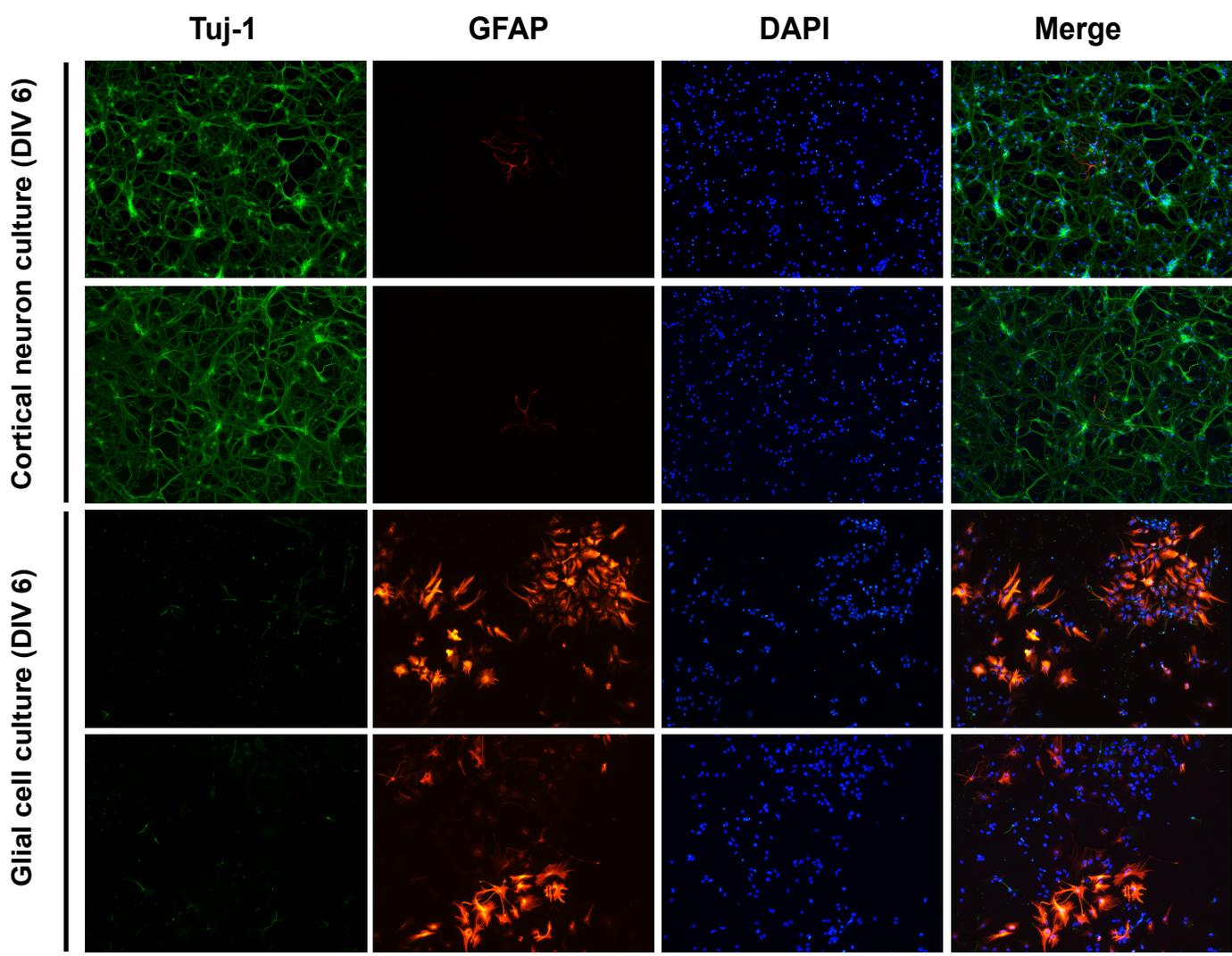
### Supplementary Figure 6

#### Expression levels of MEF2 family member mRNAs and proteins in cortical neurons.

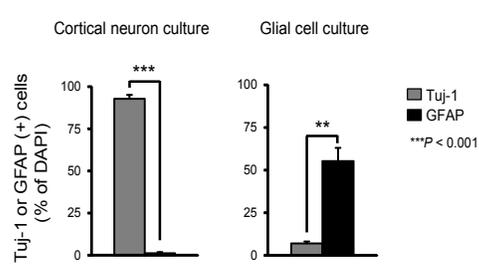
*Mef2a*, *Mef2c*, and *Mef2d* mRNAs were measured at DIV 6, and normalized to the level of *Gapdh* mRNA (top, n = 3 biological replicates). Cortical neurons infected with lentivirus encoding either a scrambled shRNA or shRNA against *Mef2a*, *Mef2c*, and *Mef2d*. Beta ( $\beta$ )-actin protein was blotted as a loading control (bottom). **Western blot experiment was only run once. The original western blot images are presented in Supplementary Figure 16.** Error bars indicate SEM. (Note that the y axes are scaled differently).

# Supplementary Figure 7

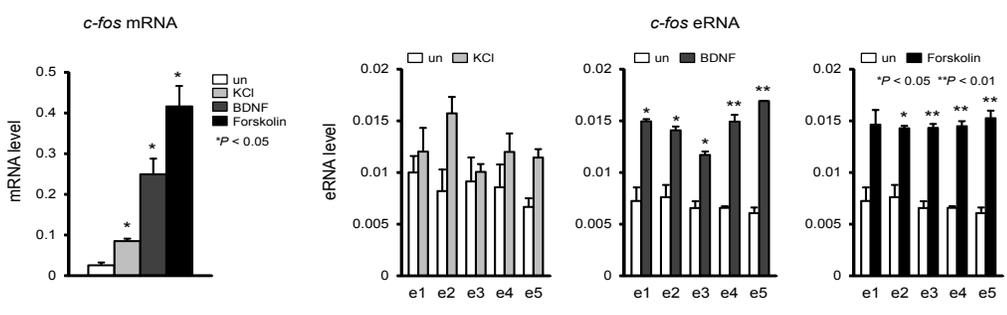
**a**



**b**



**c**

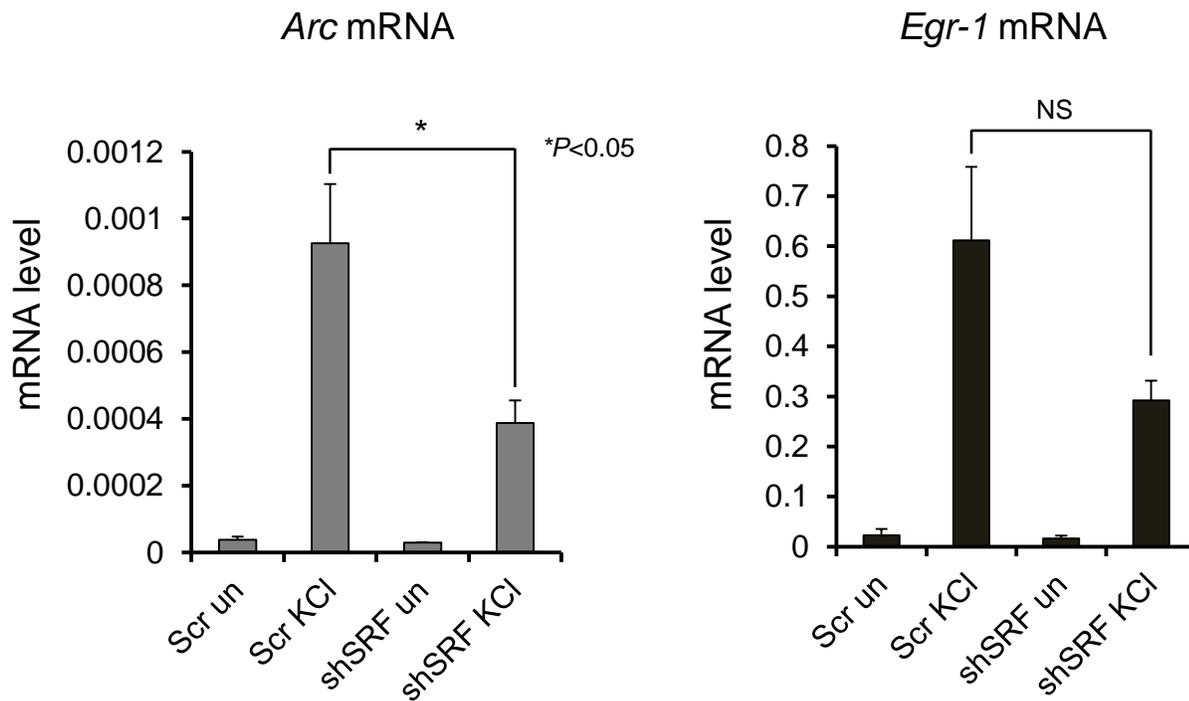


## Supplementary Figure 7

### The *c-fos* enhancer activities measured by eRNA analysis in glial cells.

(a,b) Cultured DIV 6 cortical neurons and glial cells were double immunostained by Tuj-1 and GFAP. Tuj-1 (+) or GFAP (+) cells were counted, and numbers are expressed as the percentage of DAPI (+) cells (Cortical neuron culture:  $P = 0.0001$ ,  $t(3) = 37.044$ ,  $n = 4$ ; Glial cell culture:  $P = 0.0089$ ,  $t(3) = 6.084$ ,  $n = 4$ , unpaired  $t$ -test with Welch correction unpaired  $t$ -test with Welch correction). (c) Expression of *c-fos* mRNA and eRNA in glial cells induced by KCl, BDNF, and Forskolin. The induction levels of the five *c-fos* eRNAs and mRNA were measured using RT-qPCR and normalized to *Gapdh* mRNA (KCl stimulation:  $P = 0.0239$ ,  $t(2) = 6.349$ ,  $F = 1.356$  [ $sdP = 0.4517$ ],  $n = 2$ ; BDNF stimulation:  $P = 0.0286$ ,  $t(2) = 5.787$ ,  $F = 28.667$  [ $sdP = 0.1175$ ],  $n = 2$ ; Forskolin stimulation:  $P = 0.0165$ ,  $t(2) = 7.690$ ,  $F = 50.055$  [ $sdP = 0.0894$ ],  $n = 2$ ; BDNF stimulation e1:  $P = 0.0283$ ,  $t(2) = 5.817$ ,  $F = 36.979$  [ $sdP = 0.1038$ ],  $n = 2$ , e2:  $P = 0.0345$ ,  $t(2) = 5.240$ ,  $F = 11.137$  [ $sdP = 0.1853$ ],  $n = 2$ , e3:  $P = 0.0195$ ,  $t(2) = 7.063$ ,  $F = 3.634$  [ $sdP = 0.3076$ ],  $n = 2$ , e4:  $P = 0.0064$ ,  $t(2) = 12.464$ ,  $F = 17.529$  [ $sdP = 0.1493$ ],  $n = 2$ , e5:  $P = 0.0025$ ,  $t(2) = 19.845$ ,  $F = 961.36$  [ $sdP = 0.0571$ ],  $n = 2$ ; Forskolin stimulation: e2:  $P = 0.0312$ ,  $t(2) = 5.527$ ,  $F = 26.532$  [ $sdP = 0.1221$ ],  $n = 2$ , e3:  $P = 0.0092$ ,  $t(2) = 10.368$ ,  $F = 2.896$  [ $sdP = 0.3395$ ],  $n = 2$ , e4:  $P = 0.0042$ ,  $t(2) = 15.298$ ,  $F = 10.024$  [ $sdP = 0.1948$ ],  $n = 2$ , e5:  $P = 0.0096$ ,  $t(2) = 10.109$ ,  $F = 1.775$  [ $sdP = 0.4099$ ],  $n = 2$  biological replicates, unpaired  $t$ -test). Error bars indicate SEM; p values are from a two-tailed  $t$  test.  $sdP$  is the P value from comparing the standard deviations for both groups.  $sdP > 0.05$  means the two SDs are not significantly different and can be used for an unpaired  $t$ -test.

## Supplementary Figure 8



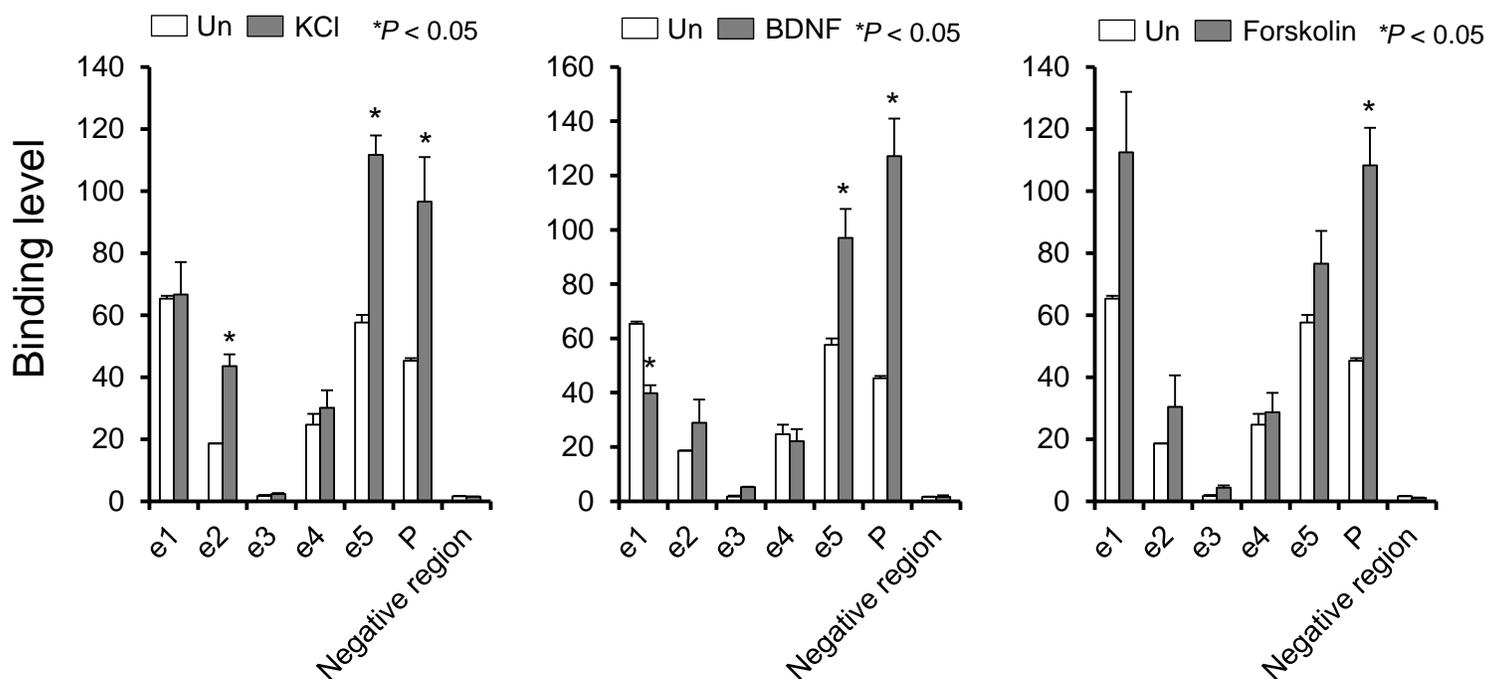
### Supplementary Figure 8

#### SRF regulates KCl-mediated induction of *Arc* and *Egr-1* mRNAs.

SRF was knocked down by shRNAs in cortical neurons, and then the induction levels of *Arc* and *Egr-1* mRNA were measured by RT-qPCR (*Arc* mRNA:  $P = 0.0481$ ,  $t(4) = 2.814$ ,  $F = 6.896$  [ $sdP = 0.1267$ ],  $n = 3$  biological replicates, unpaired  $t$ -test). Error bars indicate SEM; p values are from a two-tailed  $t$  test. NS, not significant.  $sdP$  is the P value from comparing the standard deviations for both groups.  $sdP > 0.05$  means the two SDs are not significantly different and can be used for an unpaired  $t$ -test.

# Supplementary Figure 9

## MEF2A binding

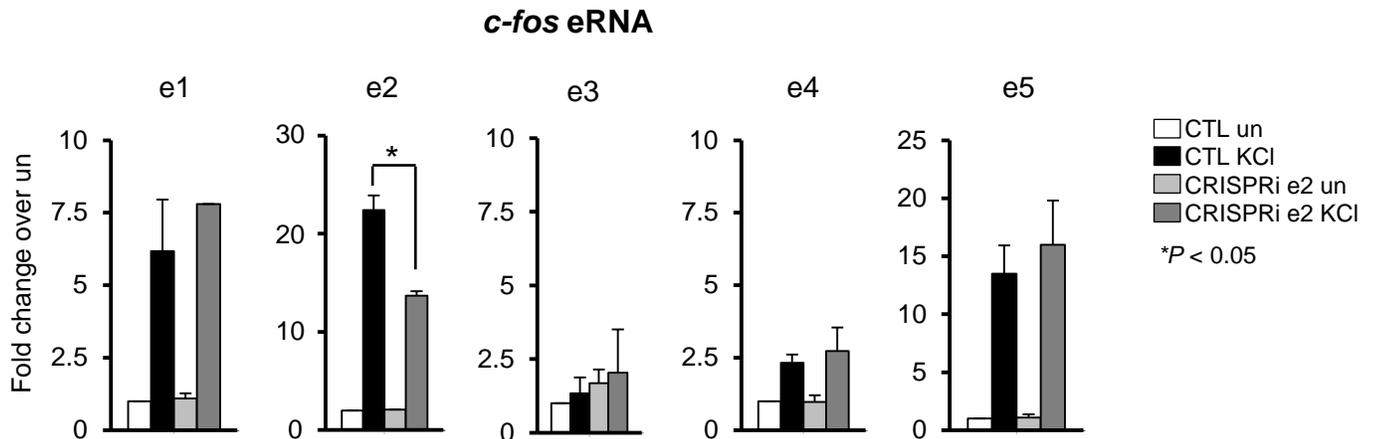


### Supplementary Figure 9

#### MEF2A binding profile at the *c-fos* promoter and five enhancers by KCl, BDNF, and Forskolin stimulation.

Binding was determined at the following locations along the *c-fos* gene: *c-fos* enhancer 1 (e1), enhancer 2 (e2), enhancer 3 (e3), enhancer 4 (e4), enhancer 5 (e5), or promoter (P) (KCl stimulation: e2,  $P = 0.0466$ ,  $t(1) = 13.632$ ,  $n = 2$ , unpaired  $t$ -test with Welch correction, e5,  $P = 0.0180$ ,  $t(2) = 7.355$ ,  $F = 6.548$  [ $sdP = 0.2372$ ],  $n = 2$ , P,  $P = 0.0228$ ,  $t(2) = 6.514$ ,  $F = 150.94$  [ $sdP = 0.0517$ ],  $n = 2$ , unpaired  $t$ -test; BDNF stimulation: e1,  $P = 0.0135$ ,  $t(2) = 8.507$ ,  $F = 10.418$  [ $sdP = 0.1913$ ],  $n = 2$ , e5,  $P = 0.0419$ ,  $t(2) = 4.733$ ,  $F = 19.313$  [ $sdP = 0.1432$ ],  $n = 2$ , P,  $P = 0.0121$ ,  $t(2) = 8.997$ ,  $F = 136.89$  [ $sdP = 0.0543$ ],  $n = 2$ , unpaired  $t$ -test; Forskolin stimulation: P,  $P = 0.0182$ ,  $t(2) = 7.313$ ,  $F = 90.055$  [ $sdP = 0.0668$ ],  $n = 2$  biological replicates, unpaired  $t$ -test). Error bars indicate SEM; p values are from a two-tailed  $t$  test.  $sdP$  is the P value from comparing the standard deviations for both groups.  $sdP > 0.05$  means the two SDs are not significantly different and can be used for an unpaired  $t$ -test.

# Supplementary Figure 10

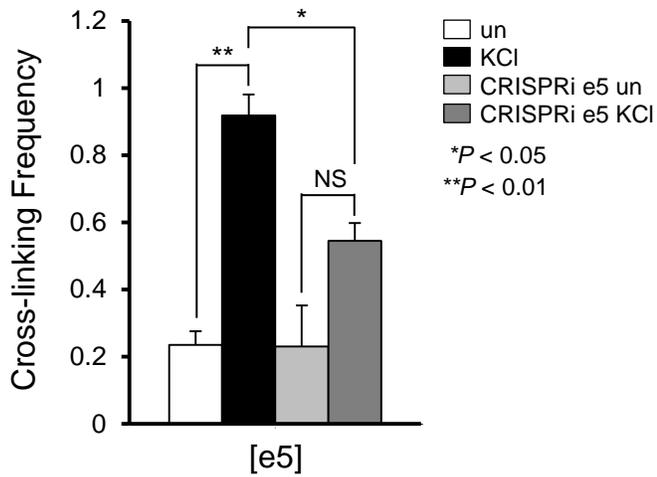


## Supplementary Figure 10

### Specific e2 enhancer-targeted CRISPRi does not affect other *c-fos* eRNA.

Expression levels of e1, e2, e3, e4, and e5 eRNA were measured using RT-qPCR (*c-fos* e2 eRNA, ,  $P = 0.0393$ ,  $t(4) = 3.015$ ,  $F = 3.461$  [ $sdP = 0.2242$ ],  $n = 3$  biological replicates). Error bars indicate SEM; p values are from a two-tailed  $t$  test.  $sdP$  is the P value from comparing the standard deviations for both groups.  $sdP > 0.05$  means the two SDs are not significantly different and can be used for an unpaired  $t$ -test.

# Supplementary Figure 11

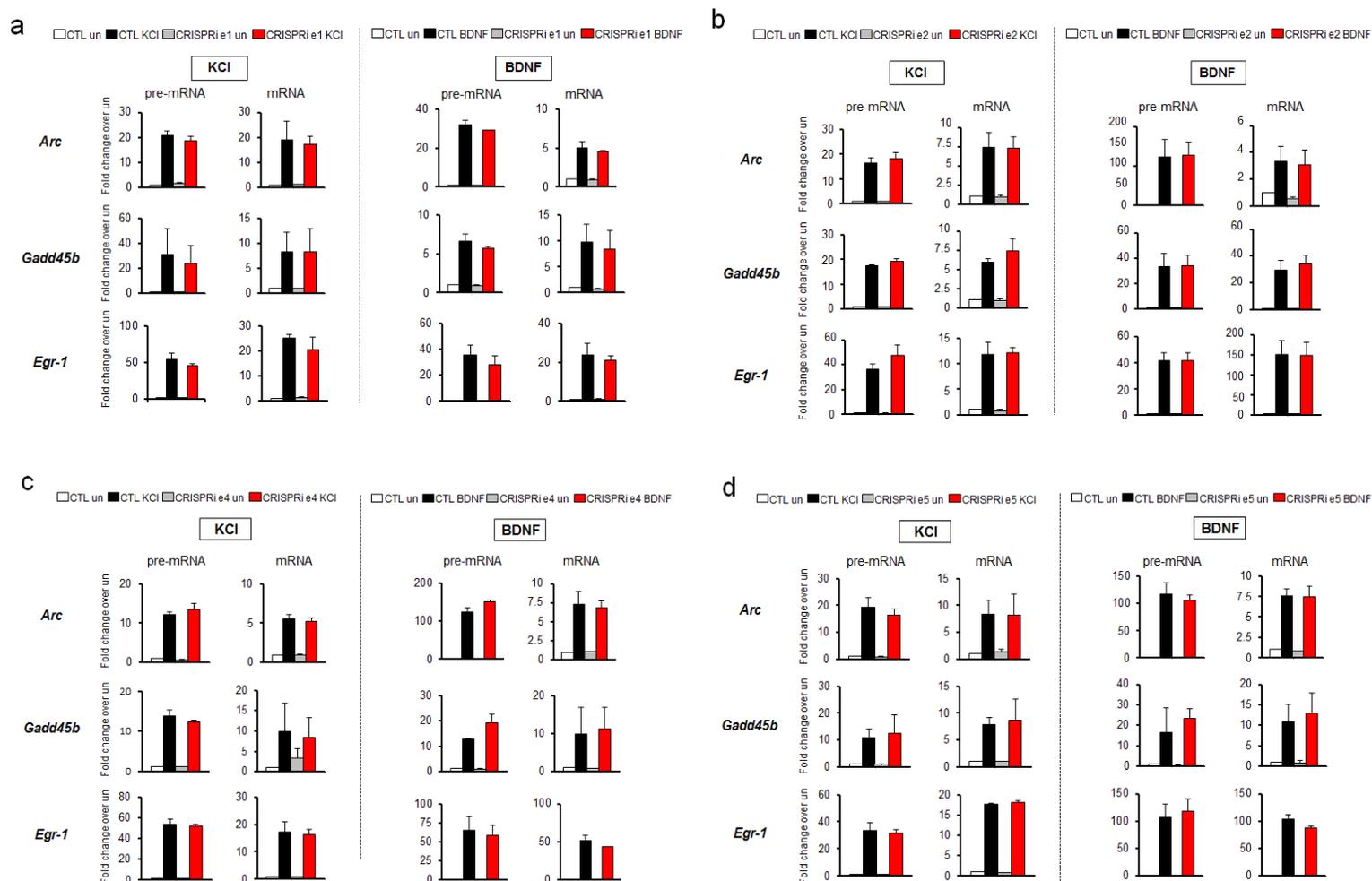


## Supplementary Figure 11

### The *c-fos* enhancer 5 and promoter interaction was decreased by suppression of the e5 enhancer

Cortical neurons were infected with an sgRNA targeting e5 and stimulated with KCl. (KCl e5:  $P = 0.0117$ ,  $t(2) = 9.144$ ,  $F = 2.420$  [ $sdP = 0.3637$ ],  $n = 2$ ; CRISPRi e5 KCl:  $P = 0.0453$ ,  $t(2) = 4.539$ ,  $F = 1.398$  [ $sdP = 0.4470$ ],  $n = 2$  biological replicates). All unpaired *t*-test. Error bars indicate SEM; p values are from a two-tailed *t* test. *sdP* is the P value from comparing the standard deviations for both groups.  $sdP > 0.05$  means the two SDs are not significantly different and can be used for an unpaired *t*-test.

# Supplementary Figure 12



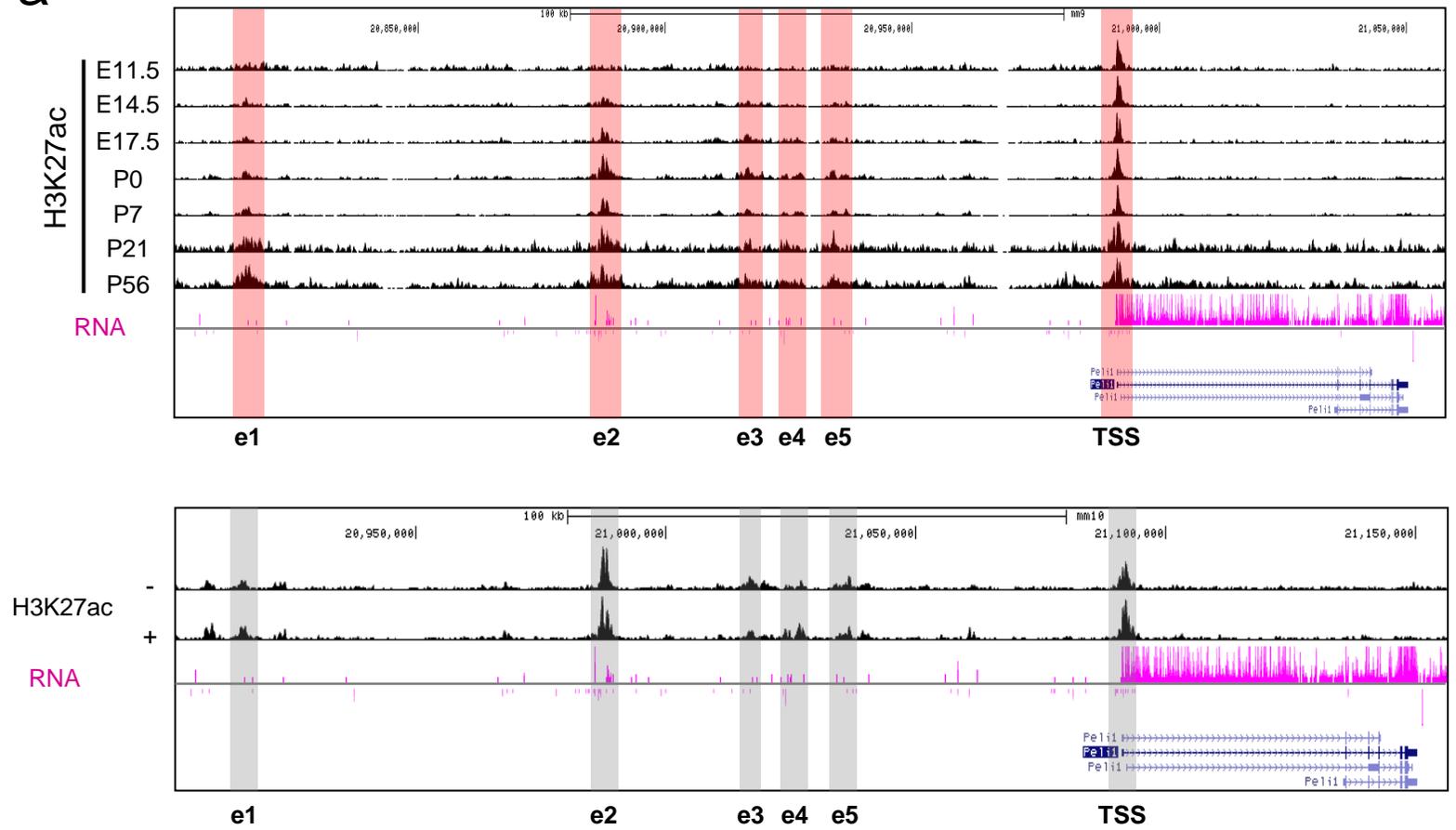
## Supplementary Figure 12

### Effect of e1, e2, e4, and e5 enhancer-targeted CRISPRi on various IEGs.

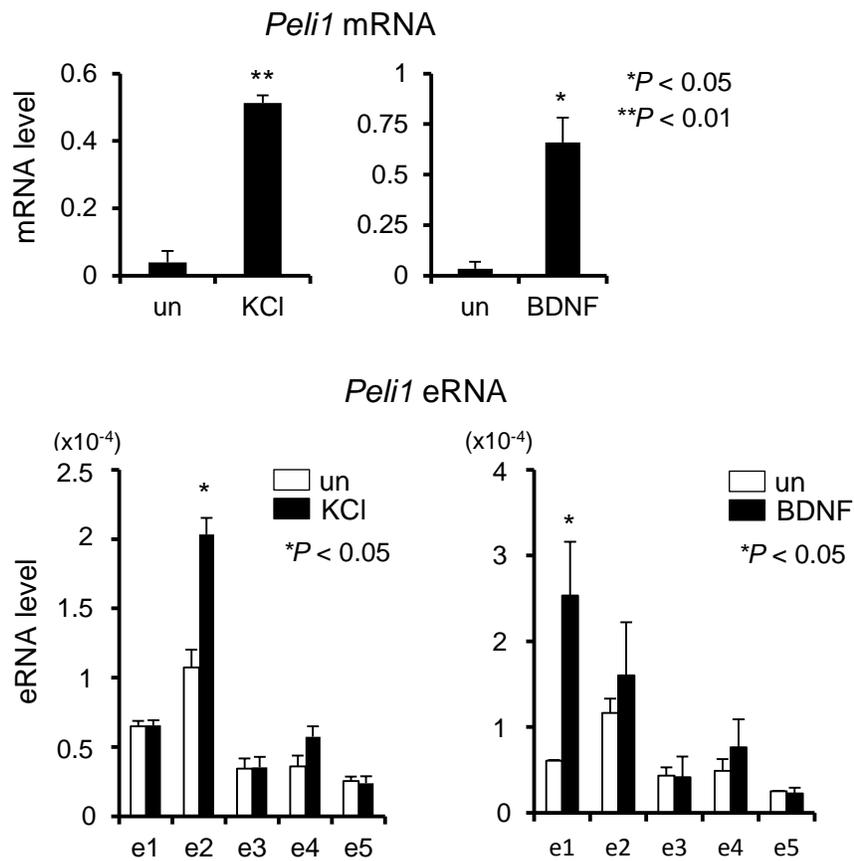
(a-d) Following the suppression of e1, e2, e4, and e5 enhancer by CRISPRi, cortical neurons were stimulated by KCI or BDNF, and expression levels of *Arc*, *Gadd45b*, and *Egr-1* pre-mRNA and mRNA were measured using RT-qPCR (KCI stimulation: n = 3 biological replicates, BDNF stimulation: n = 2 biological replicates). Error bars indicate SEM; p values are from a two-tailed *t* test.

# Supplementary Figure 13

a



b



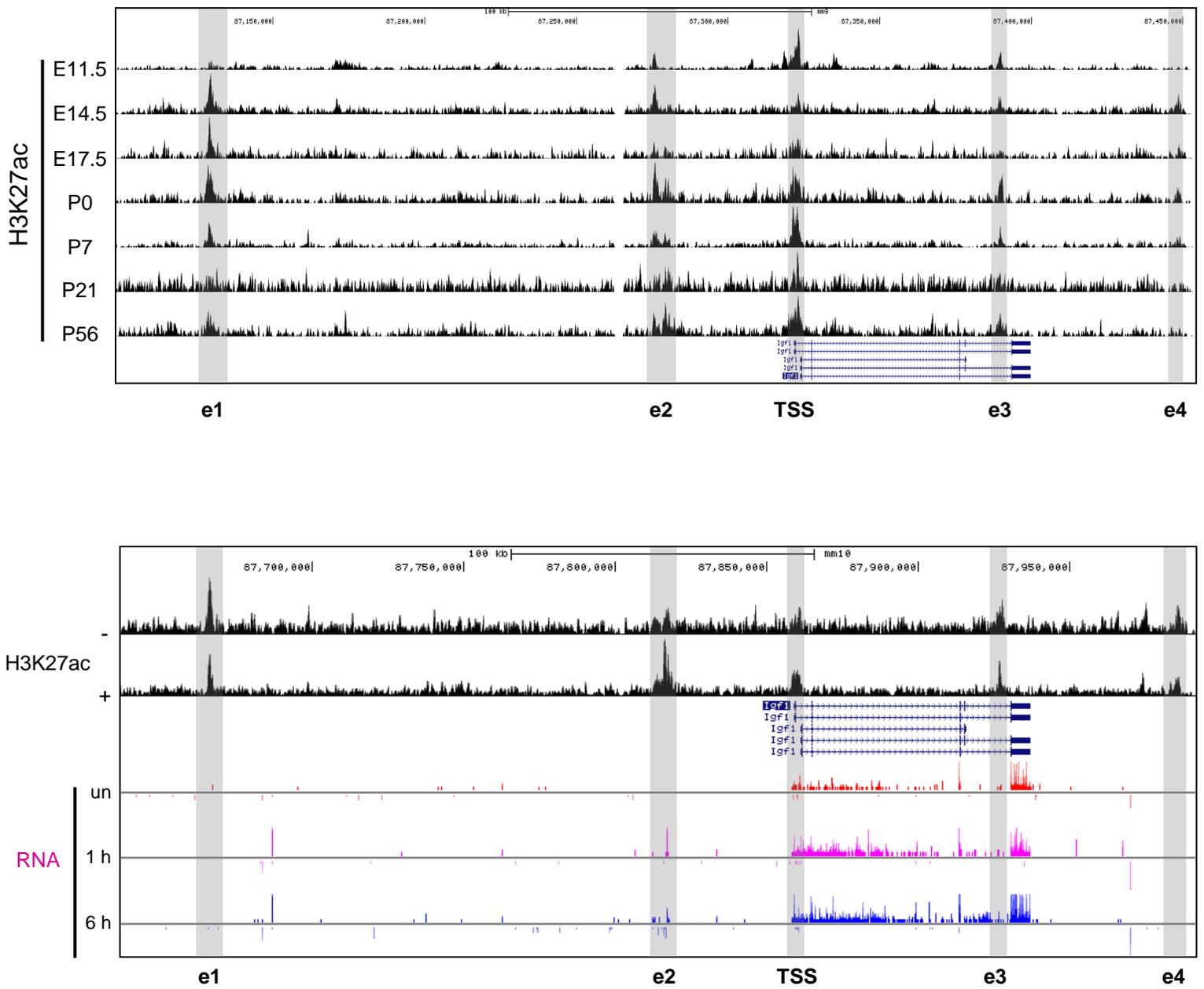
## Supplementary Figure 13

### Binding profiles of the H3K27ac mark at the *Pelil* enhancer and promoter and enhancer activities measured by eRNA analysis.

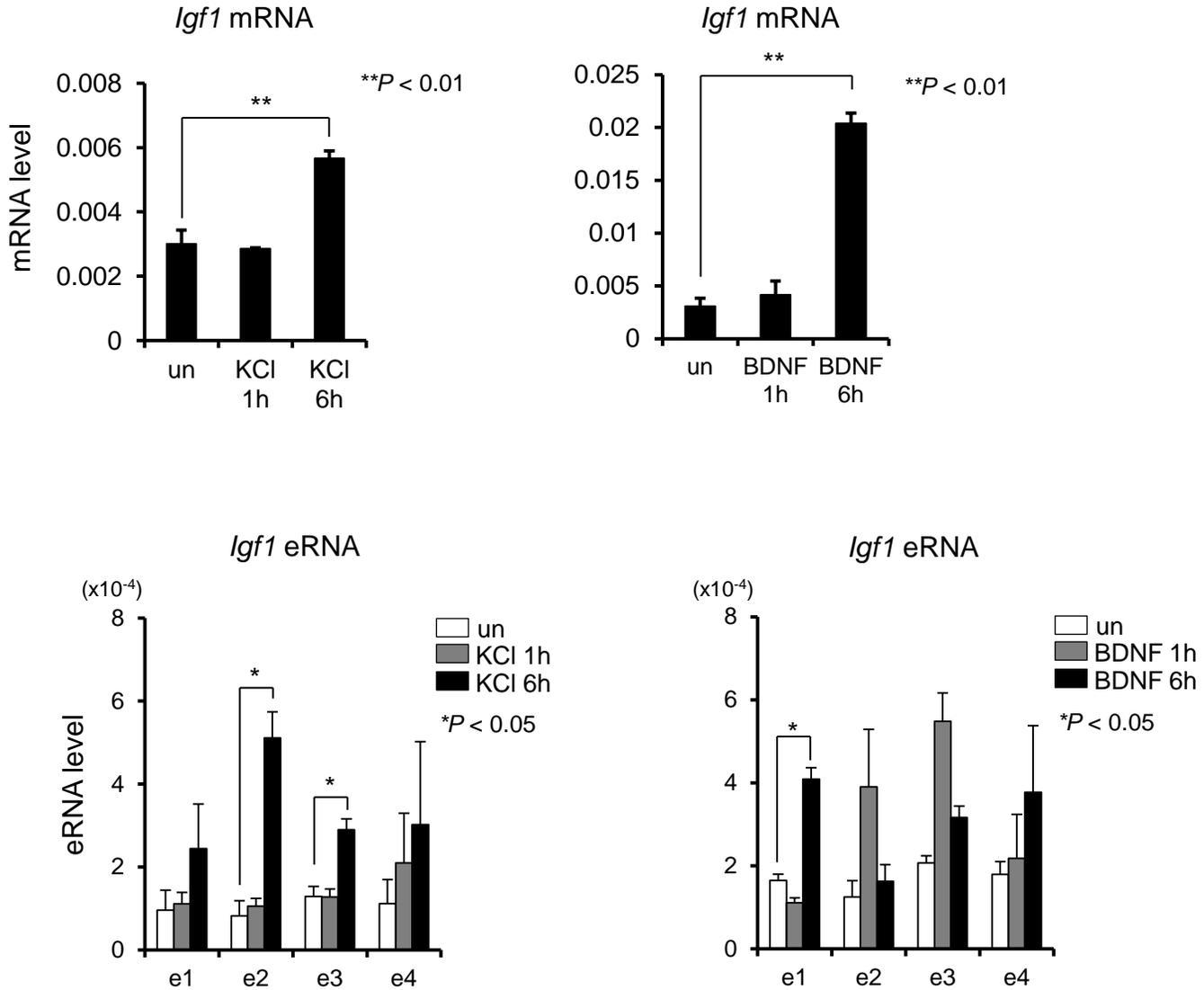
(a) H3K27ac binding profiles throughout the mouse forebrain development and KCl stimulation were adapted from the data generated by Nord *et al.* 2013 and Malik *et al.* 2014. Total RNA-seq data (1 h) was adapted from previous study by Kim *et al.* 2010. Gray vertical bars indicate the locations of the *Pelil* enhancers and the promoter, respectively. + and – indicate the presence or absence of KCl. (b) Expression of *Pelil* mRNA and eRNA in cortical neurons induced by KCl and BDNF. The induction levels of the five *Pelil* eRNAs and mRNA were measured using RT-qPCR and normalized to *Gapdh* mRNA (KCl stimulation: *Pelil* mRNA,  $P = 0.0076$ ,  $t(2) = 11.430$ ,  $F = 2.346$  [ $sdP = 0.3682$ ],  $n = 2$ , *Pelil* e2,  $P = 0.0318$ ,  $t(2) = 5.477$ ,  $F = 1.154$  [ $sdP = 0.4773$ ],  $n = 2$ ; BDNF stimulation: *Pelil* mRNA,  $P = 0.0388$ ,  $t(4) = 4.930$ ,  $F = 13.471$  [ $sdP = 0.1693$ ],  $n = 2$ , *Pelil* e1,  $P = 0.0392$ ,  $t(2) = 4.900$ ,  $F = 63.601$  [ $sdP = 0.080$ ],  $n = 2$  biological replicates). All unpaired *t*-test. Error bars indicate SEM; p values are from a two-tailed *t* test. *sdP* is the P value from comparing the standard deviations for both groups. *sdP* > 0.05 means the two SDs are not significantly different and can be used for an unpaired *t*-test.

# Supplementary Figure 14

a



b

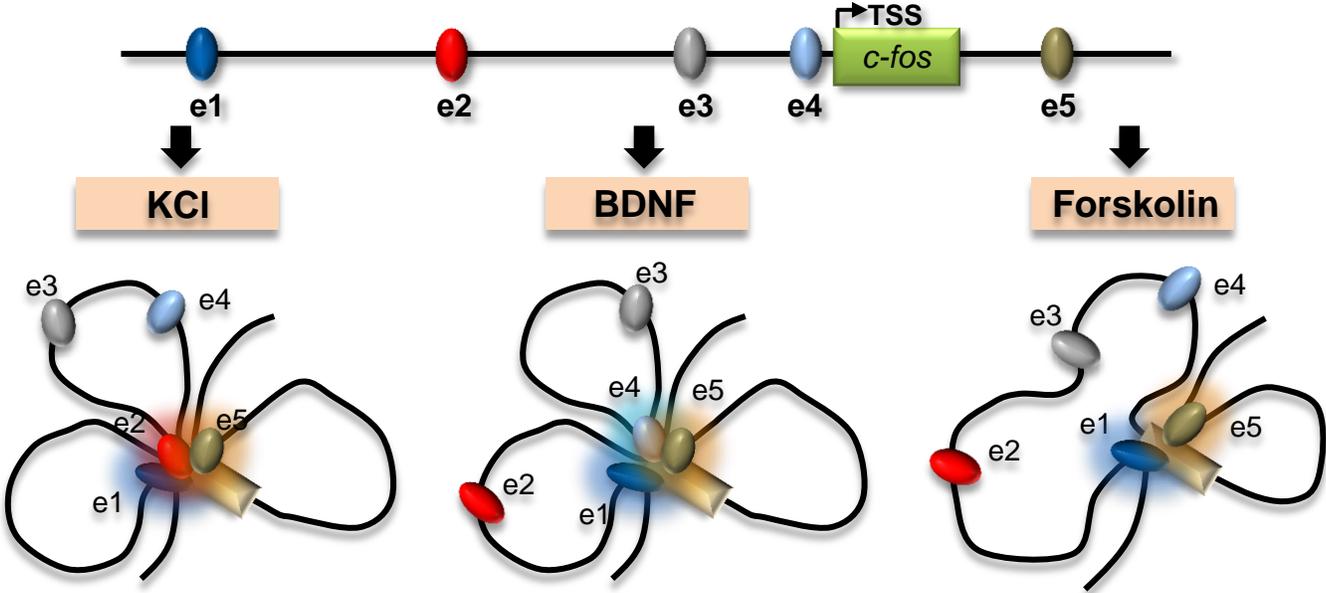


Supplementary Figure 14

**Binding profiles of the H3K27ac mark at the *Igf-1* enhancer and promoter and enhancer activities measured by eRNA analysis.**

(a) H3K27ac binding profiles throughout the mouse forebrain development and KCl stimulation were adapted from the data generated by Nord *et al.* 2013 and Malik *et al.* 2014. Total RNA-seq data (unstimulated, 1 h, 6 h) were adapted from previous study by Kim *et al.* 2010. **Gray** vertical bars indicate the locations of the *Igf-1* enhancers and the promoter, respectively. + and – indicate the presence or absence of KCl. (b) Expression of *Igf-1* mRNA and eRNA in cortical neurons induced by KCl, BDNF. The induction levels of the five *Igf-1* eRNAs and mRNA were measured using RT-qPCR and normalized to *Gapdh* mRNA (KCl stimulation: *Igf1* mRNA,  $P = 0.0317$ ,  $t(2) = 5.486$ ,  $F = 3.435$  [ $sdP = 0.3150$ ],  $n = 2$ , *Igf1* e2,  $P = 0.0280$ ,  $t(2) = 5.852$ ,  $F = 2.903$  [ $sdP = 0.3379$ ],  $n = 2$ , *Igf1* e3,  $P = 0.0465$ ,  $t(2) = 4.520$ ,  $F = 1.161$  [ $sdP = 0.4762$ ],  $n = 2$ , BDNF stimulation: *Igf1* mRNA,  $P = 0.0050$ ,  $t(2) = 14.092$ ,  $F = 1.686$  [ $sdP = 0.4178$ ],  $n = 2$ , *Igf1* e1,  $P = 0.0224$ ,  $t(2) = 6.570$ ,  $F = 3.354$  [ $sdP = 0.3182$ ],  $n = 2$  biological replicates). All unpaired *t*-test. Error bars indicate SEM; p values are from a two-tailed *t* test. *sdP* is the *P* value from comparing the standard deviations for both groups.  $sdP > 0.05$  means the two SDs are not significantly different and can be used for an unpaired *t*-test.

# Supplementary Figure 15

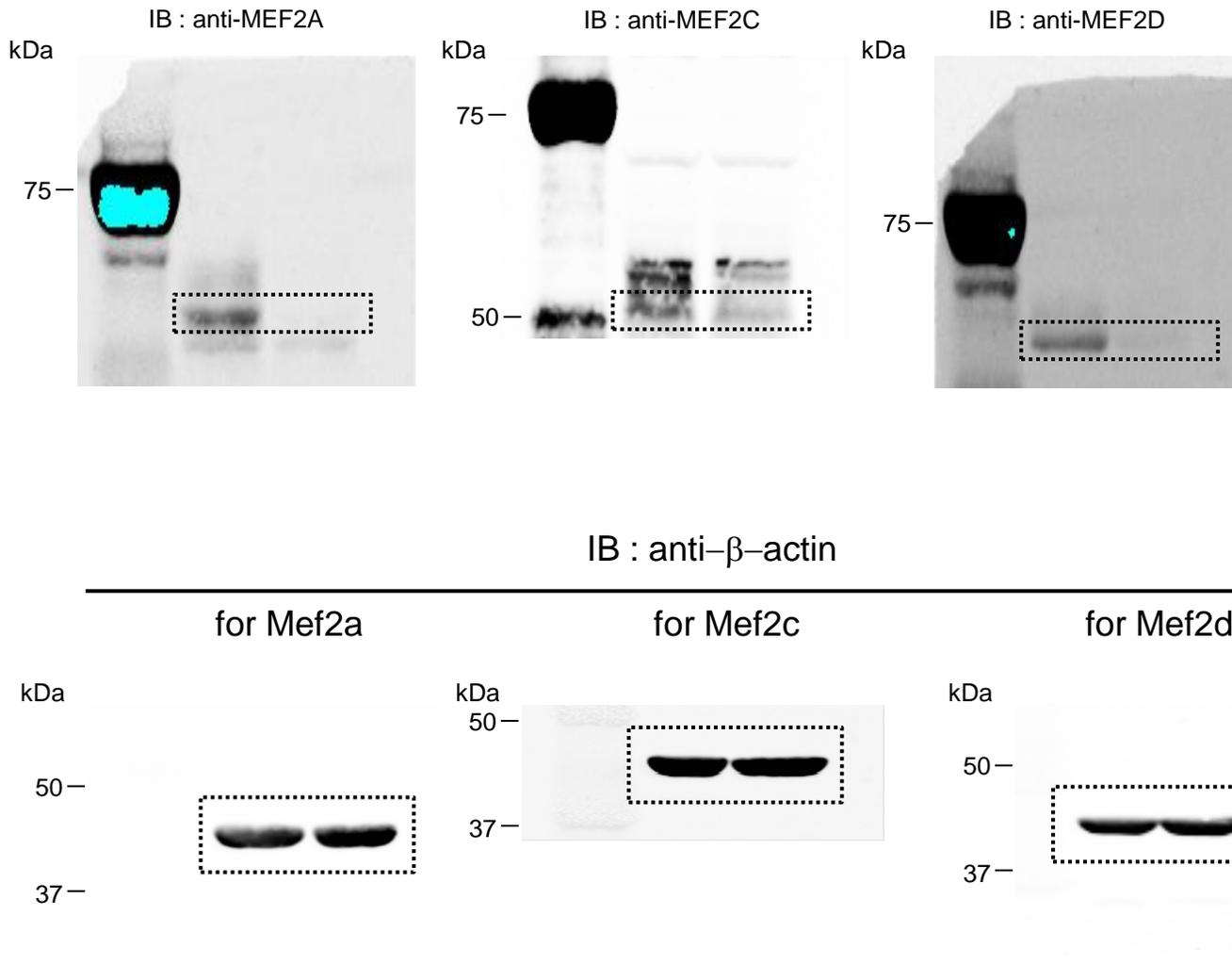


Supplementary Figure 15

**Stimulus-specific combinatorial activation of multiple *c-fos* enhancers.**

In response to different stimuli, distinct subsets of the surrounding enhancers are dynamically assembled with the promoter to mediate robust *c-fos* transcription.

# Supplementary Figure 16



## Supplementary Figure 16

Full blots of the western blots shown in the figure

This Figure contains the full blots of the western blots shown in the Supplementary Figure 6.