

Presence of unique glyoxalase III proteins in plants indicates the existence of shorter route for methylglyoxal detoxification

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Table S1 | Comparison of enzyme kinetic parameters of WT OsDJ-1C protein and its mutein C119A

Sl. no	Parameter	WT	C119A	Deviation from WT (%)
1	K_m (μM)	744	602	19
2	V_{max} ($\mu\text{mol min}^{-1}\text{mg}^{-1}$)	58	22	62
3	k_{cat} (min^{-1})	2500	945	62
4	k_{cat}/K_m ($\text{mM}^{-1}\text{min}^{-1}$)	3359	1569	53

Table S2 | List of single domain containing DJ-1 proteins in plants used in this study

Sl. No.	Name of Species	Classification	Swiss-prot Ids
1	<i>Oryza sativa subsp. japonica</i>	Monocot	A3BCD1, B9G8C7
2	<i>Hordeum vulgare var. distichum</i>	Monocot	F2D1P7, F2EG99, M0V2D0, M0WAD7, M0XZ96, M0YG87
3	<i>Triticum urartu</i>	Monocot	M7Y5X3
4	<i>Triticum aestivum</i>	Monocot	W5CKN3, W5CV43, W5DFB7, W5H8W7
5	<i>Zea mays</i>	Monocot	K7VDN4, K7VH97
6	<i>Sorghum bicolor</i>	Monocot	C5YMA2
7	<i>Arabidopsis thaliana</i>	Dicot	B3H6C6, Q56ZE0, Q56ZU8
8	<i>Medicago truncatula</i>	Dicot	B7FLB0
9	<i>Ricinus communis</i>	Dicot	B9TFX6
10	<i>Vitis vinifera</i>	Dicot	F6I594
11	<i>Glycine max</i>	Dicot	K7K833
12	<i>Brassica rapa subsp. pekinensis</i>	Dicot	Q8H6J5
13	<i>Phaseolus vulgaris</i>	Dicot	V7CEB9
14	<i>Populus trichocarpa</i>	Dicot	U7DVP5
15	<i>Genlisea aurea</i>	Angiosperms (Eudicots)	S8CKY5

Table S3 | List of *DJ-1* genes in *Arabidopsis*, along with their chromosomal locations, alternative spliced forms, CDS, polypeptide length, molecular weight, and localization (bp base pair, aa amino acid, Da daltons)

Swiss- prot ID	Gene	Protein	Chrom osome no.	Locus	CDS (bp)	Polypeptide length (aa)	Mol. Mass (Da)	pI	Localization
Q9MAH3	<i>AtDJ-1b</i>	AtDJ-1b	1	AT1G53280	1317	438	46990.1	7.95	Chloroplast
Q9ZV19	<i>AtDJ-1e</i>	AtDJ-1e.1	2	AT2G38860.1	1170	389	41668.1	5.01	Cytoplasm
		AtDJ-1e.2		AT2G38860.2	1197	398	42729.2	4.84	
		AtDJ-1e.3		AT2G38860.3	840	279	30133.6	4.37	
Q9M8R4	<i>AtDJ-1d</i>	AtDJ-1d	3	AT3G02720	1167	388	41645.3	5.08	Cytoplasm
Q9FPF0	<i>AtDJ-1a</i>	AtDJ-1a.1	3	AT3G14990.1	1179	392	41856.9	5.08	Vacuole, Plasma membrane, Chloroplast, Plasmodesmata, Cytosol
		AtDJ-1a.2		AT3G14990.2	1110	369	39478.1	5.07	
		AtDJ-1a.3		AT3G14990.3	1110	369	39478.1	5.07	
Q9MIG8	<i>AtDJ-1f</i>	AtDJ-1f	3	AT3G54600	1200	399	43141.7	5.65	Cytosol
Q56ZU8	<i>AtDJ-1c</i>	AtDJ-1c.1	4	AT4G34020.1	1419	472	50986.1	8.99	Chloroplast
		AtDJ-1c.2		AT4G34020.2	1314	437	47336.0	9.07	

Table S4 | Comparison of enzyme kinetics of conventional GLY I/II enzymes with the novel GLY III enzymes from various species

Species	Protein	K_m (μM)	k_{cat} (s^{-1})	k_{cat}/K_m ($\text{M}^{-1}\text{s}^{-1}$)	Reference
<i>Oryza sativa</i>	OsGLYI-11.2	100	70.96	7.1×10^5	1
	OsGLYII-2	254	508	2.0×10^6	2
	OsDJ-1C	744	41.66	5.6×10^4	Present study
<i>E. coli</i>	EcGLYI	27	338	1.2×10^7	3
	EcGLYII	184	53	4.7×10^5	4
	EcGLY III	1430	2.62	0.1×10^4	5
<i>Homo sapiens</i>	HsGLYI	66	500	2.3×10^7	6
	HsGLYII	187	780	4.2×10^6	7
	HsDJ-1	600	1.21	0.2×10^4	8

Table S5 | List of primer pairs and their sequence used for qRT-PCR analysis and site directed mutagenesis

Sl. no	Gene name	Primer name	Sequence (5'-3')	Product size
1	<i>OsDJ-1A</i>	DJ-1A_for1	ATGCAGATGTGTGATCTGTGA	147
		DJ-1A_rev1	TCCATCAATCAATCCATCAGCTA	
2	<i>OsDJ-1B</i>	DJ-1B_for2	GCCTGTCACCTGTGAGTGAT	177
		DJ-1B_rev2	AGACAGCGTAAACTCACCGA	
3	<i>OsDJ-1C</i>	DJ-1C_for1	CACAGCTTTGGGCTTTTCGTC	147
		DJ-1C_rev1	CAGCTCCTTCTGCTCGTACA	
4	<i>OsDJ-1D</i>	DJ-1D_for1	GCGGTTGAGAGACAGTGTGA	108
		DJ-1D_rev1	CACGGCATGAGAACAACAGC	
5	<i>OsDJ-1E</i>	DJ-1E_for1	TGAGGCATCCTTTTGCTCGT	147
		DJ-1E_rev1	ACCCAAAGGTGCAATATCCATC	
6	<i>OsDJ-1F</i>	DJ-1F_for1	CTCGCCGAGTTCATCGCTC	99
		DJ-1F_rev2	TGACAGAGAGAACAGAGAAGCA	
7	<i>eEF-1α</i>	eEF_for	TTTCACTCTTGGTGTGAAGCAGAT	100
		eEF_rev	GACTTCCTTCACGATTTTCATCGTAA	

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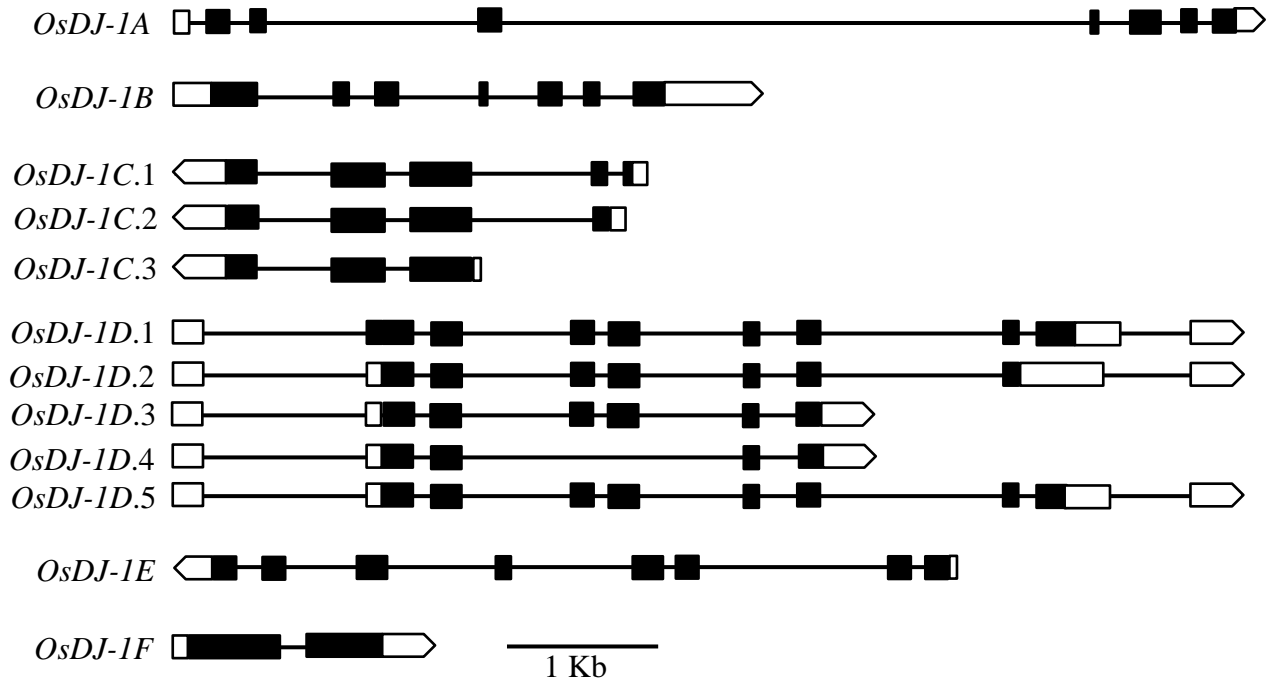


Figure S1 | Gene structures of all the *DJ-1* family members of rice including alternative spliced forms. All the exons are shown in filled black boxes and the introns are indicated by black lines. The 5'-UTR regions are shown using empty boxes and the 3'-UTR regions are shown in empty arrows that indicates the direction of the gene.

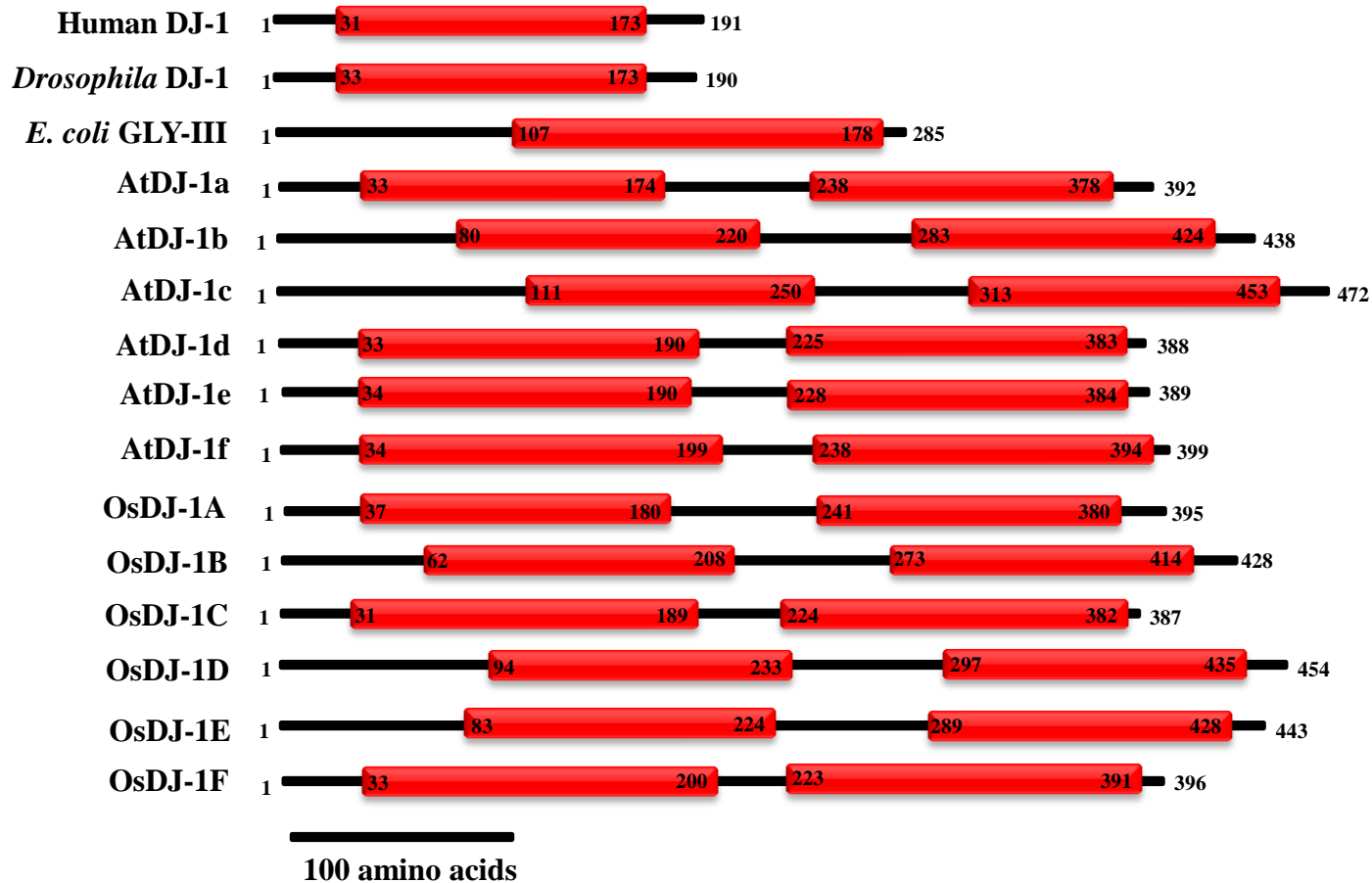


Figure S2 | Domain architecture of various GLYIII/DJ-1 proteins. All the *Arabidopsis* and rice DJ-1 proteins along with human, *Drosophila* and *E. coli* GLY-III proteins were analyzed for the presence of DJ-1 domain. All plant DJ-1 proteins possess two DJ-1 domains (represented by red box), whereas non-plant proteins possess only one DJ-1 domain. The length of full protein as well as domain(s) are indicated by exact amino acid numbers.

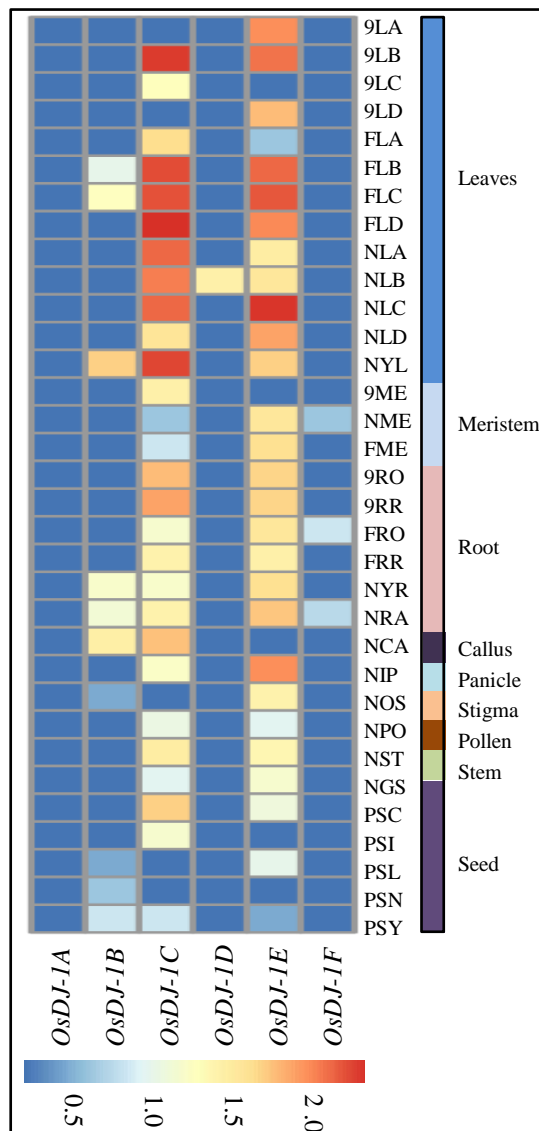


Figure S3 | Expression profiling of *OsDJ-1* genes using MPSS data in different tissues of rice. Heatmap has been generated using TIGR MeV software package and represent hierarchical clustering of average \log_2 signal values. The color bar below represents relative expression values; thereby, red color represents highest expression levels, white represents medium expression levels, and blue signifies lowest expression levels.

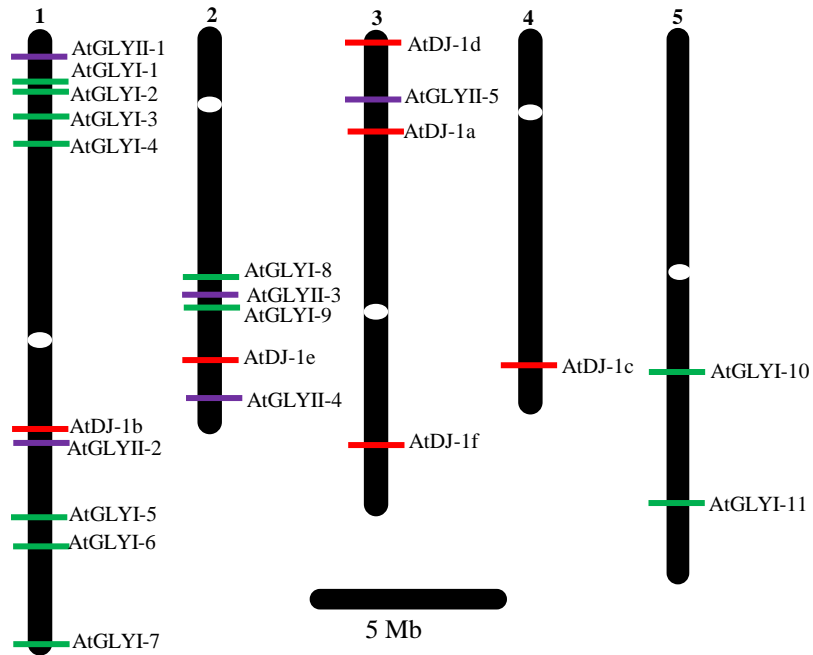


Figure S4| Chromosomal distribution of glyoxalase genes (GLYI, GLYII and DJ-1) in *Arabidopsis*. Chromosomes carrying glyoxalase genes are shown with their respective numbers at the top of the bar. Novel glyoxalase III (DJ-1) genes have been marked along with GLY I and GLY II genes. GLY I, GLY II and DJ-1 genes are marked with green, purple, and red colour boxes respectively.

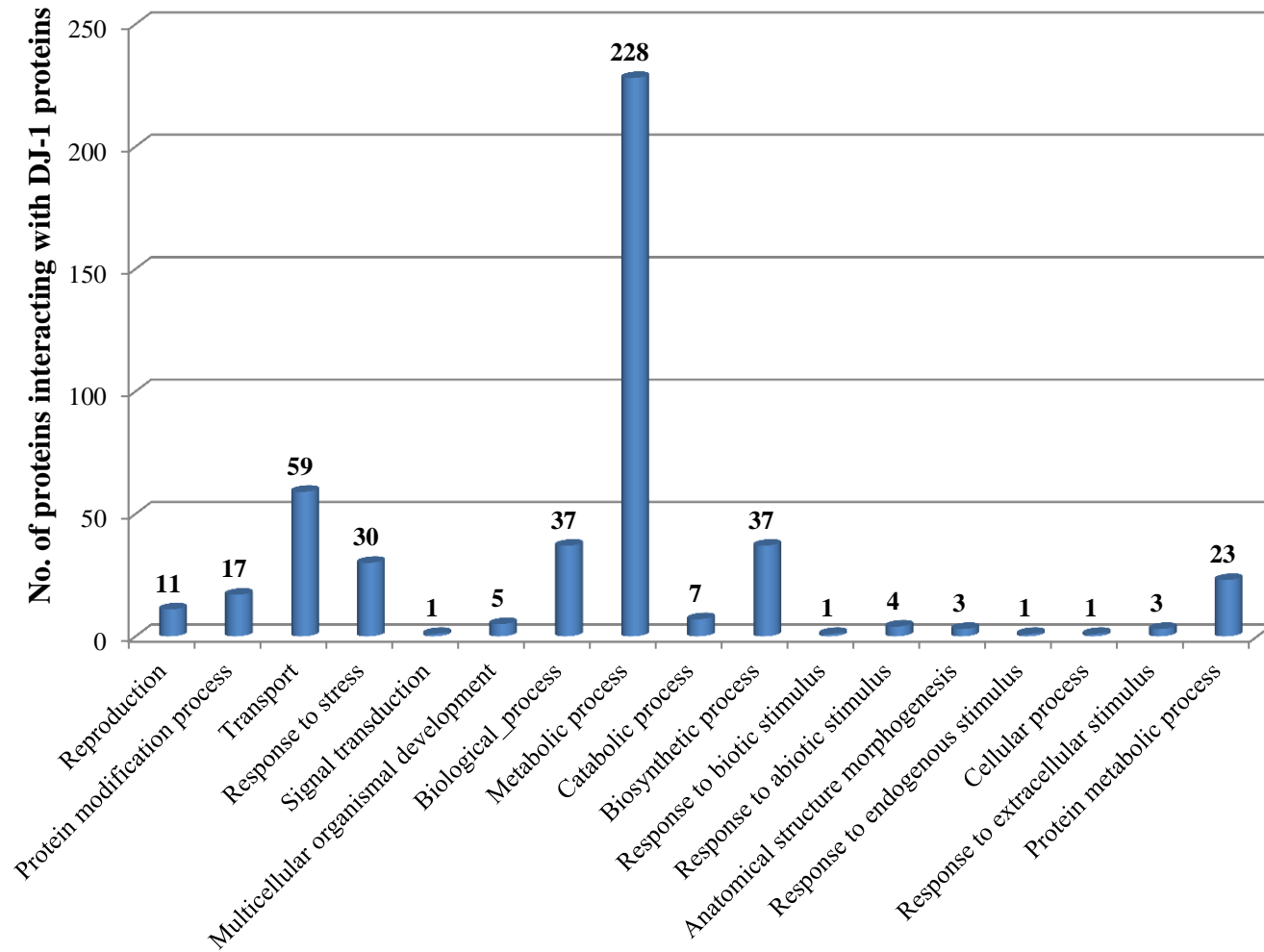


Figure S5 | *In silico* prediction of proteins interacting with OsDJ-1 proteins in rice.