

1 Comparative genome analysis of *Pseudomonas* genomes including *Populus*-
2 associated isolates

3

4 Se-Ran Jun¹, Trudy M. Wassenaar³, Intawat Nookaew², Loren Hauser², Visanu Wanchai², Miriam
5 Land², Collin Timm², Tse-Yuan S. Lu², Christopher W. Schadt², Mitchel J. Doktycz², Dale A.
6 Pelletier², David W. Ussery^{2,#}

7

8 Running Head: Comparative *Pseudomonas* genome analysis

9 ¹ Joint Institute for Computational Sciences, University of Tennessee, Knoxville, TN, USA

10 ² Biosciences Division, Oak Ridge National Laboratories, Oak Ridge, TN, USA

11 ³ Molecular Microbiology and Genomics Consultants, Zotzenheim, Germany

12

13

14 #Address correspondence to David W. Ussery

15 Oak Ridge National Laboratory

16 Biosciences Division

17 Oak Ridge, TN 37831

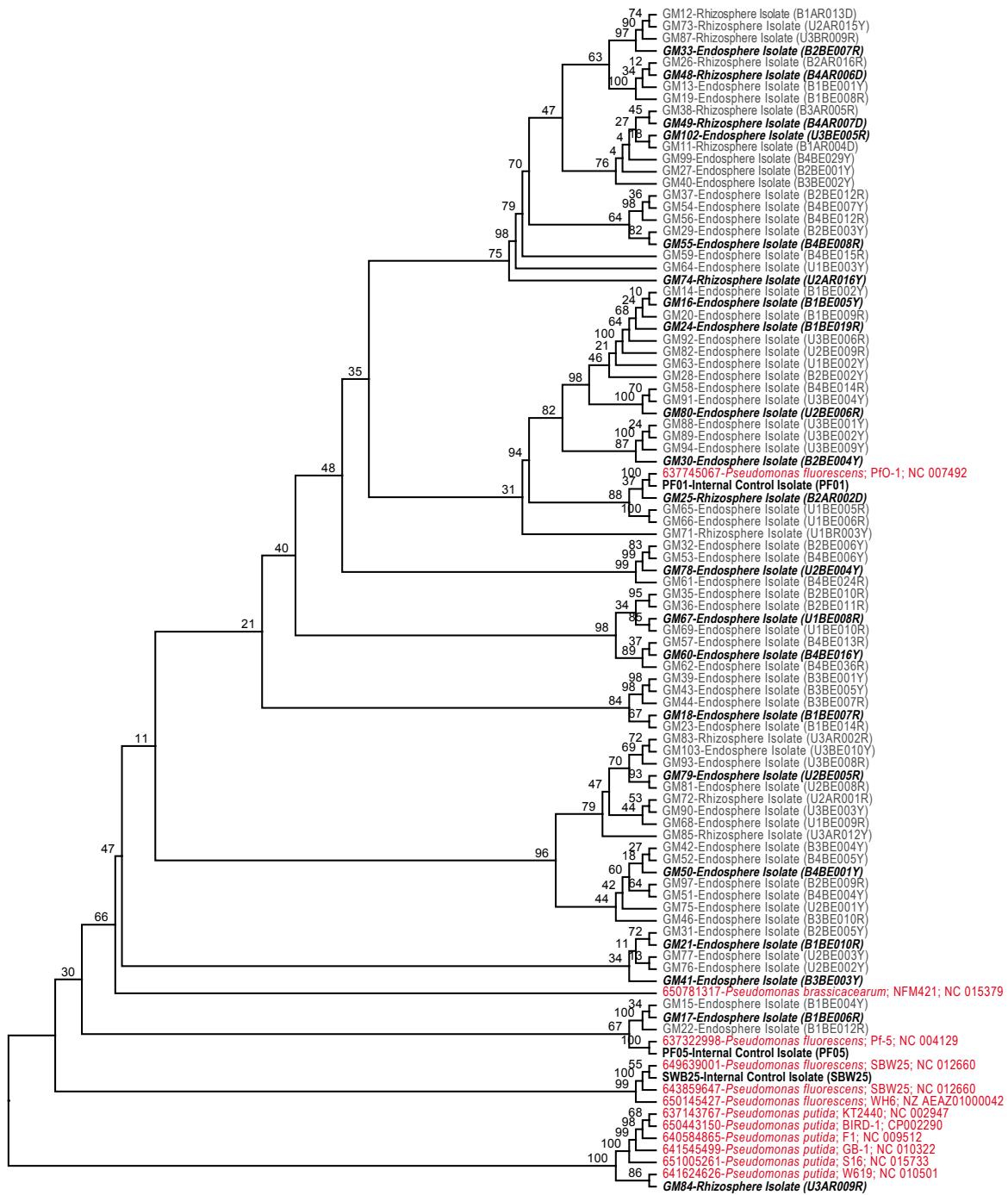
18 Phone: (865) 574-8201

19 Fax: (865) 574-3555

20 Email: usserydw@ornl.gov

21

22



25 *Populus deltoides* roots and selected references taxa (Maximum likelihood, bootstraps = 100).
26 Genes from reference isolates are listed by their GI number. Strains from the current study are
27 labeled to denote the compartment (endosphere or rhizosphere) and tree from which the
28 isolate originated. Isolates representing diverse and well-supported groups were selected for
29 complete genome sequence analysis, indicated in bold. Reference complete genomes are
30 shown in red.

31

32

33

34

35

36

37

38

39

40

41

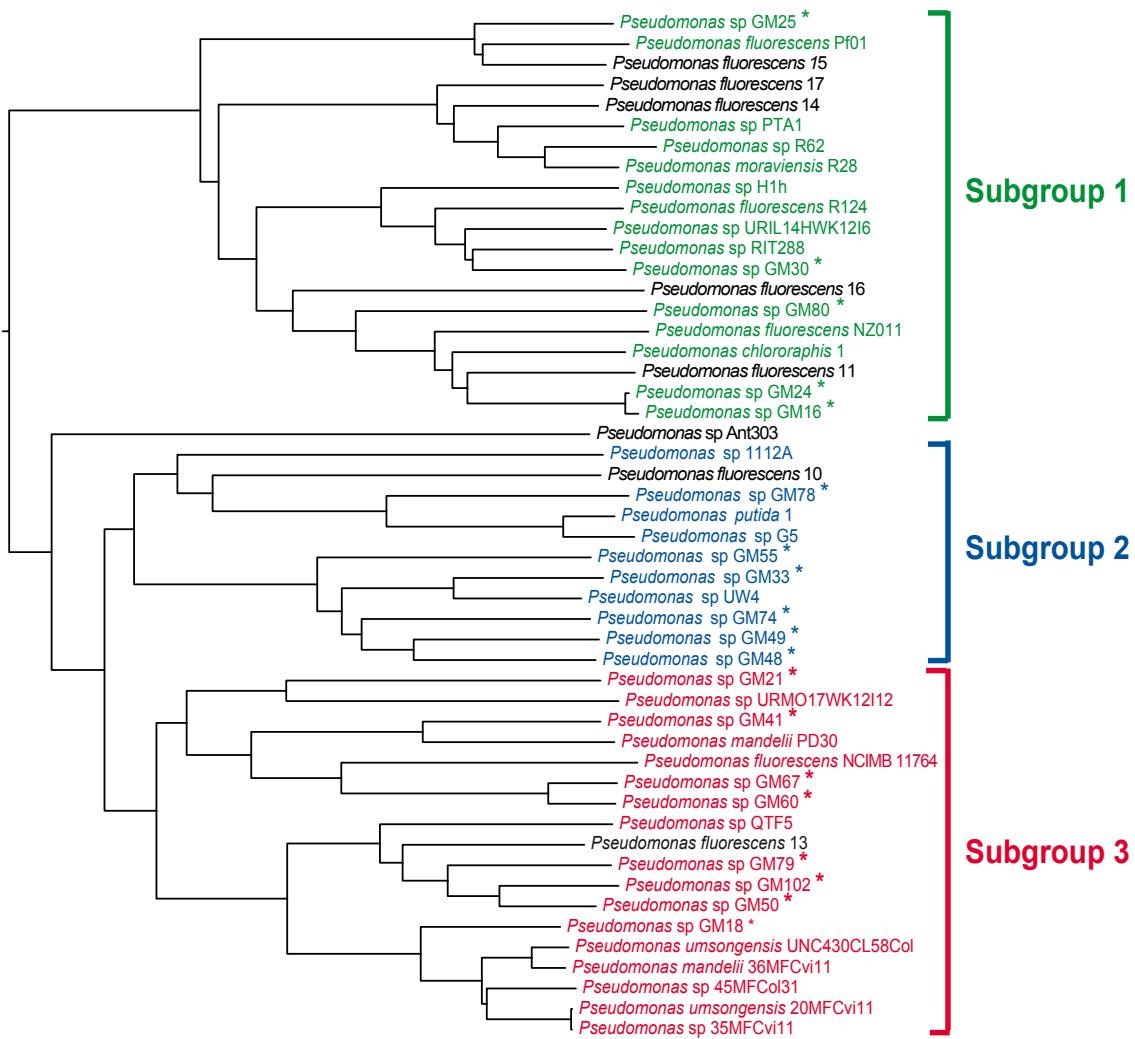
42

43

44

45

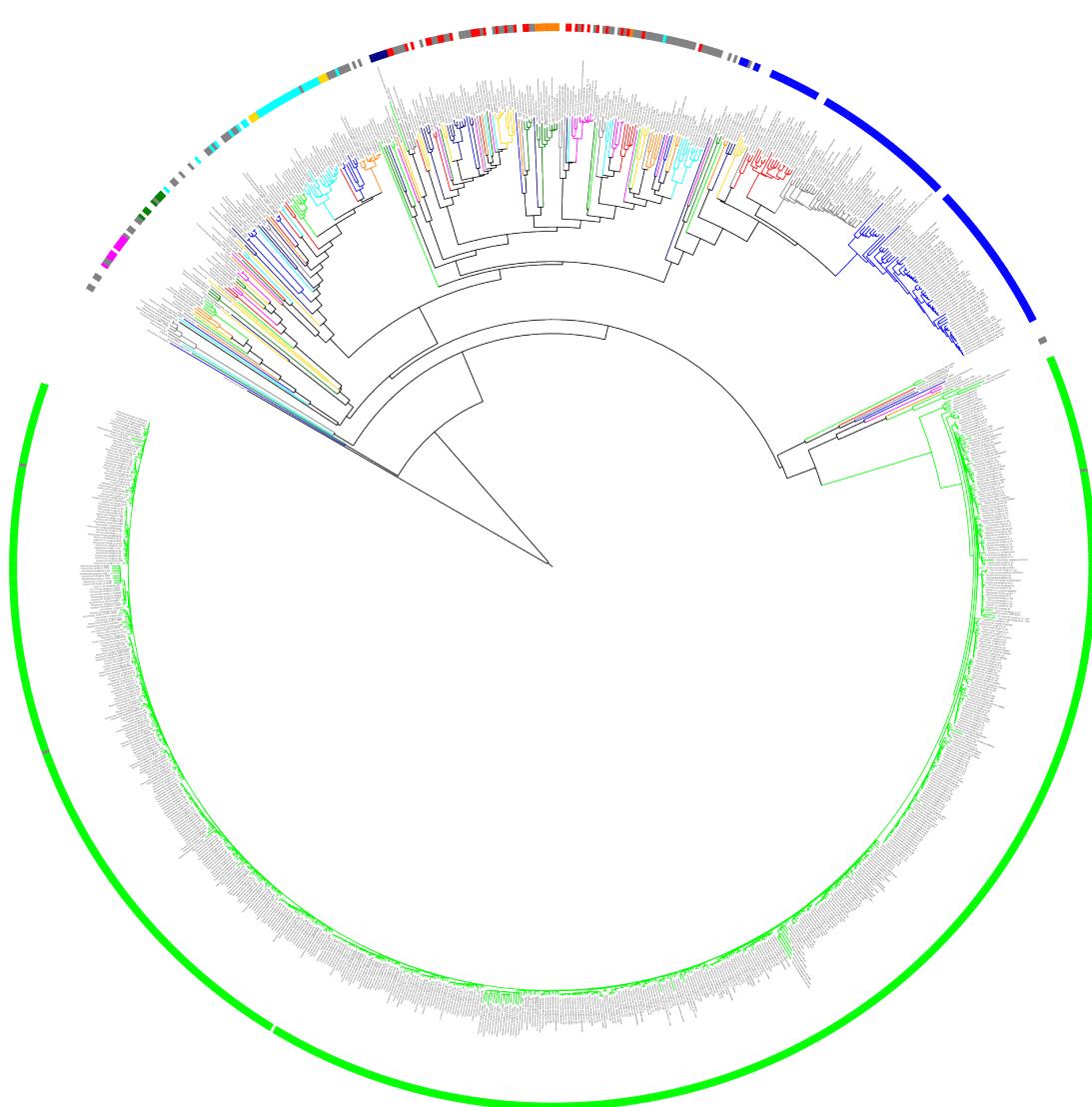
46



47

48 **Figure S2.** AAI-based tree of 19 *Populus*-associated *Pseudomonas* isolates and related
49 *Pseudomonas* species with newly sequenced *P. fluorescens* in black.

50



51

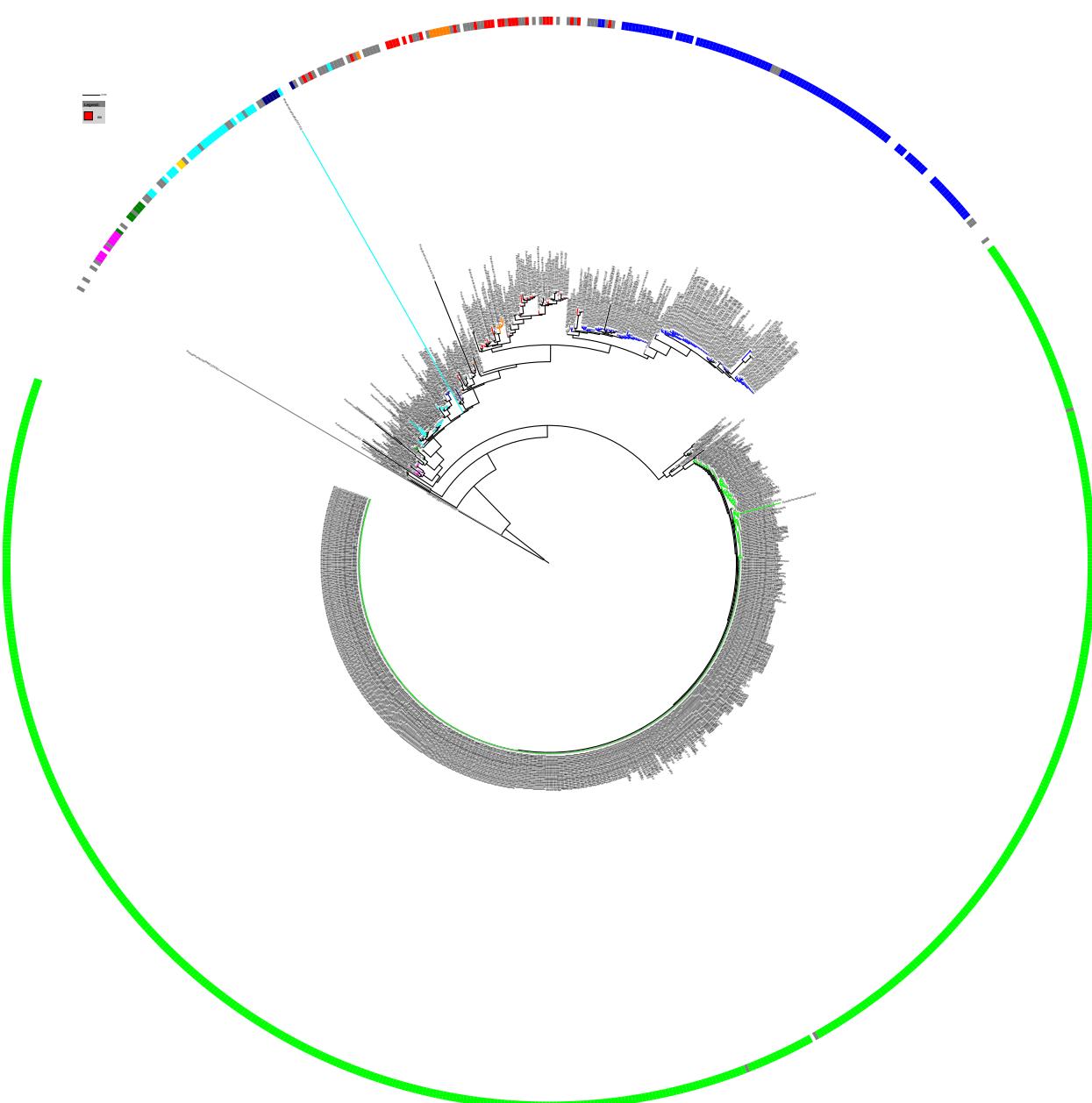
52 **Figure S3.** AAI-based tree of 1073 *Pseudomonas* genomes. The outer ring and branches are
53 colored by *Pseudomonas* species groups defined by NCBI taxonomy.

54

55

56

57



58
59 **Figure S4.** 16S-rRNA based tree. We considered only 16S rRNA genes (extracted by RNAmmer
60 v1.2 (1)) whose length is larger than 1400 nt-long, and chose one 16S rRNA gene whose length
61 is closest to 1500 when the multiple copies of 16S rRNA genes are present in the genomes,
62 resulting in 116 *Pseudomonas* without full-length 16S rRNA genes out of 1073 *Pseudomonas*
63 isolates. We aligned 16S rRNA genes using MAFFT (2), and applied FastTree (3) using a
64 combined model of generalized time-reversible and a single rate for each site (GTR+CAT model)

65 to approximate a 16S rRNA-based maximum likelihood tree. The outer ring and branches are
66 colored by *Pseudomonas* species groups defined by NCBI taxonomy.

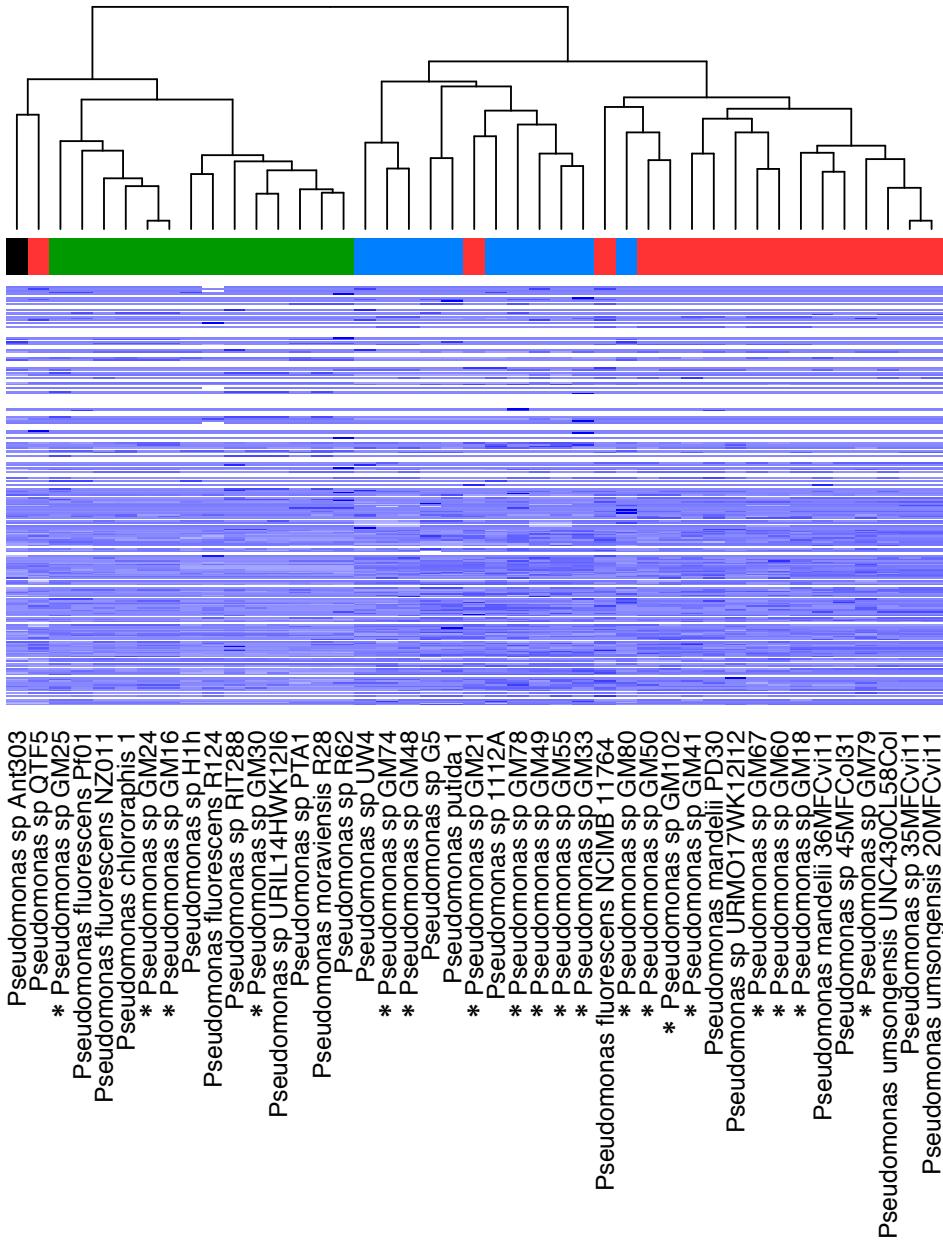
67

68

69

70

71



72

73 **Figure S5. Pathway profiles analysis of PMI *Pseudomonas* isolates.** The asterisk indicates PMI
 74 *Pseudomonas* isolates. The color code at the X-axis corresponds to three distinct subgroups
 75 identified in Figure 2.

76

77

78

79 **Table S1.** *Pseudomonas* dataset.

80

81 **Table S2.** Genomic clusters with at least five members.

Genomic cluster Index	Cluster size	Species by NCBI Taxonomy
Cluster_53	684	<i>Pseudomonas aeruginosa</i> (680) <i>Pseudomonas otitidis</i> (1) <i>Pseudomonas sp</i> (3)
Cluster_25	64	<i>Pseudomonas syringae</i> (61) <i>Pseudomonas avellanae</i> (3)
Cluster_23	33	<i>Pseudomonas syringae</i> (33)
Cluster_33	16	<i>Pseudomonas putida</i> (15) <i>Pseudomonas monteili</i> (1)
Cluster_24	16	<i>Pseudomonas syringae</i> (13) <i>Pseudomonas savastanoi</i> (3)
Cluster_32	10	<i>Pseudomonas monteili</i> (3) <i>Pseudomonas putida</i> (1) <i>Pseudomonas sp</i> (6)
Cluster_10	9	<i>Pseudomonas fluorescens</i> (4) <i>Pseudomonas brassicacearum</i> (3) <i>Pseudomonas sp</i> (2)
Cluster_13	9	<i>Pseudomonas chlororaphis</i> (8) <i>Pseudomonas sp</i> (1)
Cluster_16	8	<i>Pseudomonas fluorescens</i> (4) <i>Pseudomonas sp</i> (4)
Cluster_14	8	<i>Pseudomonas fluorescens</i> (3) <i>Pseudomonas simiae</i> (2) <i>Pseudomonas sp</i> (3)
Cluster_34	8	<i>Pseudomonas putida</i> (3) <i>Pseudomonas monteili</i> (2) <i>Pseudomonas plecoglossicida</i> (1) <i>Pseudomonas taiwanensis</i> (1) <i>Pseudomonas sp</i> (1)
Cluster_31	7	<i>Pseudomonas putida</i> (6) <i>Pseudomonas sp</i> (1)
Cluster_54	6	<i>Pseudomonas psychrotolerans</i> (1) <i>Pseudomonas oryzihabitans</i> (1) <i>Pseudomonas oleovorans</i> (1) <i>Pseudomonas sp</i> (3)
Cluster_2	6	<i>Pseudomonas umsongensis</i> (2) <i>Pseudomonas mandelii</i> (1)

		<i>Pseudomonas</i> sp (3)
Cluster_7	5	<i>Pseudomonas fluorescens</i> (1) <i>Pseudomonas</i> sp (4)
Cluster_4	5	<i>Pseudomonas</i> sp (5)

82

83 **Table S3. Membership of PMI *Pseudomonas* isolates into genomic clusters.**

PMI <i>Pseudomonas</i>	Genomic cluster index	Cluster size	<i>Pseudomonas</i> species by NCBI taxonomy
GM60	Cluster_0	2	<i>Pseudomonas</i> sp (2)
GM67	Cluster_0	2	<i>Pseudomonas</i> sp (2)
GM41	Cluster_1	2	<i>Pseudomonas mandelii</i> (1) <i>Pseudomonas</i> sp (1)
GM18	Cluster_2	6	<i>Pseudomonas umsongensis</i> (2) <i>Pseudomonas mandelii</i> (1) <i>Pseudomonas</i> sp (3)
GM50	Cluster_3	4	<i>Pseudomonas</i> sp (4)
GM79	Cluster_3	4	<i>Pseudomonas</i> sp (4)
GM102	Cluster_3	4	<i>Pseudomonas</i> sp (4)
GM33	Cluster_4	3	<i>Pseudomonas</i> sp (3)
GM48	Cluster_4	5	<i>Pseudomonas</i> sp (5)
GM49	Cluster_4	5	<i>Pseudomonas</i> sp (5)
GM74	Cluster_4	5	<i>Pseudomonas</i> sp (5)
GM78	Cluster_5	3	<i>Pseudomonas putida</i> (1) <i>Pseudomonas</i> sp (2)
GM16	Cluster_6	4	<i>Pseudomonas chlororaphis</i> (1) <i>Pseudomonas fluorescens</i> (1) <i>Pseudomonas</i> sp (2)
GM24	Cluster_6	4	<i>Pseudomonas chlororaphis</i> (1) <i>Pseudomonas fluorescens</i> (1) <i>Pseudomonas</i> sp (2)
GM30	Cluster_7	5	<i>Pseudomonas</i> sp (4) <i>Pseudomonas fluorescens</i> (1)
GM25	Cluster_8	2	<i>Pseudomonas fluorescens</i> (1) <i>Pseudomonas</i> sp (1)
GM17	Cluster_13	9	<i>Pseudomonas chlororaphis</i> (8) <i>Pseudomonas</i> sp (1)
GM55	Singleton	1	<i>Pseudomonas</i> sp (1)
GM84	Singleton	1	<i>Pseudomonas</i> sp (1)
GM80	Singleton	1	<i>Pseudomonas</i> sp (1)
GM21	Singleton	1	<i>Pseudomonas</i> sp (1)

84

85 **Table S4.** Subgroup-specific genes with annotation.

86

87 **REFERENCES**

88

89 1. **Lagesen K, Hallin P, Rodland EA, Staerfeldt H-H, Rognes T, Ussery DW.** 2007.

90 RNAmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic acids
91 research **35**:3100-3108.

92 2. **Katoh K, Standley DM.** 2013. MAFFT multiple sequence alignment software version 7:
93 improvements in performance and usability. Molecular biology and evolution **30**:772-
94 780.

95 3. **Price MN, Dehal PS, Arkin AP.** 2010. FastTree 2--approximately maximum-likelihood
96 trees for large alignments. PloS one **5**:e9490.

97