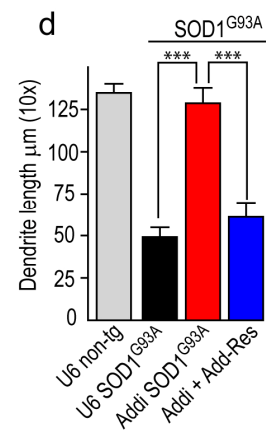
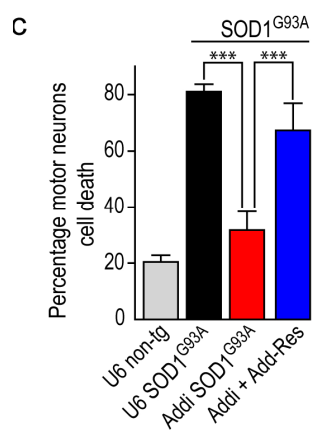
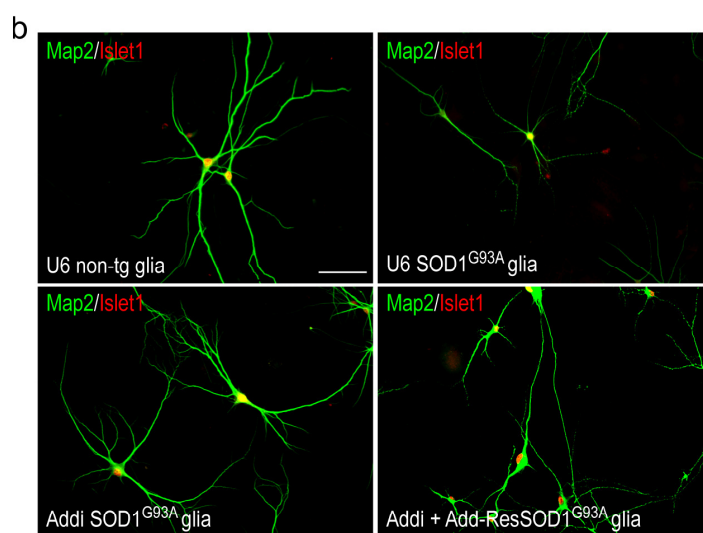
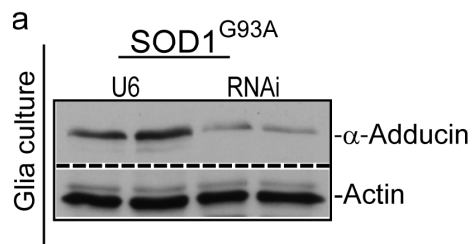


Supplementary Figure 1

α -Adducin in spinal cord is upregulated in symptomatic SOD1^{G93A} spinal cord within astrocytes.

(a) Immunoblots for α -Adducin protein at 60 and 90 day old SOD1^{G93A} mice show α -Adducin is upregulated at 90 days. (b) Immunohistochemistry with in sections of the lumbar spinal cord from symptomatic SOD1^{G93A} mice displays Ser436-phosphorylated α -Adducin does not co-localize with the motor neuron marker (SMi32). Arrowheads indicate motor neurons; scale bar 50 μ m. (c) Immunohistochemistry from sections of control wild type lumbar spinal cord displays Ser436-phosphorylated α -Adducin does not co-localize with the motor neuron marker (SMi32); *upper panels*. Arrowheads indicate motor neurons. Immunohistochemistry from sections of control wild type lumbar spinal cord displays Ser436-phosphorylated α -Adducin co-localize with the astrocyte marker (GFAP) lower panels. Scale bar 50 μ m. (a) are cropped; full length images are presented in **Supplementary Figure 11**.

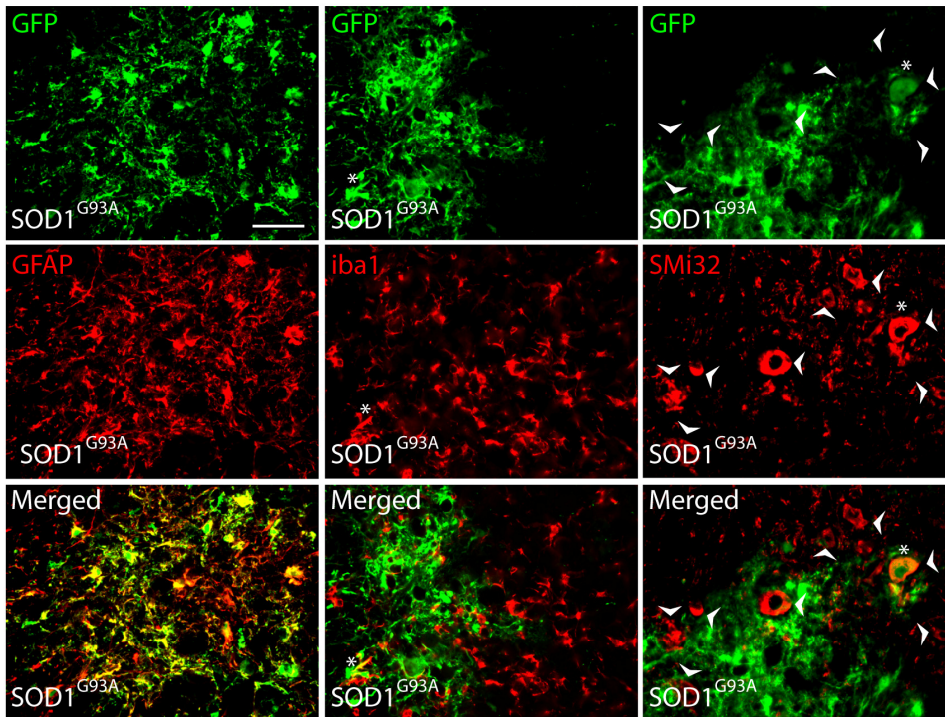


Supplementary Figure 2

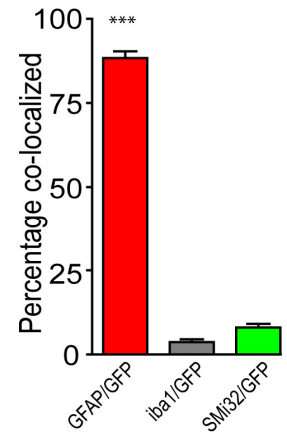
Expression of a RNAi-resistant form of α -Adducin in SOD1^{G93A} astrocytes restores the ability of SOD1^{G93A} astrocytes to induce non-cell autonomous motor neuron cell death.

(a) Knockdown of α -Adducin relative to control U6 in astrocytes (b) Co-cultured astrocytes and motor neurons were subjected to immunocytochemistry with the motor neuron nuclear protein Islet1 (red) and the dendrite protein MAP2 (green); scale bar 50 μ m. Wild type astrocytes transfected with the control U6 or α -Adducin RNAi plasmid had little or no effect on motor primary motor neurons cell death or dendrite abnormalities (*upper left panel*); quantified (c and d). Control U6 SOD1^{G93A} astrocytes induced non-cell autonomous motor neuron cell death and dendrite abnormalities (*upper right panel*); quantified (c and d). Knockdown of α -Adducin in SOD1^{G93A} astrocytes protected motor neurons against the non-cell autonomous cell death and dendrite abnormalities (*lower left panel*); quantified (c and d). Expressions of an RNAi-resistant form of α -Adducin (Add-Res) in the background of α -Adducin RNAi in SOD1^{G93A} astrocytes restored the ability of the SOD1^{G93A} astrocytes to induce non-cell autonomous cell death and dendrite abnormalities in motor neurons (*lower right panel*); quantified (d and e). All data in bar charts show mean \pm s.e.m (***p<0.001; unpaired t-test). (a) are cropped; full length images are presented in **Supplementary Figure 11**.

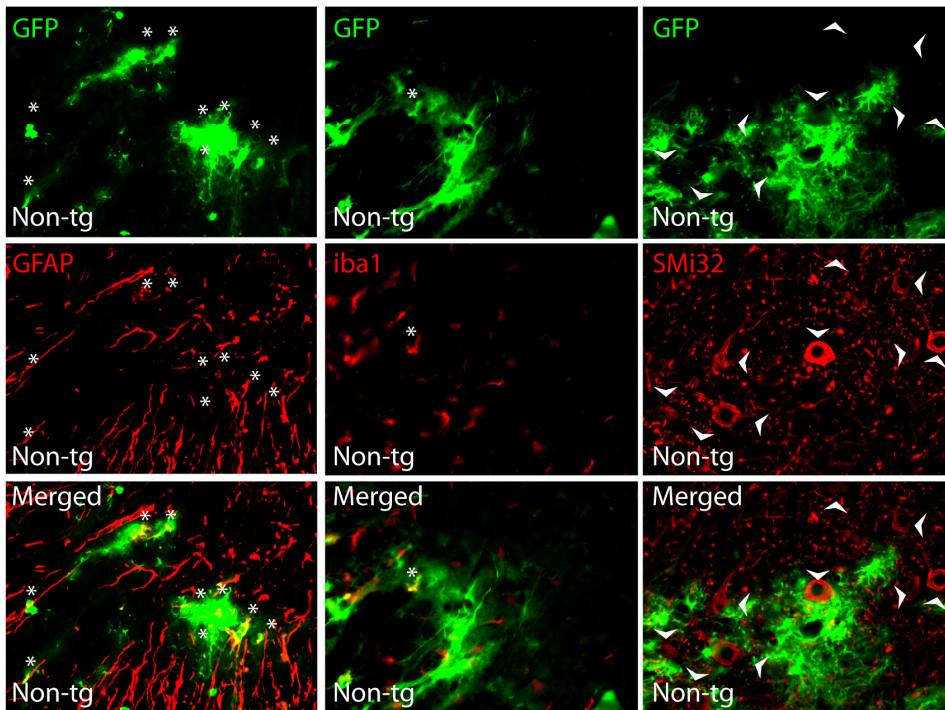
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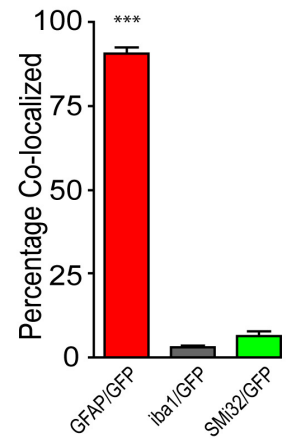
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c



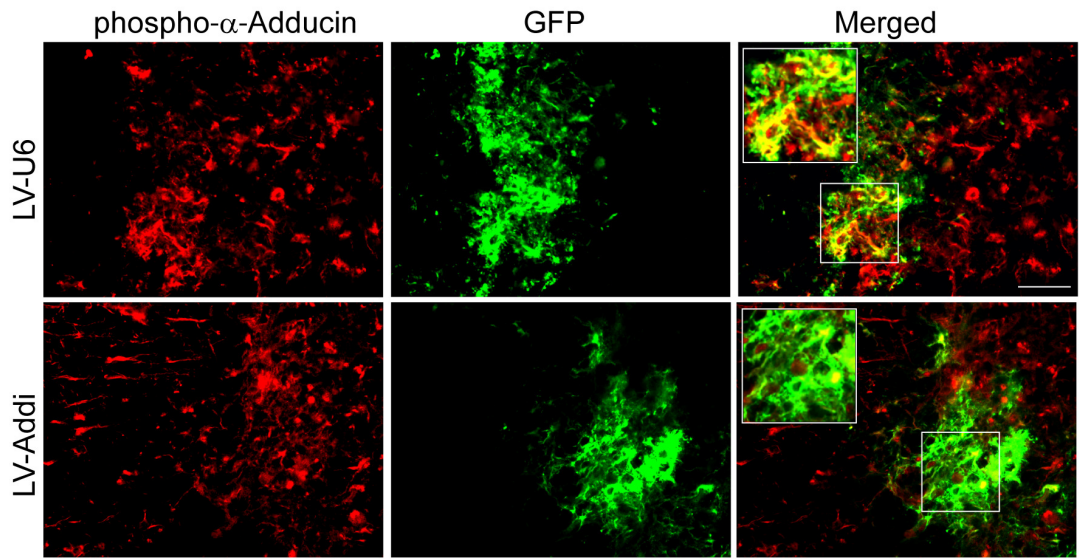
d



Supplementary Figure 3

Lentiviral mediated knockdown *in vivo* predominately target astrocytes.

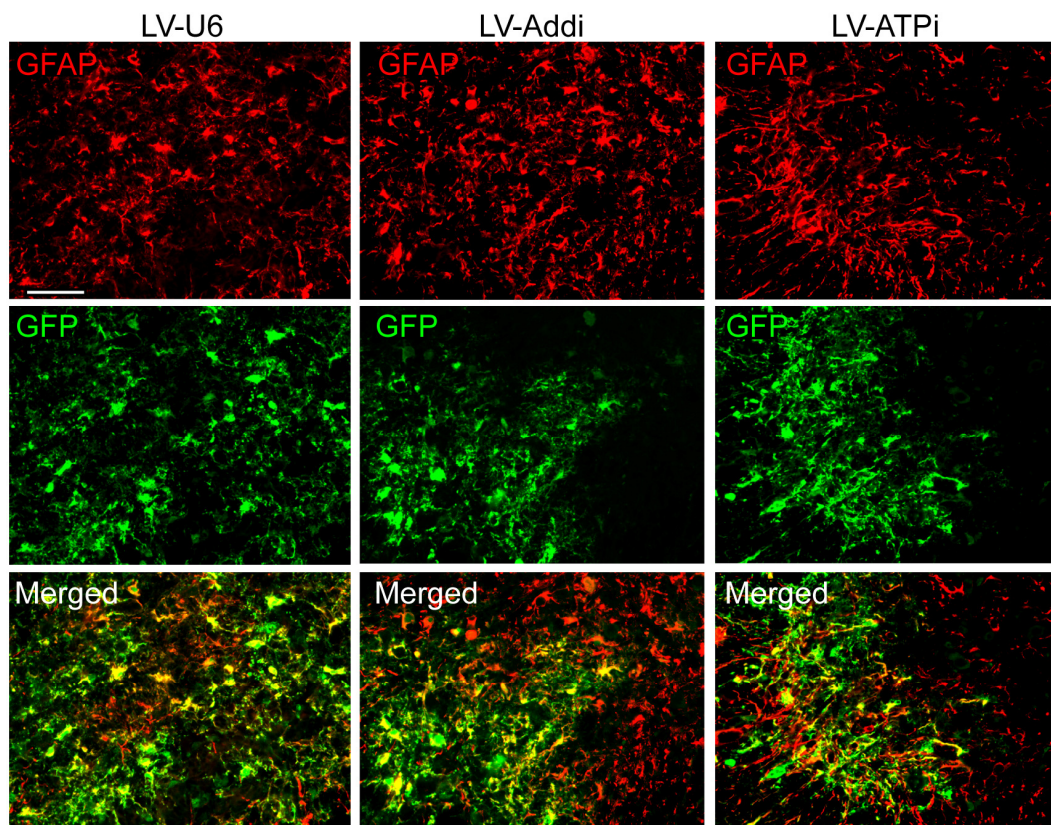
(a) Immunohistochemistry with GFP in sections of the lumbar spinal cord from SOD1^{G93A} mice displaying percent GFP positive astrocytes (GFAP), microglia (Iba1) and motor neurons (SMi32) 30 days post-injection. Arrowheads indicate motor neurons; scale bar 50 μ m. **(b)** Quantifications of percent GFP positive cells revealed lentivirus predominately target astrocytes; n= \sim 300 per cell type/three. All data in bar charts show \pm s.e.m (***)p<0.001; ANOVA). **(c)** Immunohistochemistry with GFP in sections of the lumbar spinal cord from wild type mice displaying percent GFP positive astrocytes (GFAP), microglia (Iba1) and motor neurons (SMi32) 30 days post-injection. Arrowheads indicate motor neurons; scale bar 50 μ m. **(d)** Quantifications of percent GFP positive cells revealed lentivirus predominately target astrocytes; n= \sim 300 per cell type/three. All data in bar charts show \pm s.e.m (***)p<0.001; unpaired t-test).



Supplementary Figure 4

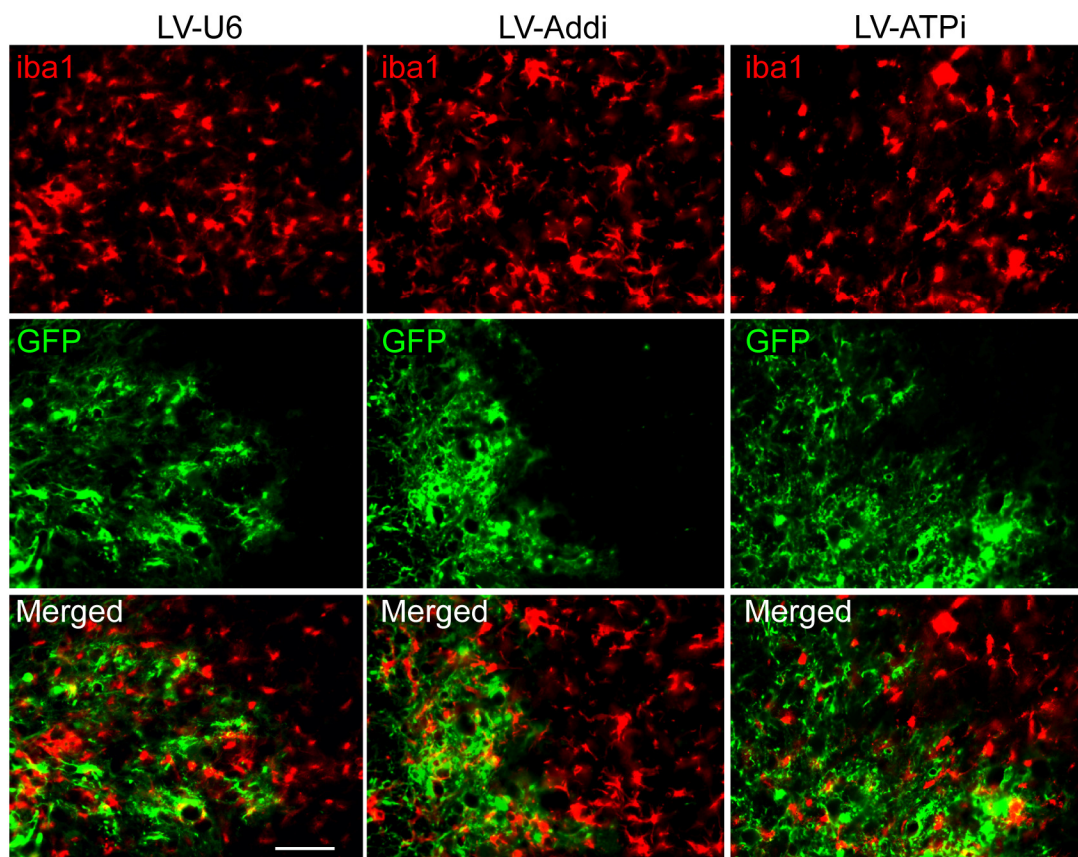
Knockdown of α -Adducin in SOD1^{G93A} mice decreases immunoreactivity for phosphorylated Ser436- α -Adducin.

Spinal cord from SOD1^{G93A} mice injected intraspinally with lentivirus expressing short hairpin RNAs targeting α -Adducin and encoding GFP (LV-Addi) or the corresponding control U6 (LV-U6) were subjected to immunohistochemistry using GFP and phospho- α -Adducin (red) antibodies. Knockdown of α -Adducin (LV-Addi) led to a decreased in immunoreactivity of phospho- α -Adducin within the GFP-labeled ventral horn as compared to control U6 (LV-U6) injected ventral horn; scale bar 50 μ m.



Supplementary Figure 5

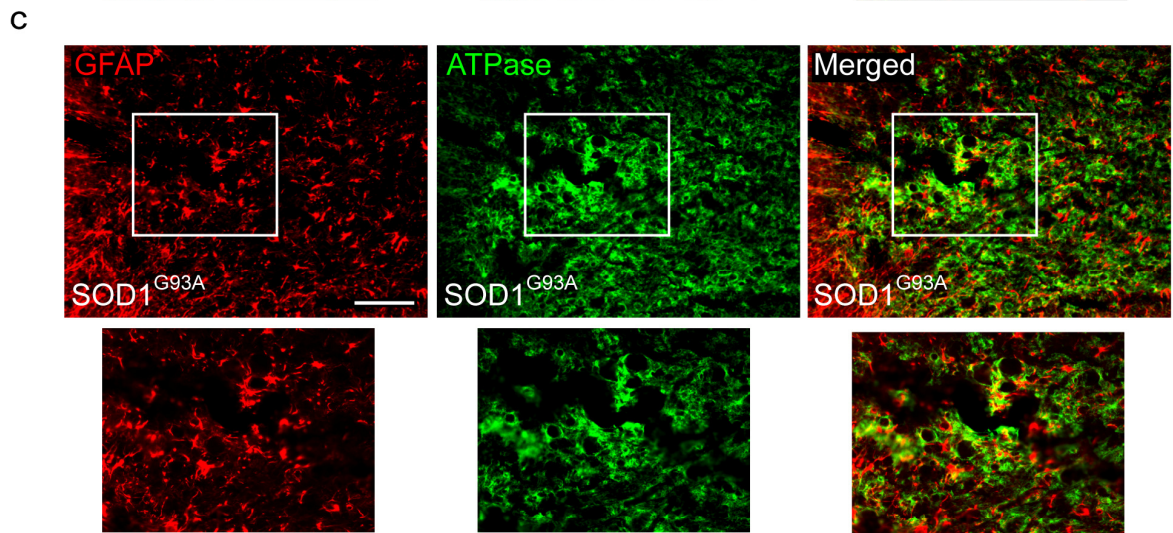
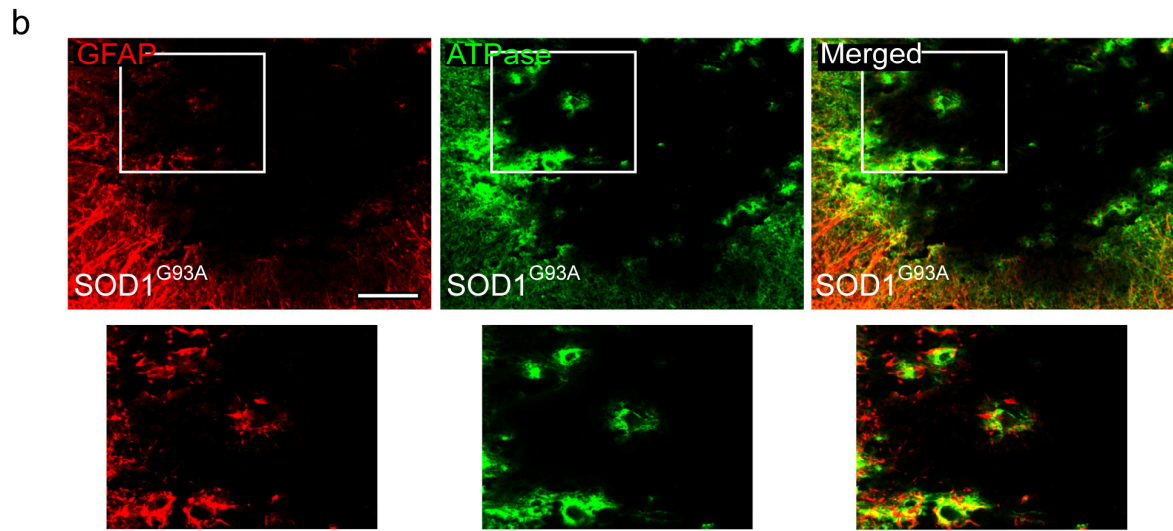
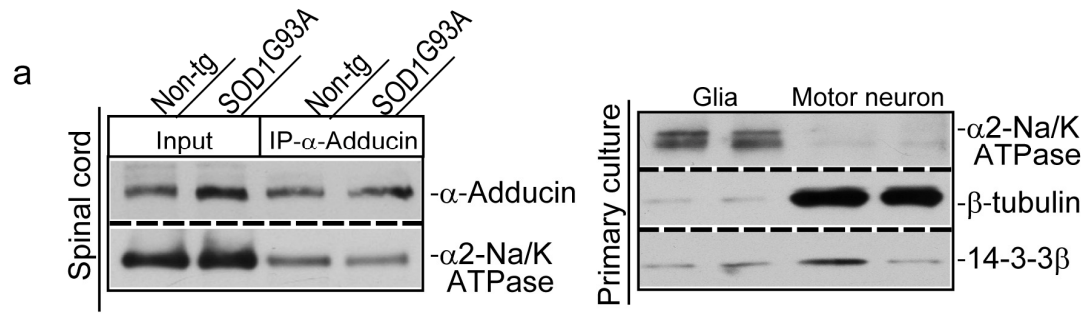
Knockdown of α -Adducin or α 2-Na/K ATPase in SOD1^{G93A} mice do not alter gliosis in the spinal cord. Spinal cord from SOD1^{G93A} mice injected intraspinally with lentivirus expressing short hairpin RNAs targeting α -Adducin or α 2-Na/K ATPase or the corresponding control U6 virus were subjected to immunohistochemistry using GFP and the GFAP (red) antibodies. Knockdown of α -Adducin (LV-Addi) or α 2-Na/K ATPase (LV-ATPi) had little or no effect on the presence or abundance of astrocytes within the GFP-labeled ventral horn; scale bar 50 μ m.



Supplementary Figure 6

Knockdown of α -Adducin or α 2-Na/K ATPase in SOD1^{G93A} mice do not alter microgliosis in spinal cord. Spinal cord from SOD1^{G93A} mice injected intraspinally with lentivirus expressing short hairpin RNAs targeting

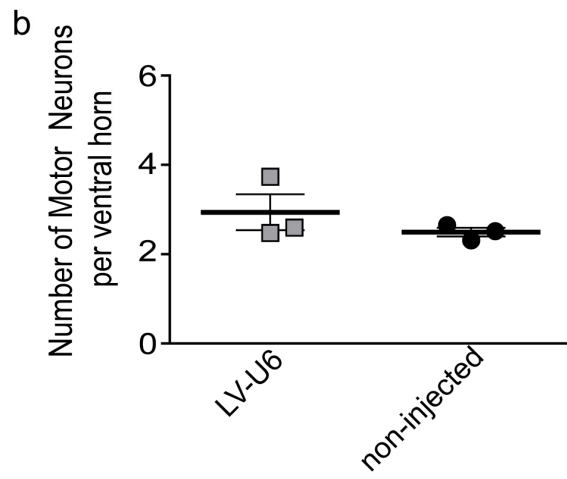
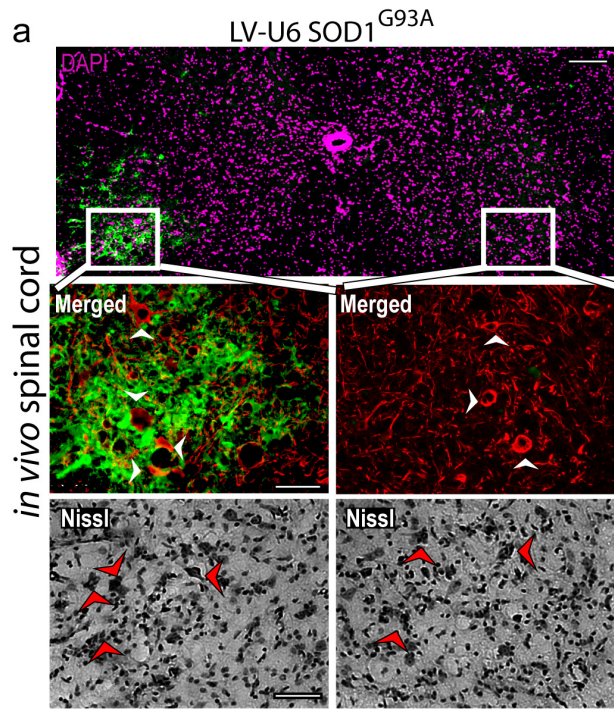
α -Adducin or α 2-Na/K ATPase or the corresponding control U6 virus were subjected to immunohistochemistry using GFP and the Iba1 (red) antibodies. Knockdown of α -Adducin (LV-Addi) or α 2-Na/K ATPase (LV-ATPi) had little or no effect on the presence or abundance of microglia within the GFP-labeled ventral horn; scale bar 50 μ m.



Supplementary Figure 7

α 2-Na/K ATPase co-immunoprecipitates with α -Adducin in spinal cord lysates and is specifically upregulated in astrocytes in symptomatic SOD1^{G93A} mice.

(a) Immunoblots show immunoprecipitated α -Adducin from SOD1^{G93A} and control wild type spinal cord lysates subjected to immunoblotting with the α -Adducin and α 2-Na/K ATPase antibodies following glycine elution, confirming α 2-Na/K ATPase as an interactor of α -Adducin (*left panels*). Immunoblots show α 2-Na/K ATPase is predominately expressed in primary glial cultures relative to primary motor neuron cultures enriched with the neuron marker β -tubulin. 14-3-3 β is used as an internal control (*right panel*). (b) Immunohistochemistry with astrocyte marker GFAP and α 2-Na/K ATPase antibody in sections of the lumbar spinal cord from SOD1^{G93A} mice at 60 days displays α 2-Na/K ATPase expression within astrocytes; scale bar 50 μ m. (c) Immunohistochemistry with GFAP and α 2-Na/K ATPase antibody in sections of the lumbar spinal cord from symptomatic SOD1^{G93A} mice at 120 days displays upregulation of α 2-Na/K ATPase expression within astrocytes; scale bar 50 μ m. (a) are cropped; full length images are presented in **Supplementary Figure 11**.



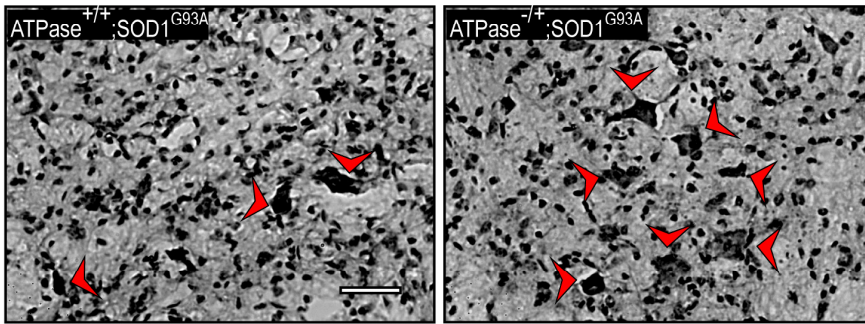
Supplementary Figure 8

Intraspinal injection of control lentivirus in SOD1^{G93A} mice had no effect on motor neuron survival.

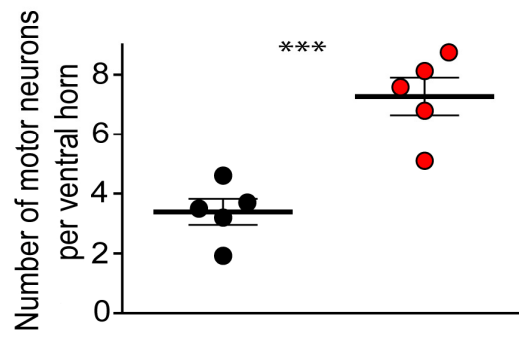
(a) Spinal cord from end stage SOD1^{G93A} mice injected at age 90 days intraspinaly with the control lentivirus encoding GFP (LV-U6 SOD1^{G93A}) was subjected to immunohistochemistry at end stage. End stage was defined as a time point at which the animal was unable to upright itself within 30s of placement on its side.

Immunohistochemistry with GFP in SOD1^{G93A} lumbar sections revealed delivery of control injected virus (LV-U6) into the ventral horn; scale bar 100 μ m. Alternating GFP positive sections were subjected to immunohistochemistry using the GFP antibody and the neurofilament-SMi32 antibody (red), a motor neuron marker, or Nissl stained (*lower panels*) for quantification of surviving motor neurons within GFP-labeled injected ventral horn and contralateral non-injected ventral horn ($n \geq 20$ sections per animal); scale bar 50 μ m. Control LV-U6 SOD1^{G93A} mice ($n=3$) displayed equivalent degeneration of motor neurons within injected GFP-labeled ventral horn and non-injected contralateral ventral horn. Arrowheads indicate surviving motor neurons; quantification shown in (b).

a



b

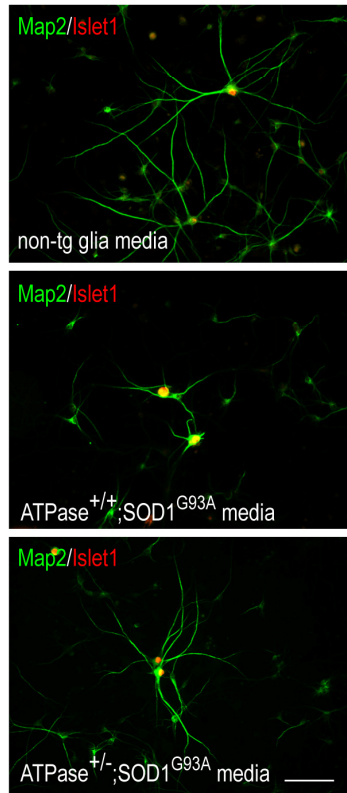


Supplementary Figure 9

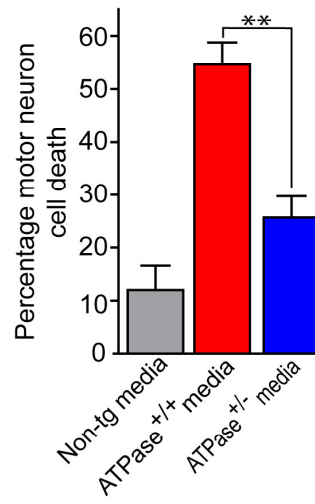
Heterozygous disruption of the $\alpha 2$ -Na/K ATPase gene in SOD1^{G93A} mice delays motor neuron degeneration.

(a) Nissl stained sections from endstage control SOD1^{G93A} mice (n=5) and aged-matched SOD1^{G93A} littermates heterozygous-null for the $\alpha 2$ -Na/K ATPase allele (n=5) displayed more than twice the number of motor neurons in ATPase^{+/-};SOD1^{G93A} than control SOD1^{G93A} mice. Arrow heads indicate surviving motor neurons; quantification shown in (b); scale bar 50 μ m (**p<0.001; unpaired t-test).

a



b



Supplementary Figure 10

Condition media from heterozygous-null from $\alpha 2$ -Na/K ATPase SOD1^{G93A} astrocytes is neuroprotective.

(a) Precondition media from wild type, SOD1^{G93A}, and heterozygous-null $\alpha 2$ -Na/K ATPase; SOD1^{G93A} astrocytes were exposed to motor neurons and subjected to immunocytochemistry with antibodies recognizing the motor neuron nuclear protein Islet1 (red) and the dendrite protein MAP2 (green); scale bar 50 μ m.

Precondition media from wild type astrocytes had little or no effect on motor neuron survival (*upper panels*); quantified (b). Preconditioned medium from SOD1^{G93A} astrocytes induced non-cell autonomous motor neuron cell death (*middle*); quantified (b). Preconditioned medium from heterozygous-null $\alpha 2$ -Na/K ATPase; SOD1^{G93A} astrocytes protected motor neurons against the non-cell autonomous cell death (*lower panel*); All data in bar charts show mean \pm s.e.m (***) $p < 0.001$; unpaired t-test).

Fig. 1a

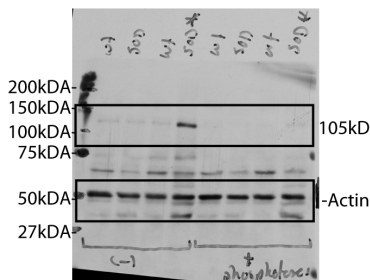


Fig. 1b

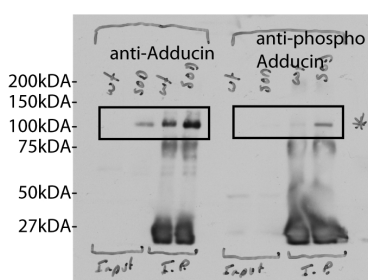


Fig. 1c

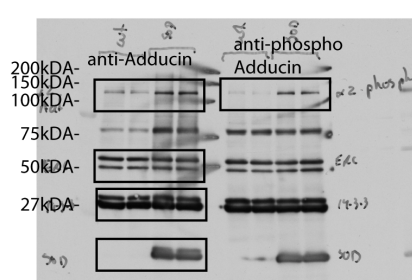


Fig. 1d

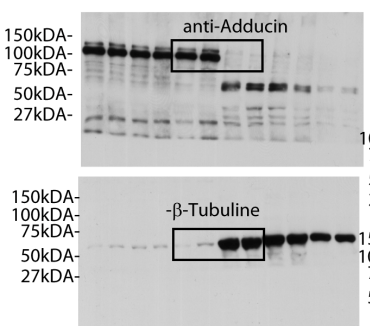


Fig. 3a

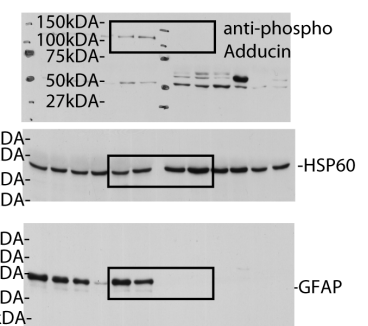


Fig. 3a

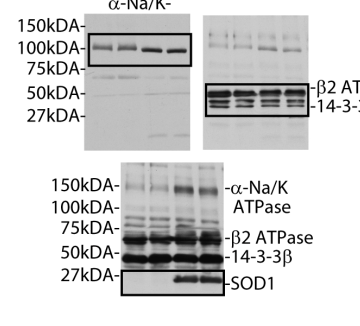


Fig. 3b

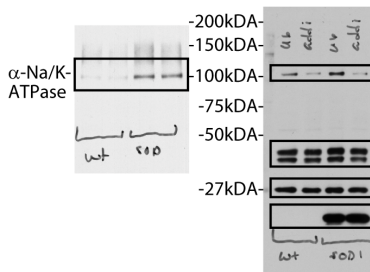


Fig. 6a

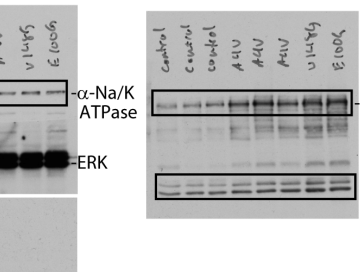
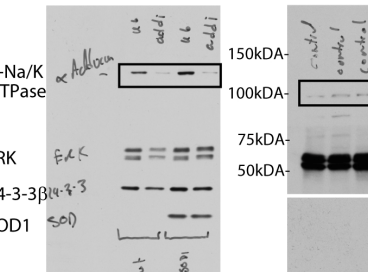
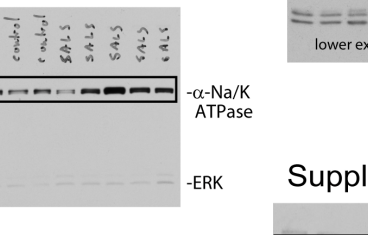
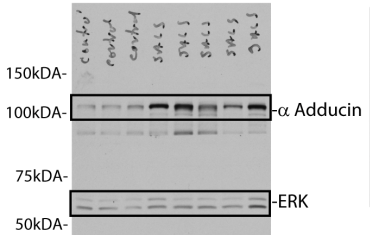
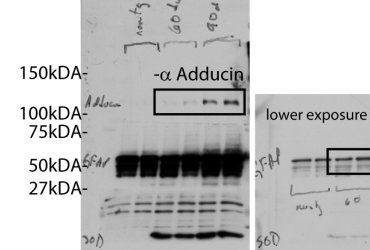


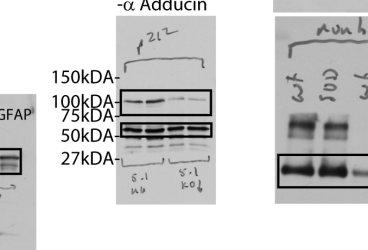
Fig. 6d



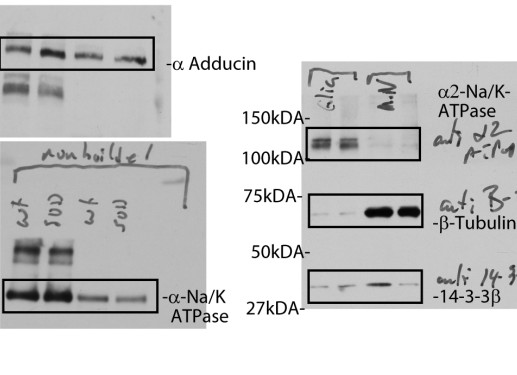
Supplementary Fig. 1



Supplementary Fig. 2



Supplementary Fig. 7a



Supplementary Figure 11

Full scans of key Western blot data. In many experiments, membranes were stripped and reblotted with a second antibody.