Supportive Information

Multifunctional MRI/PET Nanobeacons Derived from the In situ Self-Assembly of Translational Polymers and Clinical Cargo through Coalescent Intermolecular Forces

Charalambos Kaittanis^{1,2}, Travis M. Shaffer^{1,3}, Alexander Bolaender¹, Zachary Appelbaum¹, Jeremy Appelbaum¹, Gabriela Chiosis¹, Jan Grimm^{1, 4, 5, 6*}

¹Molecular Pharmacology Program, ⁴Department of Radiology, Memorial Sloan Kettering Cancer Center, 1275 York Avenue, New York, NY 10065, USA ² Center for Advanced Medical Imaging Sciences, Department of Radiology, Massachusetts General Hospital, 55 Fruit Street, Boston, MA 02114. ³Department of Chemistry, Hunter College and Graduate Center of the City University of New York, New York, NY 10065, USA ⁵Department of Pharmacology and ⁶Radiology, Weill Cornell Medical College, New York, NY 10065, USA

Figure S1. Retention of gadolinium (Gd) by carbohydrate PNPs. **A)** The PNPs that chelated Gd at room temperature slowly released the tracer in serum, indicated by increases in the PNP solution's T1 signal. **B)** When the Gd chelation at room temperature, the PNPs unloaded gadolinium once the pH reached 6.0. **C)** PNP of highmolecular-weight dextran chelated Gd more efficiently, **D)** whereas Gd chelation did not affect the nanoparticles size and distribution. Means \pm SEM.

Figure S2. Chelation and imaging with ⁸⁹Zr-PNPs. **A**) The PNPs effectively chelated ⁸⁹Zr after 30-min heating at 70 $^{\circ}$ C, as shown by the radio-ITLC chromatograms. **B**) After chelation at 70 °C, the stability of the PNP – ${}^{89}Zr$ association was evaluated, which showed that the PNPs retained the radiotracer under EDTA challenge, and in fetal bovine and mouse sera. Means \pm SEM. **C**) CT/PET (left) and PET only (right) of draining lymph nodes (white arrows) after peritumoral administration of ${}^{89}Zr$ -PNPs, revealing proximal and distal drainage through the afferent lymphatics.

Figure S3. Atomic force microscopy performed at liquid state showed the structural stability of the PNPs after cargo loading and release. **A)** Distribution of the unloaded PNPs. Doxorubicin-loaded PNPs after incubation at **B)** pH 7.4 and **C)** pH 6.8. (**insets** show higher dilutions of the two samples, with the PNP being monodispersed).

Figure S4. A) Doxorubicin-loaded dextran nanophores cause more cell death than free doxorubicin in the androgen-receptor-negative human prostate cancer cell line PC3. (PNP: unloaded nanoparticles; Doxo: Doxorubicin; Doxo-PNP: doxorubicin-loaded PNPs) The unloaded PNP are not toxic to the cells. **B)** Serum stability of ¹³¹I-PU-H71 determined through HPLC, after 1 and 24-hour incubation in mouse serum. Intestinal excretion of free and PNP-loaded 131 I-PU-H71 **C**) 4 h and **D**) 24 h post i.v. administration (%ID/g: % injected dose/tissue mass). Means \pm SEM.

Figure S5. Tumor growth response after clodrosome-based therapy of athymic, male nude mice with LNCaP xenografts on their flanks ($n_{\text{vehicle}} = n_{\text{Cloudrosomes}} = 5$). Means \pm SEM.

Figure S6. The loading of molecular cargos into PNP affects the nanoparticles' magnetic resonance signal. Gd-retaining PNP were co-loaded with Doxorubicin (λ_{ex} = 485 nm, λ_{em}) = 590 nm) and Flutax1 (λ_{ex} = 495 nm, λ_{em} = 520 nm), which caused increased in the T1 relaxation times. Means ± SEM.