

**Supporting Figures S1-S3 for:**

**Title: Enhancer turnover is associated with a divergent transcriptional response to glucocorticoid in mouse and human macrophages.**

**Running Title: The genomic response of macrophages to glucocorticoids**

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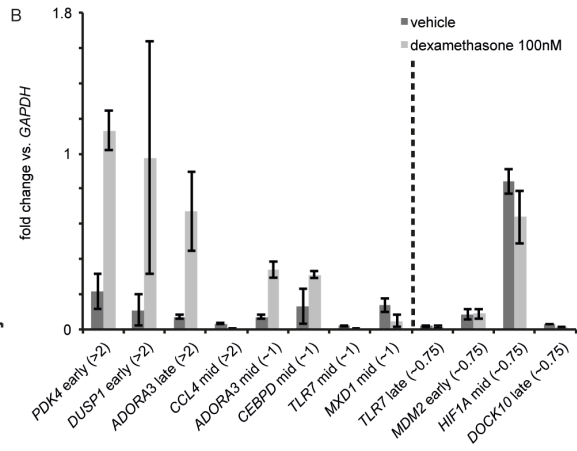
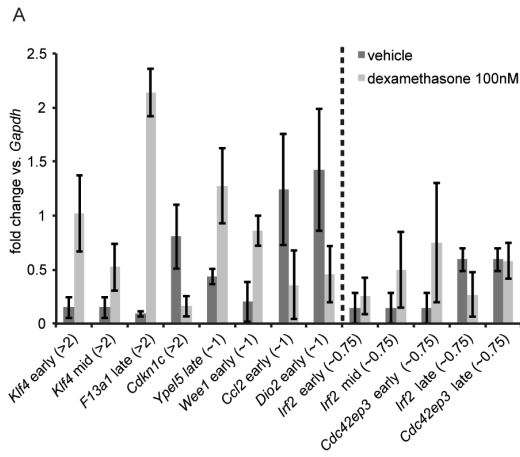
Fax: +44 131 651 9105

**Keywords: enhancer, evolution, glucocorticoid, macrophage**

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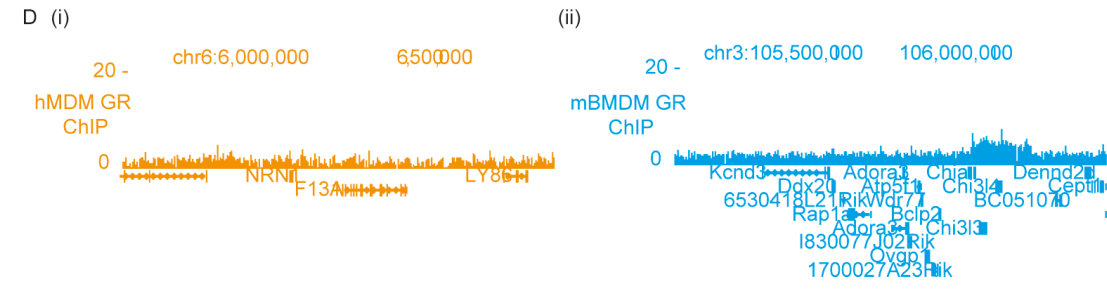
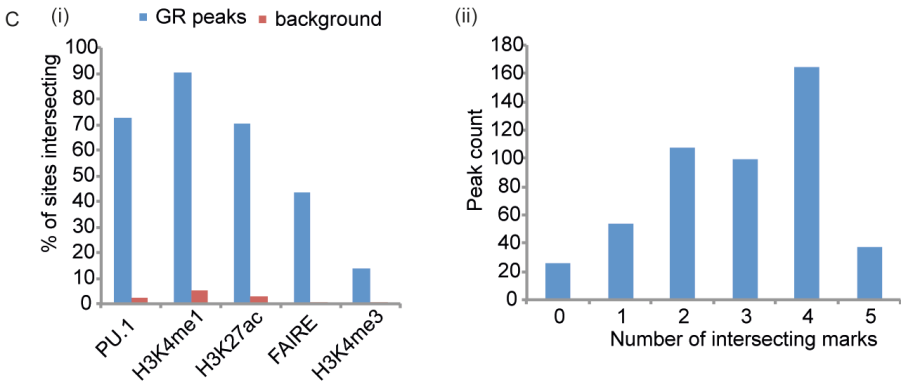
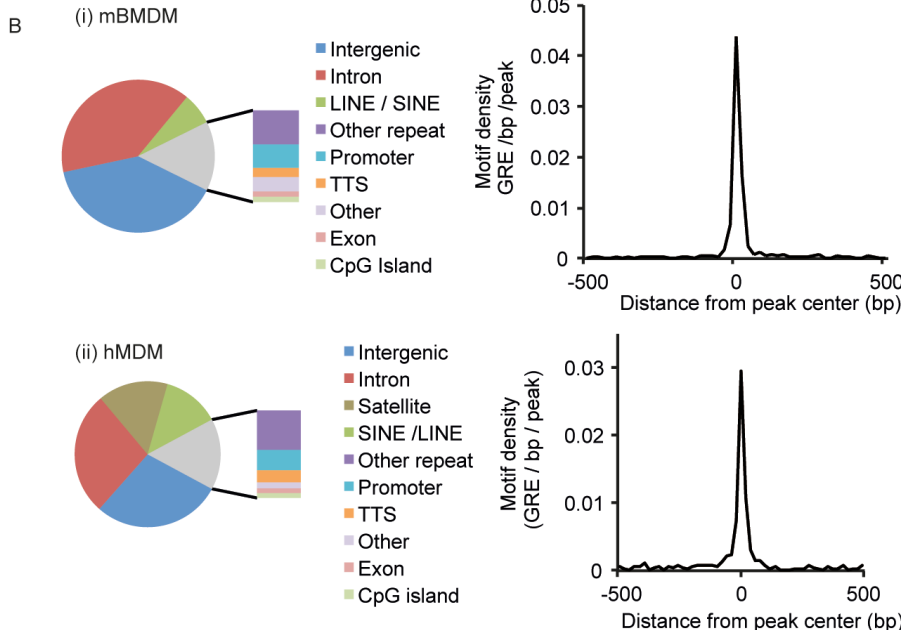
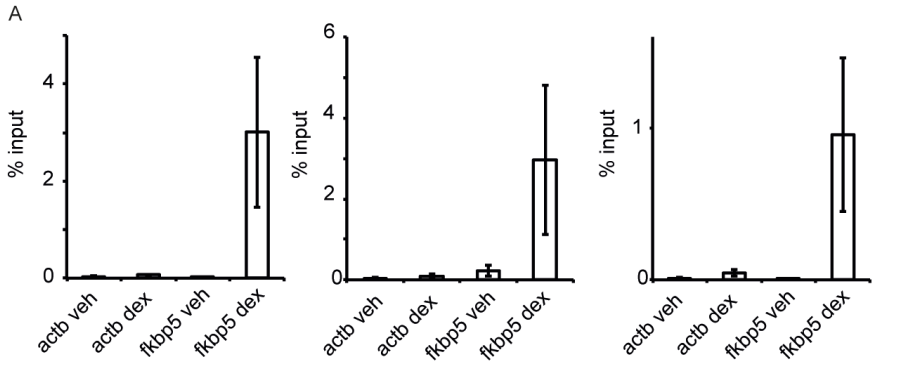
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**Figure S1.** Additional validation and analysis of microarray expression data.

Combinations of genes and time points from our microarray data were selected to represent a range of fold change responses at several time points (early, <2h; mid, 4-10h; late, >10h) in (A) mBMDM and (B) hMDM treated with dexamethasone (see methods). Expression was measured using RT-qPCR to establish a threshold where all tested conditions were confirmed, represented by the vertical dashed line. Values in brackets represent the approximate log<sub>2</sub> fold change for each gene at the given time point using our microarray data. Error bars represent the standard deviation of the mean for 3 biological replicates in mBMDM and 4 biological replicates in hMDM.

(C) Functional annotation of gene expression data from mBMDM and hMDM. Filtered results from mining multiple public functional annotation databases are shown for induced and repressed gene sets. Terms enriched in mBMDM data in yellow and hMDM in blue. Duplicate terms have been removed. All terms shown are significant to  $-\log p\text{-value} > 6.5$ . \* = Enriched in genes regulated 0-2h. Abbreviations: KEGG = Kyoto Encyclopedia of Genes and Genomes, GO = Gene Ontology, BP = Biological Process, MF = Molecular Function, CC = Cellular Compartment.



**Figure S2. Additional data supporting GR ChIP-seq data.** (A) Replication of GR ChIP-qPCR in hMDM. ChIP-qPCR was performed for GR in hMDM, each panel shows an independent biological replicate. Signal from vehicle treated (veh) and 100nM dexamethasone treated (dex) samples is shown for a known GR binding site at +88kb in the *FKBP5* locus (*fkbp5*) and the promoter of *ACTB* (*actb*). Error bars represent 2xSEM for 3 technical replicates of each condition.

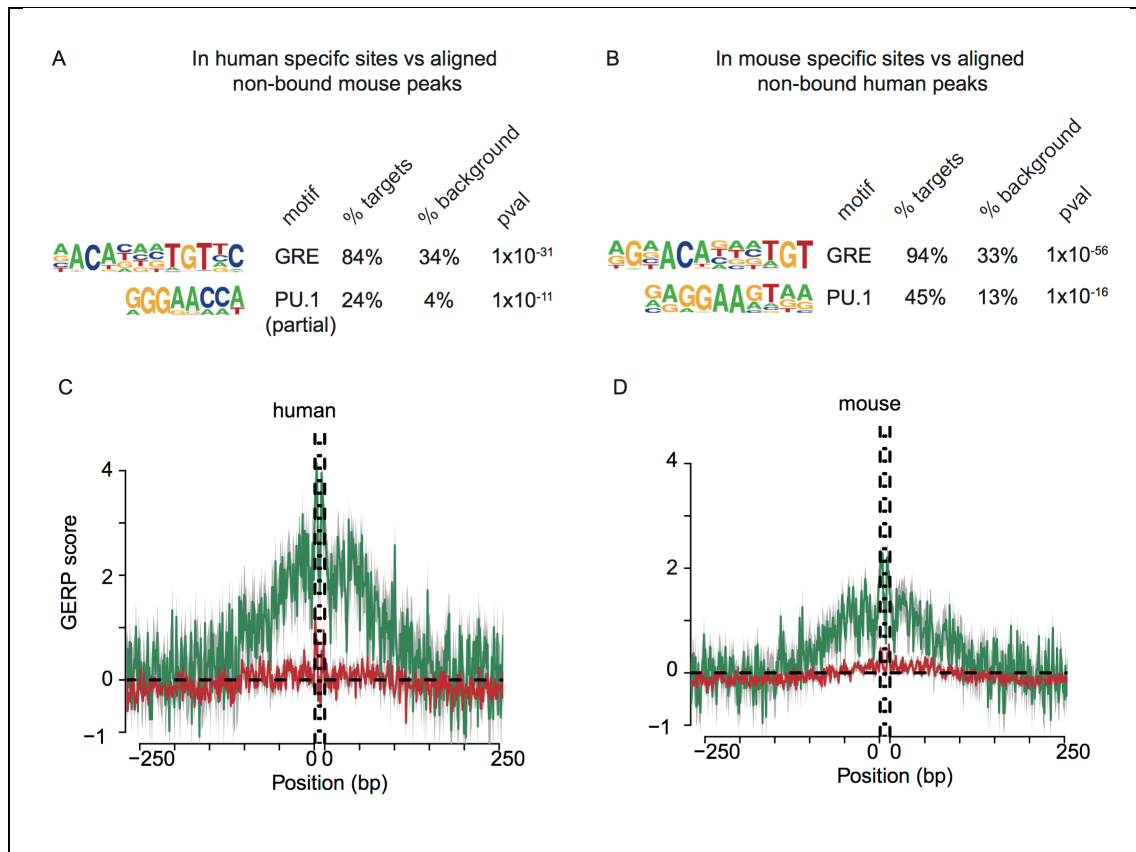
(B) GR binding occurs at sites that are enriched for the GRE and lie distant from gene promoters in both mBMDM and hMDM. The classification of genomic sites where GR is bound is shown from (i) mBMDM and (ii) hMDM, when treated with 100nM dexamethasone. GRE motif density (GRE/bp/peak) for the GR bound sites for each species is shown alongside.

(C) GR bound sites in mBMDM display marks consistent with enhancers.

Comparison was made with published data(57) from non-stimulated mBMDM. (i) Blue = counts of GR bound sites co incident with the previously reported ChIP peaks for PU.1, H3K4me1, H3K27ac, H3K4me3 and open chromatin measured by formaldehyde assisted identification of regulatory elements (FAIRE-seq). A 1000x permuted background for each is shown in red, p values are reported in the main text.

(ii) Percentage of GR peaks overlapping 0-5 of the marks described in (i).

(D) Chip-seq data showing absence of GR binding in a wider area for (i) the human F13A1 locus which is mouse specific (Figure 4C) and (ii) mouse Adora3 which is human specific (Figure 4E).



**Figure S3.** (A) Motifs enriched in the human specific peaks compared to the aligned mouse sites that are not bound by GR in hMDM. (B) motifs enriched in the mouse specific peaks compared to the aligned human sites that are not bound by GR in mBMDM. (C) Mean per base constraint scores calculated using GERP (72) across a 500bp region surrounding the the GRE in shared (green) and species-specific (red) peaks found in hMDM, where the grey bars represents the standard error of the mean. Vertical dashed lines delineate the GRE, as derived de novo from our human data. (D) Analogous to (C) for GR bound peaks and GRE motif found in mBMDM.