

Supporting Information

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1. General Considerations

TLC analysis was performed (fluorescence quenching or KMnO_4 stain) with silica gel HL TLC plates with UV254 purchased from Sorbent Technologies.

Silica gel used for column chromatography (60 Å porosity, 230 x 400 mesh, standard grade) was purchased from Sorbent Technologies (catalog # 30930M-25).

$\text{Pd}(\text{PPh}_3)_4$ and (R,R)-ANDEN-Phenyl Trost ligand were purchased from Sigma Aldrich and stored in an Ar filled glovebox. Pd_2dba_3 was purchased from Strem Chemicals and stored in a glovebox.

The photocatalyst $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbpy})]\text{PF}_6$ was either synthesized according to a literature procedure¹ or purchased from Sigma Aldrich and stored in a glove box.

Anhydrous DMSO and Toluene were purchased from Sigma Aldrich and stored in a glove box.

GC/MS data was obtained on a Shimadzu GCMS-QP2010 SE. ^1H and ^{13}C NMR spectra were obtained on a Bruker ADVANCE 500 DRX equipped with a QNP cryoprobe. ^1H and ^{13}C NMR spectra were referenced to residual protio solvent signals. HRMS data was obtained on a ESI LC-TOF Micromass LCT (Waters).

Final decarboxylative allylation reactions were run in screw threaded tubes from Chemglass (CLS-4208). Two different blue LED light baths were employed. Room temperature reactions were run in a beaker wrapped with blue LED strips which was then covered in aluminum foil. A fan was placed directly above the beaker (photo 1). Reactions performed at elevated temperatures were done in a glass crystallization dish with the outside walls wrapped in blue LED strips. The dish was covered in aluminum foil and filled with water, which was heated to a preset temperature while stirring using a stir plate. The reaction tubes were held in place by a cardboard lid (photo 2).

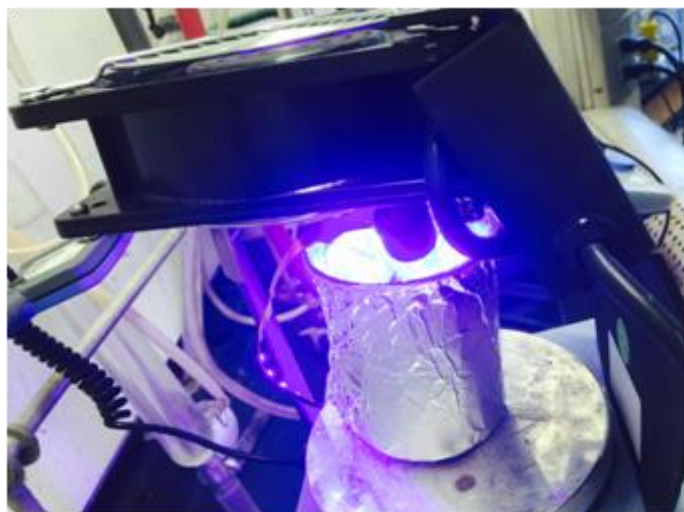


Photo 1

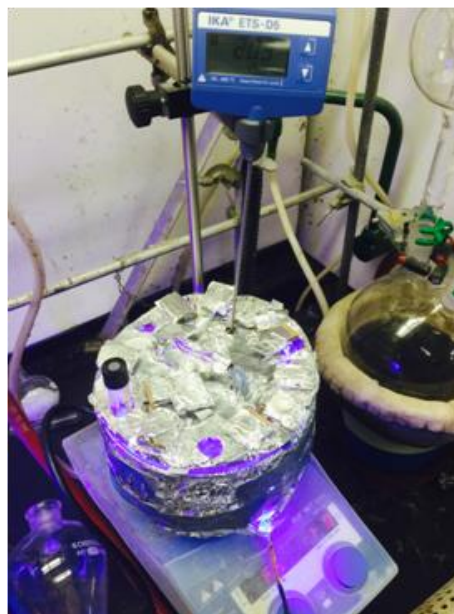
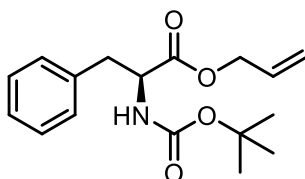


Photo 2

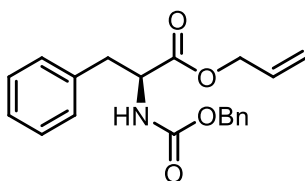
2. Synthesis and Characterization of Allylic Esters

The allylic esters corresponding to products **1a** – **1i** have been previously described by our group.²

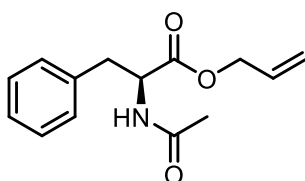
Representative procedure for the synthesis of allylic esters: The N-protected amino acid (5.0 mmol) was added to DMF (15 mL) and the solution cooled to 0 °C (ice bath). Hunig's base (6.5 mmol) was added followed by allyl bromide (6.5 mmol) and the solution was allowed to warm to room temperature and stir overnight. The solution was diluted with EtOAc (40 mL) and then washed with water (2 x 30 mL) and brine (30 mL). The organic phase was dried with MgSO₄, concentrated *in vacuo*, and purified via flash column chromatography.



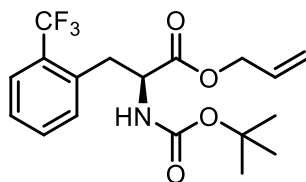
Prepared according to the representative procedure (7.5 mmol scale, 98% yield). The spectroscopic data matched that already reported in the literature.³



Prepared according to the representative procedure (6.7 mmol scale, 99% yield). The spectroscopic data matched that already reported in the literature.⁴



Prepared according to the representative procedure (9.7 mmol scale, 81% yield). The spectroscopic data matched that already reported in the literature.⁵



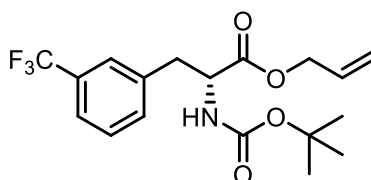
Prepared according to the representative procedure (3 mmol scale, 69% yield). A mixture of rotamers. Major resonances reported.

^1H NMR (500 MHz, CDCl_3) δ 7.64 (d, J = 7.95 Hz, 1H), 7.48 (t, J = 7.52 Hz, 1H), 7.36 (m, 2H), 5.81 (ddt, J = 16.5, 10.9, 5.70, 1H), 5.23 (m, 2H), 5.07 (d, J = 8.75 Hz, 1H), 4.61 (m, 3H), 3.34 (dd, J = 14.5, 5.95 Hz, 1H), 3.09 (dd, J = 14.4, 9.01 Hz, 1H), 1.36 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 171.7, 155.1, 135.4, 131.9, 131.7, 131.4, 129.2 (q, J = 29.6 Hz), 127.1, 126.2 (q, J = 5.59 Hz), 124.6 (q, J = 273.7 Hz), 118.8, 80.1, 66.1, 54.6, 35.6, 28.3.

IR (film) 3367, 2980, 2935, 1745, 1715, 1504, 1454, 1367, 1315, 1251, 1171, 1120, 1060, 1039, 989, 935, 767 cm^{-1} .

HRMS: Calc'd $\text{C}_{18}\text{H}_{22}\text{F}_3\text{NO}_4\text{Na}$ ($\text{M}+\text{Na}$) = 396.1399, found 396.1389.



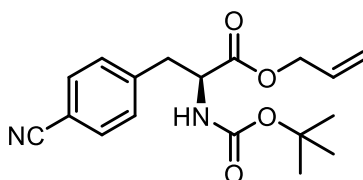
Prepared according to the representative procedure (3 mmol scale, 61% yield).

^1H NMR (500 MHz, CDCl_3) δ 7.50 (d, J = 7.77 Hz, 1H), 7.38 (m, 3H), 5.85 (ddt, J = 16.5, 10.4, 5.95 Hz, 1H), 5.28 (m, 2H), 5.04 (d, J = 7.90 Hz, 1H), 4.60 (m, 3H), 3.22 (dd, J = 13.8, 5.95 Hz, 1H), 3.12 (dd, J = 13.9, 6.07 Hz, 1H), 1.42 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 171.2, 155.1, 137.2, 132.9, 131.3, 130.9 (q, J = 32.3 Hz), 129.1, 126.3 (q, J = 3.36 Hz), 124.1 (q, J = 273 Hz), 124.0 (q, J = 3.61 Hz), 119.6, 80.3, 66.3, 54.4, 38.3, 28.4.

IR (film) 3365, 2979, 2935, 1745, 1714, 1504, 1454, 1367, 1328, 1251, 1164, 1126, 1074, 991, 937, 794, 703, 659 cm^{-1} .

HRMS: Calc'd $\text{C}_{18}\text{H}_{22}\text{F}_3\text{NO}_4\text{Na}$ ($\text{M}+\text{Na}$)⁺ = 396.1399, found = 396.1397.



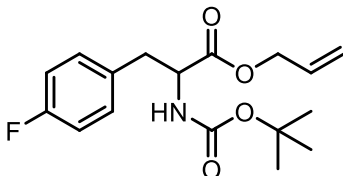
Prepared according to the representative procedure (3.4 mmol scale, 59% yield).

^1H NMR (500 MHz, CDCl_3) δ 7.58 (d, J = 7.91 Hz, 2H), 7.26 (d, J = 7.99 Hz, 2H), 5.85 (ddt, J = 16.6, 11.1, 5.91 Hz, 1H), 5.28 (m, 2H), 5.04 (d, J = 8.05 Hz, 1H), 4.61 (m, 3H), 3.22 (dd, J = 13.8, 5.86 Hz, 1H), 3.08 (dd, J = 13.8, 6.51 Hz, 1H), 1.40 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 171.1, 155.0, 142.0, 132.3, 131.3, 130.4, 119.5, 118.9, 111.1, 80.4, 66.4, 54.2, 38.8, 28.4.

IR (film) 3367, 2977, 2933, 2227, 1743, 1714, 1650, 1608, 1504, 1454, 1367, 1274, 1251, 1166, 1054, 1022, 991, 937, 825, 748, 557 cm^{-1} .

HRMS: Calc'd $\text{C}_{18}\text{H}_{22}\text{N}_2\text{O}_4\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 353.1477, found = 353.1474.



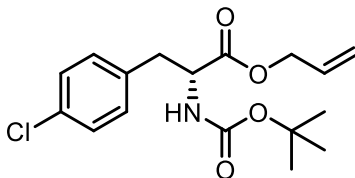
The free amino acid (5.5 mmol) was added to a 1:1 mixture of THF/ H_2O (15 mL each) and the mixture was cooled to 0 $^\circ\text{C}$ (ice bath). Triethylamine (9.0 mmol) was added to the reaction mixture followed by Boc_2O (7.0 mmol). The reaction was allowed to warm to room temperature and stirred overnight. Next, the volatiles were removed *in vacuo*, the pH was adjusted to 3 with 1M HCl and the reaction mixture extracted with EtOAc (40 mL). The organic phase was washed with brine (20 mL), dried with MgSO_4 and concentrated *in vacuo*. The crude product was subjected to the representative procedure to yield the desired allyl ester after flash column chromatography (66% yield over two steps).

^1H NMR (500 MHz, CDCl_3) δ 7.10 (m, 2H), 6.97 (t, J = 8.67 Hz, 2H), 5.86 (ddt, J = 17.2, 10.3, 5.85 Hz, 1H), 5.29 (m, 2H), 4.98 (d, J = 8.31 Hz, 1H), 4.60 (m, 3H), 3.11 (dd, J = 14.0, 5.86 Hz, 1H), 3.03 (dd, J = 14.0, 6.19 Hz, 1H), 1.42 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 171.6, 162.1 (d, J = 245 Hz), 155.1, 131.8 (d, J = 3.12 Hz), 131.5, 131.0 (d, J = 7.95 Hz), 119.2, 115.5 (d, J = 21.3 Hz), 80.2, 66.2, 55.6, 37.8, 28.4.

IR (film) 3434, 3365, 2977, 2933, 1745, 1714, 1602, 1510, 1446, 1367, 1222, 1168, 1097, 1056, 1016, 991, 935, 825, 748 cm^{-1} .

HRMS: Calc'd $\text{C}_{17}\text{H}_{22}\text{FNO}_4\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 346.1431, found = 346.1412.



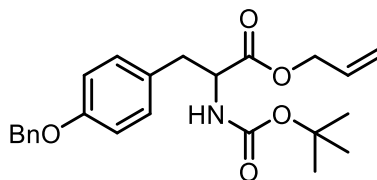
Prepared according to the representative procedure (2.2 mmol scale, 95% yield).

^1H NMR (500 MHz, CDCl_3) δ 7.26 (m, 2H), 7.07 (d, J = 8.38 Hz, 2H), 5.86 (ddt, J = 17.2, 10.4, 5.90 Hz, 1H), 5.29 (m, 2H), 4.97 (d, J = 8.27 Hz, 1H), 4.60 (m, 3H), 3.11 (dd, J = 13.9, 5.80 Hz, 1H), 3.03 (dd, J = 13.9, 6.10 Hz, 1H), 1.42 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 171.4, 155.1, 134.6, 133.1, 131.5, 130.9, 128.8, 119.3, 80.2, 66.2, 54.5, 37.9, 28.4.

IR (film) 3355, 2977, 2933, 1741, 1714, 1492, 1454, 1367, 1253, 1166, 1091, 1056, 1016, 989, 933, 748 cm^{-1} .

HRMS: Calc'd $\text{C}_{17}\text{H}_{22}\text{ClNO}_4\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 362.1135, found = 362.1128.



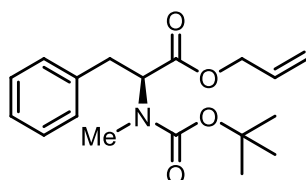
Prepared according to the representative procedure (6.5 mmol scale, 32% yield).

^1H NMR (500 MHz, CDCl_3) δ 7.40 (m, 4H), 7.33 (m, 1H), 7.05 (d, $J = 8.59$ Hz, 2H), 6.90 (m, 2H), 5.87 (ddt, $J = 17.2$, 10.4, 5.84 Hz, 1H), 5.31 (dq, $J = 17.2$, 1.49 Hz, 1H), 5.24 (dd, $J = 10.4$, 1.31 Hz, 1H), 5.04 (s, 2H), 4.96 (d, $J = 8.35$ Hz, 1H), 4.58 (m, 3H), 3.04 (m, 2H), 1.42 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 171.8, 158.0, 155.2, 137.1, 131.7, 130.5, 128.7, 128.3, 128.1, 127.6, 119.1, 115.0, 80.0, 70.1, 66.1, 54.7, 37.6, 28.5.

IR (film) 3367, 3031, 2977, 2931, 1741, 1714, 1610, 1583, 1512, 1454, 1367, 1244, 1174, 1056, 1018, 991, 931, 862, 821, 738, 696 cm^{-1} .

HRMS: Calc'd $\text{C}_{24}\text{H}_{29}\text{NO}_5\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 434.1943, found = 434.1940.



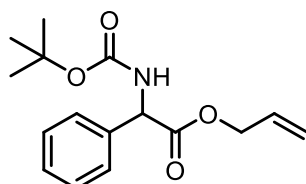
Prepared according to the representative procedure (3.6 mmol scale, 51% yield). A mixture of rotamers.

^1H NMR (500 MHz, CDCl_3) δ 7.27 (m, 2H), 7.19 (m, 3H), 5.91 (ddt, $J = 16.4$, 10.8, 5.65 Hz, 1H), 5.29 (m, 2H), 4.92 (dd, $J = 10.7$, 5.33 Hz, 0.5H), 4.63 (m, 2.5H), 3.31 (m, 1H), 3.03 (m, 1H), 2.74 (s, 1.6 H), 2.71 (s, 1.4H), 1.37 (s, 3.8H), 1.33 (s, 5.2H).

^{13}C NMR (126 MHz, CDCl_3) δ 171.1 (br s), 155.5 (br s), 137.7 (br s), 131.9 (br s), 129.1 (br s), 128.6 (br s), 126.7 (br s), 118.6 (br s), 80.3 (br s), 65.9 (br s), 60.8 (br s), 35.4 (br s), 32.4 (br s), 28.3 (br s).

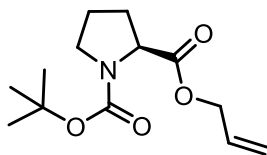
IR (film) 3064, 3028, 2975, 2931, 1745, 1697, 1479, 1454, 1392, 1367, 1326, 1274, 1257, 1218, 1170, 1143, 1080, 1029, 987, 933, 869, 750, 700 cm^{-1} .

HRMS: Calc'd $\text{C}_{18}\text{H}_{25}\text{NO}_4\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 342.1681, found = 342.1678.

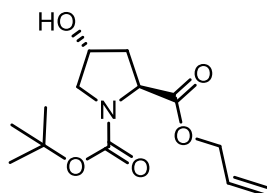


The free amino acid (25.3 mmol) was added to 1M NaOH (35.0 mL). To this solution was added Boc₂O (28.1 mmol) in *t*BuOH (20.0 mL) and the mixture was stirred for 1 h. After removing volatiles *in vacuo* the pH was adjusted to 3 with 1M HCl and the solution was extracted with DCM (3 x 50 mL) which was dried with MgSO₄, and concentrated *in vacuo*. The crude material was subjected to the representative reaction conditions to furnish the desired allyl

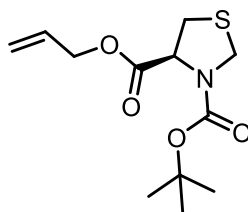
ester after flash column chromatography (76% yield over two steps). The spectroscopic data matched that already reported in the literature.⁶



Prepared according to the representative procedure (3 mmol scale, 95% yield). The spectroscopic data matched that already reported in the literature.⁷



Prepared according to a literature procedure with matching spectroscopic data.⁸



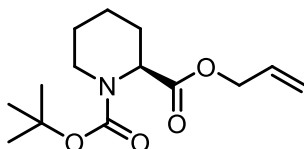
Prepared according to the representative procedure (2 mmol scale, 82% yield). A mixture of rotamers.

¹H NMR (500 MHz, CDCl₃) δ 5.91 (m, 1H), 5.31 (m, 2H), 4.91 (m, 0.5H), 4.63 (m, 3H), 4.50 (m, 1.5H), 3.32 (m, 1H), 3.20 (m, 1H), 1.47 (s, 4.5H), 1.43 (s, 4.5H).

¹³C NMR (126 MHz, CDCl₃) δ 170.5 (br s), 153.3 (br s), 131.6 (br s), 118.8 (br s), 81.4, 66.2, 61.7 (br s), 48.7 (br s), 34.2 (br s), 28.4 (br s).

IR (film) 3429, 2977, 2935, 1749, 1697, 1650, 1456, 1384, 1367, 1336, 1274, 1257, 1163, 1108, 985, 914, 889, 864, 763, 748 cm⁻¹.

HRMS: Calc'd C₁₂H₁₉NO₄SNa (M+Na)⁺ = 296.0933, found = 296.0919.



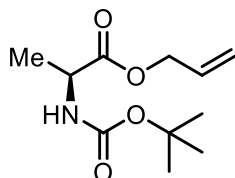
Prepared according to the representative procedure (2.2 mmol scale, 96% yield). A mixture of rotamers.

¹H NMR (500 MHz, CDCl₃) δ 5.90 (ddt, *J* = 16.6, 10.8, 5.54 Hz, 1H), 5.32 (d, *J* = 17.0 Hz, 1H), 5.23 (d, *J* = 10.5 Hz, 1H), 4.91 (s, 0.5H), 4.73 (s, 0.5H), 4.63 (m, 2H), 4.02 (d, *J* = 11.9 Hz, 0.5H), 3.91 (d, *J* = 11.9 Hz, 0.5H), 2.98 (t, *J* = 13.2 Hz, 0.5H), 2.87 (t, *J* = 13.3 Hz, 0.5H), 2.22 (t, *J* = 13.8 Hz, 1H), 1.67 (m, 3H), 1.44 (m, 10H), 1.22 (q, *J* = 14.0 Hz, 1H).

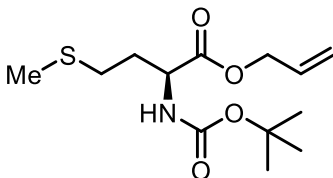
^{13}C NMR (126 MHz, CDCl_3) δ 171.8 (br s), 155.8 (br s), 132.0 (br s), 118.4 (br s), 80.1, 65.6, 54.5 (br s), 41.7 (br s), 28.5, 26.9, 24.8 (br s), 20.8 (br s).

IR (film) 2974, 2941, 2862, 1743, 1693, 1650, 1475, 1448, 1392, 1365, 1338, 1323, 1269, 1247, 1186, 1157, 1124, 1091, 1043, 1000, 977, 931, 871, 771, 559 cm^{-1} .

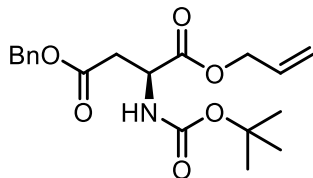
HRMS: Calc'd $\text{C}_{14}\text{H}_{23}\text{NO}_4\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 292.1525, found = 292.1523.



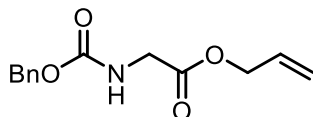
Prepared according to the representative procedure (5 mmol scale, 99% yield). The spectroscopic data matched that already reported in the literature.⁶



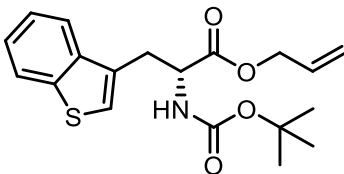
The free amino acid (16.8 mmol) was added to a solution of 1M NaOH (16.0 mL) and dioxane (20.0 mL). The solution was cooled to 0 °C (ice bath) and Boc_2O (17.6 mmol) was added and allowed to stir at room temperature overnight. Next, the volatiles were removed *in vacuo*, the pH was adjusted to 3 with 1M HCl, and the solution was extracted with EtOAc (2 x 20 mL). The organic phase was washed with brine (15 mL), dried with MgSO_4 , and concentrated *in vacuo*. The crude product was subjected to the representative reaction conditions to provide the desired product after flash column chromatography (63% yield over two steps). The spectroscopic data matched that already reported in the literature.⁴



Prepared according to the representative procedure (6.2 mmol scale, 99% yield). The spectroscopic data matched that already reported in the literature.⁹



Prepared according to the representative procedure (5 mmol scale, 86% yield). The spectroscopic data matched that already reported in the literature.¹⁰



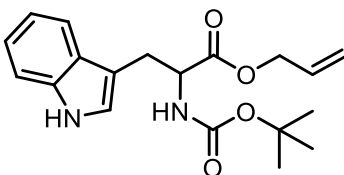
Prepared according to the representative procedure (3.1 mmol scale, 41% yield).

^1H NMR (500 MHz, CDCl_3) δ 7.85 (d, J = 7.52 Hz, 1H), 7.76 (m, 1H), 7.36 (m, 2H), 7.17 (s, 1H), 5.80 (ddt, J = 16.4, 11.1, 5.82 Hz, 1H), 5.24 (m, 2H), 5.10 (d, J = 8.20 Hz, 1H), 4.73 (q, J = 6.13 Hz, 1H), 4.56 (m, 2H), 3.41 (dd, J = 14.7, 5.89 Hz, 1H), 3.34 (dd, J = 14.7, 6.12 Hz, 1H), 1.43 (s, 7.5H, Boc), 1.25 (s, 1.5H, Boc).

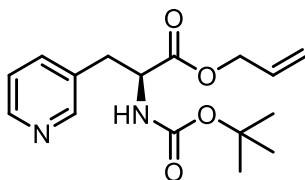
^{13}C NMR (126 MHz, CDCl_3) δ 171.7, 155.2, 140.4, 139.1, 131.5, 131.0, 124.5, 124.2, 124.0, 123.0, 121.8, 119.2, 80.2, 66.3, 53.8, 31.4, 28.3 (br s).

IR (film) 3357, 2977, 2931, 1739, 1712, 1504, 1458, 1365, 1274, 1251, 1166, 1058, 1020, 989, 933, 854, 761, 732 cm^{-1} .

HRMS: Calc'd $\text{C}_{19}\text{H}_{23}\text{NO}_4\text{SNa}$ ($\text{M}+\text{Na}$) $^+$ = 384.1246, found = 384.1241.



The free amino acid (7.3 mmol) was added to a solution of H_2O (15 mL) and THF (15 mL). NaOH (8.0 mmol) was added followed by Boc_2O (8.0 mmol). After stirring overnight the THF was removed *in vacuo* and the pH adjusted to 4 using 1M HCl. The aqueous phase was extracted with DCM (2 x 20 mL) which was dried with MgSO_4 and concentrated *in vacuo*. The crude material was subjected to the representative reaction conditions to furnish the desired allyl ester after flash column chromatography in a 16% yield. The spectroscopic data matched that already reported in the literature.^{11,12}



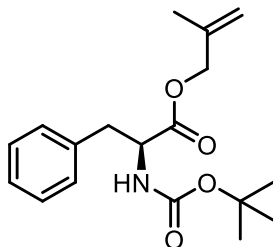
Prepared according to the representative procedure (3.8 mmol scale, 34% yield).

^1H NMR (500 MHz, CDCl_3) δ 8.49 (d, J = 4.64 Hz, 1H), 8.39 (br s, 1H), 7.48 (dt, J = 7.78, 1.99 Hz, 1H), 7.22 (dd, J = 7.84, 4.81 Hz, 1H), 5.86 (m, 1H), 5.30 (m, 2H), 5.06 (br s, 1H), 4.62 (m, 3H), 3.17 (dd, J = 14.0, 5.72 Hz, 1H), 3.05 (dd, J = 14.0, 6.17 Hz, 1H), 1.41 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 171.2, 155.1, 150.8, 148.6, 137.0, 131.8, 131.4, 123.5, 119.6, 80.3, 66.4, 54.3, 35.8, 28.4.

IR (film) 3350, 3217, 2977, 2933, 1743, 1712, 1649, 1577, 1519, 1481, 1446, 1425, 1367, 1276, 1253, 1168, 1054, 1027, 991, 935, 750, 713 cm^{-1} .

HRMS: Calc'd $\text{C}_{16}\text{H}_{23}\text{N}_2\text{O}_4$ ($\text{M}+\text{H}$) $^+$ = 307.1658, found = 307.1664.



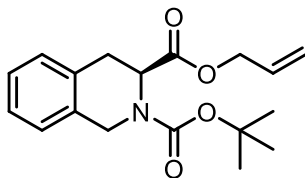
Boc-Phe-OH (5.7 mmol) was added to 20 mL DCM under Ar. The solution was cooled to 0 °C (ice bath) and β -Methallyl alcohol (7.3 mmol) and triethylamine (14 mmol) were added. After stirring for 5 mins, PyBOP (7.3 mmol) was added and the solution was allowed to warm to room temperature and stir overnight. Next, the reaction mixture was washed with water (1 x 10 mL), brine (1 x 10 mL), concentrated *in vacuo*, and purified via flash column chromatography to provide the desired allyl ester in a 96% yield.

^1H NMR (500 MHz, CDCl_3) δ 7.29 (m, 2H), 7.25 (m, 1H), 7.14 (d, $J = 7.03$ Hz, 2H), 4.95 (m, 3H), 4.63 (q, $J = 6.18$ Hz, 1H), 4.52 (s, 2H), 3.14 (dd, $J = 13.8, 5.88$ Hz, 1H), 3.07 (dd, $J = 13.9, 6.28$ Hz, 1H), 1.71 (s, 3H), 1.41 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 171.8, 155.2, 139.4, 136.1, 129.5, 128.7, 127.2, 113.9, 80.0, 68.7, 54.6, 38.5, 28.4, 19.7.

IR (film) 3369, 3064, 3029, 2977, 2931, 1745, 1693, 1604, 1487, 1454, 1367, 1251, 1164, 1054, 1022, 960, 912, 860, 744, 700, 568 cm^{-1} .

HRMS: Calc'd $\text{C}_{18}\text{H}_{25}\text{NO}_4\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 342.1681, found = 342.1670.



The free amino acid (14.1 mmol) was added to a solution of 1,4 dioxane (30 mL) and 1M NaOH (18 mL). The solution was cooled to 0°C (ice bath), and Boc_2O (18.0 mmol) was added. The solution was allowed to warm to room temperature and stirred overnight. Next, the dioxane was removed *in vacuo*, the pH was adjusted to 3 with 1 M HCl and the reaction mixture extracted with EtOAc (50 mL). The organic phase was washed with brine (30 mL), dried with MgSO_4 , and concentrated *in vacuo*. The crude product was subjected to the representative procedure to yield the desired allyl ester after flash column chromatography (58% yield over two steps) as a mixture of rotamers.

^1H NMR (500 MHz, CDCl_3) δ 7.15 (m, 4H), 5.76 (dddd, $J = 18.0, 12.9, 6.31, 3.70$ Hz, 1H), 5.14 (m, 2.5H), 4.81 (t, $J = 5.40$ Hz, 0.5H), 4.70 (dd, $J = 16.3, 9.82$ Hz, 1H), 4.51 (m, 3H), 3.27 (dd, $J = 15.8, 3.11$ Hz, 0.5H), 3.18 (m, 1.5H), 1.53 (s, 4.5H), 1.45 (s, 4.5H).

^{13}C NMR (126 MHz, CDCl_3) δ 171.5 (br s), 155.3 (br s), 134.1, 133.0, 132.1 (br s), 131.8 (br s), 128.3 (br s), 127.0 (br s), 126.8 (br s), 126.4 (br s), 118.2 (br s), 80.7, 65.7 (br s), 53.6 (br s), 44.5 (br s), 31.6 (br s), 28.5 (br s).

IR (film) 3336, 2975, 2931, 1737, 1697, 1456, 1392, 1367, 1357, 1332, 1251, 1220, 1164, 1122, 997, 923, 757 cm^{-1} .

HRMS: Calc'd $\text{C}_{18}\text{H}_{23}\text{NO}_4\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 340.1525, found = 340.1517.

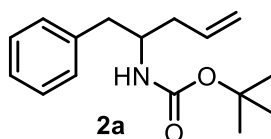
3. General Experimental Procedures

For conversion of allylic esters to homoallylic amines: An oven dried 16 x 125 mm threaded glass tube charged with a stirbar was taken into the glovebox. Pd(PPh₃)₄ and [Ir{dF(CF₃)ppy}₂(dtbpy)]PF₆ (5.0 and 1.0 mol %, respectively) were added to the vial. If the starting material was a solid, 0.25 mmol was added at this time. 2.0 mL DMSO was added to the vial which was then capped and removed from the glovebox. If the starting material was a liquid, it was added via syringe at this time. The reaction was placed in a room temperature blue LED light bath (photo 1, S-2) and analyzed by GC/MS. The crude reaction mixture was directly loaded onto a silica gel column and purified using EtOAc/Hexanes as the eluant.

For conversion of carboxylic acids to homoallylic amines: An oven dried 16 x 125 mm threaded glass tube charged with a stirbar was taken into the glovebox. Pd(PPh₃)₄ and [Ir{dF(CF₃)ppy}₂(dtbpy)]PF₆ (5.0 and 1.0 mol %, respectively) were added to the vial followed by the carboxylic acid (0.25 mmol), and 2.0 mL DMSO. The vial was capped, removed from the glovebox, and 0.25 mmol of allyl methyl carbonate was added via syringe. The reaction was placed in a room temperature blue LED light bath (photo 1, S-2) and analyzed by GC/MS. The crude reaction mixture was directly loaded onto a silica gel column and purified using EtOAc/Hexanes as the eluant.

4. Characterization of Allylated Products

The products **1a** – **1i** have been previously described by our group.²

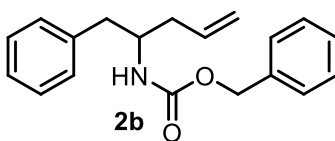


¹H NMR (500 MHz, CDCl₃) δ 7.29 (m, 2H), 7.21 (m, 3H), 5.80 (ddt, *J* = 17.3, 10.5, 7.04 Hz, 1H), 5.10 (m, 2H), 4.40 (s, 1H), 3.91 (s, 1H), 2.78 (m, 2H), 2.26 (m, 1H), 2.11 (m, 1H), 1.41 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 155.5, 138.3, 134.6, 129.6, 128.5, 126.5, 118.1, 79.3, 51.2, 40.6, 38.2, 28.5.

IR (film) 3348, 3076, 2977, 2927, 1697, 1641, 1523, 1496, 1454, 1438, 1388, 1363, 1272, 1251, 1230, 1172, 1045, 1024, 910, 856, 742, 698 cm⁻¹.

HRMS: Calc'd C₁₆H₂₃NO₂Na (M+Na)⁺ = 284.1626, found = 284.1616.

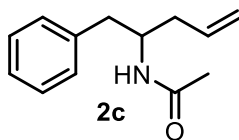


¹H NMR (500 MHz, CDCl₃) δ 7.33 (m, 7H), 7.19 (m, 3H), 5.79 (ddt, *J* = 17.3, 10.4, 7.04 Hz, 1H), 5.09 (m, 4H), 4.63 (d, *J* = 8.34 Hz, 1H), 3.99 (m, 1H), 2.81 (m, 2H), 2.29 (dt, *J* = 12.7, 6.09 Hz, 1H), 2.14 (dt, *J* = 14.2, 7.13 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 155.9, 137.9, 136.7, 134.3, 129.6, 128.6, 128.5, 128.2, 128.1, 126.6, 118.4, 66.7, 51.7, 40.5, 38.2.

IR (film) 3325, 3062, 3029, 2943, 1697, 1531, 1514, 1454, 1440, 1342, 1257, 1222, 1041, 1027, 993, 914, 746, 698 cm⁻¹.

HRMS: Calc'd C₁₉H₂₁NO₂Na (M+Na)⁺ = 318.1470, found = 318.1469.

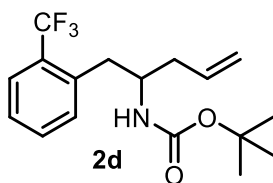


¹H NMR (500 MHz, CDCl₃) δ 7.29 (m, 2H), 7.20 (m, 3H), 5.79 (ddt, *J* = 17.3, 10.3, 7.07 Hz, 1H), 5.30 (d, *J* = 7.47 Hz, 1H), 5.10 (m, 2H), 4.26 (m, 1H), 2.79 (dd, *J* = 14.0, 6.27 Hz, 2H), 2.28 (m, 1H), 2.13 (m, 1H), 1.92 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 169.6, 137.9, 134.5, 129.5, 128.6, 126.6, 118.2, 49.6, 40.1, 37.9, 23.6.

IR (film) 3274, 3076, 2927, 1645, 1556, 1496, 1438, 1373, 1298, 991, 748, 700 cm⁻¹.

HRMS: Calc'd C₁₃H₁₇NONa (M+Na)⁺ = 226.1208, found = 226.1187.

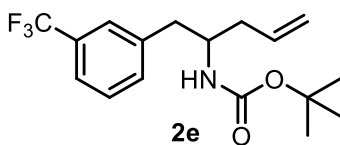


¹H NMR (500 MHz, CDCl₃) δ 7.62 (d, *J* = 7.83 Hz, 1H), 7.46 (m, 2H), 7.30 (m, 1H), 5.81 (ddt, *J* = 17.7, 9.66, 7.10 Hz, 1H), 5.12 (m, 2H), 4.42 (d, *J* = 9.23 Hz, 1H), 4.00 (m, 1H), 2.96 (m, 2H), 2.28 (m, 2H), 1.33 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 155.4, 137.5, 134.3, 131.8, 131.6, 129.0 (q, *J* = 29.7 Hz), 126.5, 126.0 (q, *J* = 6.32 Hz), 124.8 (q, *J* = 274 Hz), 118.3, 79.3, 51.3, 39.6, 37.4, 28.4.

IR (film) 3363, 2981, 2931, 1681, 1606, 1523, 1454, 1367, 1307, 1271, 1251, 1163, 1107, 1049, 914, 767 cm⁻¹.

HRMS: Calc'd C₁₇H₂₂F₃NO₂Na (M+Na)⁺ = 352.1500, found = 352.1505.

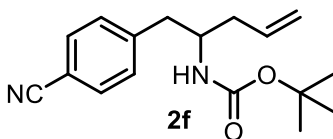


¹H NMR (500 MHz, CDCl₃) δ 7.48 (m, 1H), 7.41 (m, 3H), 5.79 (ddt, *J* = 17.3, 10.3, 7.05 Hz, 1H), 5.11 (m, 2H), 4.39 (m, 1H), 3.91 (m, 1H), 2.85 (d, *J* = 6.65 Hz, 2H), 2.26 (m, 1H), 2.13 (m, 1H), 1.39 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 155.4, 139.3, 134.2, 133.0, 130.7 (q, *J* = 30.9 Hz), 128.9, 126.3, 124.4 (q, *J* = 272 Hz), 123.4 (q, *J* = 3.65 Hz), 118.5, 79.5, 51.1, 40.4, 38.3, 28.4.

IR (film) 3363, 2979, 2935, 1681, 1523, 1446, 1392, 1367, 1332, 1159, 1116, 1074, 918, 800, 750, 703 cm⁻¹.

HRMS: Calc'd C₁₇H₂₂F₃NO₂Na (M+Na)⁺ = 352.1500, found = 352.1470.

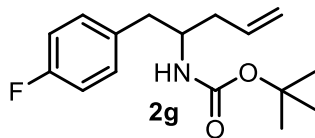


¹H NMR (500 MHz, CDCl₃) δ 7.58 (m, 2H), 7.31 (d, *J* = 8.07 Hz, 2H), 5.77 (ddt, *J* = 17.2, 10.3, 7.04 Hz, 1H), 5.13 (m, 2H), 4.38 (d, *J* = 8.68 Hz, 1H), 3.91 (m, 1H), 2.84 (d, *J* = 6.74 Hz, 2H), 2.25 (m, 1H), 2.14 (m, 1H), 1.38 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 155.3, 144.2, 133.9, 132.3, 130.3, 119.1, 118.7, 110.4, 79.6, 51.0, 41.0, 38.5, 28.5.

IR (film) 3363, 2979, 2962, 2929, 2225, 1681, 1645, 1608, 1523, 1461, 1442, 1390, 1367, 1350, 1272, 1251, 1174, 1049, 1029, 919, 842, 823, 750, 655, 567 cm^{-1} .

HRMS: Calc'd $\text{C}_{17}\text{H}_{22}\text{N}_2\text{O}_2\text{Na}$ ($\text{M}+\text{Na}$)⁺ = 309.1579, found = 309.1585.

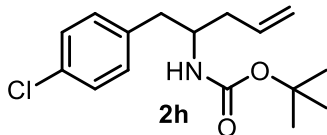


¹H NMR (500 MHz, CDCl_3) δ 7.14 (m, 2H), 6.97 (m, 2H), 5.78 (ddt, J = 17.3, 10.4, 7.07 Hz, 1H), 5.10 (m, 2H), 4.37 (s, 1H), 3.86 (s, 1H), 2.73 (m, 2H), 2.24 (m, 1H), 2.10 (m, 1H), 1.40 (s, 9H).

¹³C NMR (126 MHz, CDCl_3) δ 161.6 (d, J = 244 Hz), 155.4, 134.4, 134.0 (d, J = 3.03 Hz), 130.9 (d, J = 7.65 Hz), 118.2, 115.3 (d, J = 21.1 Hz), 79.4, 51.2, 39.9, 38.2, 28.5.

IR (film) 3342, 2979, 2929, 1687, 1600, 1504, 1454, 1365, 1271, 1253, 1218, 1172, 1045, 914, 831, 750 cm^{-1} .

HRMS: Calc'd $\text{C}_{16}\text{H}_{22}\text{FNO}_2\text{Na}$ ($\text{M}+\text{Na}$)⁺ = 302.1532, found = 302.1546.

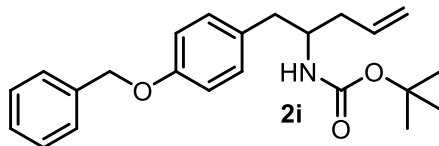


¹H NMR (500 MHz, CDCl_3) δ 7.27 (m, 2H), 7.13 (d, J = 8.26 Hz, 2H), 5.79 (ddt, J = 17.2, 10.3, 7.06 Hz, 1H), 5.11 (m, 2H), 4.38 (s, 1H), 3.88 (s, 1H), 2.76 (m, 2H), 2.25 (m, 1H), 2.11 (m, 1H), 1.41 (s, 9H).

¹³C NMR (126 MHz, CDCl_3) δ 155.4, 136.8, 134.3, 132.3, 130.9, 128.6, 118.3, 79.4, 51.1, 40.0, 38.2, 28.5.

IR (film) 3344, 2977, 2929, 1687, 1643, 1519, 1492, 1444, 1390, 1365, 1269, 1251, 1170, 1091, 1043, 1016, 667 cm^{-1} .

HRMS: Calc'd $\text{C}_{16}\text{H}_{22}\text{ClNO}_2\text{Na}$ ($\text{M}+\text{Na}$)⁺ = 318.1237, found = 318.1220.

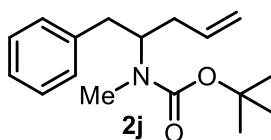


¹H NMR (500 MHz, CDCl_3) δ 7.43 (m, 2H), 7.39 (ddd, J = 7.65, 6.80, 1.29 Hz, 2H), 7.33 (m, 1H), 7.11 (d, J = 8.35 Hz, 2H), 6.91 (m, 2H), 5.79 (ddt, J = 17.5, 10.5, 7.02 Hz, 1H), 5.09 (m, 2H), 5.04 (s, 2H), 4.39 (s, 1H), 3.87 (s, 1H), 2.72 (m, 2H), 2.45 (m, 1H), 2.10 (m, 1H), 1.41 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 157.6, 155.5, 137.2, 134.6, 130.6, 128.7, 128.1, 127.6, 118.0, 114.9, 79.2, 70.2, 51.3, 39.7, 38.1, 28.5.

IR (film) 3365, 2977, 2929, 1687, 1610, 1510, 1454, 1390, 1365, 1242, 1170, 1024, 914, 742, 696, 667 cm^{-1} .

HRMS: Calc'd $\text{C}_{23}\text{H}_{29}\text{NO}_3\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 390.2045, found = 390.2039.



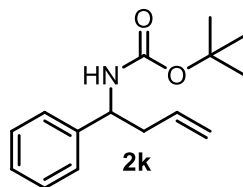
A mixture of rotamers.

^1H NMR (500 MHz, CDCl_3) δ 7.26 (m, 2H), 7.17 (m, 3H), 5.73 (br m, 1H), 5.06 (br m, 2H), 4.47 (br m, 0.4H), 4.29 (br m, 0.6H) 2.77 (d, J = 7.76 Hz, 1H), 2.71 (t, J = 3.39 Hz, 3H), 2.62 (s, 1H), 2.27 (br m, 2H), 1.36 (s, 4H), 1.28 (s, 5H).

^{13}C NMR (126 MHz, CDCl_3) δ 156.0, 139.0 (br s), 135.2 (br s), 129.1 (br s), 128.4 (br s), 126.3 (br s), 116.9 (br s), 79.2 (br s), 56.5 (br s), 38.9 (br s), 36.9 (br s), 29.5 (br s), 28.4 (br s).

IR (film) 3028, 2975, 2927, 1693, 1643, 1479, 1454, 1392, 1365, 1298, 1253, 1224, 1172, 1143, 991, 914, 873, 769, 746, 700 cm^{-1} .

HRMS: Calc'd $\text{C}_{17}\text{H}_{25}\text{NO}_2\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 298.1783, found = 298.1772.

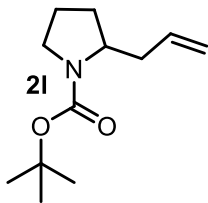


^1H NMR (500 MHz, CDCl_3) δ 7.32 (m, 2H), 7.26 (m, 3H), 5.68 (ddt, J = 17.2, 10.2, 7.05 Hz, 1H), 5.09 (m, 2H), 4.87 (s, 1H), 4.74 (s, 1H), 2.52 (m, 2H), 1.42 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 155.3, 142.5, 134.1, 128.6, 127.3, 126.4, 118.3, 79.6, 54.1, 41.4, 28.5.

IR (film) 3338, 3064, 2977, 2931, 1693, 1519, 1494, 1454, 1367, 1251, 1170, 1018, 916, 756, 700 cm^{-1} .

HRMS: Calc'd $\text{C}_{15}\text{H}_{21}\text{NO}_2\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 270.1470, found = 270.1468.



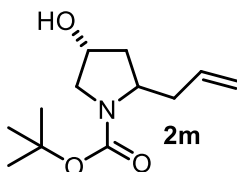
A mixture of rotamers.

^1H NMR (500 MHz, CDCl_3) δ 5.73 (br m, 1H), 5.04 (m, 2H), 3.80 (br m, 1H), 3.35 (br m, 2H), 2.45 (br m, 1H), 2.11 (br m, 1H), 1.80 (br m, 4H), 1.46 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 154.7, 135.4, 117.1, 79.1 (br s), 56.9, 46.6 (br s), 38.8 (br s), 29.8 (br s), 28.70, 23.4 (br s).

IR (film) 3076, 2974, 2875, 1693, 1641, 1479, 1454, 1392, 1365, 1253, 1172, 1108, 995, 914, 771 cm^{-1} .

HRMS: Calc'd $\text{C}_{12}\text{H}_{21}\text{NO}_2\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 234.1470, found = 234.1452.



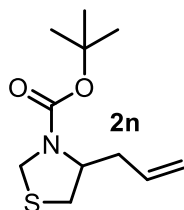
A mixture of diastereomers and rotamers.

^1H NMR (500 MHz, CDCl_3) δ 5.73 (br m, 1H), 5.07 (br m, 2H), 4.38 (br m, 1H), 3.43 (br m, 3H), 2.34 (br m, 2H), 2.09 (br m, 2H), 1.84 (br m, 1H), 1.46 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3 , major resonances reported) δ 154.6, 135.4, 117.6, 79.6, 70.1 (br s), 56.6, 55.4 (br s), 39.4 (br s), 38.2 (br s), 28.6.

IR (film) 3419, 2975, 2931, 1693, 1668, 1407, 1365, 1255, 1166, 1118, 1080, 914, 765 cm^{-1} .

HRMS: Calc'd $\text{C}_{12}\text{H}_{21}\text{NO}_3\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 250.1419, found = 250.1411.



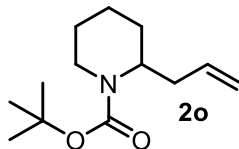
A mixture of rotamers.

^1H NMR (500 MHz, CDCl_3) δ 5.76 (ddt, J = 17.1, 10.1, 7.01 Hz, 1H), 5.09 (m, 2H), 4.62 (br s, 1H), 4.22 (br m, 2H), 3.07 (m, 1H), 2.80 (dd, J = 11.2, 2.53 Hz, 1H), 2.46 (br s, 1H), 2.33 (m, 1H), 1.47 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 153.4, 134.7, 118.0, 80.6, 59.2, 48.0 (br s), 37.7 (br s), 34.6 (br s), 28.5.

IR (film) 3367, 2977, 2931, 1735, 1693, 1384, 1369, 1330, 1255, 1151, 1041, 918, 852, 767, 667 cm^{-1} .

HRMS: $[\text{M} - \text{isobutylene} + \text{H}]^+ = 174.0589$, found = 174.0581. GC/MS: $[\text{M}]^+ = 229.1$, found = 229.2.

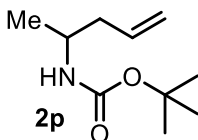


^1H NMR (500 MHz, CDCl_3) δ 5.74 (ddt, $J = 17.2, 10.1, 7.15$ Hz, 1H), 5.01 (m, 2H), 4.27 (s, 1H), 3.96 (d, $J = 12.3$ Hz, 1H), 2.75 (m, 1H), 2.39 (m, 1H), 2.22 (dt, $J = 14.1, 7.19$ Hz, 1H), 1.57 (m, 5H), 1.44 (s, 9H), 1.39 (m, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 115.2, 135.8, 116.7, 79.2, 50.2, 39.0, 34.6, 28.6, 27.8, 25.6, 19.0.

IR (film) 3076, 2975, 2933, 2858, 1693, 1643, 1446, 1409, 1365, 1319, 1267, 1170, 1147, 1068, 1033, 995, 912, 869, 806, 767 cm^{-1} .

HRMS: Calc'd $\text{C}_{13}\text{H}_{23}\text{NO}_2\text{Na}$ ($\text{M}+\text{Na}$) $^+ = 248.1626$, found = 248.1615.

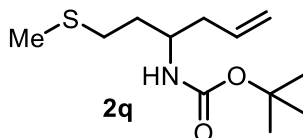


^1H NMR (500 MHz, CDCl_3) δ 5.77 (m, 1H), 5.08 (m, 2H), 4.38 (s, 1H), 3.73 (m, 1H), 2.19 (m, 2H), 1.43 (s, 9H), 1.12 (d, $J = 6.63$ Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 155.4, 134.6, 117.8, 79.2, 46.0, 41.4, 28.6, 20.7.

IR (film) 3338, 3078, 2977, 2931, 1693, 1643, 1519, 1454, 1390, 1365, 1251, 1174, 1056, 993, 914, 781, 634 cm^{-1} .

HRMS: Calc'd $\text{C}_{10}\text{H}_{19}\text{NO}_2\text{Na}$ ($\text{M}+\text{Na}$) $^+ = 208.1313$, found = 208.1293.

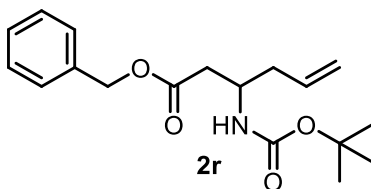


^1H NMR (500 MHz, CDCl_3) δ 5.76 (ddt, $J = 17.6, 9.55, 7.13$ Hz, 1H), 5.09 (m, 2H), 4.39 (d, $J = 9.23$ Hz, 1H), 3.74 (m, 1H), 2.52 (m, 2H), 2.23 (m, 2H), 2.10 (s, 3H), 1.79 (m, 1H), 1.63 (m, 1H), 1.43 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 155.7, 134.2, 118.2, 79.3, 49.7, 39.7, 34.6, 30.9, 28.5, 15.8.

IR (film) 3336, 3076, 2977, 2918, 1693, 1641, 1519, 1442, 1390, 1365, 1290, 1251, 1170, 1049, 1020, 993, 914, 856, 779, 748, 649 cm^{-1} .

HRMS: Calc'd $\text{C}_{12}\text{H}_{23}\text{NO}_2\text{SNa}$ ($\text{M}+\text{Na}$)⁺ = 268.1347, found = 268.1338.

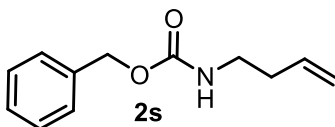


¹H NMR (500 MHz, CDCl_3) δ 7.36 (m, 5H), 5.74 (ddt, J = 17.4, 10.5, 7.15 Hz, 1H), 5.08 (m, 4H), 4.95 (m, 1H), 4.01 (m, 1H), 2.58 (d, J = 5.73 Hz, 2H), 2.29 (m, 2H), 1.42 (s, 9H).

¹³C NMR (126 MHz, CDCl_3) δ 171.6, 155.3, 135.8, 134.1, 128.8, 128.5, 128.4, 118.4, 79.5, 66.6, 47.3, 38.9, 38.5, 28.5.

IR (film) 3359, 3066, 2977, 1714, 1697, 1643, 1504, 1456, 1390, 1365, 1249, 1170, 1047, 1026, 993, 918, 750, 698 cm^{-1} .

HRMS: Calc'd $\text{C}_{18}\text{H}_{25}\text{NO}_4\text{Na}$ ($\text{M}+\text{Na}$)⁺ = 342.1681, found = 342.1674.

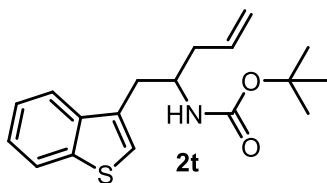


¹H NMR (500 MHz, CDCl_3) δ 7.35 (m, 5H), 5.75 (ddt, J = 17.2, 10.4, 6.82 Hz, 1H), 5.09 (m, 4H), 4.79 (s, 1H), 3.28 (q, J = 6.48 Hz, 2H), 2.27 (q, J = 6.92 Hz, 2H).

¹³C NMR (126 MHz, CDCl_3) δ 156.4, 136.7, 135.2, 128.6, 128.3, 128.2, 117.5, 66.8, 40.2, 34.2.

IR (film) 3330, 3074, 3031, 2935, 1697, 1641, 1523, 1454, 1253, 1217, 1134, 1026, 914, 748, 695 cm^{-1} .

HRMS: Calc'd $\text{C}_{12}\text{H}_{15}\text{NO}_2\text{Na}$ ($\text{M}+\text{Na}$)⁺ = 228.1000, found = 228.0989.

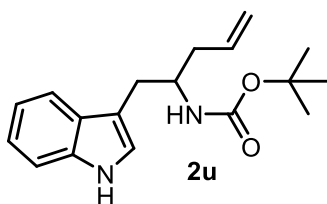


^1H NMR (500 MHz, CDCl_3) δ 7.86 (m, 2H), 7.38 (m, 2H), 7.17 (s, 1H), 5.82 (ddt, $J = 17.3, 10.4, 7.02$ Hz, 1H), 5.12 (m, 2H), 4.48 (d, $J = 8.89$ Hz, 1H), 4.08 (m, 1H), 3.10 (m, 1H), 3.00 (dd, $J = 14.3, 6.99$ Hz, 1H), 2.32 (m, 1H), 2.18 (m, 1H), 1.43 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 155.5, 140.4, 139.3, 134.4, 132.9, 124.4, 124.2, 123.3, 122.9, 122.2, 118.3, 79.4, 49.8, 38.5, 33.6, 28.5.

IR (film) 3425, 3340, 3074, 2975, 2929, 1697, 1641, 1502, 1458, 1429, 1390, 1365, 1251, 1168, 1047, 1020, 914, 852, 759, 732 cm^{-1} .

HRMS: Calc'd $\text{C}_{18}\text{H}_{23}\text{NO}_2\text{SNa}$ ($\text{M}+\text{Na}$) = 340.1347, found 340.1330.

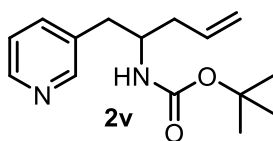


^1H NMR (500 MHz, CDCl_3) δ 8.10 (s, 1H), 7.65 (d, $J = 7.92$ Hz, 1H), 7.36 (dt, $J = 8.16, 0.96$ Hz, 1H), 7.20 (ddd, $J = 8.14, 6.97, 1.20$ Hz, 1H), 7.13 (ddd, $J = 8.04, 7.03, 1.06$ Hz, 1H), 7.03 (s, 1H), 5.83 (m, 1H), 5.09 (m, 2H), 4.48 (s, 1H), 4.02 (m, 1H), 2.95 (m, 2H), 2.32 (m, 1H), 2.15 (m, 1H), 1.43 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 155.7, 136.3, 134.9, 128.1, 122.8, 122.1, 119.6, 119.3, 117.8, 112.3, 111.1, 79.2, 50.5, 38.6, 30.1, 28.6.

IR (film) 3411, 3325, 3056, 2975, 2929, 1693, 1641, 1504, 1438, 1392, 1365, 1249, 1168, 1099, 1060, 1010, 914, 742 cm^{-1} .

HRMS: Calc'd $\text{C}_{18}\text{H}_{24}\text{N}_2\text{O}_2\text{Na}$ ($\text{M}+\text{Na}$)⁺ = 323.1735, found = 323.1725.

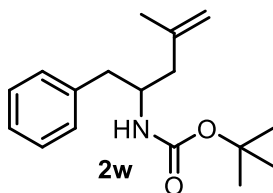


^1H NMR (500 MHz, CDCl_3) δ 8.47 (dd, $J = 4.89, 1.67$ Hz, 1H), 8.43 (s, 1H), 7.54 (m, 1H), 7.22 (dd, $J = 7.63, 4.92$ Hz, 1H), 5.78 (ddt, $J = 17.2, 10.3, 7.05$ Hz, 1H), 5.12 (m, 2H), 4.42 (m, 1H), 3.90 (m, 1H), 2.80 (m, 2H), 2.25 (m, 1H), 2.13 (m, 1H), 1.38 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 155.4, 150.8, 148.0, 137.0, 134.0, 133.8, 123.4, 118.5, 79.5, 50.9, 38.4, 37.8, 28.5.

IR (film) 3336, 3240, 2977, 2929, 1693, 1519, 1440, 1425, 1390, 1365, 1249, 1170, 1045, 1027, 914, 844, 715 cm^{-1} .

HRMS: Calc'd $C_{15}H_{23}N_2O_2$ (M+H)⁺ = 263.1760, found = 263.1764.

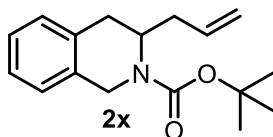


¹H NMR (500 MHz, CDCl₃) δ 7.29 (m, 2H), 7.21 (m, 3H), 4.81 (s, 1H), 4.73 (s, 1H), 4.32 (s, 1H), 3.99 (s, 1H), 2.79 (m, 2H), 2.18 (dd, *J* = 14.3, 5.57 Hz, 1H), 2.06 (dd, *J* = 14.2, 8.97 Hz, 1H), 1.72 (s, 3H), 1.40 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 155.5, 142.6, 138.3, 129.7, 128.4, 126.4, 113.3, 79.2, 49.6, 42.8, 41.1, 28.5, 22.2.

IR (film) 3350, 3072, 3028, 2975, 2929, 1701, 1496, 1454, 1390, 1365, 1247, 1170, 1049, 1016, 889, 746, 700, 667 cm⁻¹.

HRMS: Calc'd $C_{17}H_{25}NO_2Na$ (M+Na)⁺ = 298.1783, found = 298.1786.



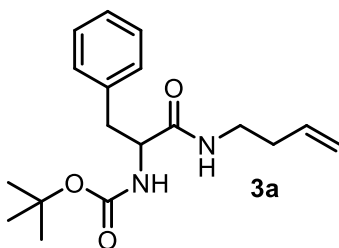
A mixture of rotamers.

¹H NMR (500 MHz, CDCl₃) δ 7.18 (m, 2H), 7.11 (m, 2H), 5.78 (m, 1H), 5.00 (m, 2H), 4.48 (br m, 1H), 4.54 (br m, 1H), 4.26 (s, 0.5H), 4.23 (s, 0.5H), 3.03 (dd, *J* = 15.9, 5.93 Hz, 1H), 2.69 (d, *J* = 15.3 Hz, 1H), 2.24 (m, 1H), 2.06 (m, 1H), 1.49 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 155.1, 135.2 (br s), 133.2 (br s), 132.9 (br s), 129.4 (br s), 126.7, 126.3 (br s), 117.3 (br s), 79.9, 49.2 (br s), 42.9 (br s), 36.7 (br s), 32.6 (br s), 28.6.

IR (film) 3350, 2975, 2931, 1697, 1456, 1392, 1367, 1245, 1168, 1124, 1010, 993, 914, 862, 753, 648 cm⁻¹.

HRMS: Calc'd $C_{17}H_{23}NO_2Na$ (M+Na)⁺ = 296.1626, found = 296.1620.

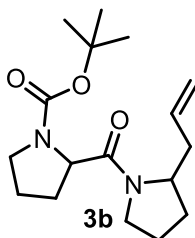


¹H NMR (500 MHz, CDCl₃) δ 7.29 (tt, *J* = 6.98, 1.11 Hz, 2H), 7.24 (m, 1H), 7.20 (m, 2H), 5.72 (br s, 1H), 5.61 (ddt, *J* = 17.1, 10.6, 6.89 Hz, 1H), 5.08 (br s, 1H), 4.97 (m, 2H), 4.26 (q, *J* = 7.51 Hz, 1H), 3.22 (m, 2H), 3.04 (m, 2H), 2.09 (m, 2H), 1.41 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 171.1, 155.5, 137.0, 135.0, 129.4, 128.8, 127.0, 117.4, 80.2, 56.2, 38.9, 38.4, 33.5, 28.4.

IR (film) 3415, 2256, 2129, 1650, 1519, 1496, 1454, 1365, 1249, 1170, 1049, 1026, 1000, 825, 763, 700, 632 cm^{-1} .

HRMS: Calc'd $\text{C}_{18}\text{H}_{26}\text{N}_2\text{O}_3\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 341.1841, found = 341.1835.



A mixture of diastereomers and rotamers.

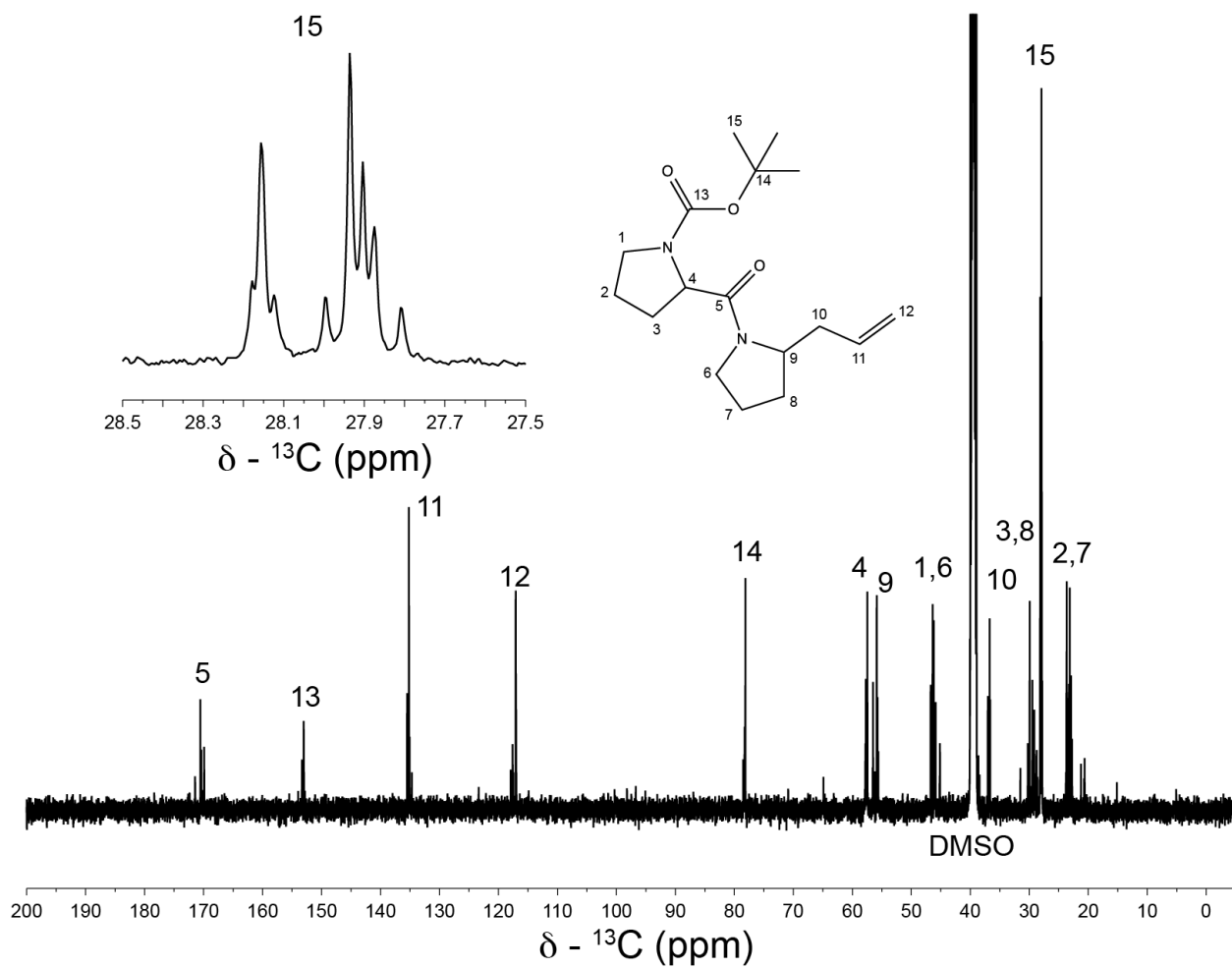
^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 5.86 – 5.68 (m, 1H), 5.18 – 4.98 (m, 2H), 4.43 – 4.29 (m, 1H), 4.09 – 3.79 (m, 1H), 3.60 – 3.22 (m, 4H), 2.47 – 2.29 (m, 1H), 2.29 – 1.95 (m, 2H), 1.94 – 1.61 (m, 7H), 1.48 – 1.22 (m, 9H).

In the 1D ^{13}C spectrum of **3b**, there are 8 peaks for each carbon, corresponding to 2 diastereomers and 4 rotamers for each diastereomer (see below).

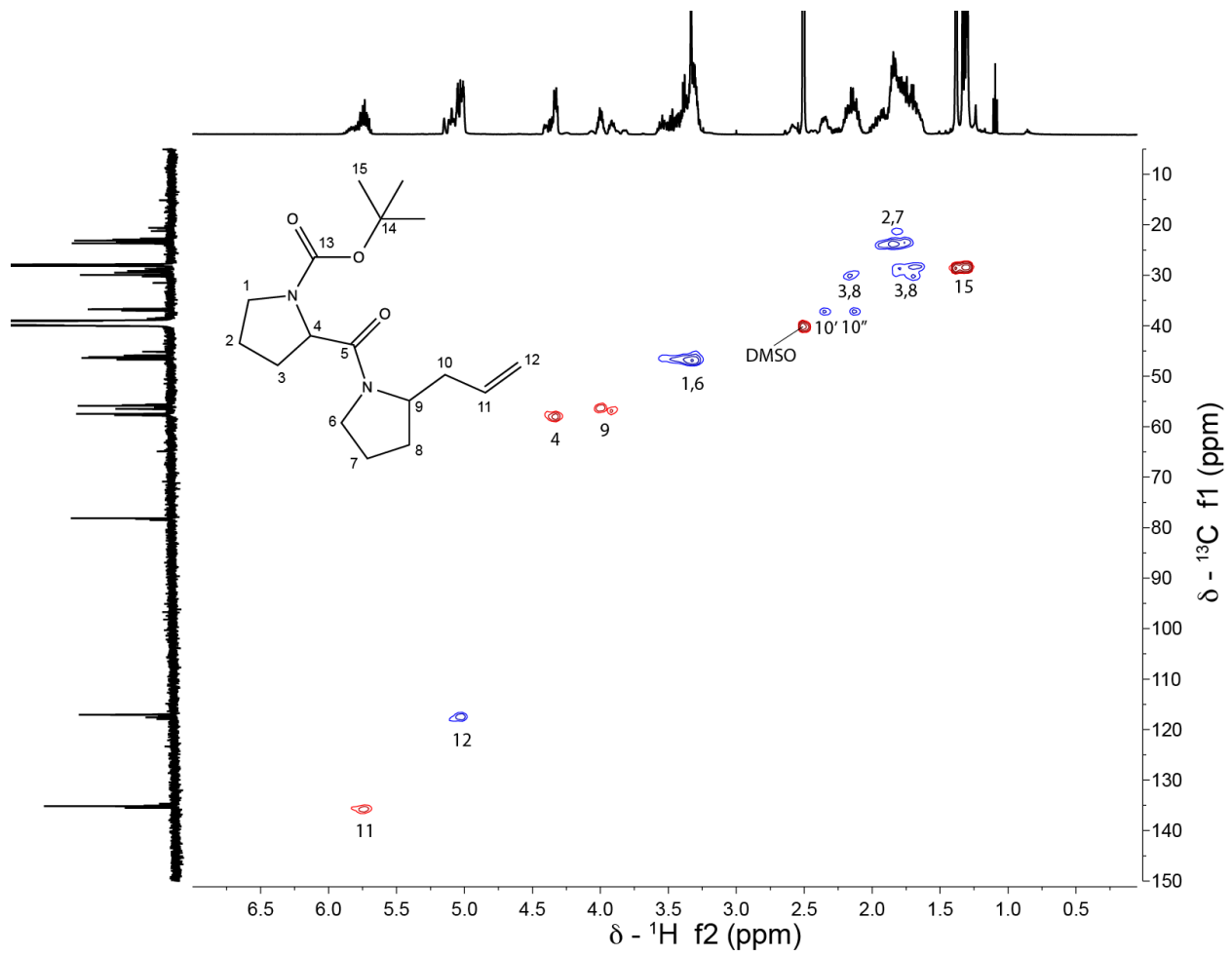
^{13}C NMR (126 MHz, $\text{DMSO}-d_6$) δ 171.4, 170.5, 170.4, 169.9, 153.3, 153.0, 153.0, 135.5, 135.5, 135.2, 135.1, 134.7, 117.9, 117.6, 117.5, 117.1, 117.0, 117.0, 78.5, 78.3, 78.3, 78.1, 64.9, 57.8, 57.7, 57.7, 57.6, 57.5, 57.5, 56.6, 56.5, 56.2, 55.9, 55.7, 55.7, 55.6, 46.7, 46.6, 46.5, 46.4, 46.4, 46.2, 46.1, 46.1, 46.0, 45.9, 45.2, 45.1, 40.1, 40.0, 39.9, 39.8, 39.7, 39.6, 39.6, 39.5, 39.4, 39.3, 39.1, 39.0, 38.6, 38.4, 37.0, 36.7, 36.6, 31.5, 30.3, 29.9, 29.5, 29.4, 29.1, 29.0, 28.7, 28.6, 28.2, 28.2, 28.1, 28.0, 27.9, 27.9, 27.9, 27.8, 23.8, 23.7, 23.7, 23.6, 23.6, 23.4, 23.3, 23.2, 23.1, 22.9, 22.7, 21.2, 20.7, 20.6, 15.1.

IR (film) 3456, 2974, 2875, 1693, 1645, 1427, 1398, 1259, 1163, 1120, 1085, 997, 912, 748 cm^{-1} .

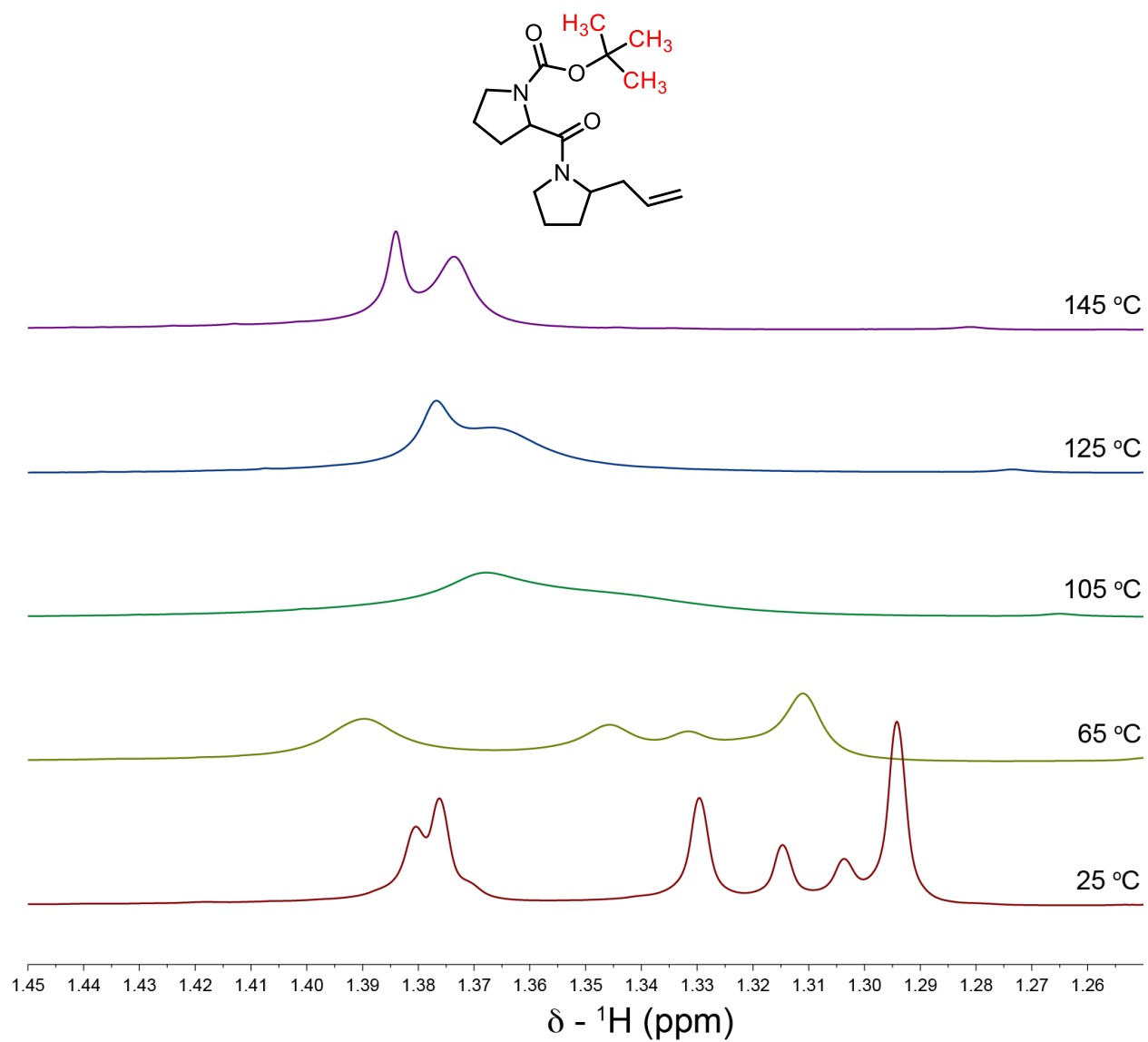
HRMS: Calc'd $\text{C}_{17}\text{H}_{29}\text{N}_2\text{O}_3$ ($\text{M}+\text{H}$) $^+$ = 309.2178, found = 309.2163.

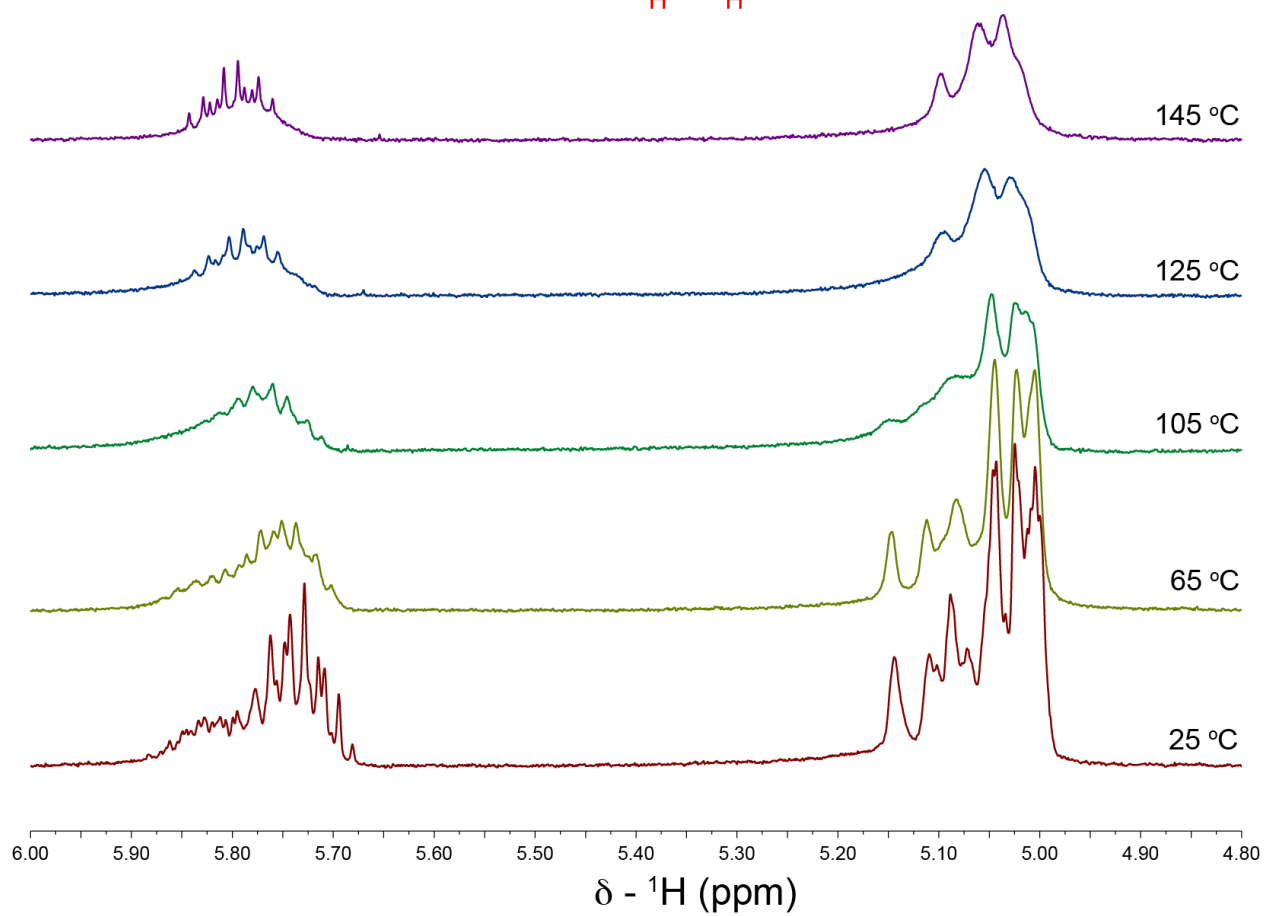
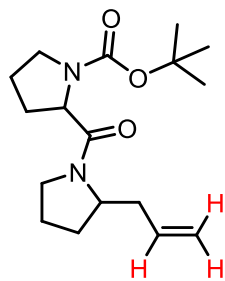


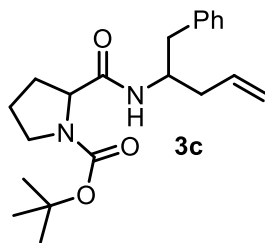
To assign this spectrum, we used a combination of 1D and 2D homo- and heteronuclear NMR (see 2D ^1H - ^{13}C HSQC spectrum below).



To confirm the presence of rotamers, variable temperature NMR was performed. At 145 °C the rotamers coalesce and only two peaks are present in 1D ¹H and 1D ¹³C spectra.







Mixture of diastereomers and rotamers

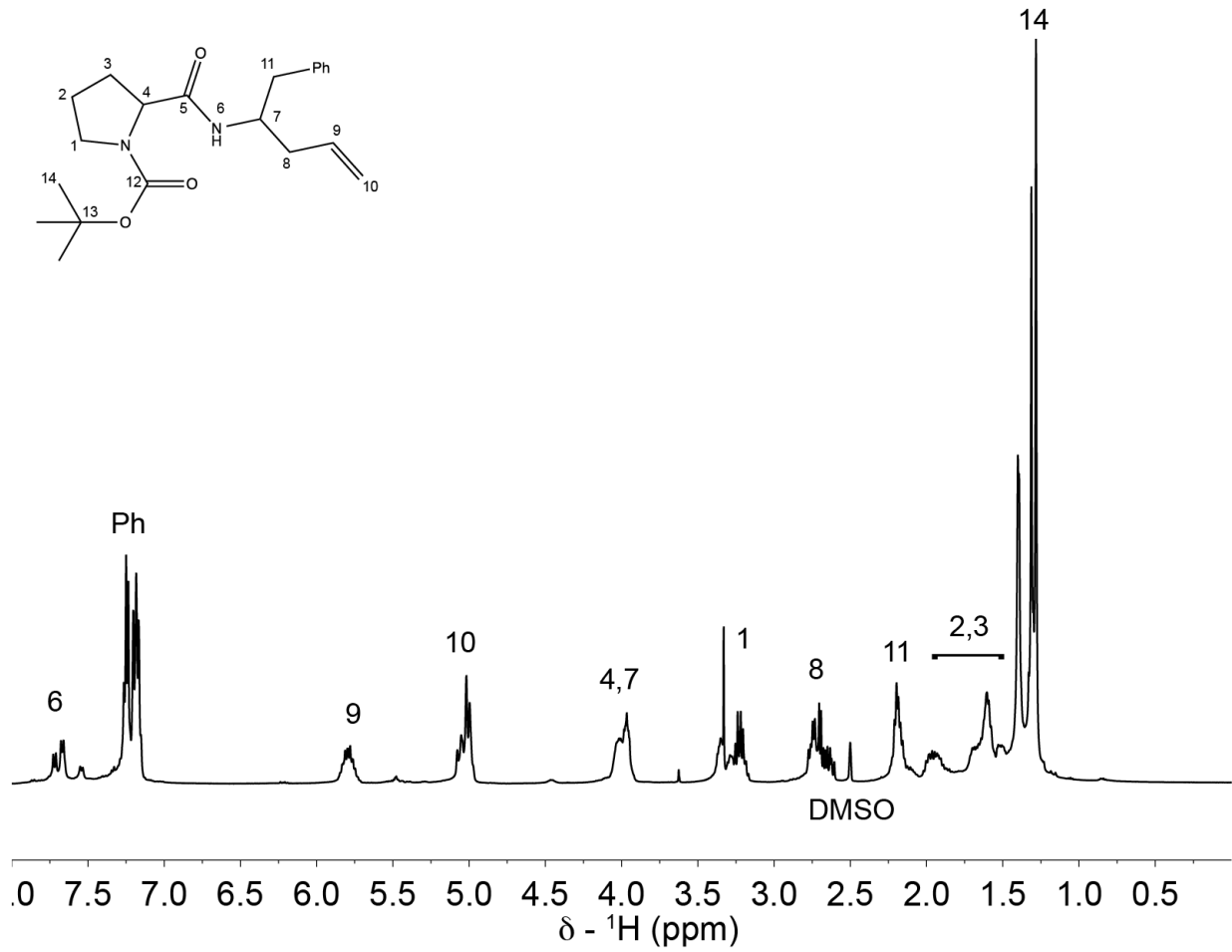
^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ 7.78 – 7.49 (m, 1H), 7.36 – 7.11 (m, 5H), 5.90 – 5.69 (m, 1H), 5.13 – 4.92 (m, 2H), 4.12 – 3.88 (m, 2H), 3.41 – 3.16 (m, 2H), 2.82 – 2.58 (m, 2H), 2.30 – 2.08 (m, 2H), 2.05 – 1.84 (m, 1H), 1.78 – 1.47 (m, 3H), 1.45 – 1.24 (m, 9H).

^{13}C NMR at 105 $^\circ\text{C}$ (126 MHz, $\text{DMSO-}d_6$) δ 172.0, 154.2, 139.4, 139.4, 135.9, 135.8, 129.5, 129.5, 128.5, 128.4, 126.4, 126.3, 117.0, 117.0, 79.6, 79.0, 78.9, 60.5, 60.3, 50.1, 50.0, 47.0, 47.0, 40.4, 40.3, 38.9, 38.6, 31.0, 28.6, 28.6, 23.6.

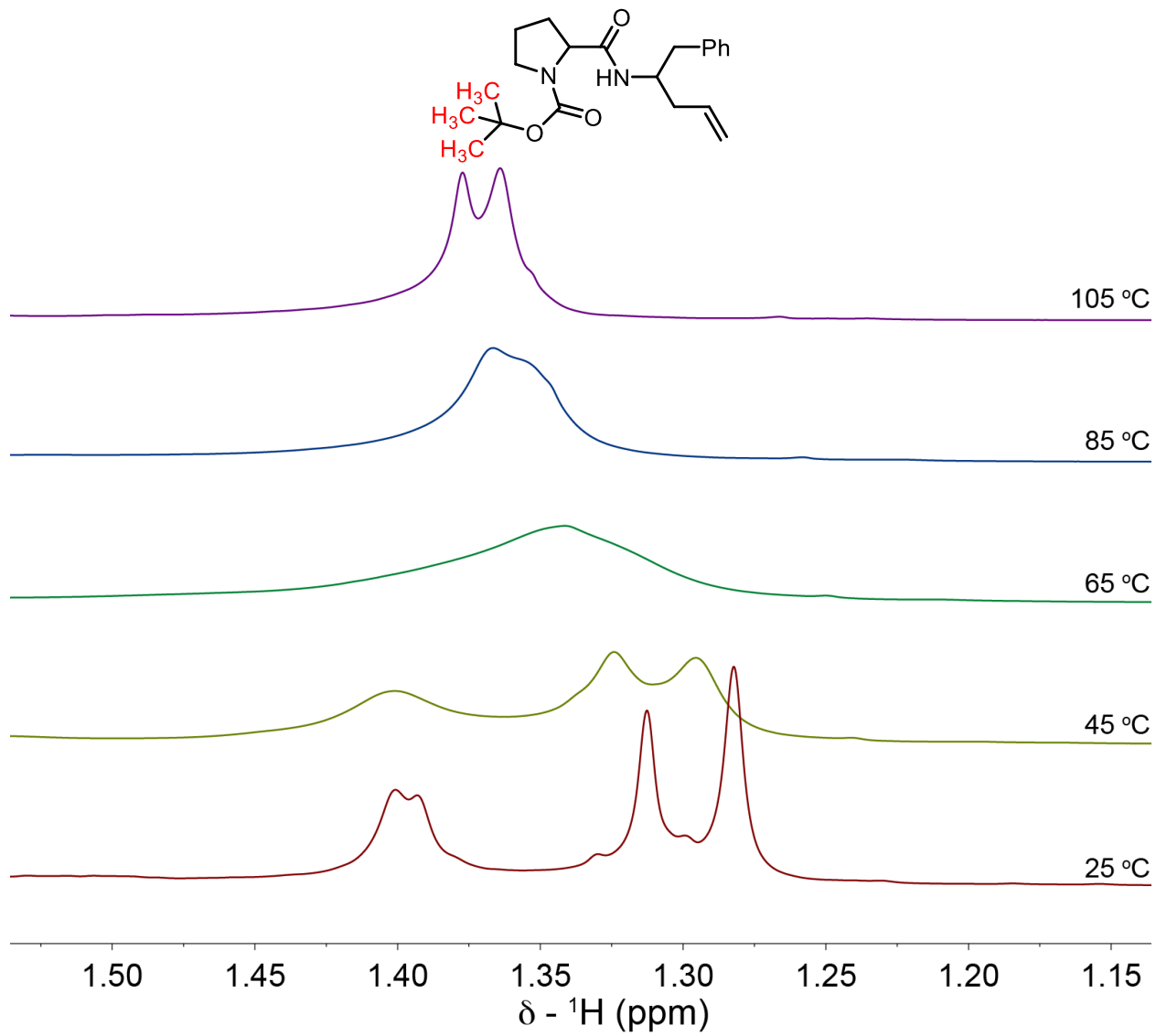
IR (film) 3303, 3074, 2975, 2877, 1697, 1660, 1552, 1454, 1392, 1365, 1257, 1163, 1122, 912, 748 cm^{-1} .

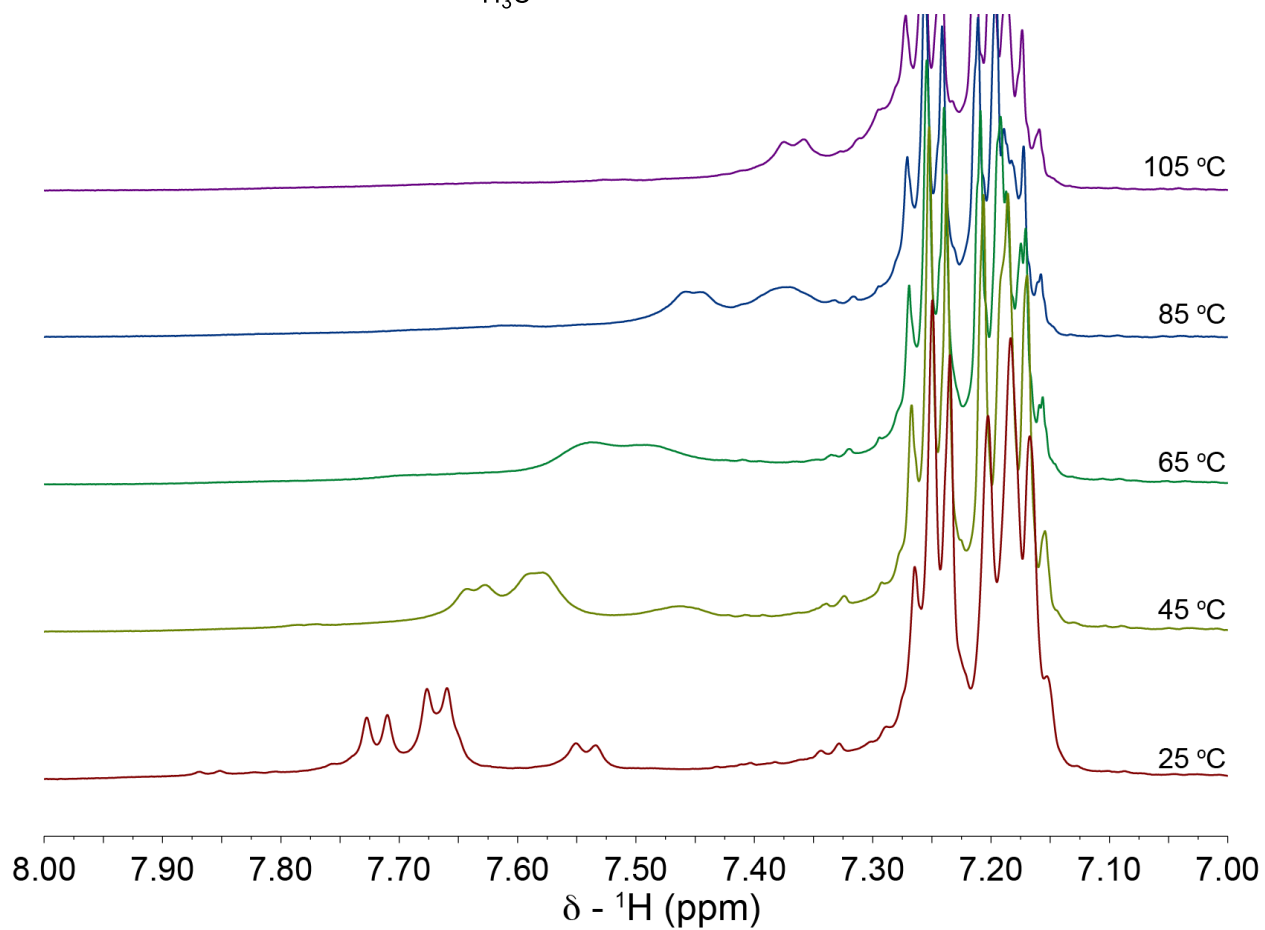
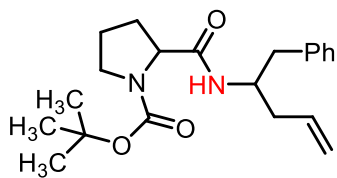
HRMS: Calc'd $\text{C}_{21}\text{H}_{30}\text{N}_2\text{O}_3\text{Na}$ ($\text{M}+\text{Na}$) $^+$ = 381.2154, found = 381.2149.

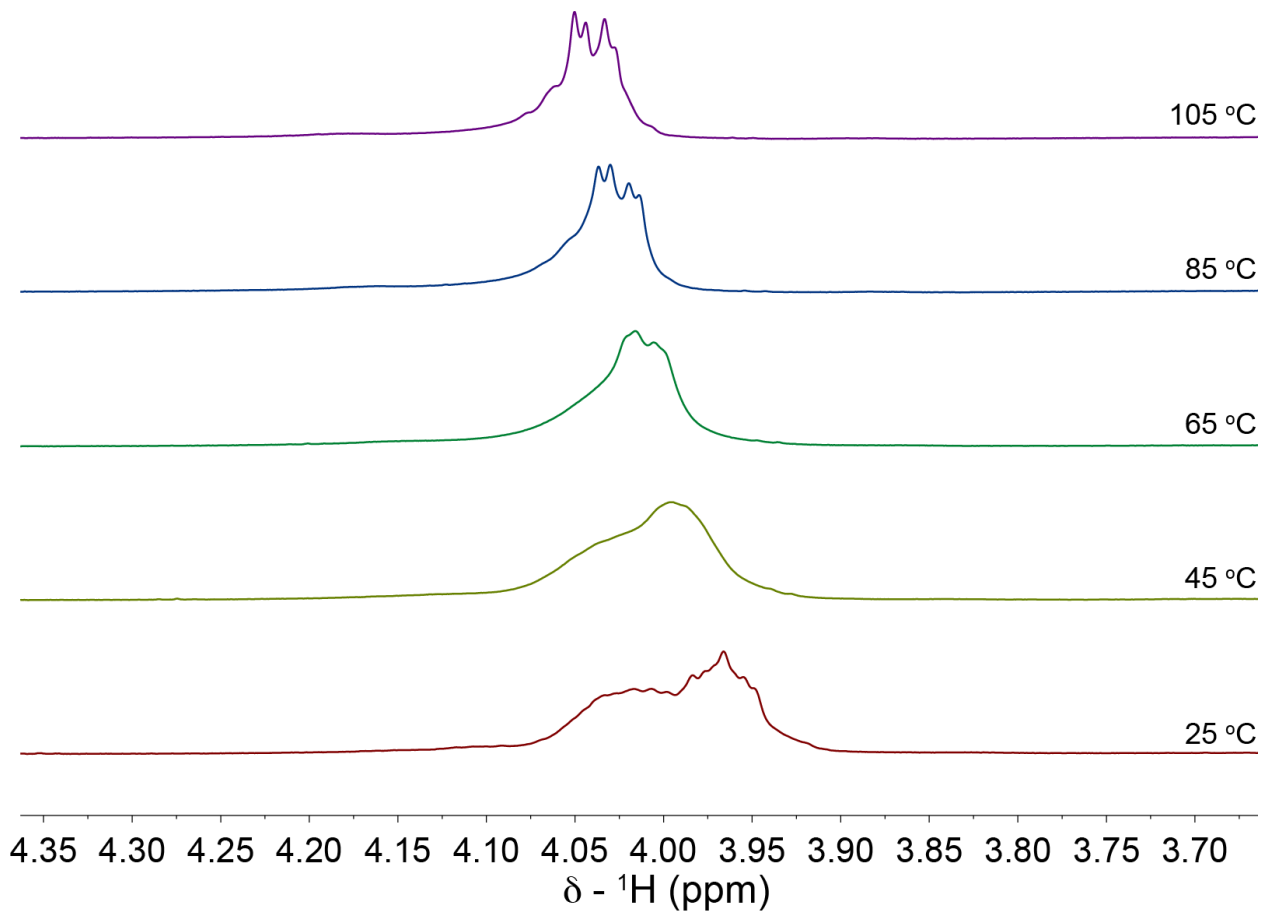
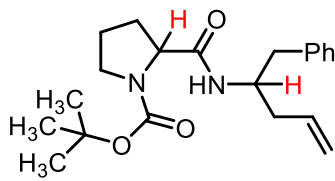
At room temperature the 1D ^1H of **3c** includes a mixture of diastereomers and rotamers. Upon raising the temperature to 105 °C, these peaks coalesce into discernable signals in the 1D ^1H and 1D ^{13}C spectra (see below)

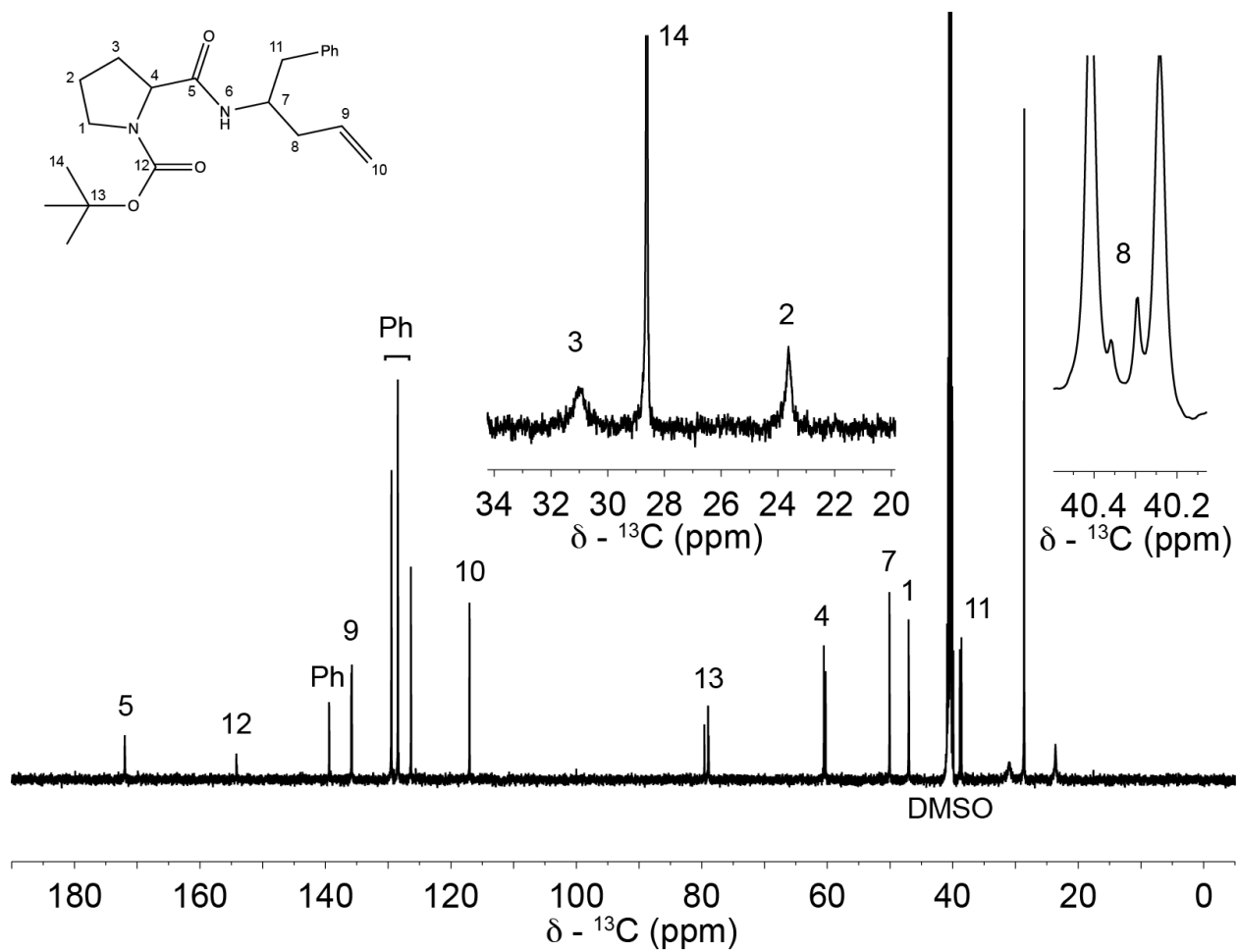


Room Temperature 1D ^1H





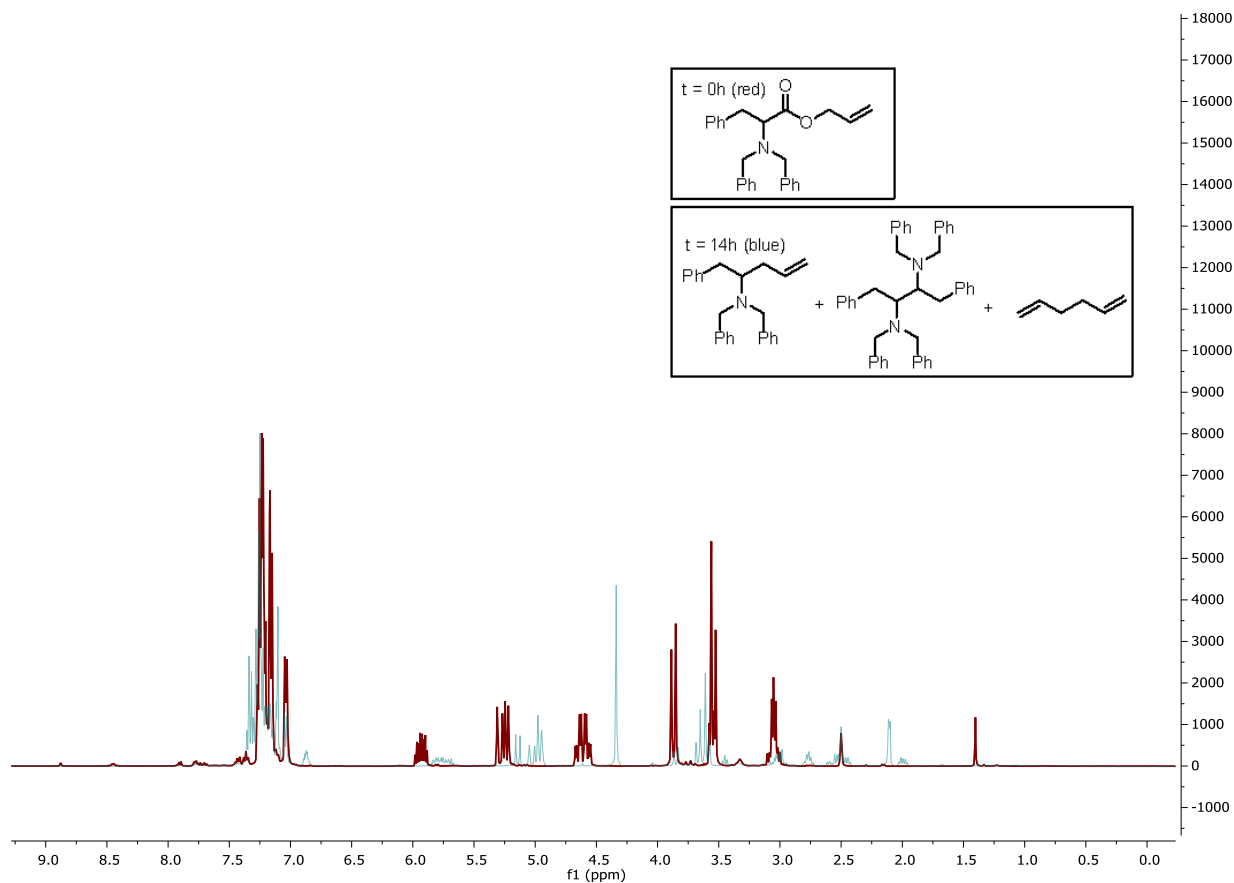


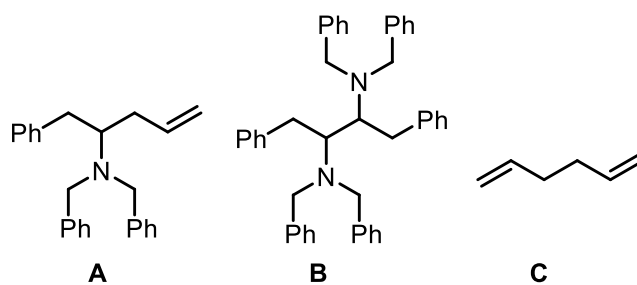
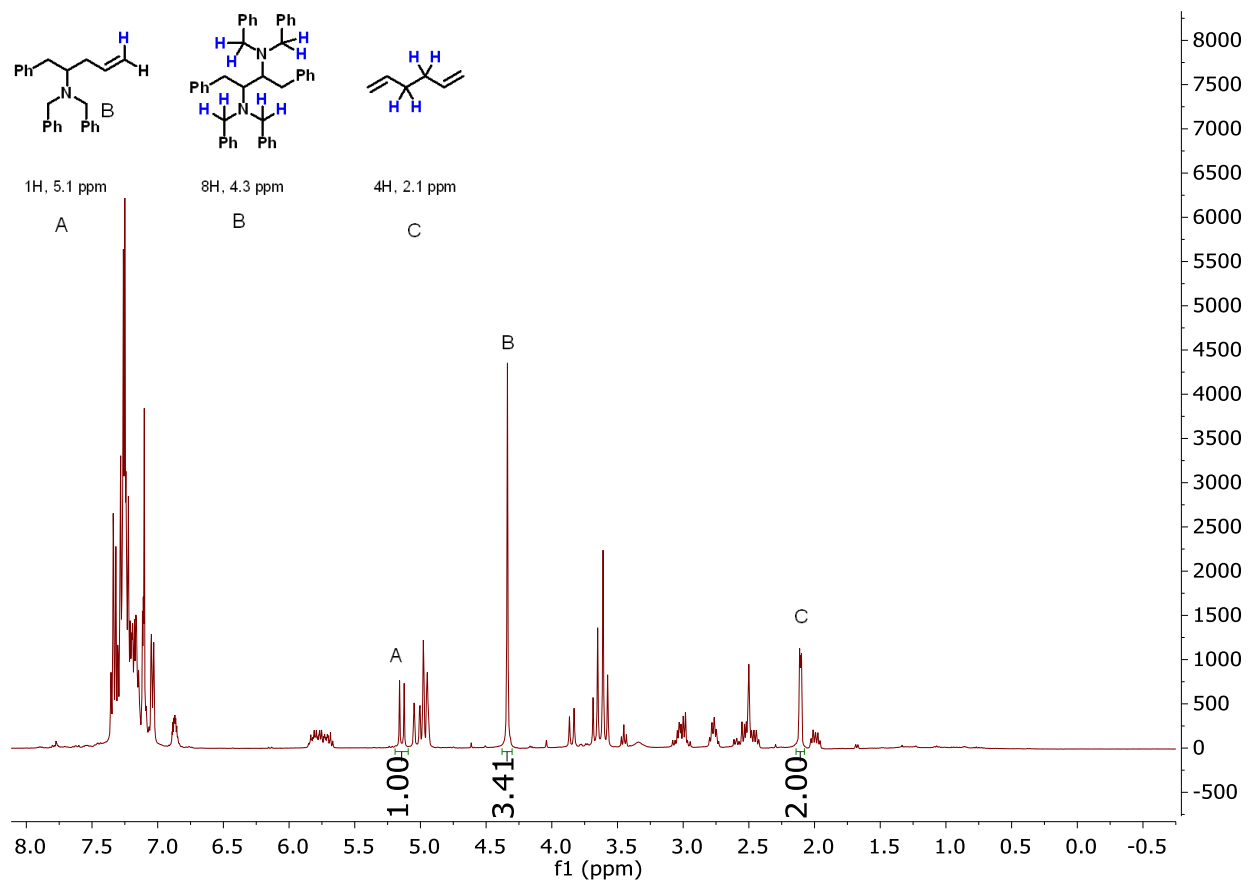


1D ^{13}C at 105 °C

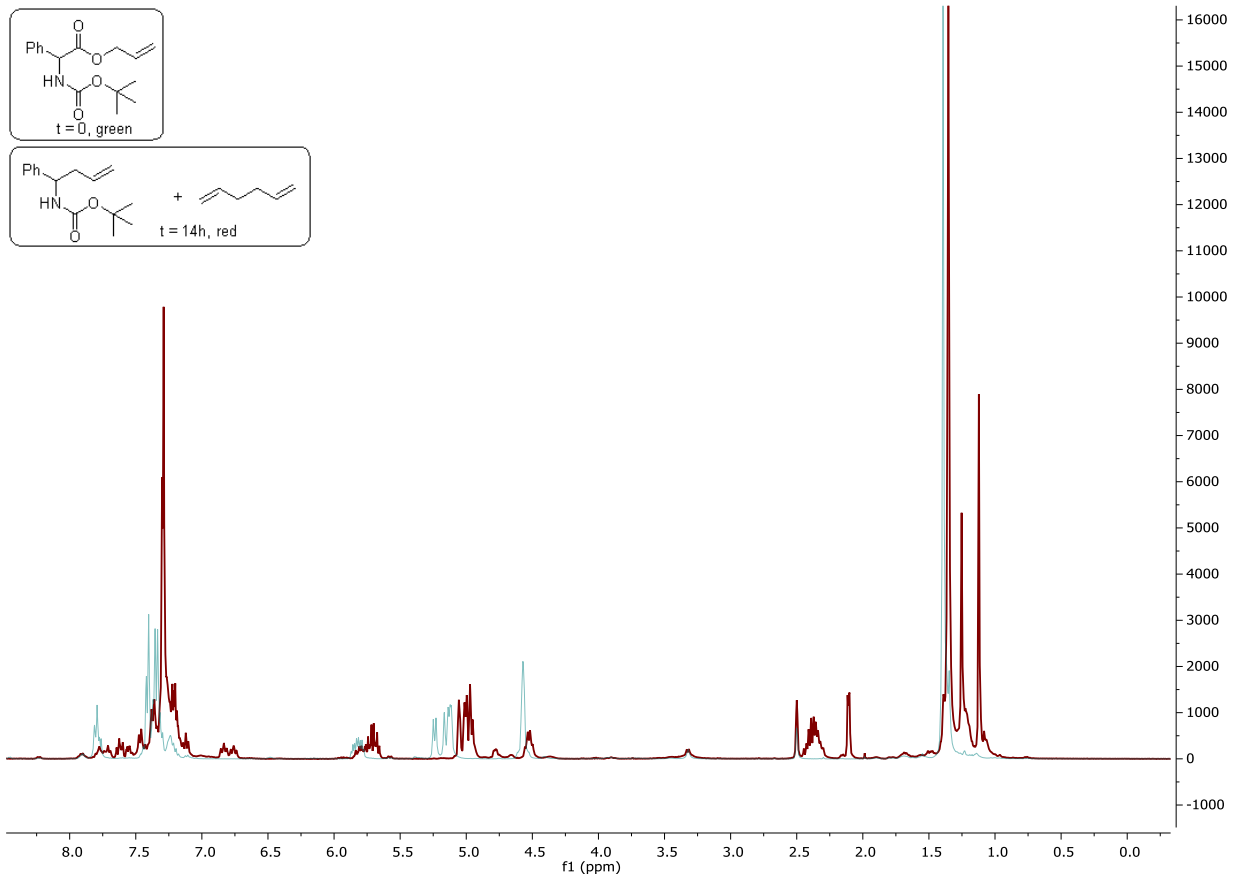
5. ^1H NMR Spectroscopy Experiments

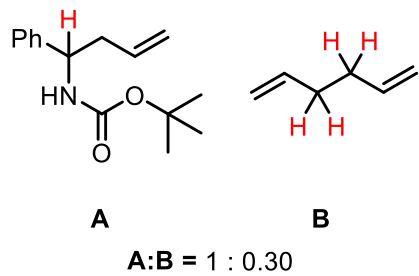
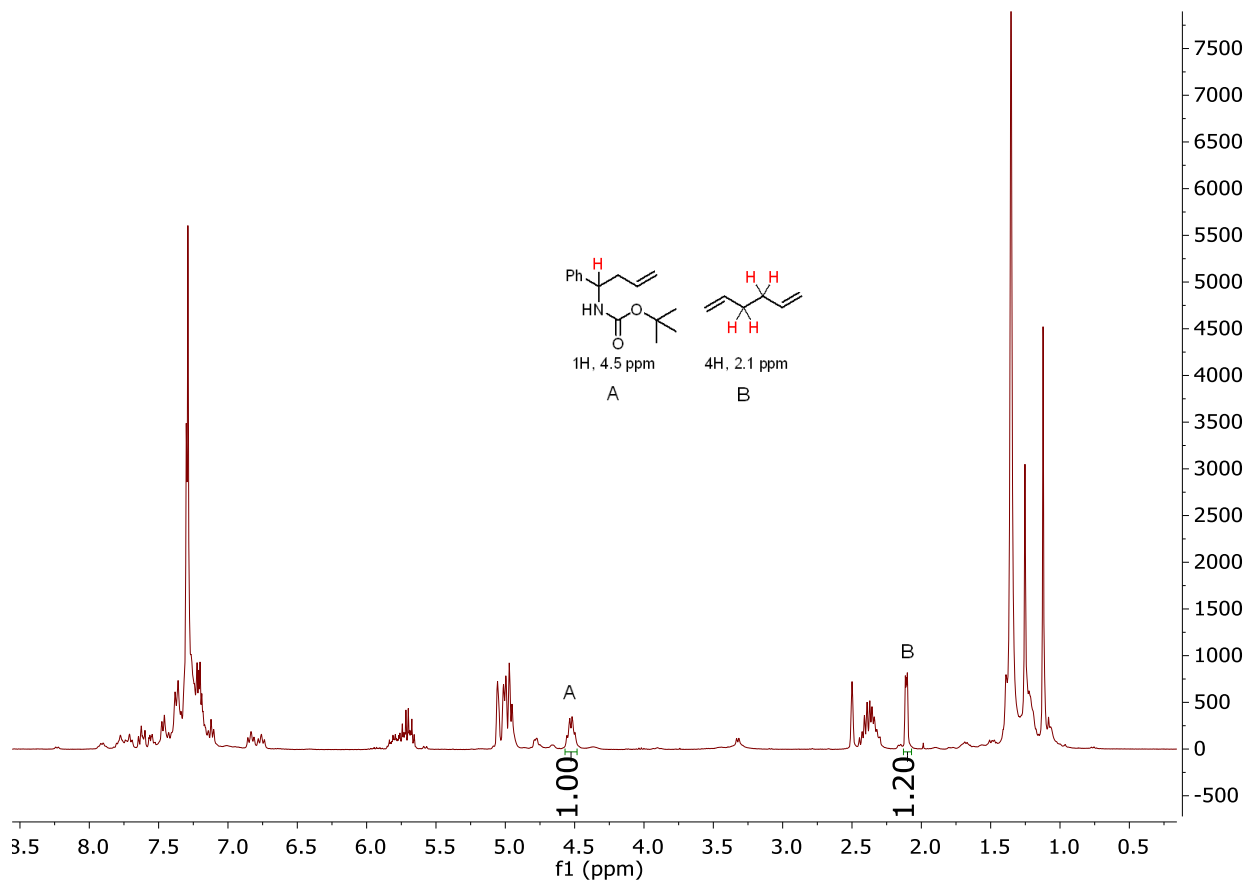
General Procedure: An oven dried 16 x 125 mm threaded glass tube and an oven dried NMR tube were taken into a glovebox. Allyl ester (0.25 mmol), was added to the threaded glass tube followed by $\text{Pd}(\text{PPh}_3)_4$ and $[\text{Ir}\{\text{dF}(\text{CF}_3)\text{ppy}\}_2(\text{dtbpy})]\text{PF}_6$ (5.0 and 1.0 mol %, respectively). DMSO-d_6 (2.0 mL) was added to the glass tube and the reaction mixture was gently shaken for 5 mins. After this time, approximately 0.8 mL of the reaction mixture was transferred via syringe to the NMR tube, which was then capped and removed from the glovebox. A $t = 0\text{h}$ ^1H NMR spectrum was recorded and the tube was placed in a blue LED light bath at rt for 14h (photo 1). A $t = 14\text{h}$ ^1H NMR spectrum was then recorded.

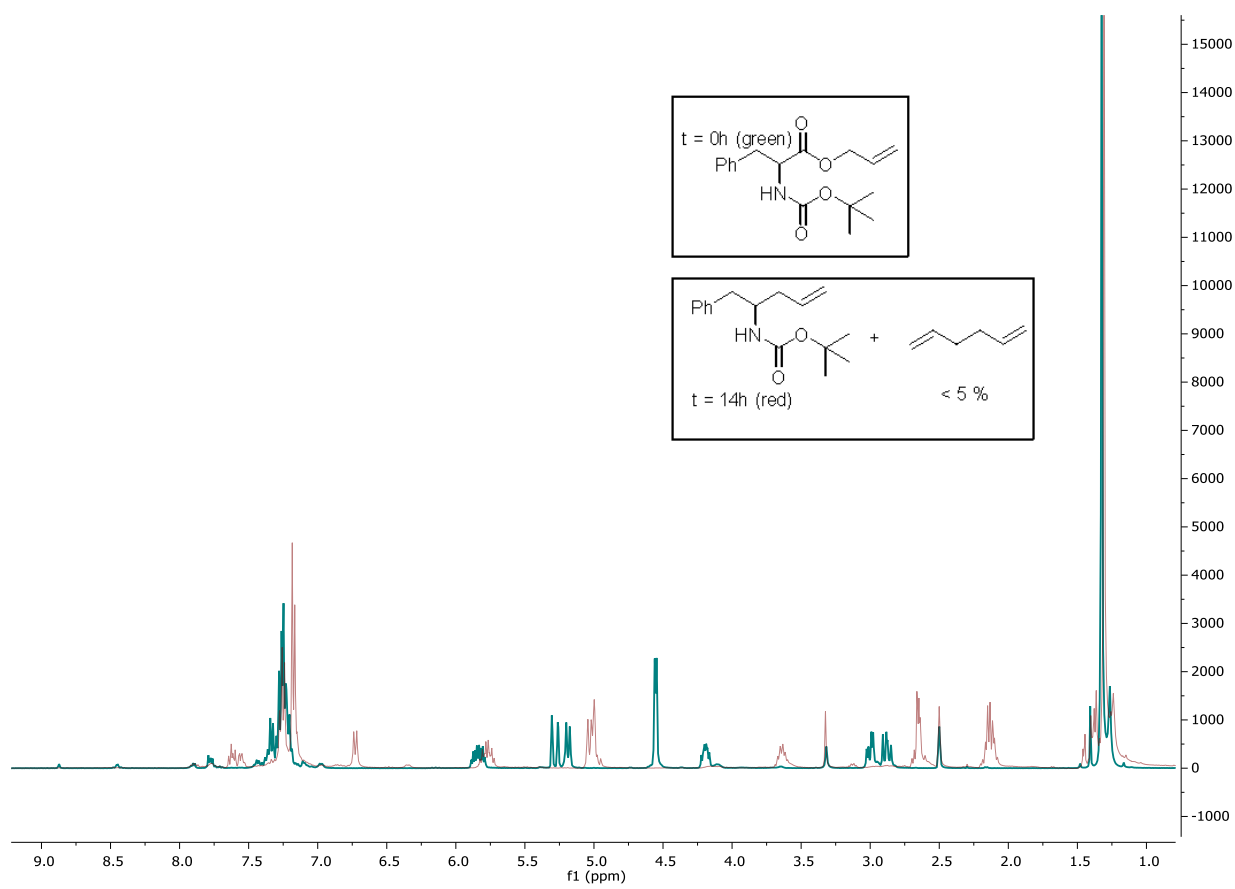


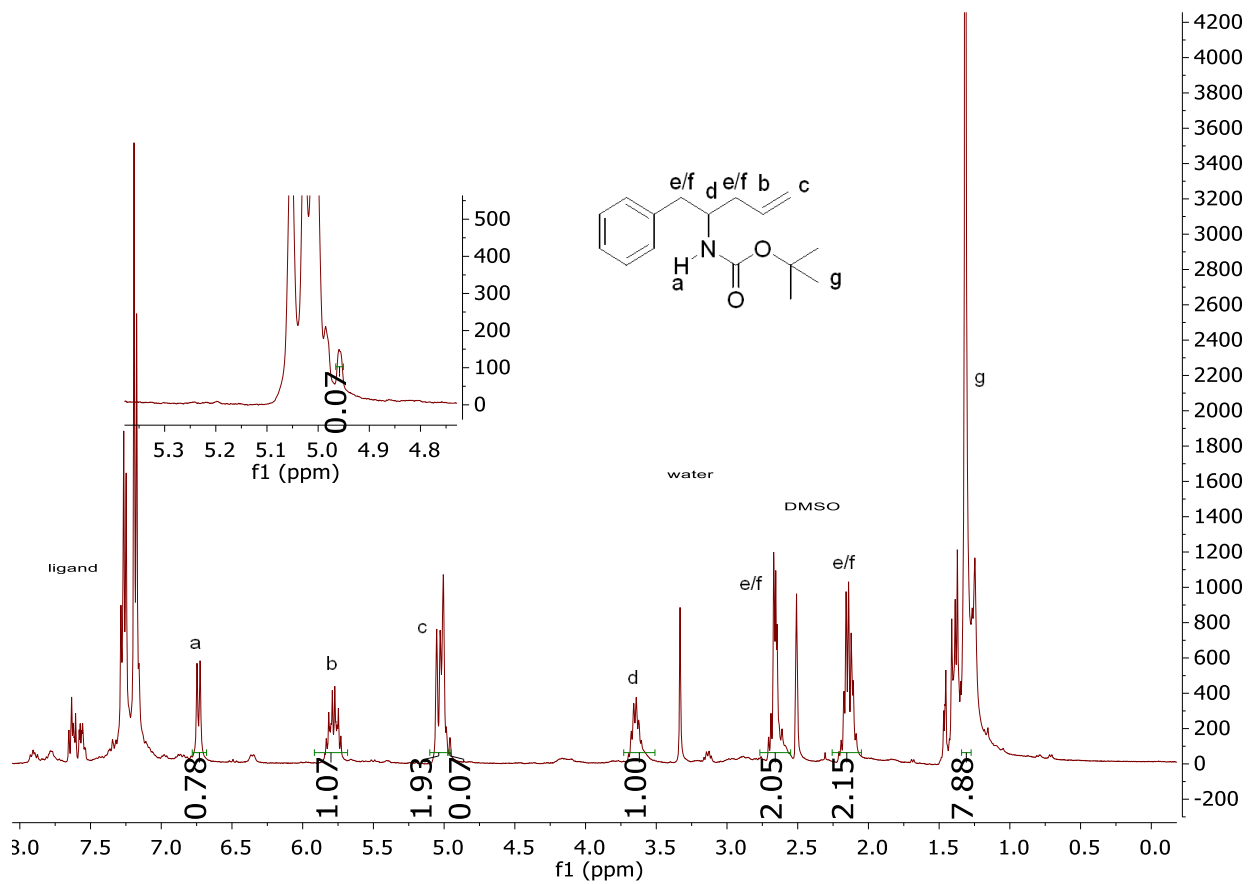


A:B:C = 1.0 : 0.43 : 0.50

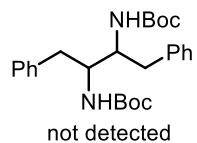




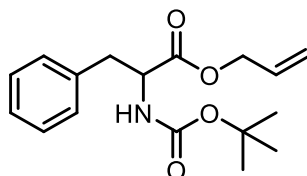




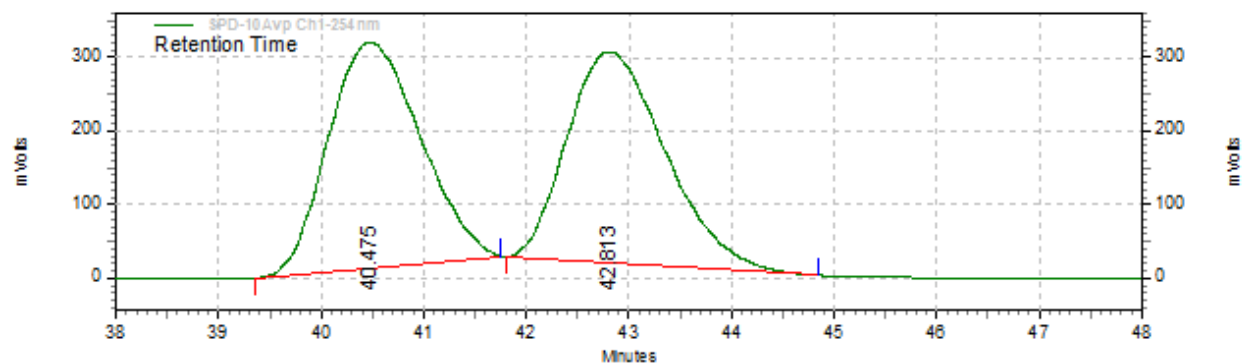
No significant amount of dibenzyl dicarbamate dimer was detected. ~ 7% 1,5 hexadiene was detected.



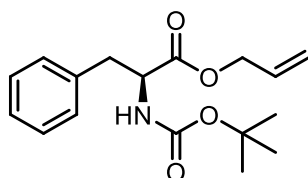
6. Chiral HPLC Analysis



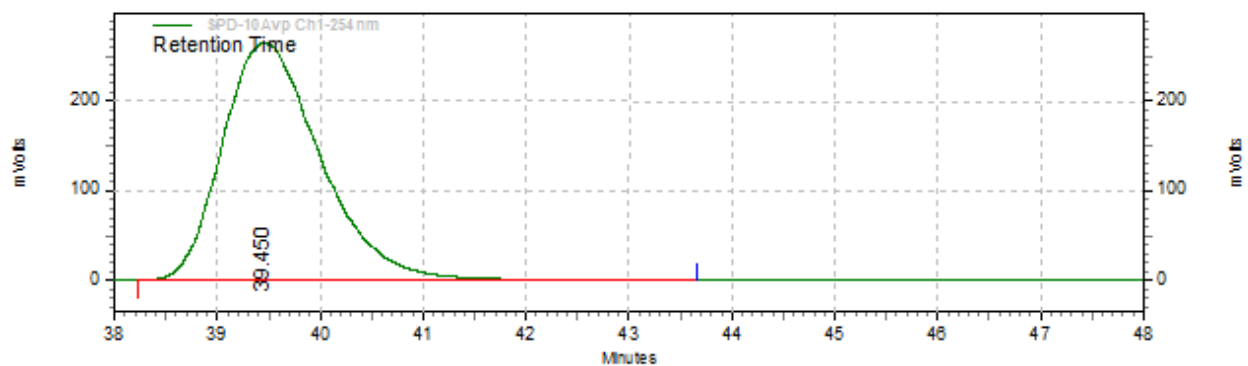
Racemic, ADH column, 30:70 *i*PrOH:hexanes, 0.38 mL/min



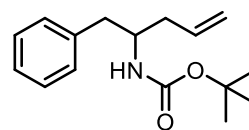
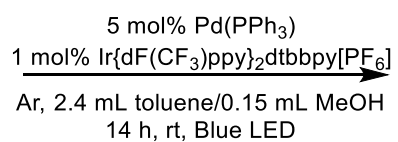
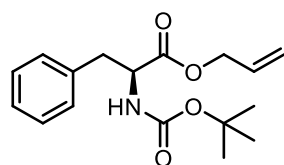
SPD-10Avp Ch1-254nm Results				
Retention Time	Area	Area %	Height	Height %
40.475	19100552	50.01	306410	51.65
42.813	19095979	49.99	286811	48.35
Totals	38196531	100.00	593221	100.00



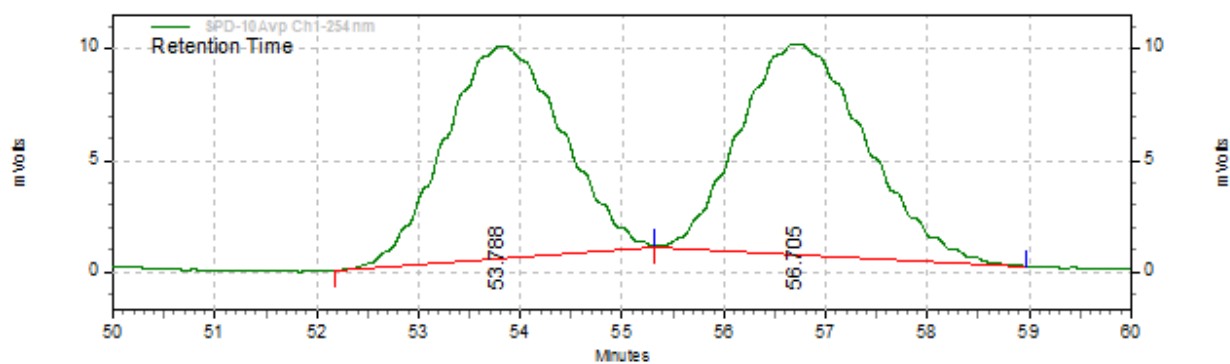
Enantioenriched, ADH column, 30:70 *i*PrOH:hexanes, 0.38 mL/min



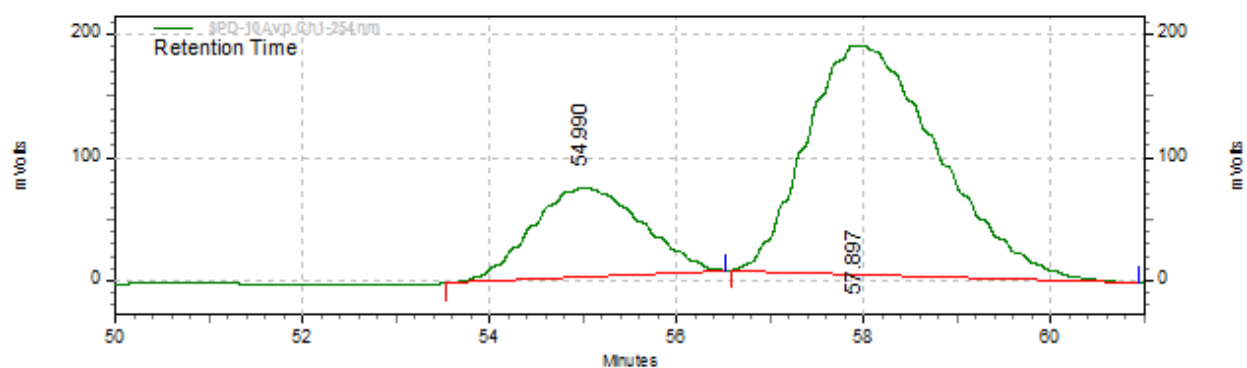
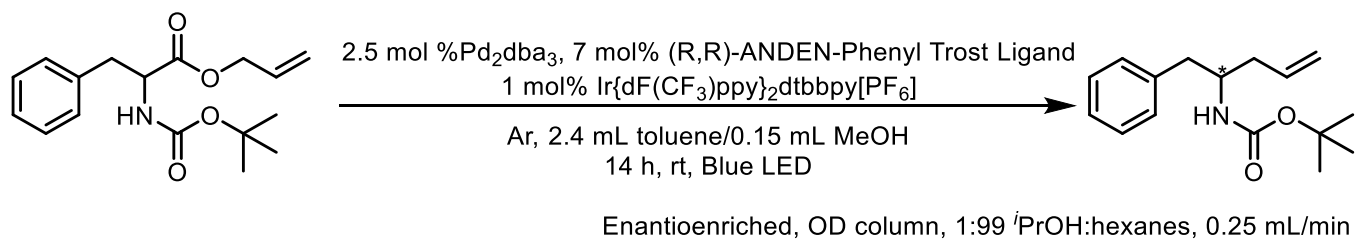
SPD-10Avp Ch1-254nm Results				
Retention Time	Area	Area %	Height	Height %
39.450	17593284	100.00	265087	100.00
Totals	17593284	100.00	265087	100.00



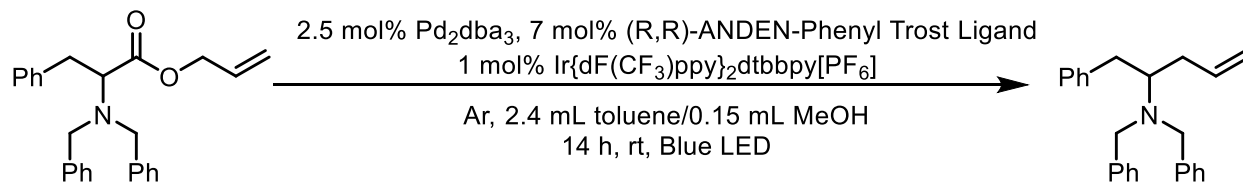
Racemic, OD column, 1:99 ⁱPrOH:hexanes, 0.25 mL/min



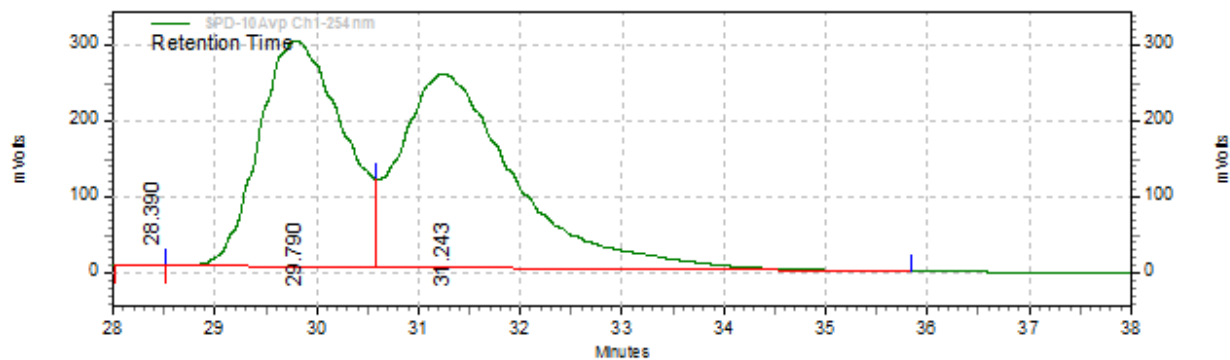
SPD-10Avp Ch1-254nm Results				
Retention Time	Area	Area %	Height	Height %
53.788	760590	48.35	9481	50.18
56.705	812548	51.65	9414	49.82
Totals	1573138	100.00	18895	100.00



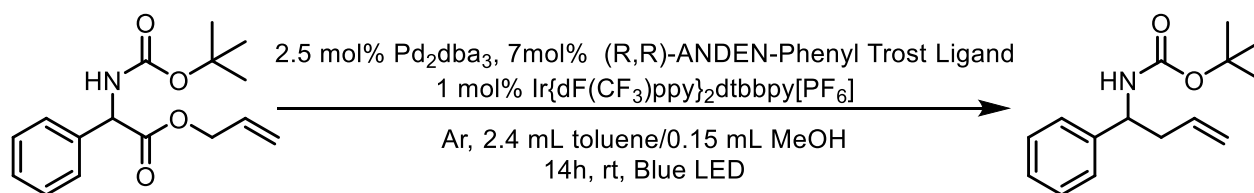
SPD-10Avp Chl-254nm Results				
Retention Time	Area	Area %	Height	Height %
54.990	5916249	24.54	72540	28.10
57.897	18195938	75.46	185612	71.90
Totals	24112187	100.00	258152	100.00



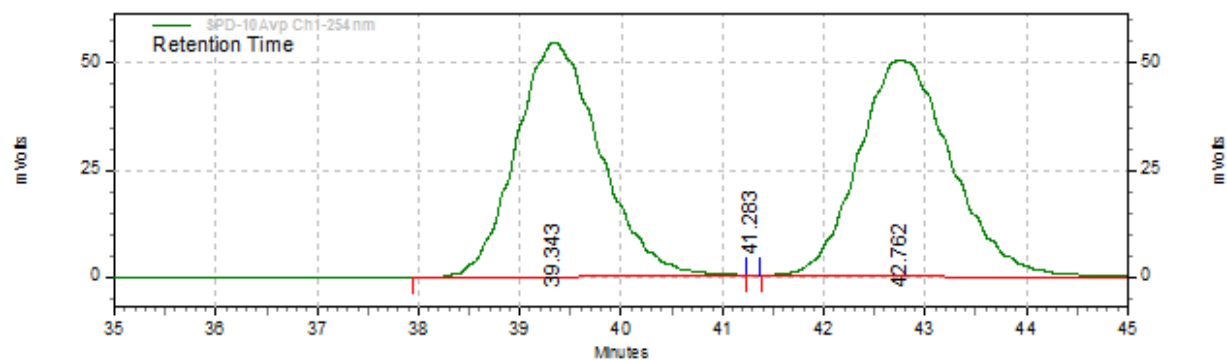
ADH column, 0.1:99.9 *i*PrOH:hexanes, 0.30 mL/min



SPD-10Avp Ch1-254nm Results				
Retention Time	Area	Area %	Height	Height %
28.390	13335	0.04	656	0.12
29.790	17275395	46.03	298344	53.78
31.243	20238562	53.93	255717	46.10
Totals	37527292	100.00	554717	100.00



ADH column, 0.1:99.9 iPrOH:hexanes, 0.30 mL/min

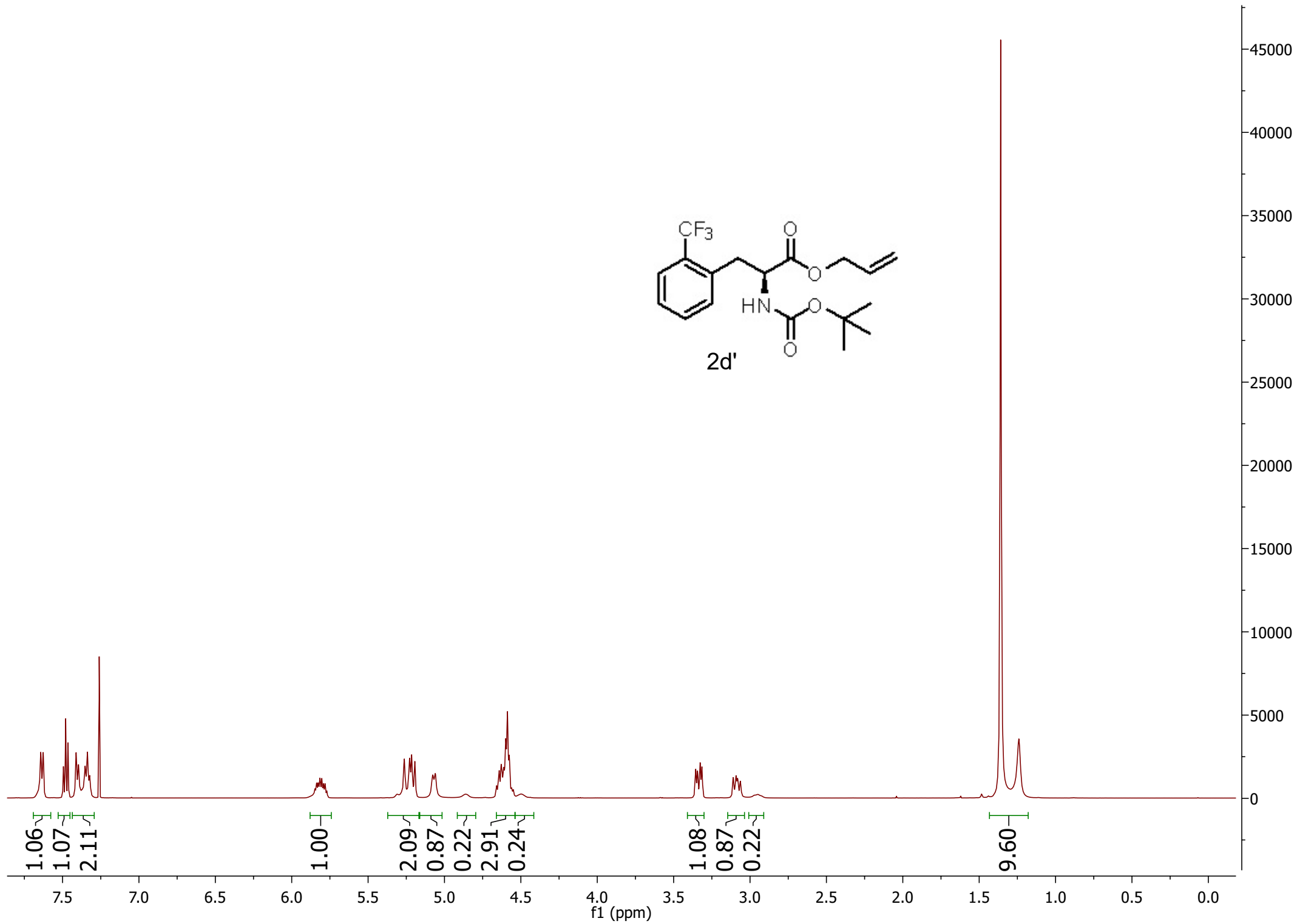
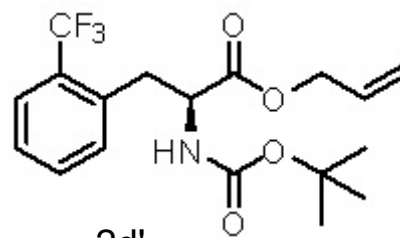


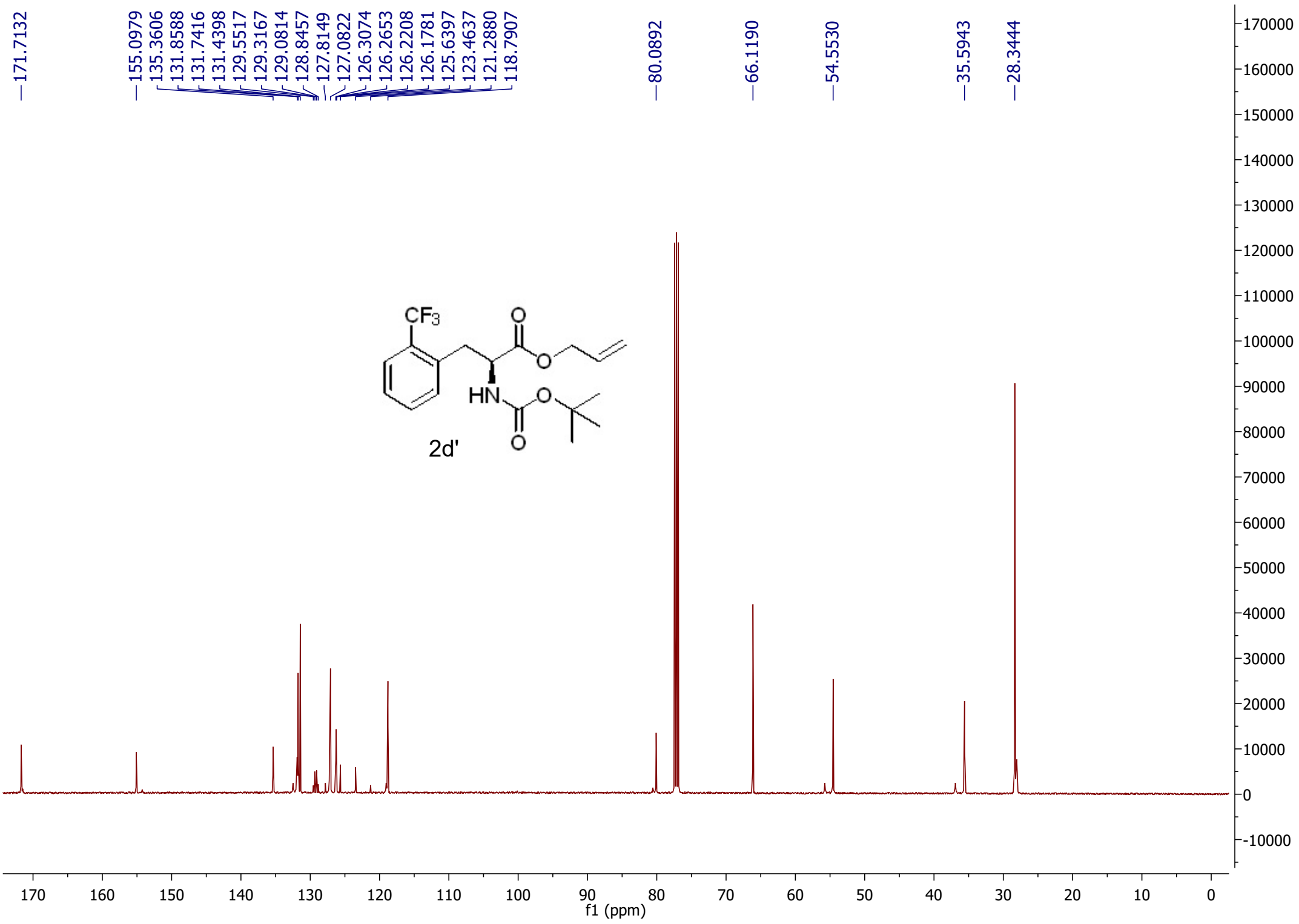
SPD-10Avp Ch1-254nm Results				
Retention Time	Area	Area %	Height	Height %
39.343	3146874	49.67	54437	52.09
41.283	380	0.01	70	0.07
42.762	3187722	50.32	49991	47.84
Totals	6334976	100.00	104498	100.00

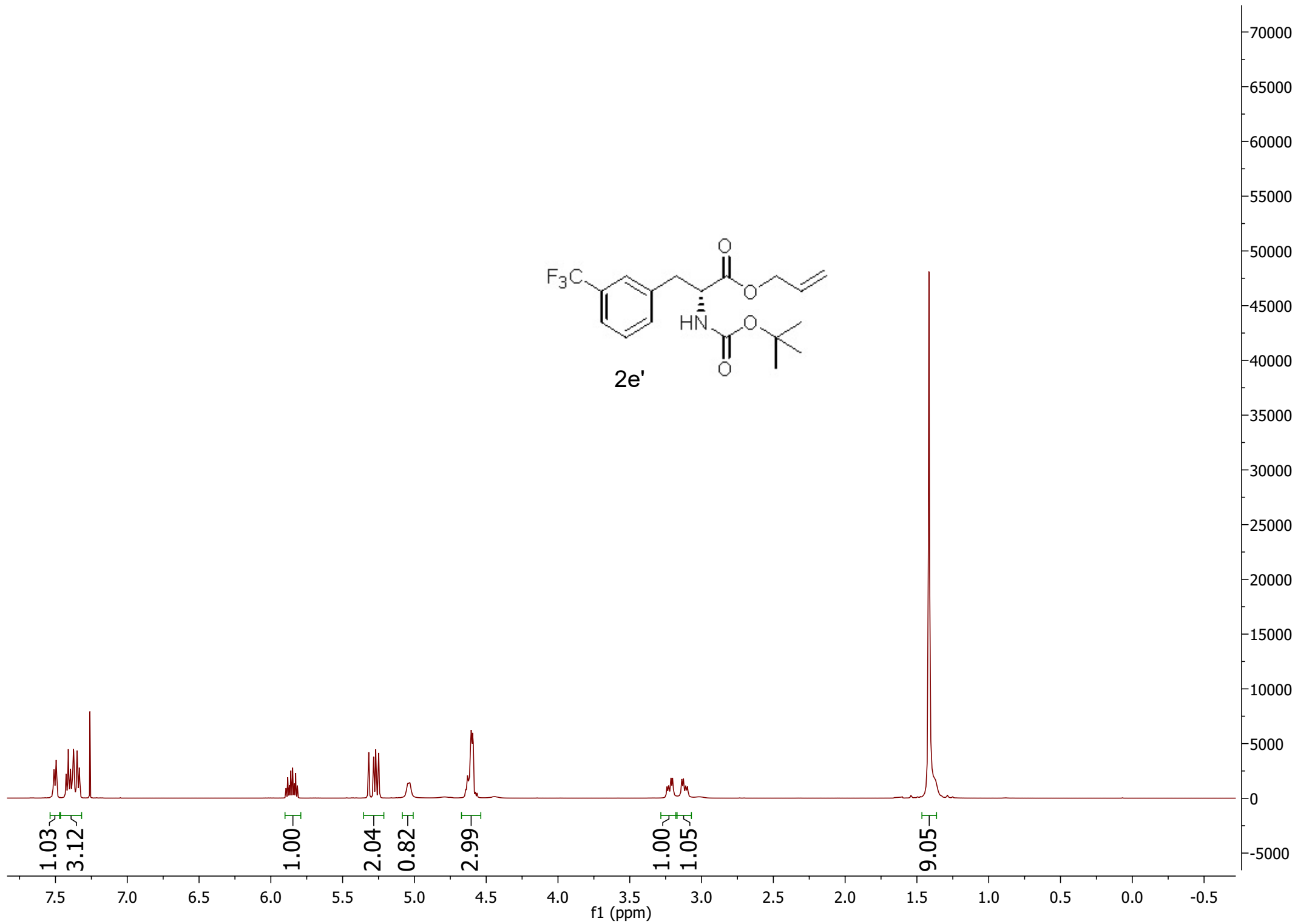
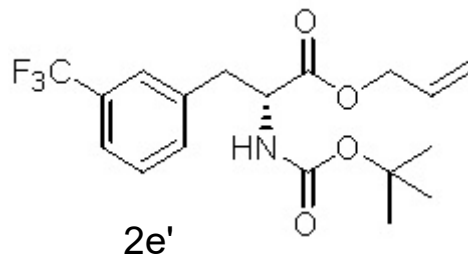
7. References

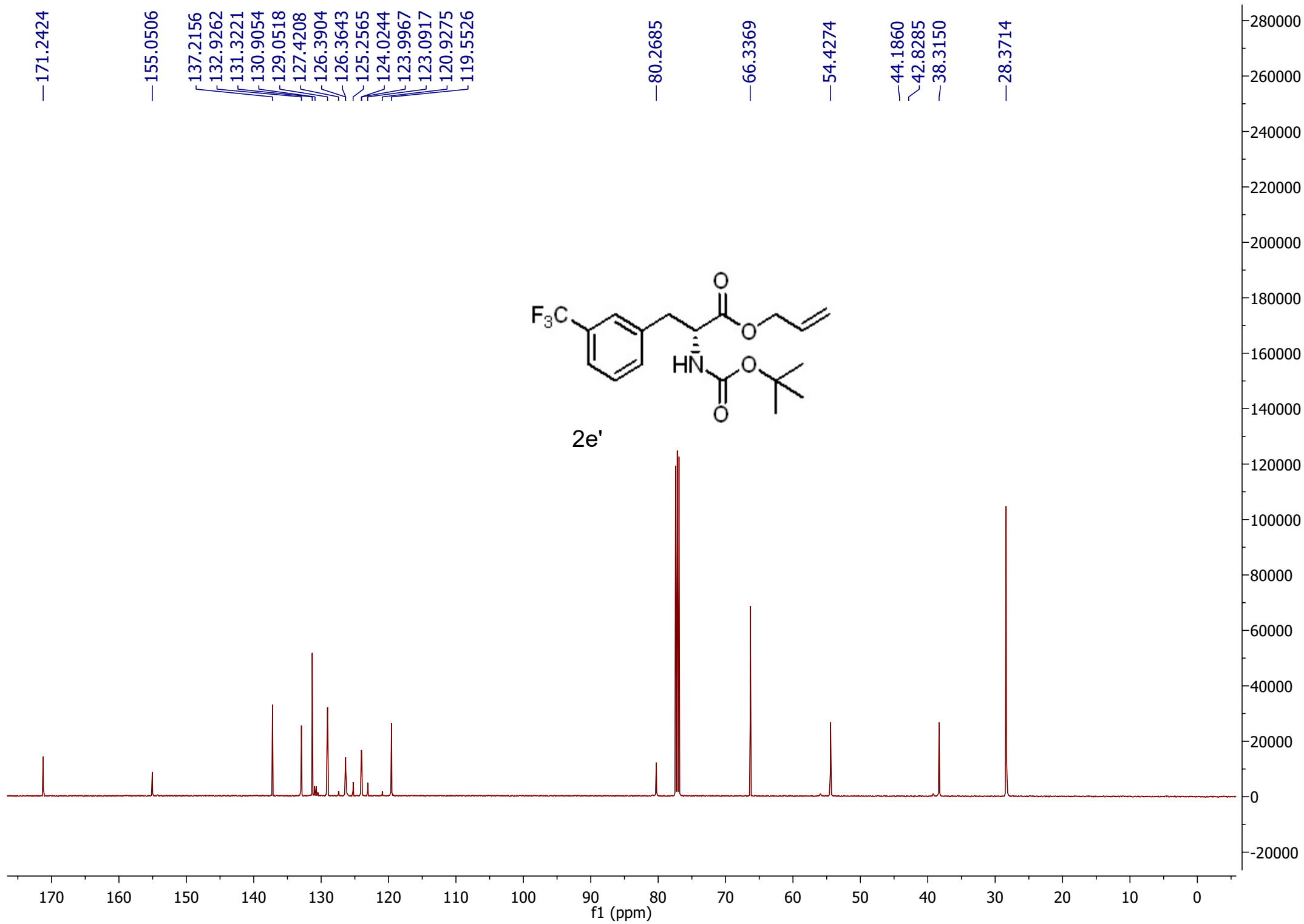
- (1) Singh, A.; Teegardin, K.; Kelly, M.; Prasad, K. S.; Krishnan, S.; Weaver, J. D. *J. Organomet. Chem.* **2015**, *776*, 51.
- (2) Lang, S. B.; O'Nele, K. M.; Tunge, J. A. *J. Am. Chem. Soc.* **2014**, *136*, 13606.
- (3) Enholm, E.; Low, T. *J. Org. Chem.* **2006**, *71*, 2272.
- (4) Konnert, L.; Lamaty, F.; Martinez, J.; Colacino, E. *J. Org. Chem.* **2014**, *79*, 4008.
- (5) da Silva, M. R.; de Mattos, M. C.; de Oliveira, M. da C. F.; de Lemos, T. L. G.; Ricardo, N. M. P. S.; de Gonzalo, G.; Lavandera, I.; Gotor-Fernández, V. *Tetrahedron.* **2014**, *70*, 2264.
- (6) Hupp, C. D.; Tepe, J. J. *J. Org. Chem.* **2009**, *74*, 3406.
- (7) Bugde, S.; Majik, M.; Mandrekar, V.; Nadkarni, V.; Tilve, S. *Synth. Commun.* **2013**, 2536.
- (8) Xie, N.; Taylor, C. M. *Org. Lett.* **2010**, *12*, 4968.
- (9) Arnaud, O.; Koubeissi, A.; Ettouati, L.; Terreux, R.; Alamé, G.; Grenot, C.; Dumontet, C.; Di Pietro, A.; Paris, J.; Flason, P. *J. Med. Chem.* **2010**, *53*, 6720.
- (10) Maegawa, Y.; Agura, K.; Hayashi, Y.; Ohshima, T.; Mashima, K. *Synlett.* **2012**, *23*, 137.
- (11) Crosignani, S.; Gonzalez, J.; Swinnen, D. *Org. Lett.* **2004**, *6*, 4579.
- (12) Crosignani, S.; White, P. D.; Linclau, B. *J. Org. Chem.* **2004**, *69*, 5897.

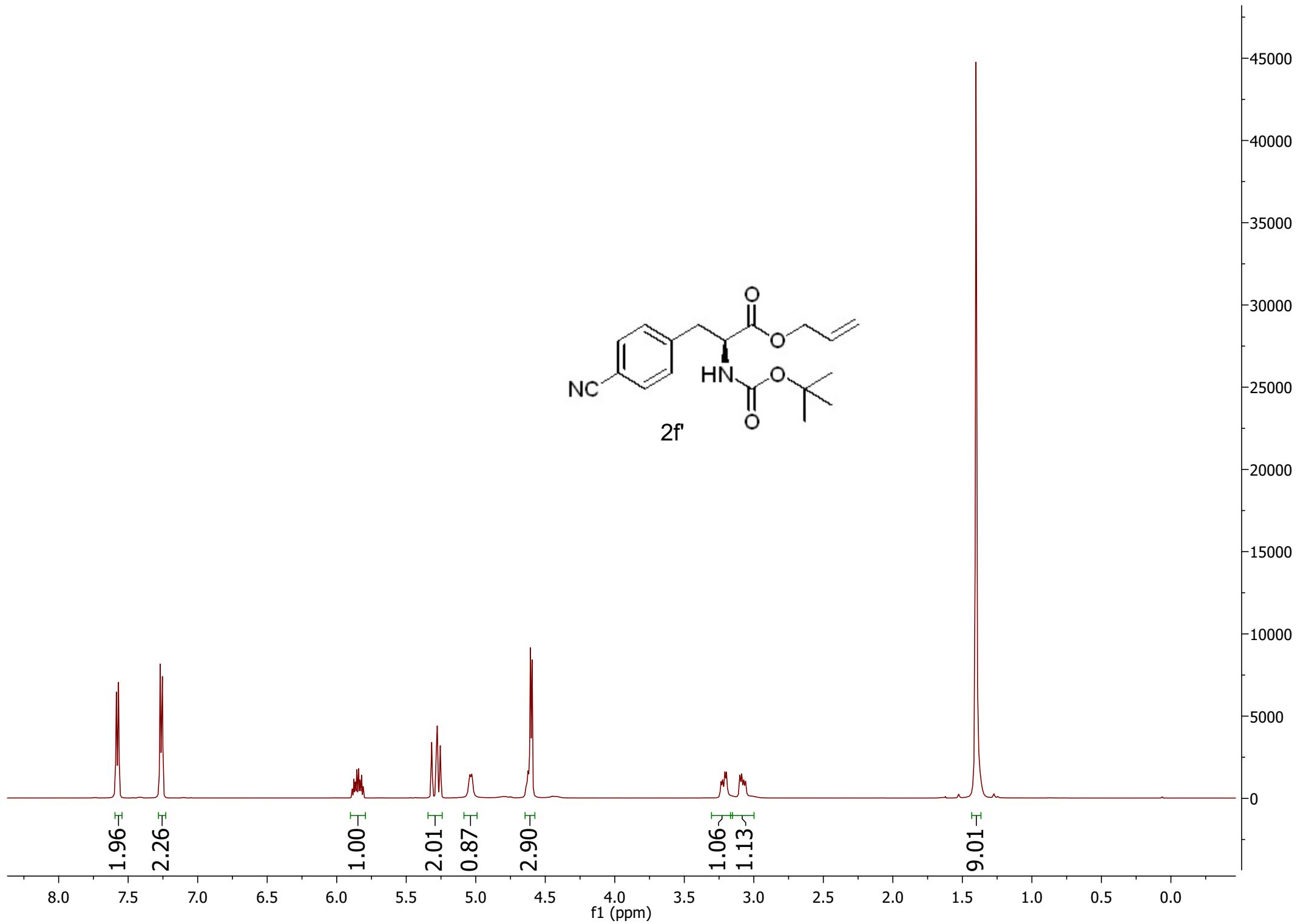
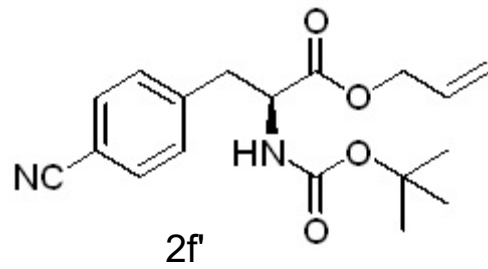
8. ^1H and ^{13}C NMR Spectra

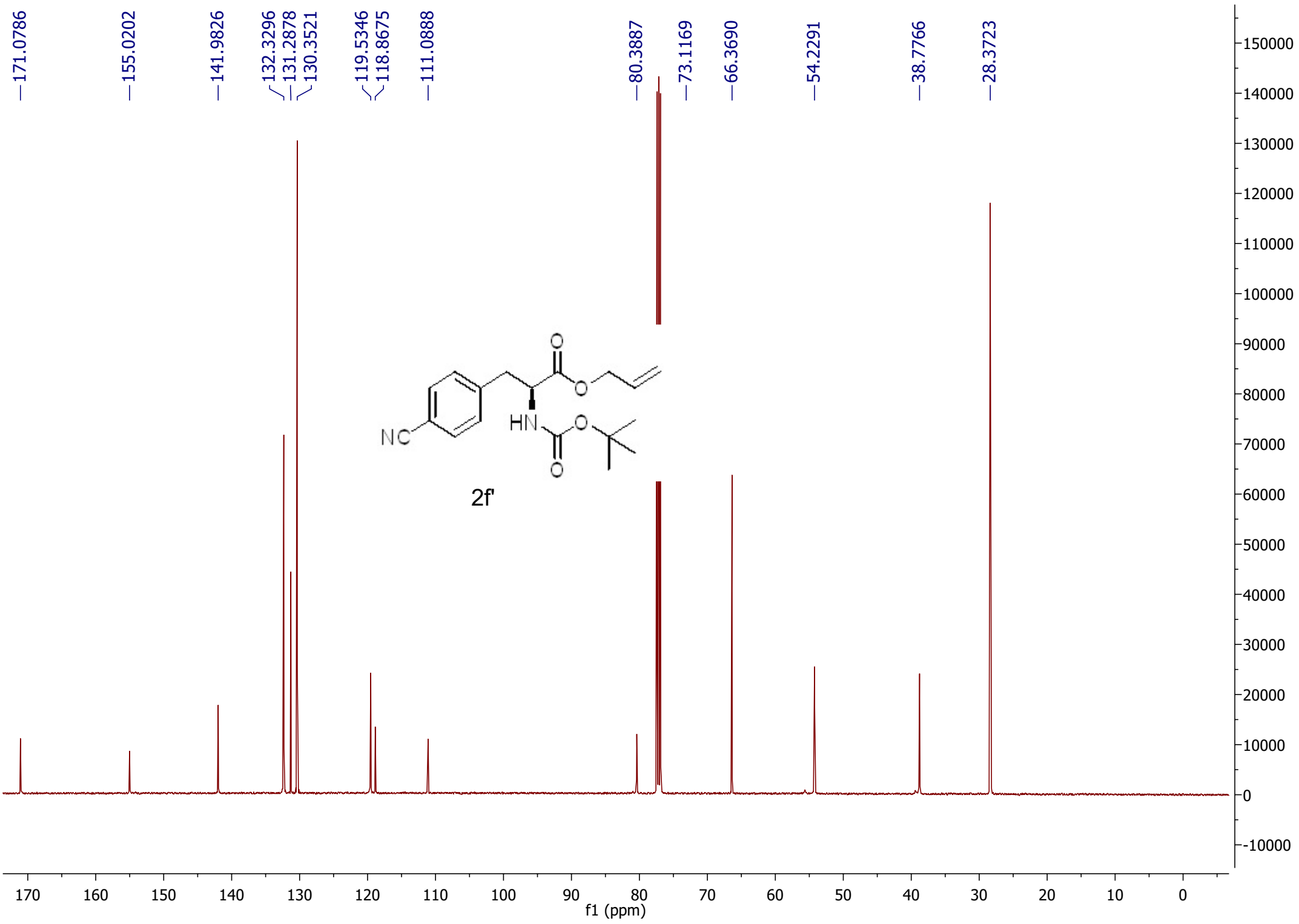


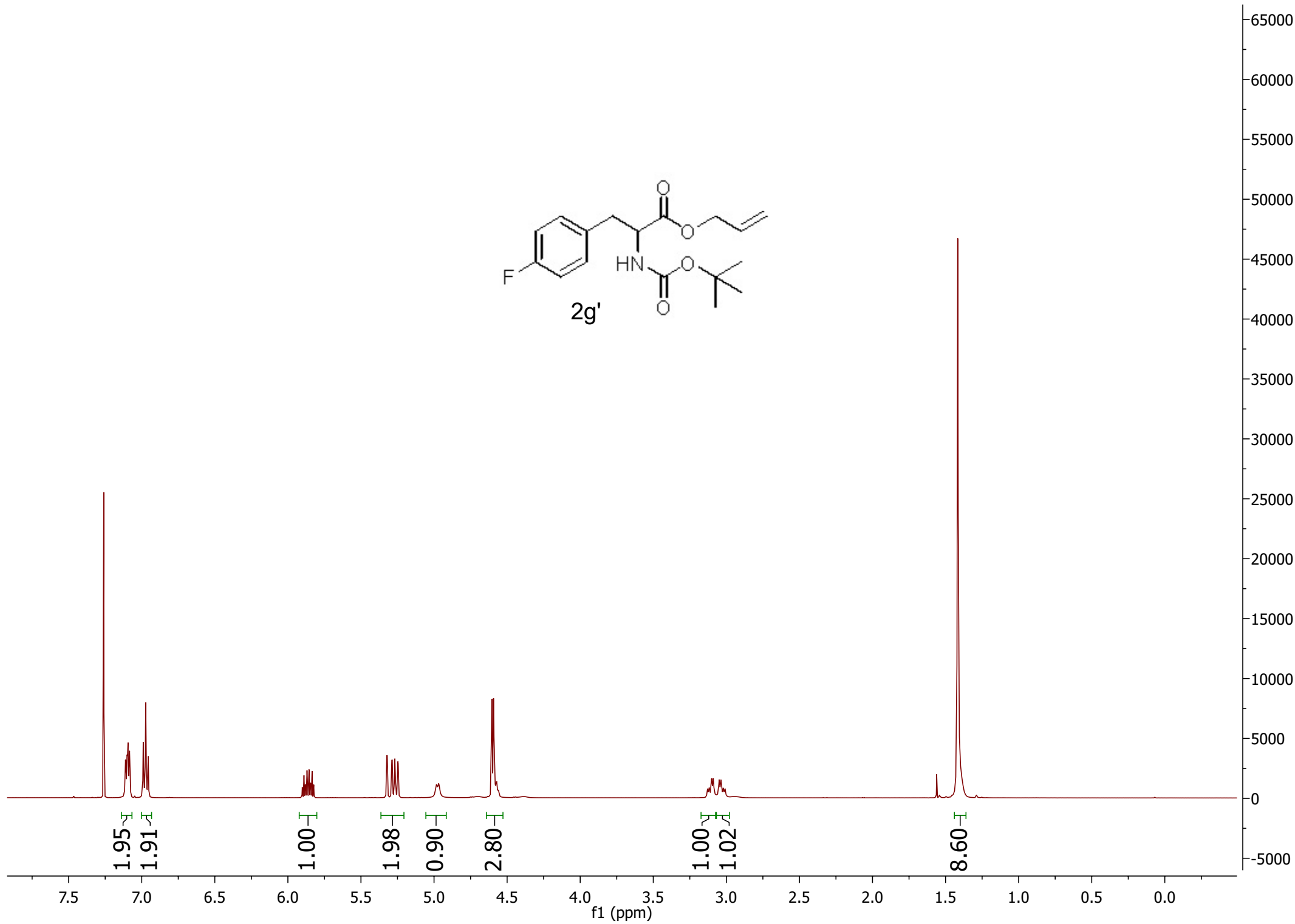
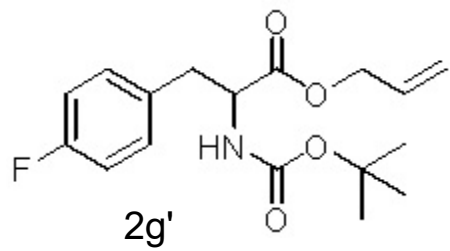












—171.5571

—163.1088

—161.1600

—155.1472

—131.8498

—131.8250

—131.5346

—131.0354

—130.9722

—119.2423

—115.5831

—115.4140

—80.1643

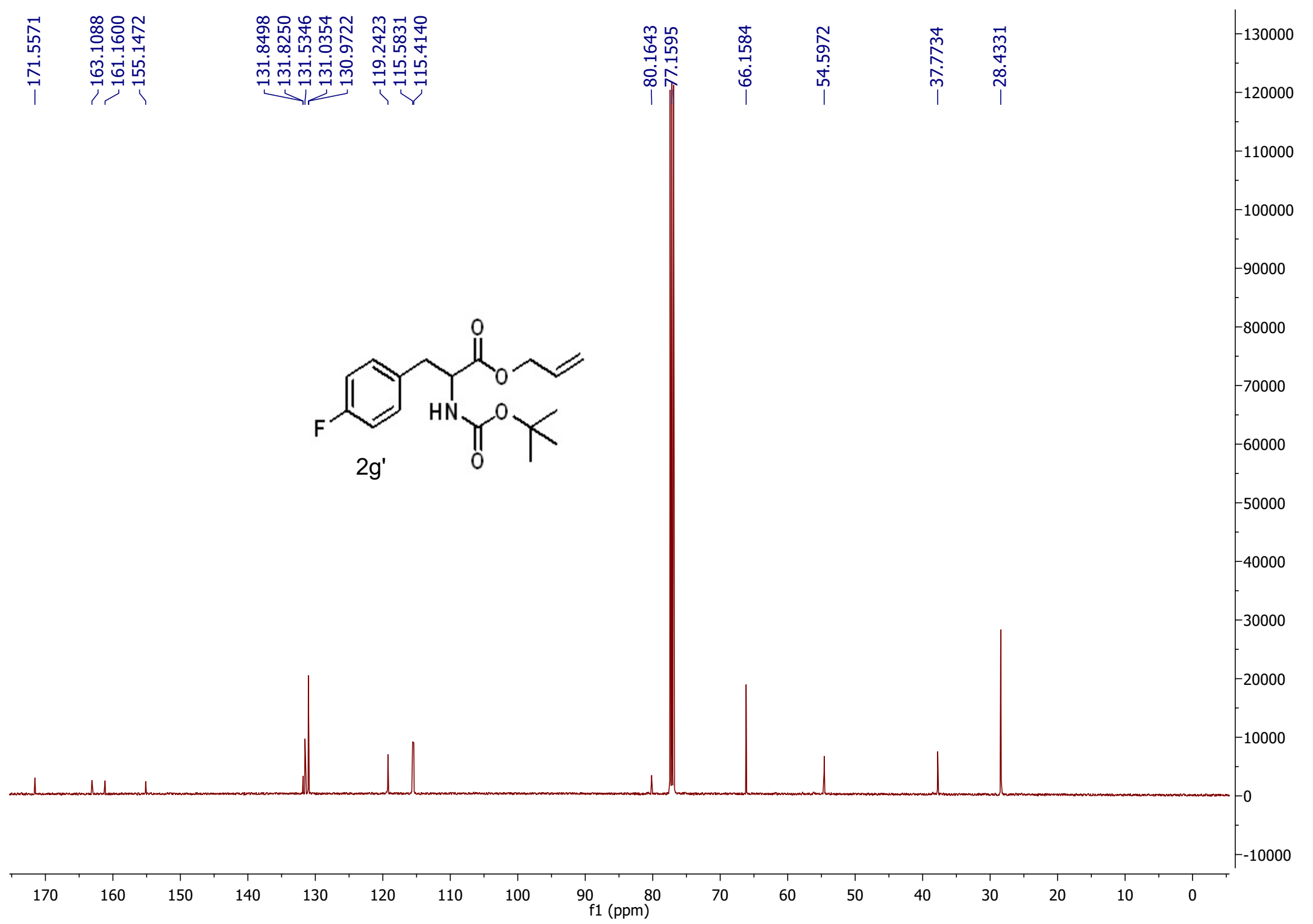
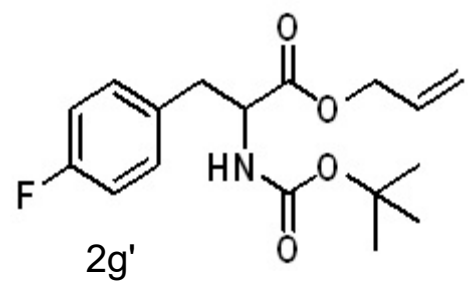
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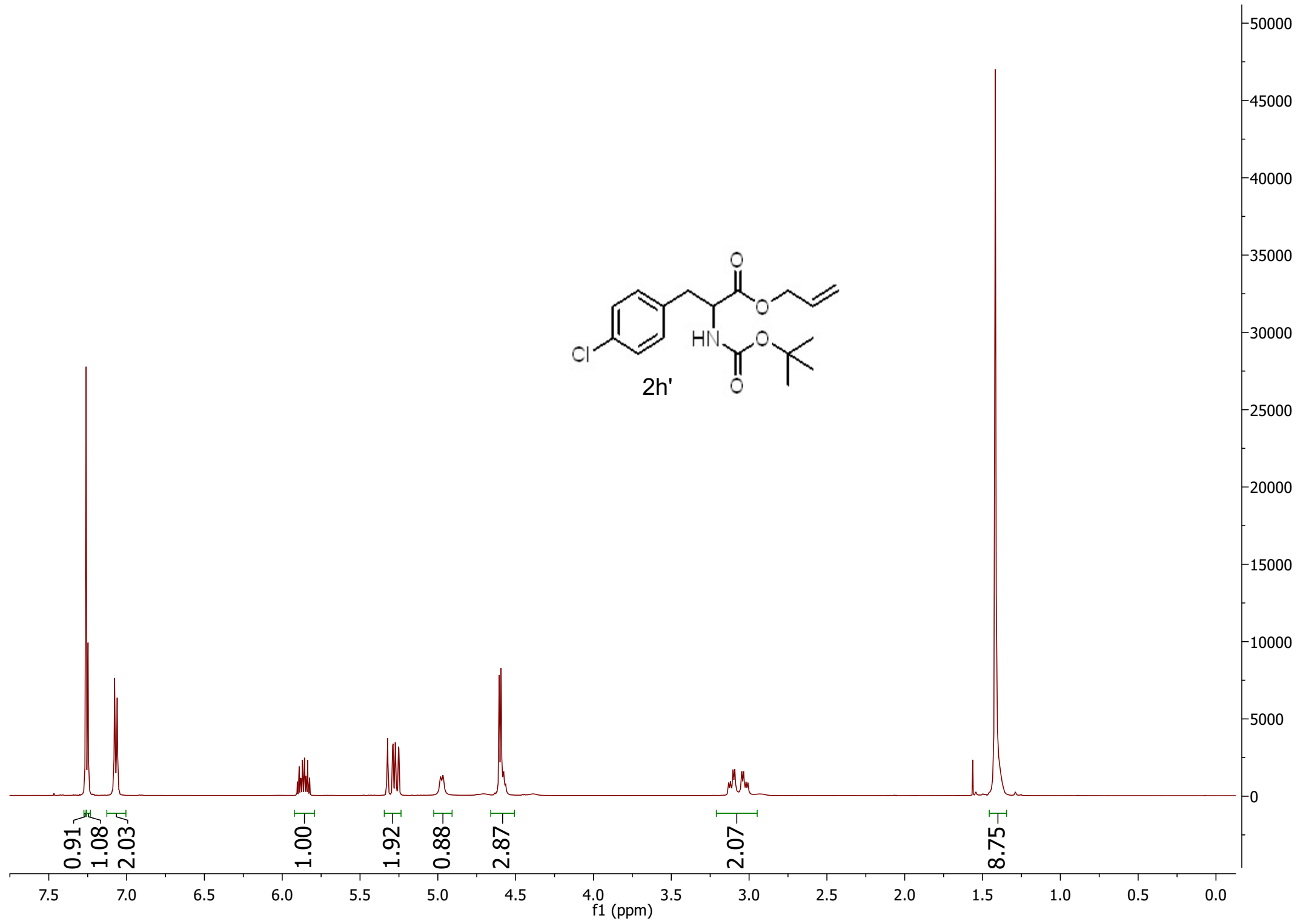
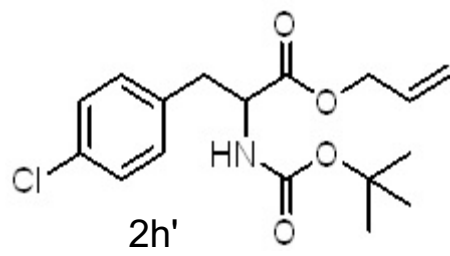
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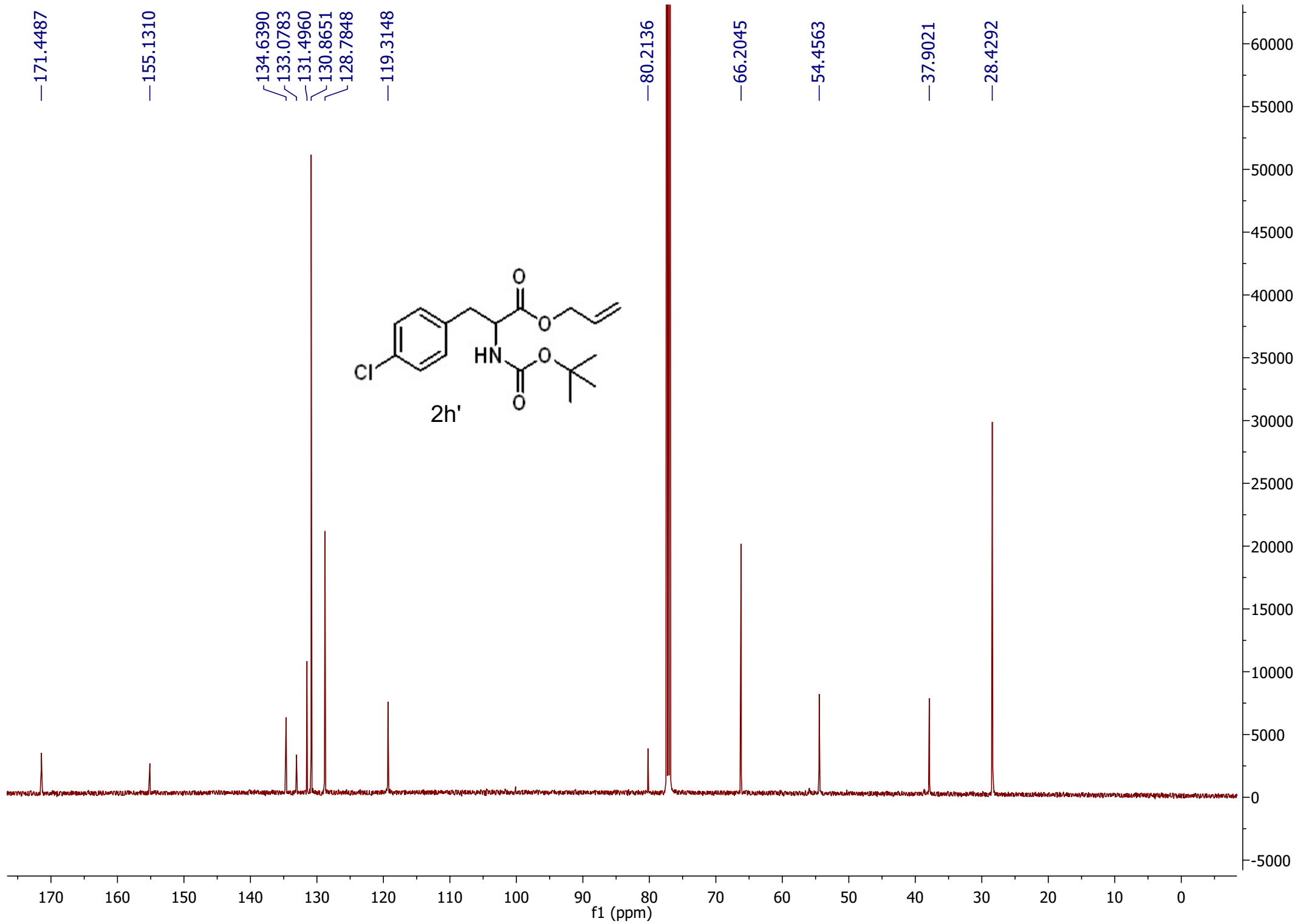
—54.5972

—37.7734

—28.4331







—171.4487

—155.1310

134.6390

133.0783

131.4960

130.8651

128.7848

—119.3148

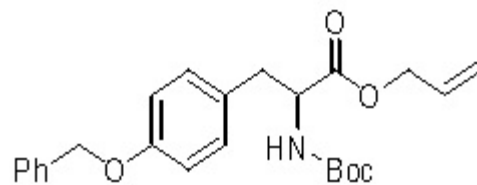
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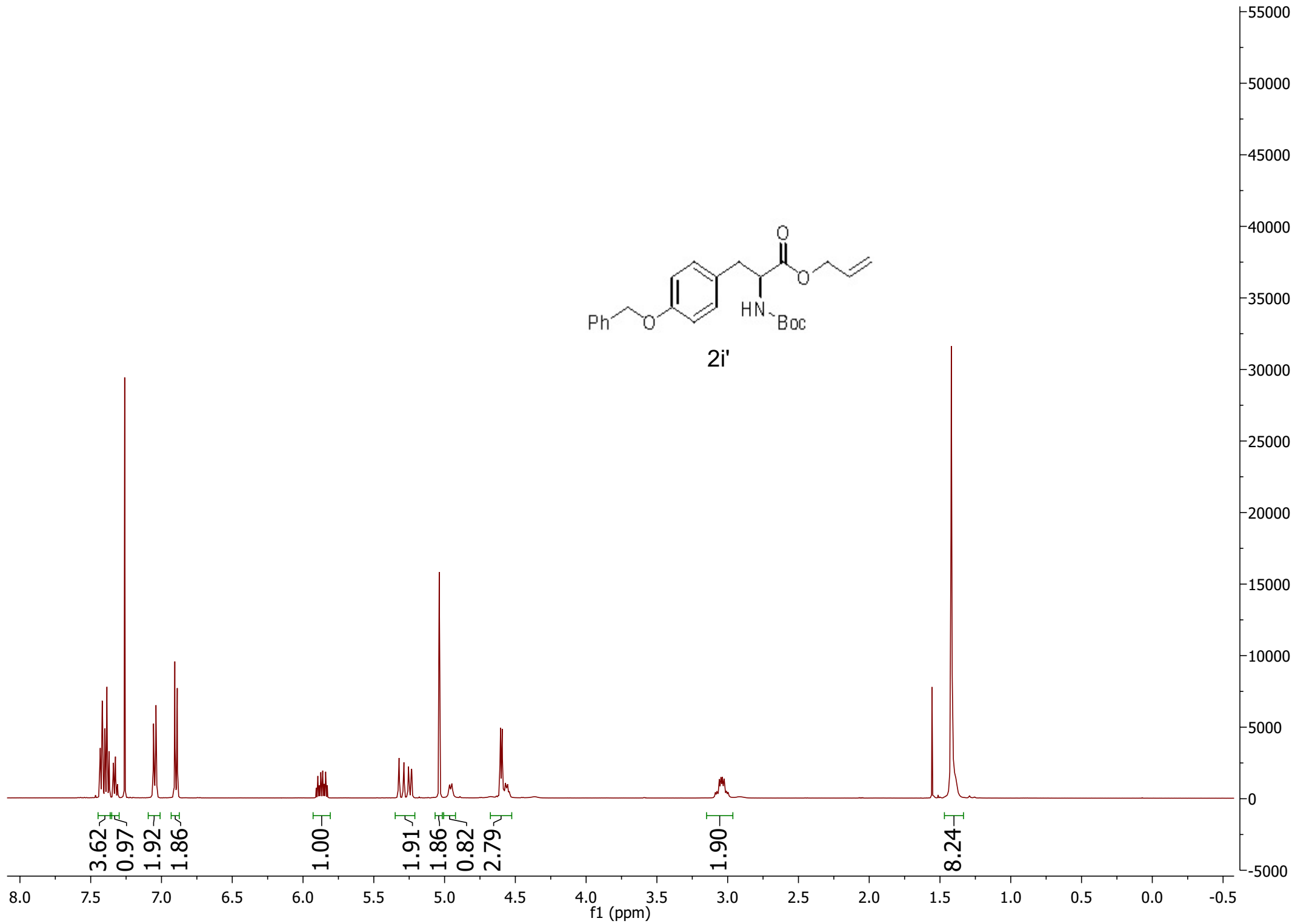
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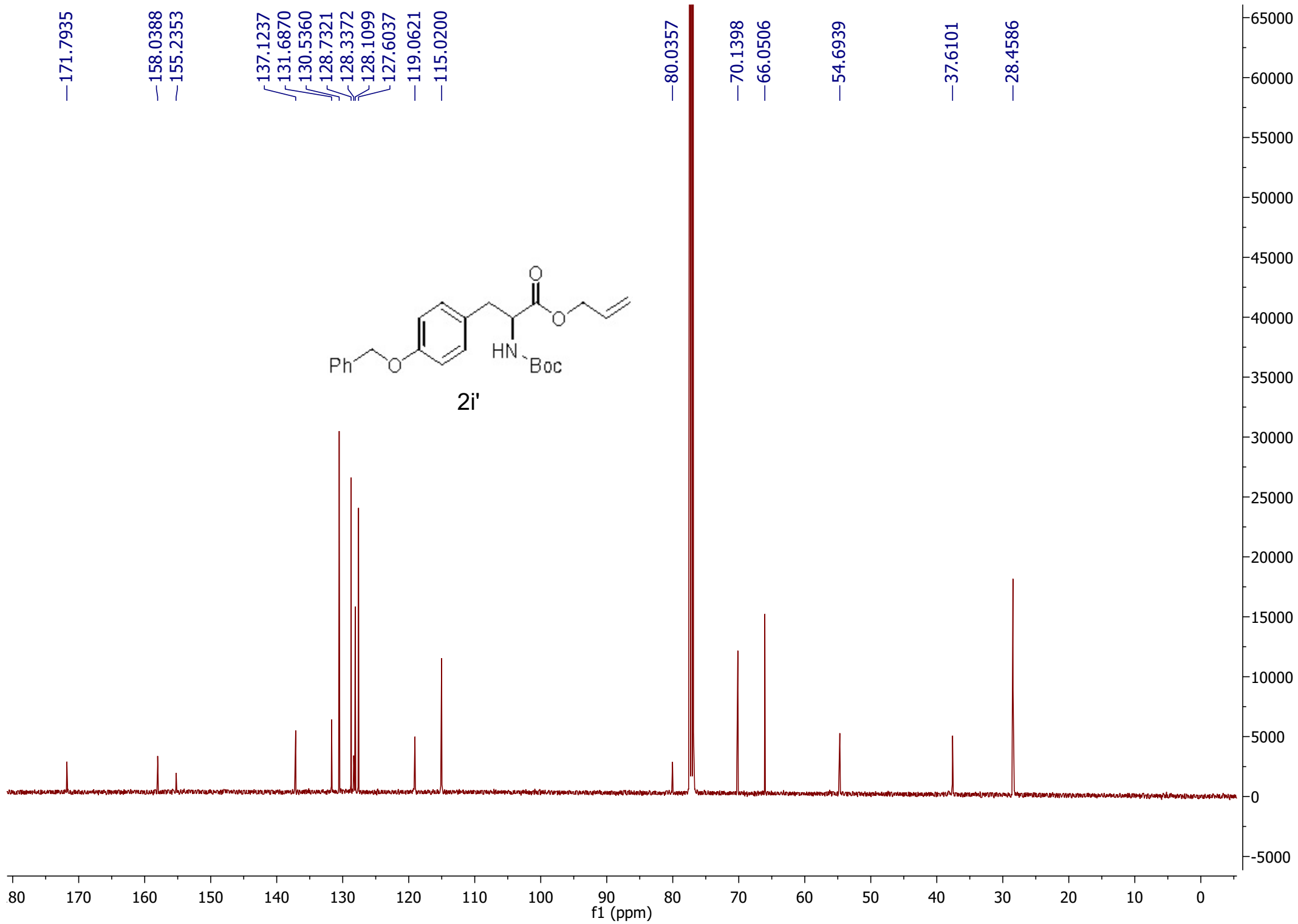
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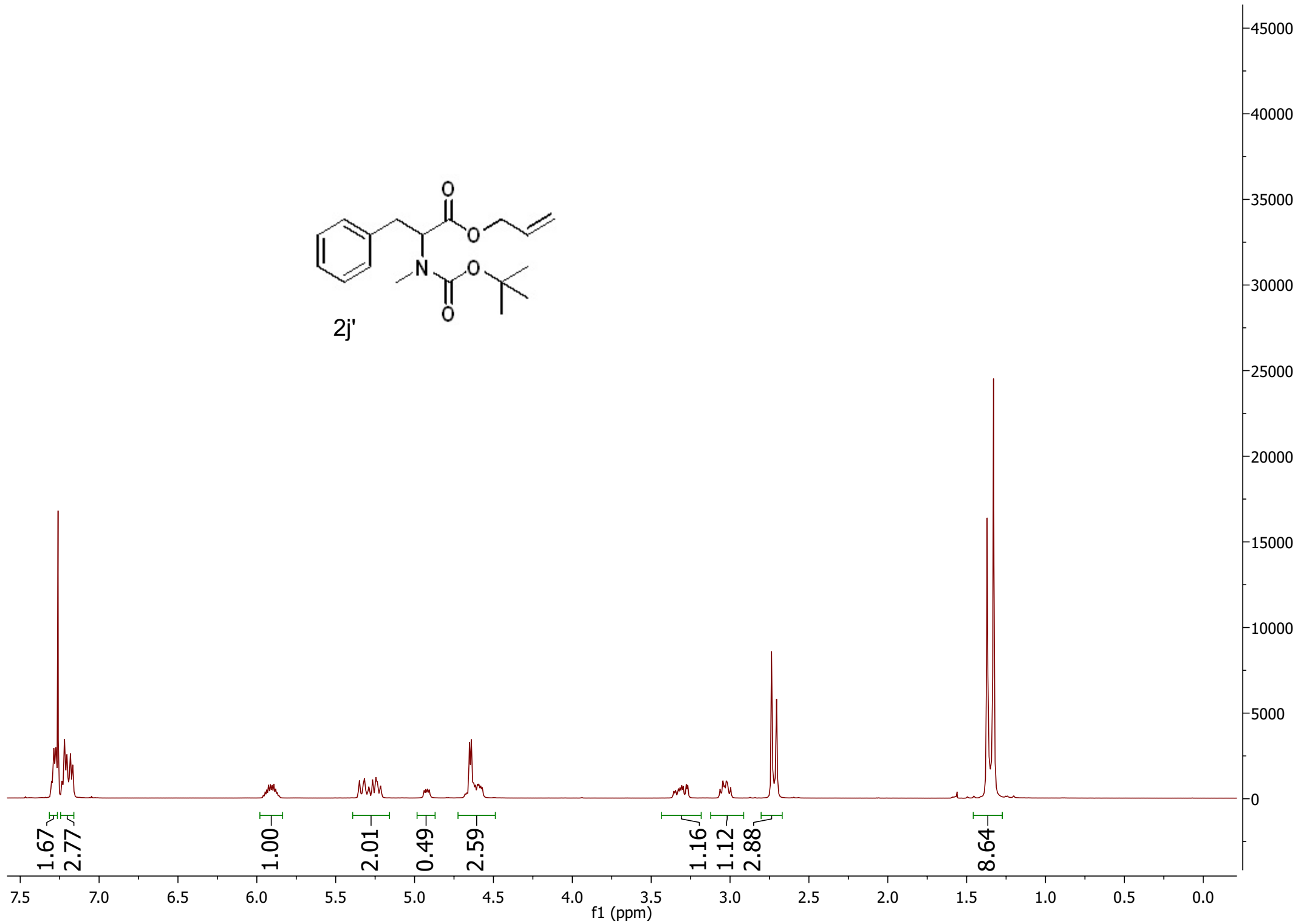
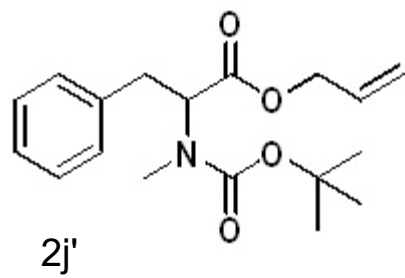
—28.4292



2i'







—171.0954

—155.5135

~137.7037

~131.9140

~129.1166

~128.5804

~126.7094

—118.5824

—80.2616

77.4137

—65.8686

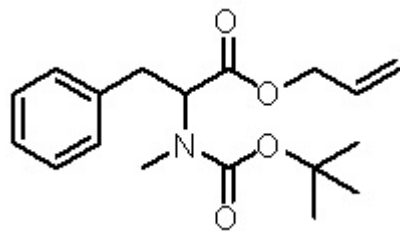
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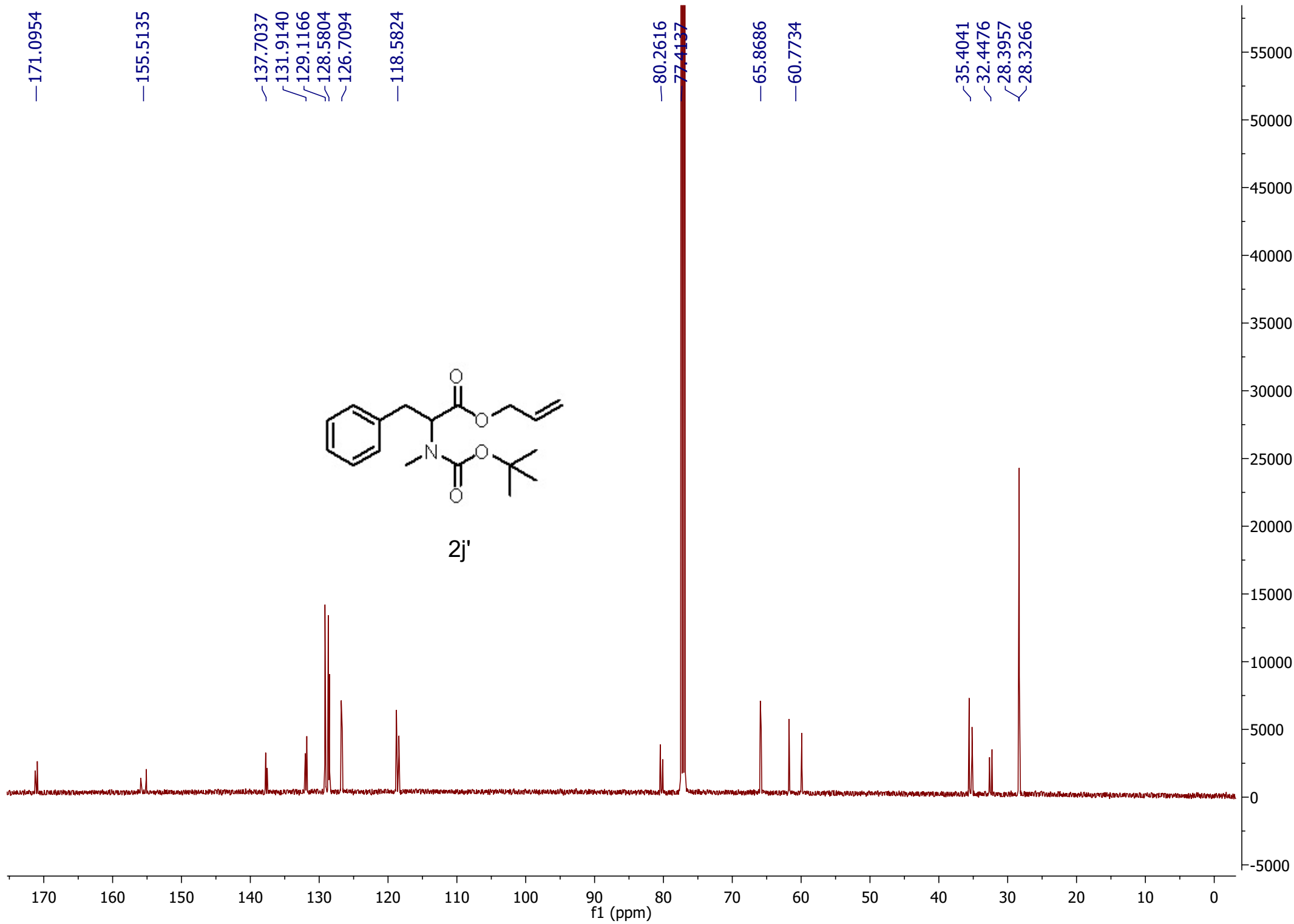
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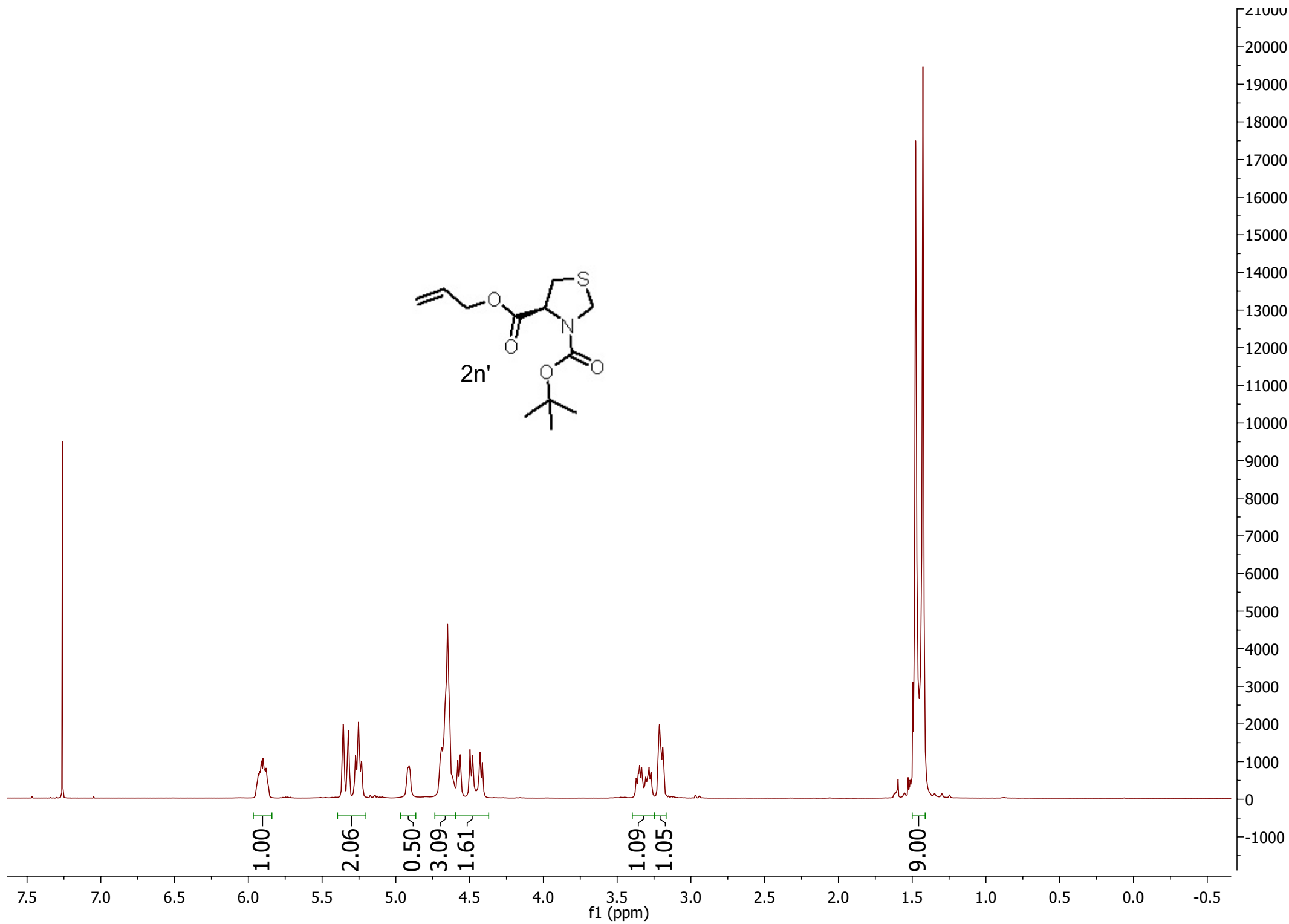
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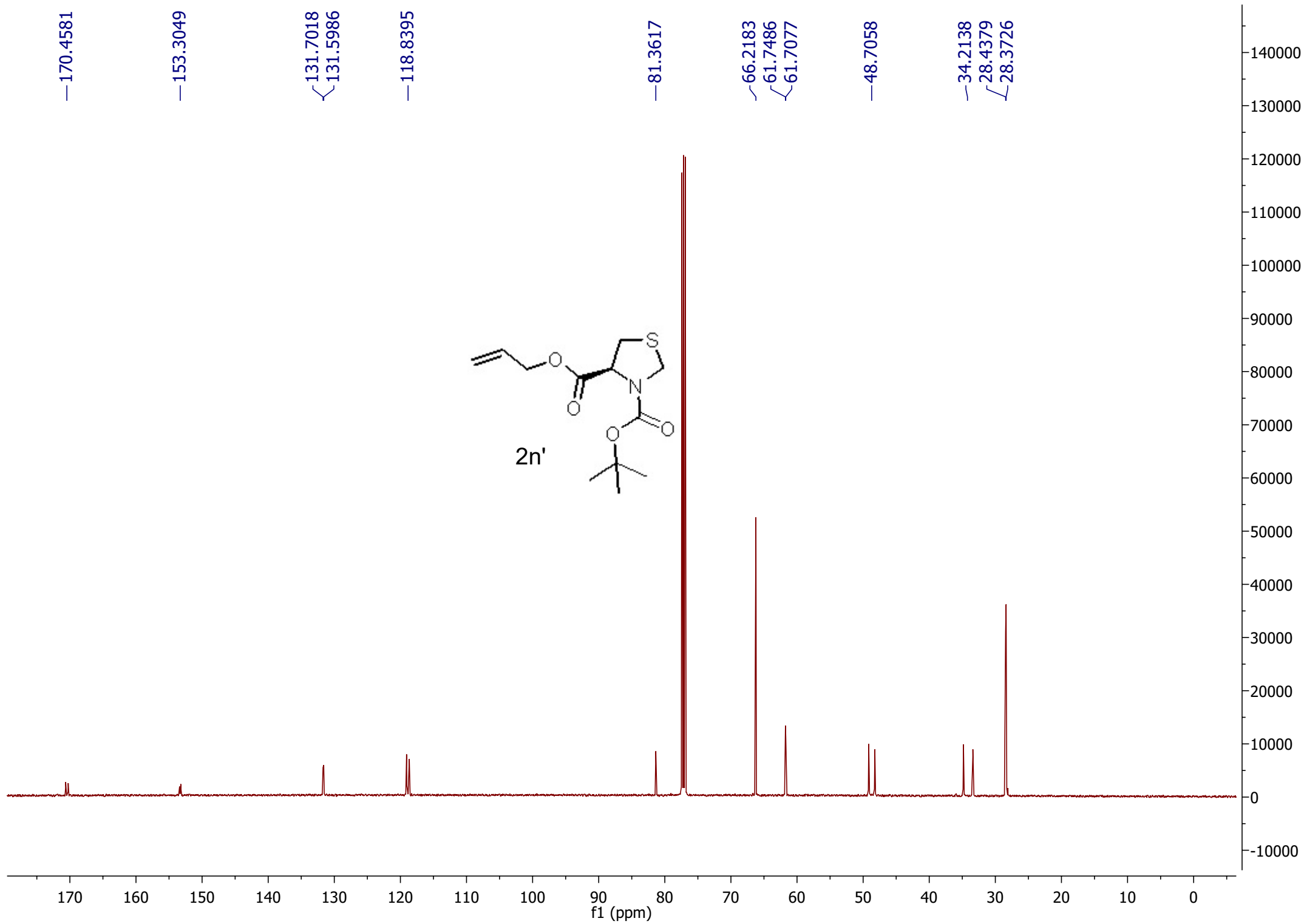
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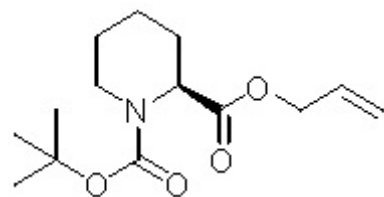


2j'

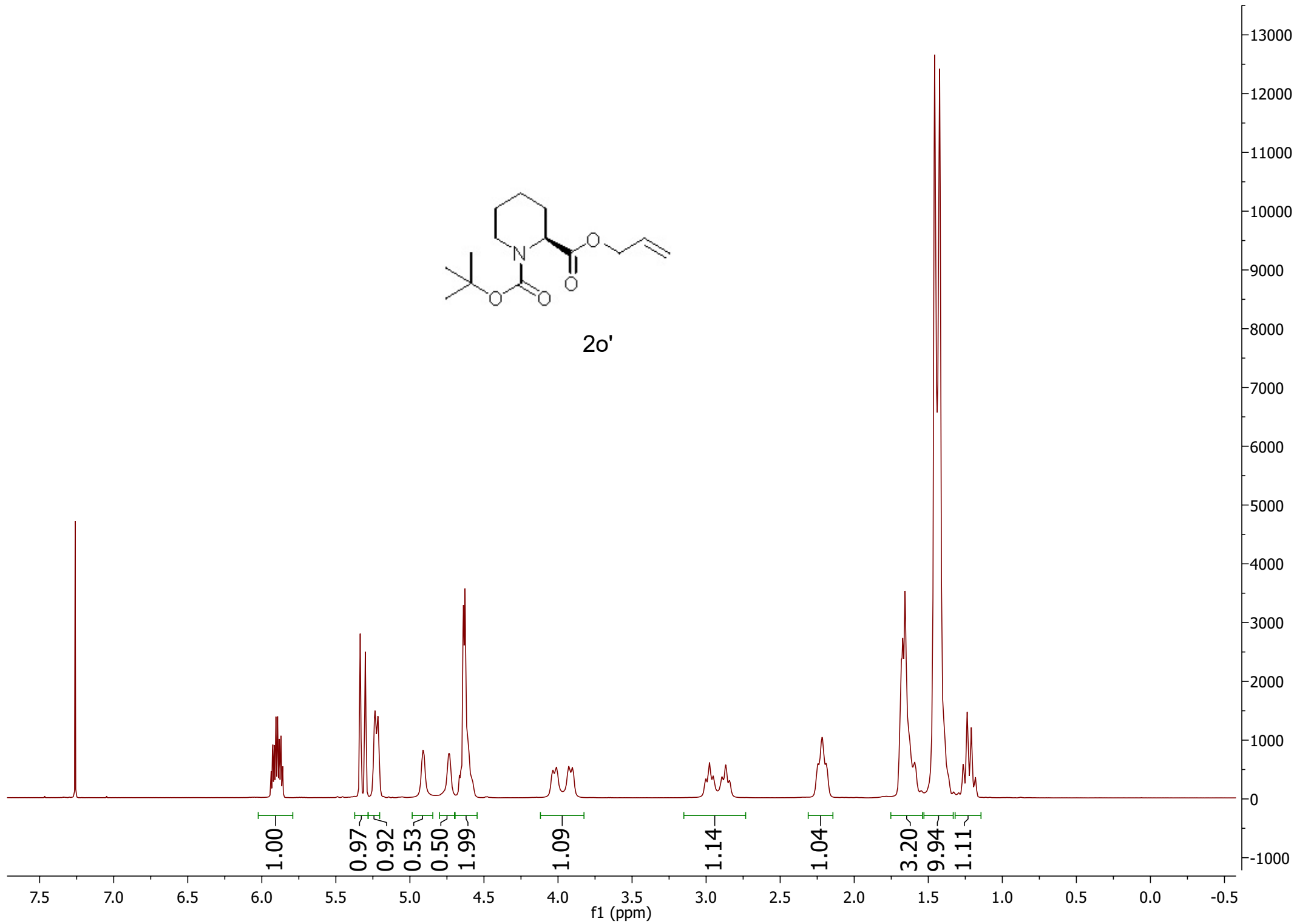


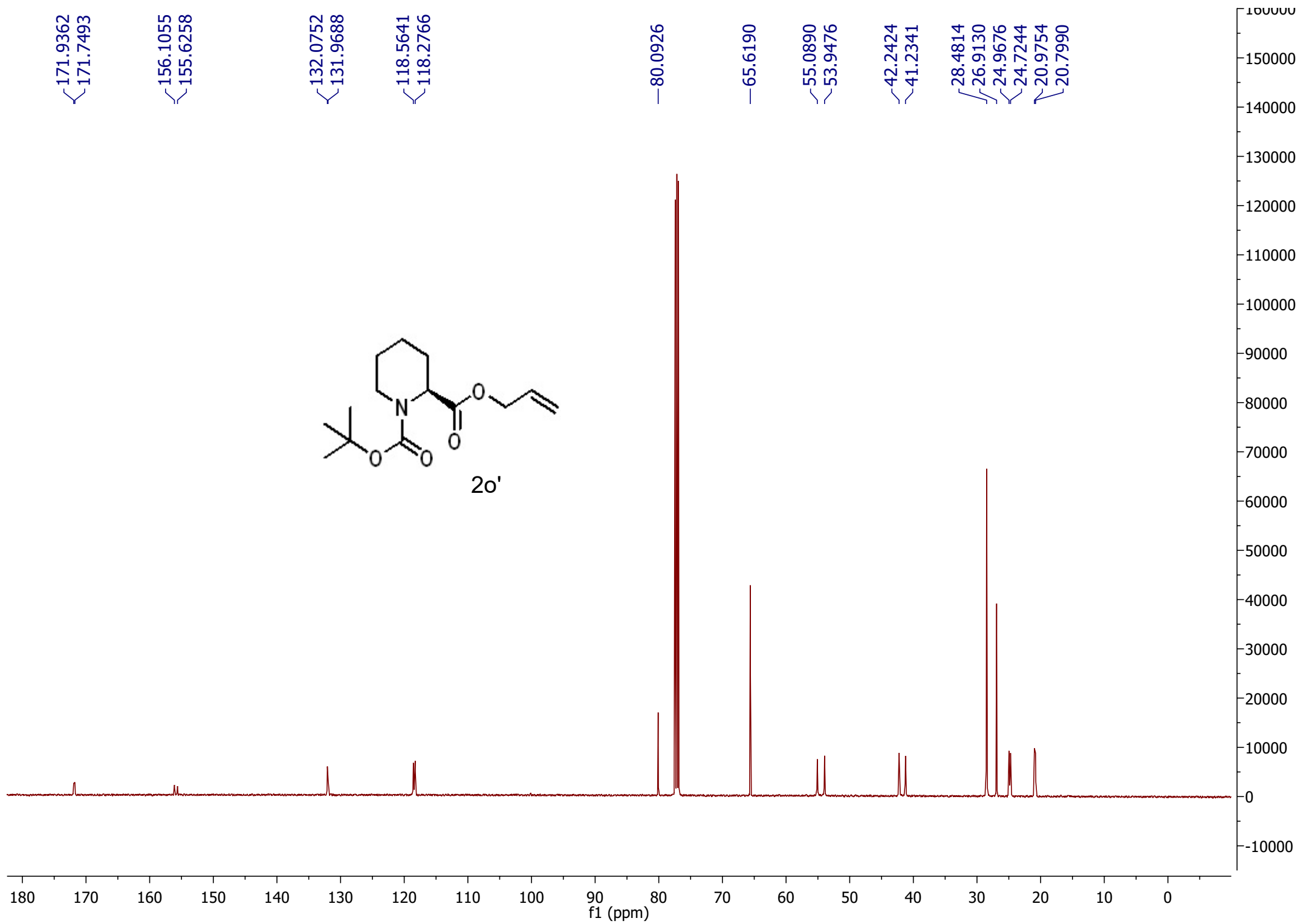


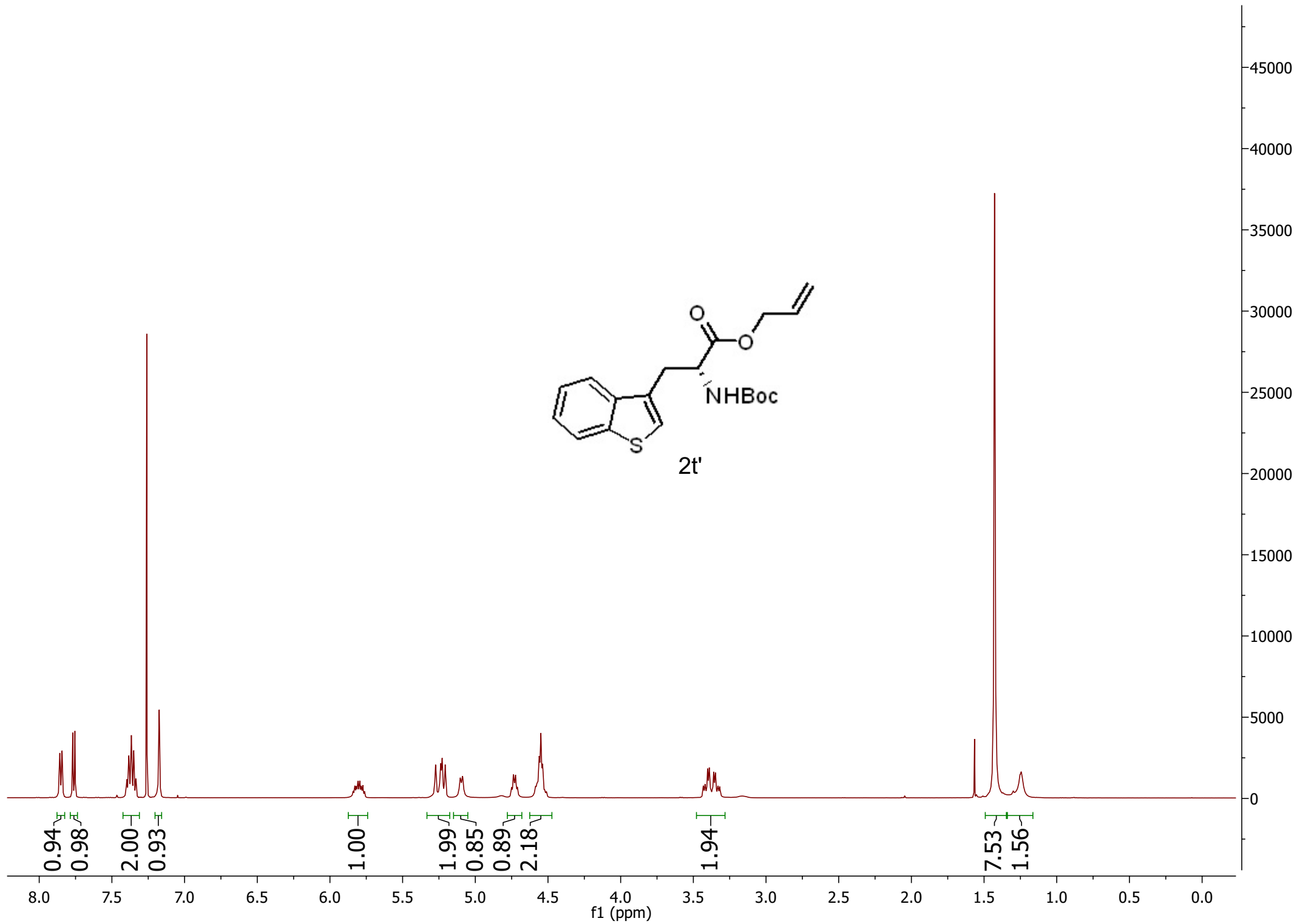


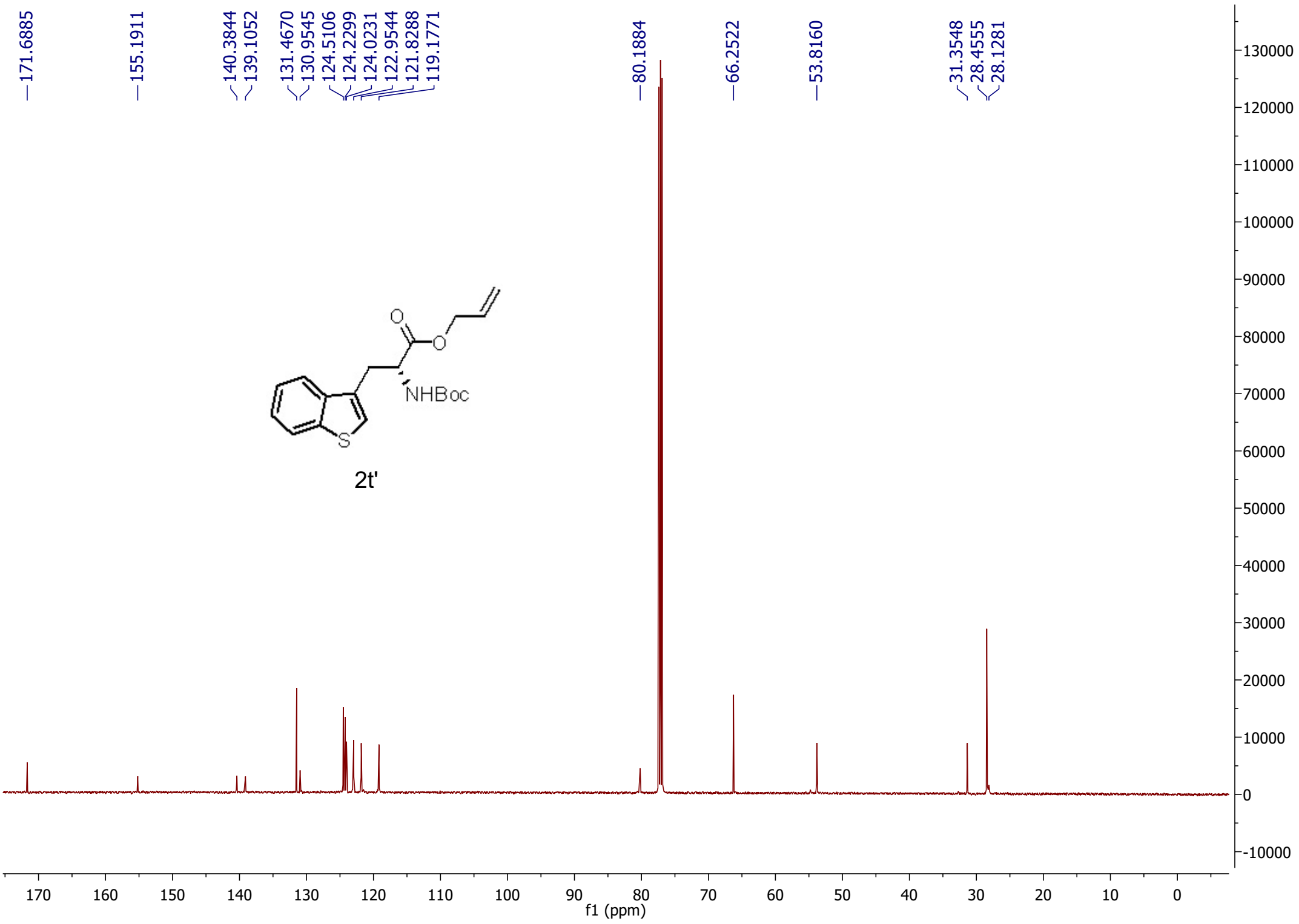


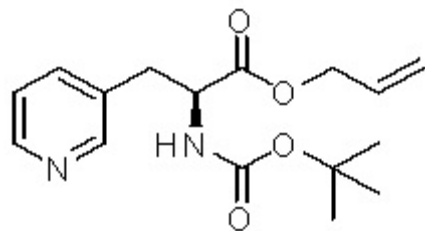
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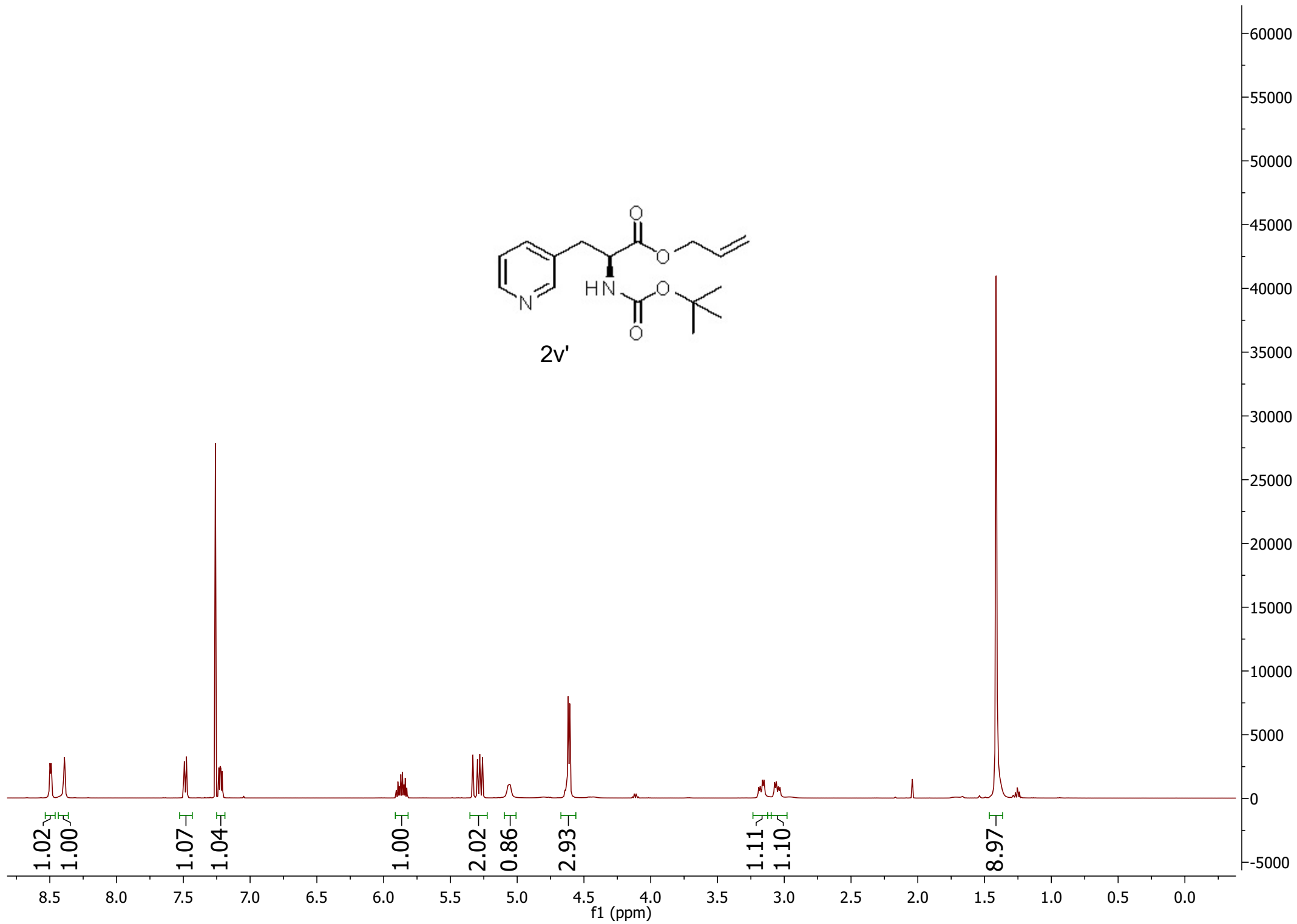


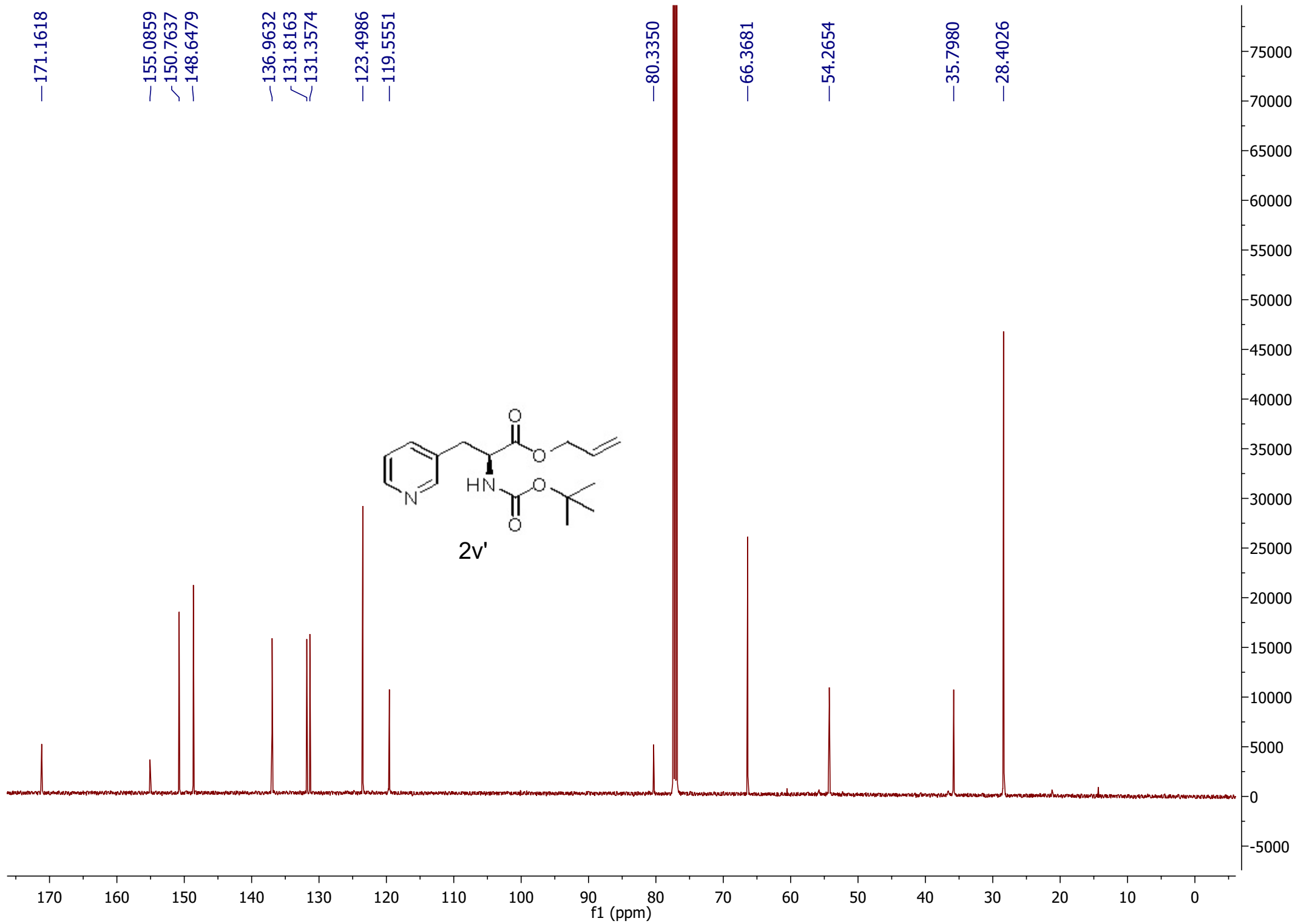


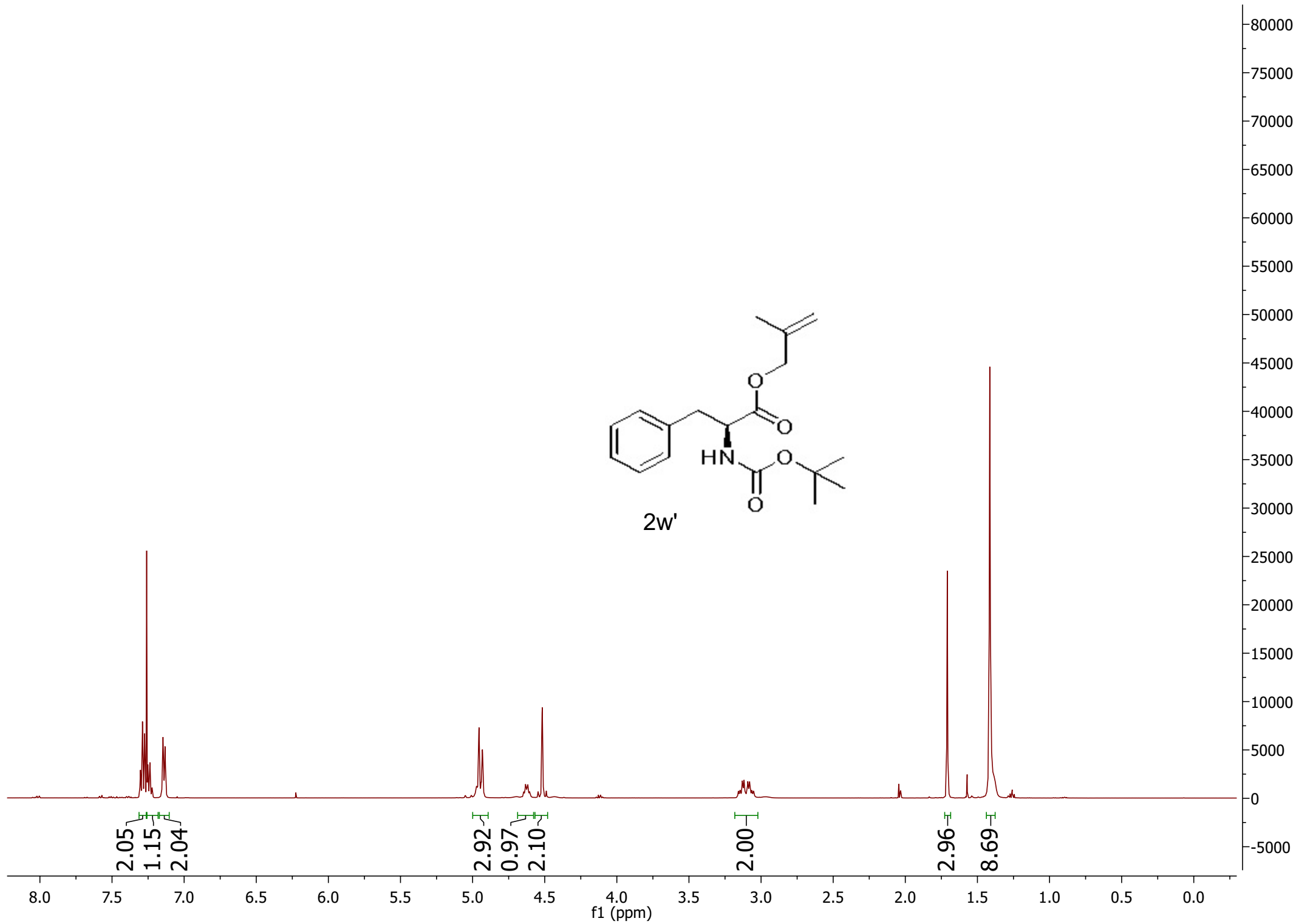




2v'







—171.7797

—155.2061

—139.3892

—136.1119

—129.4614

—128.6933

—127.1524

—113.8833

—80.0470

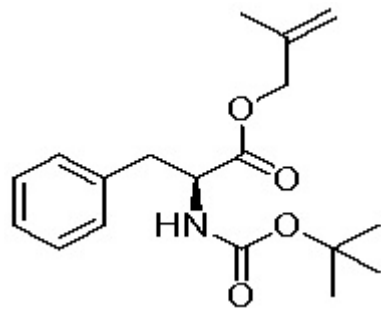
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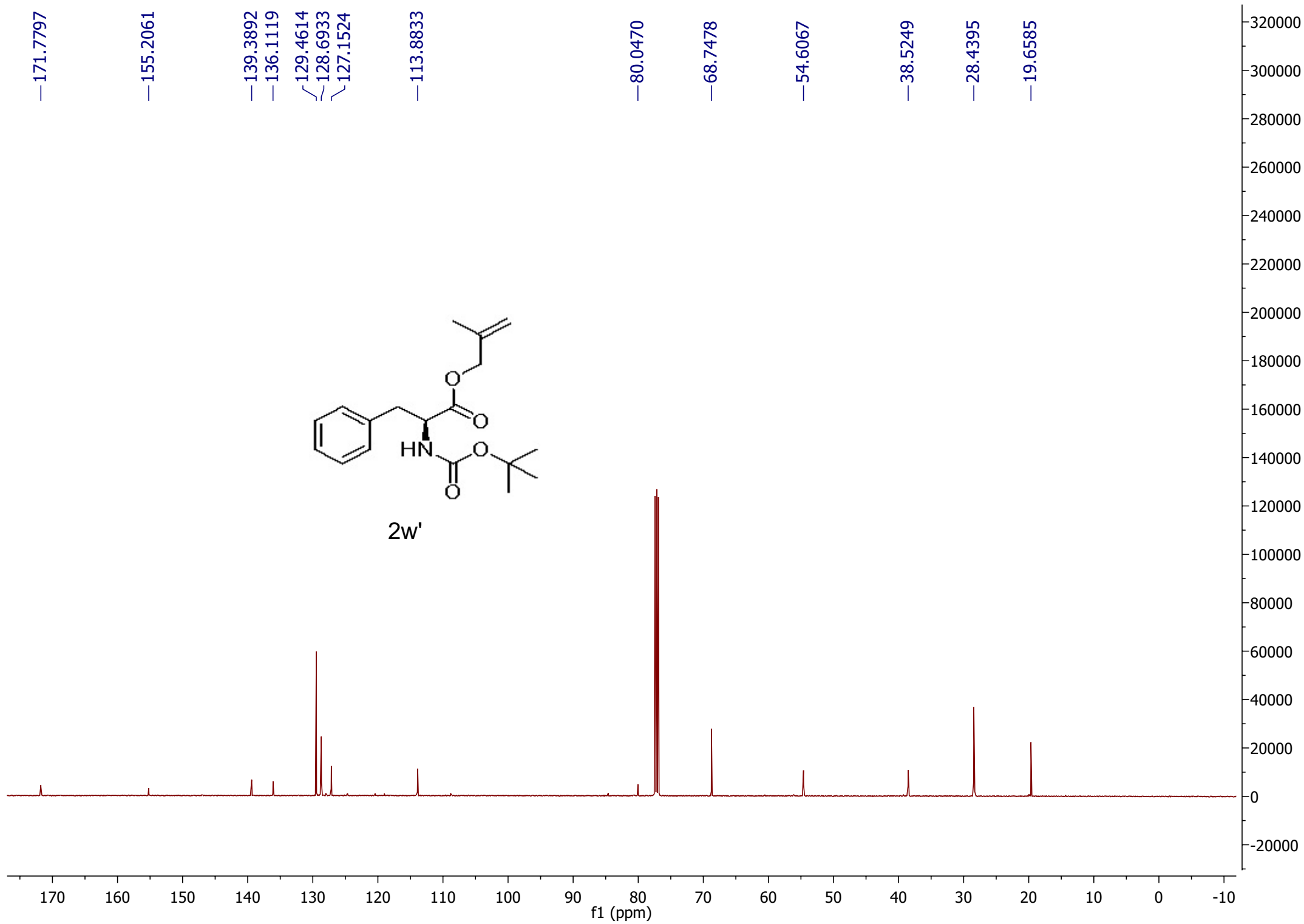
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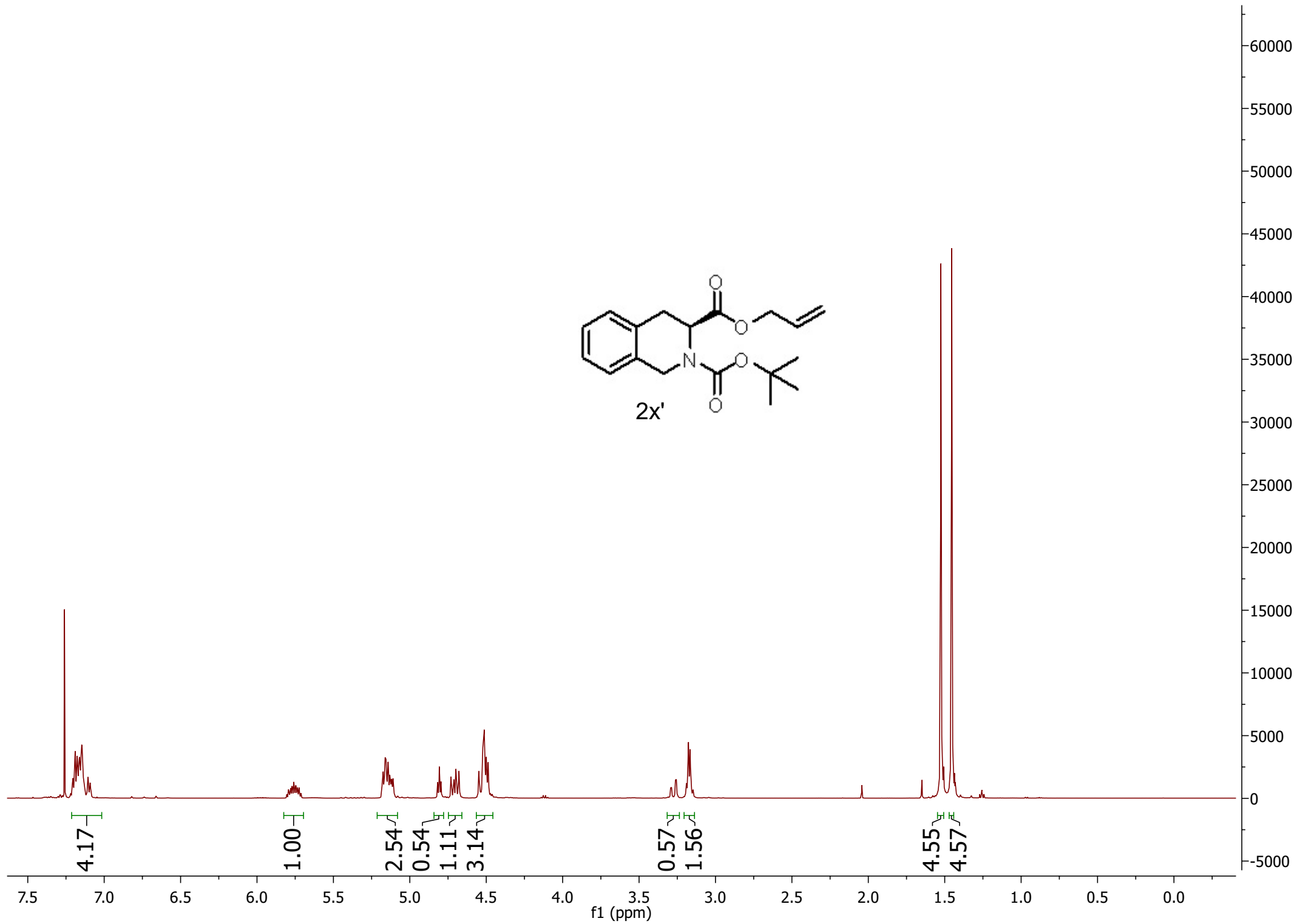
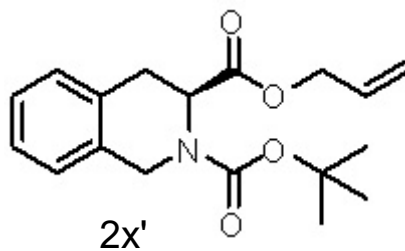
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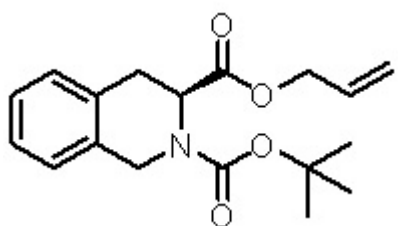
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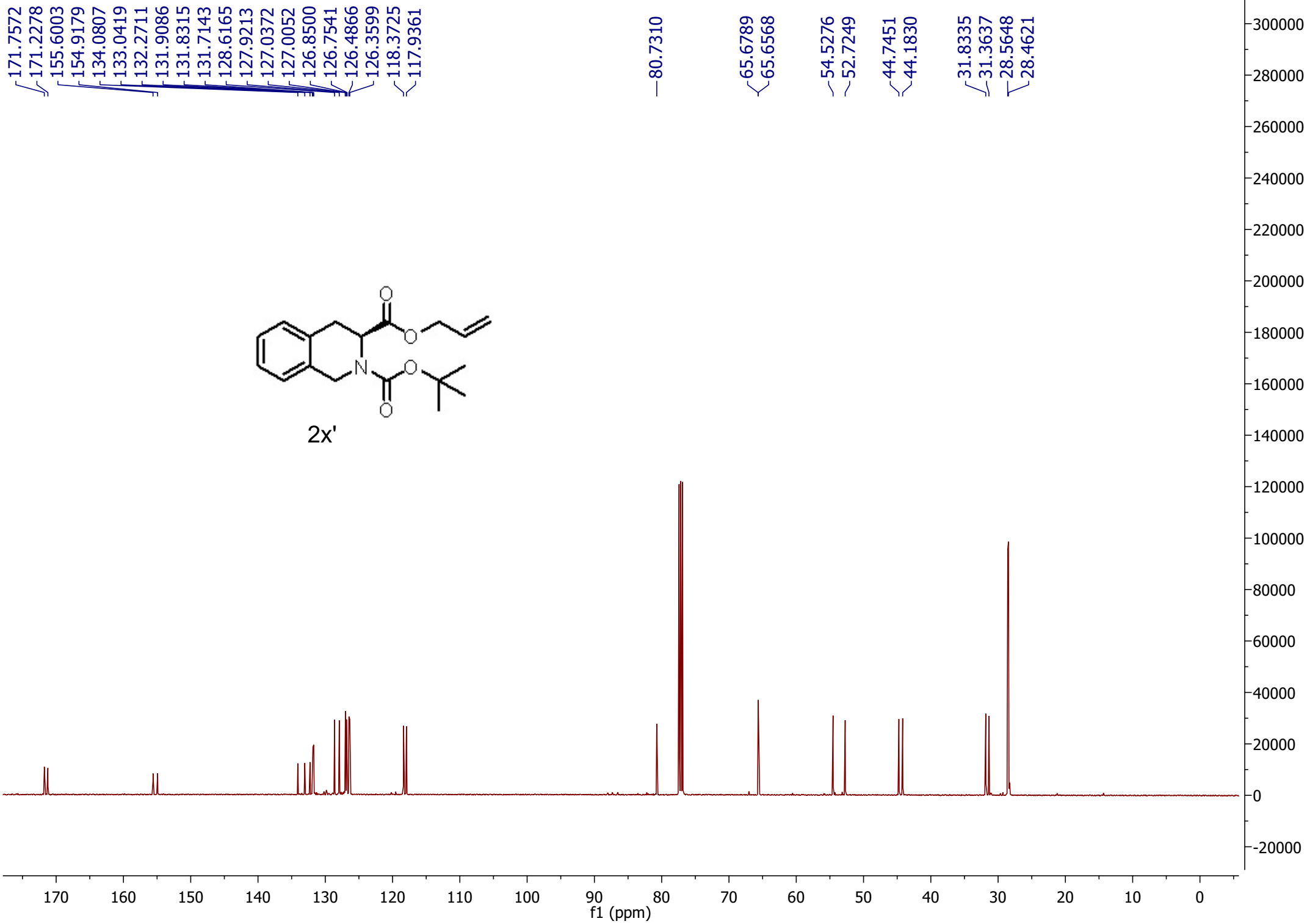
2w'



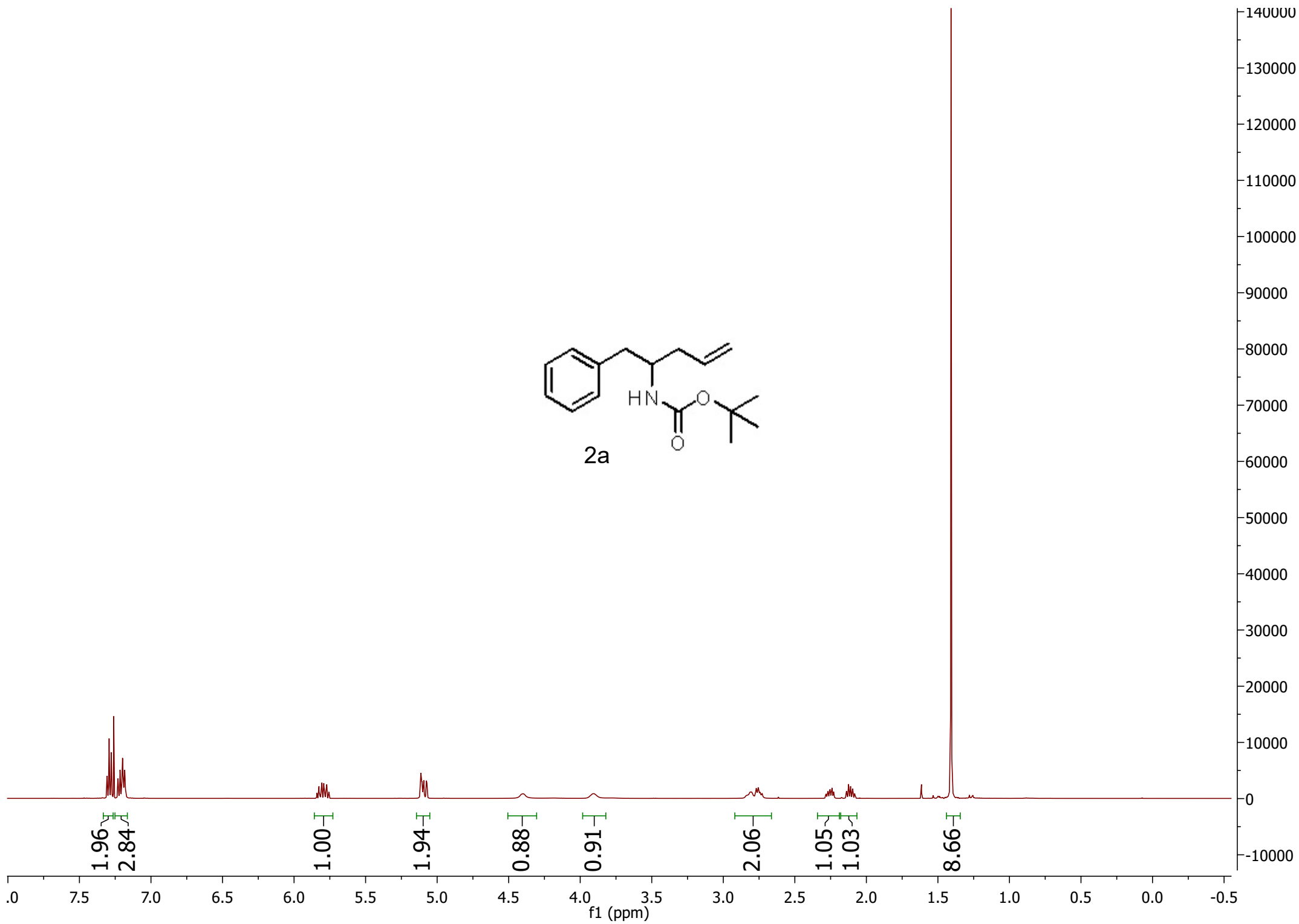
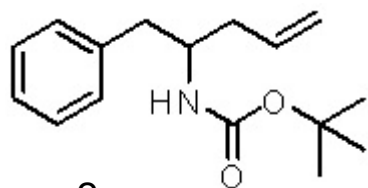


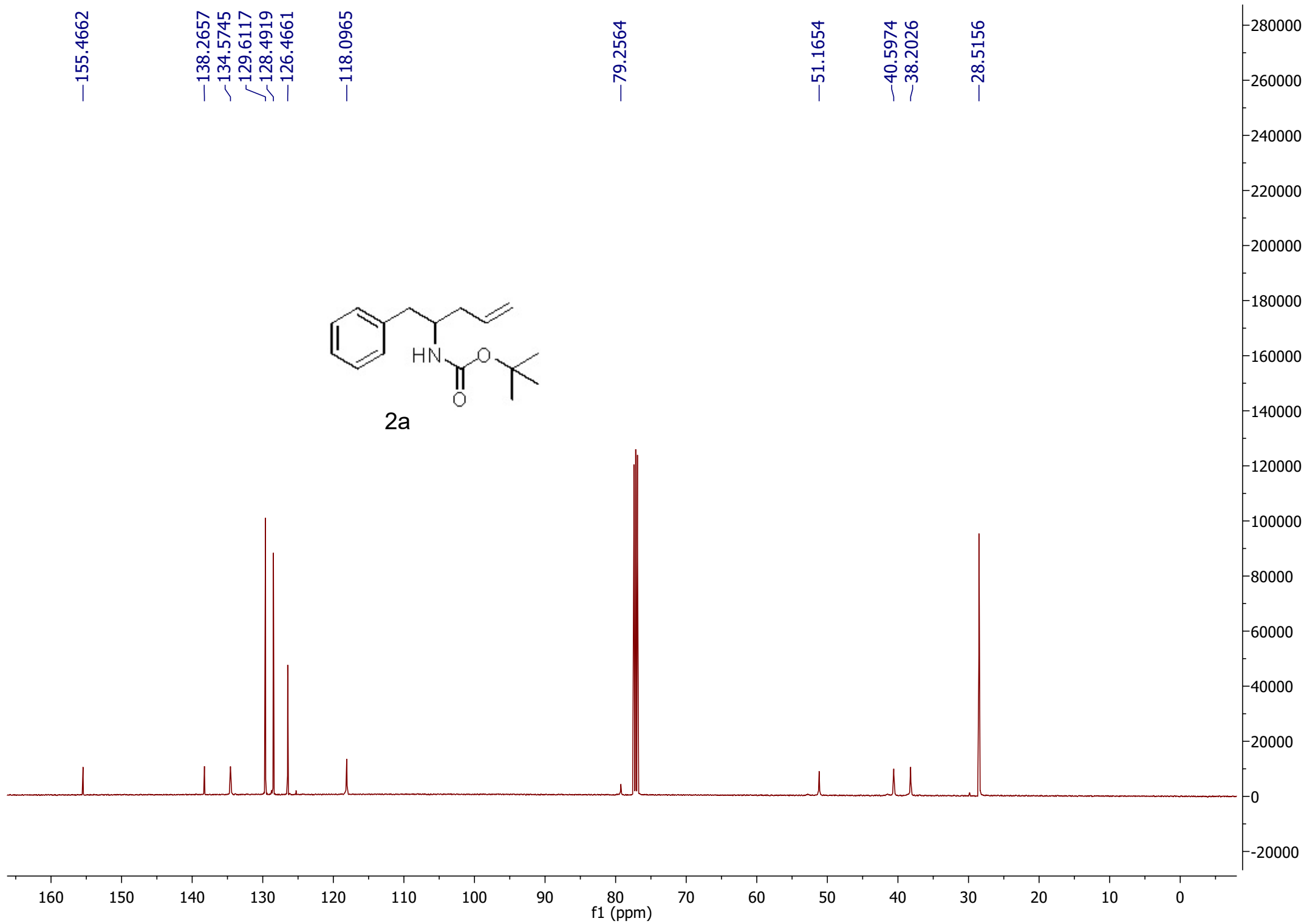


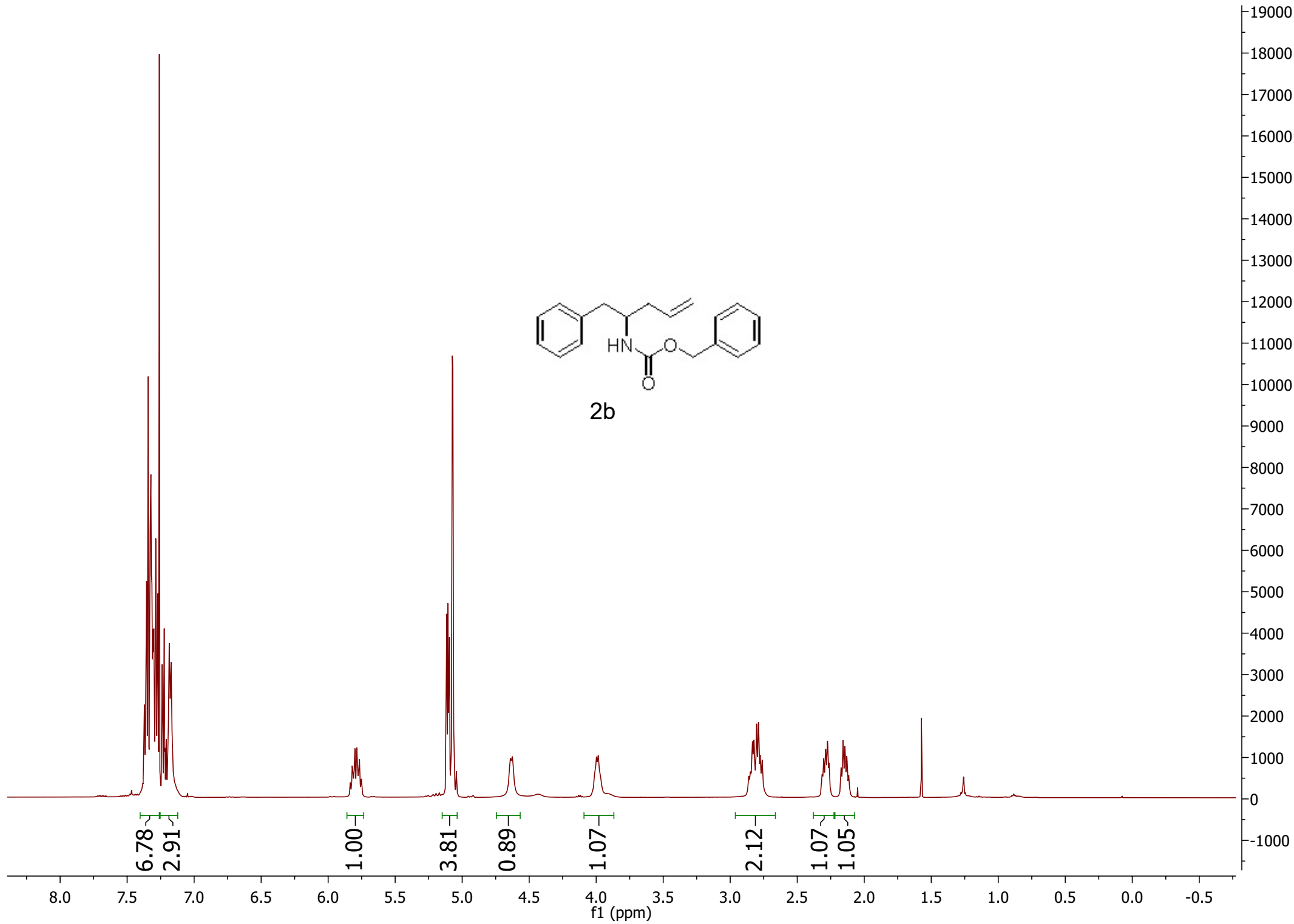
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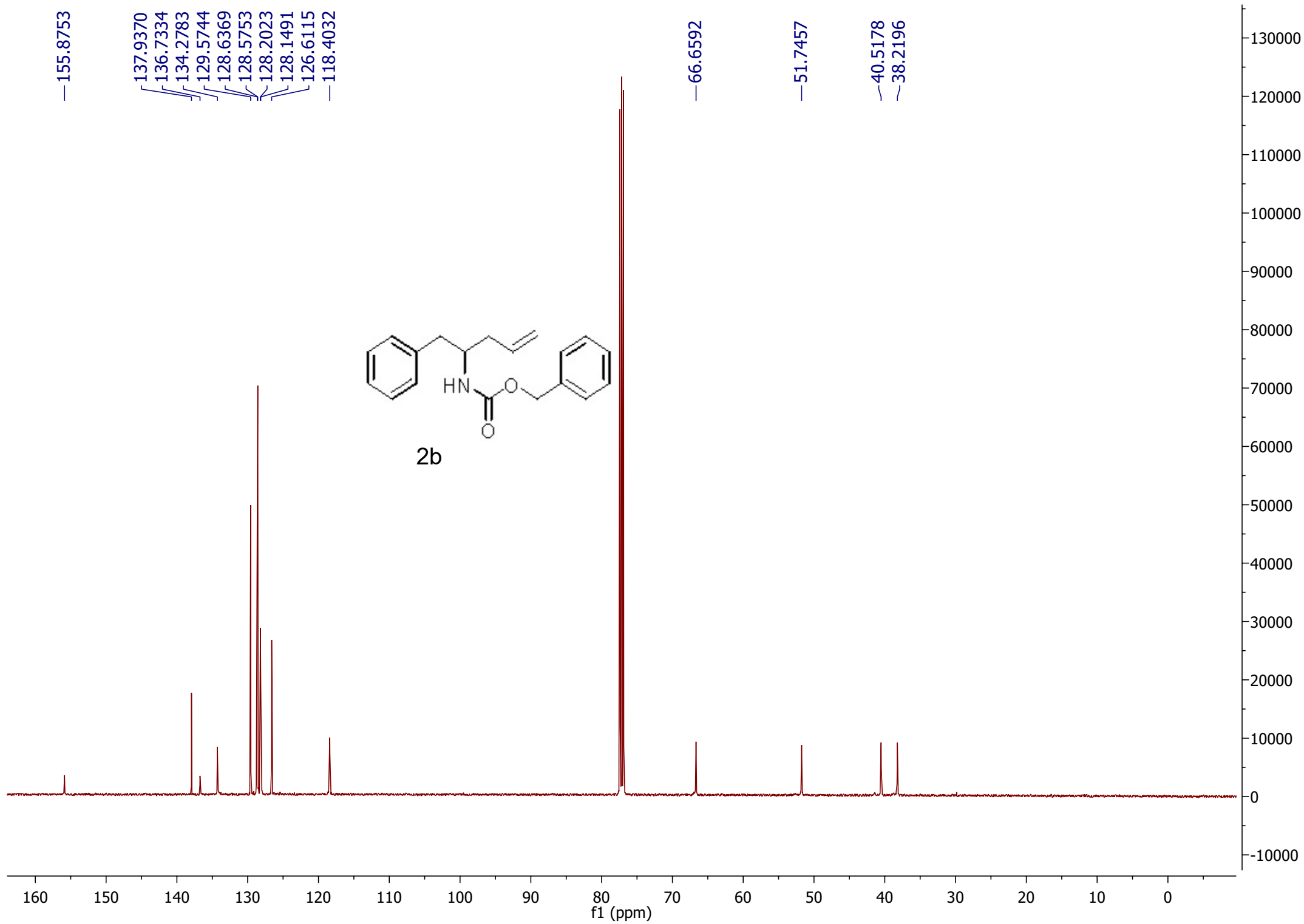


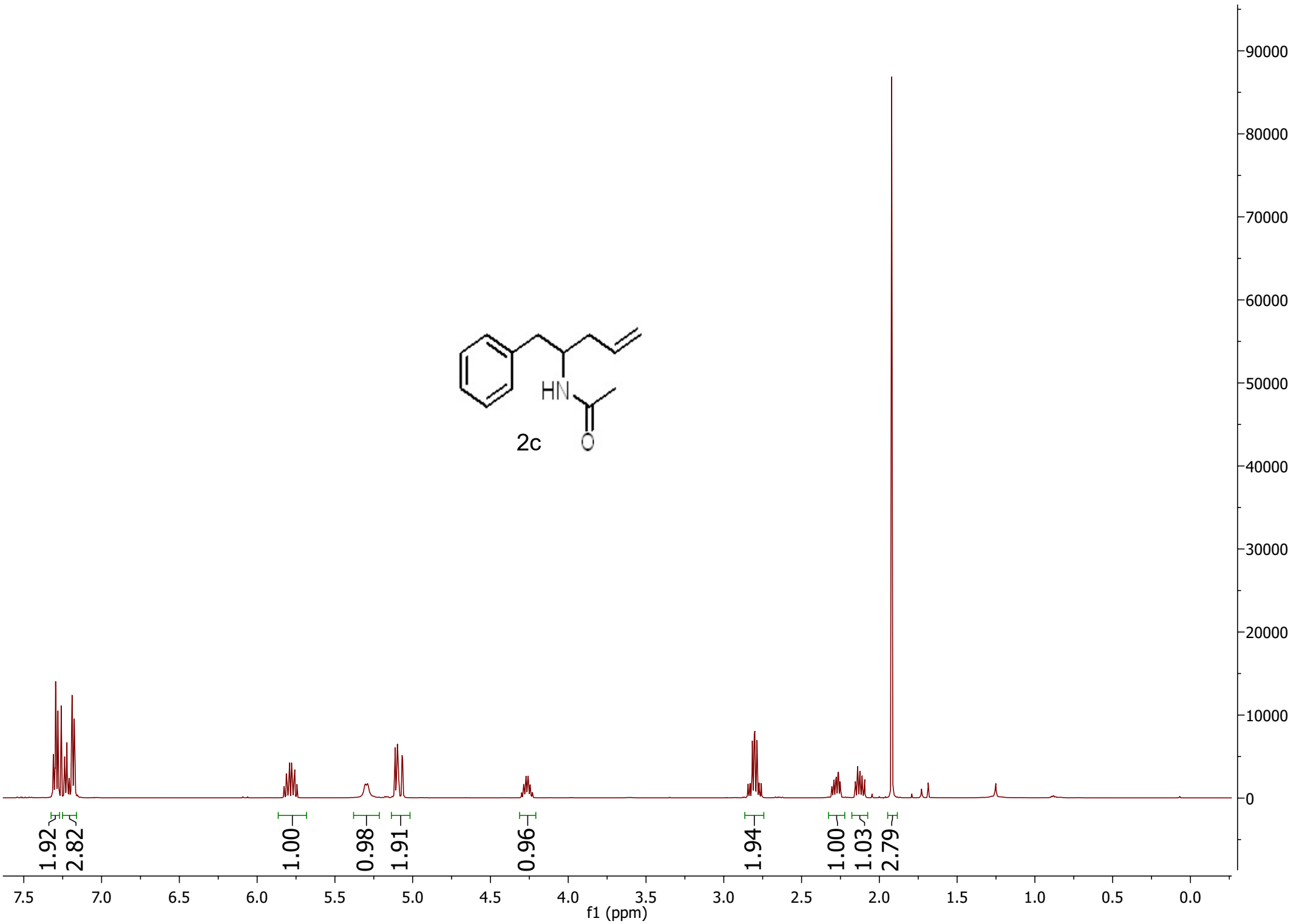
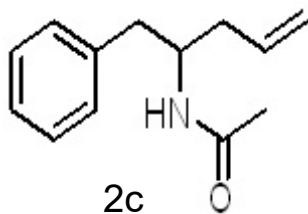
Product NMRs











—169.6053

—137.9284

—134.4656

—129.5335

—128.5575

—126.6129

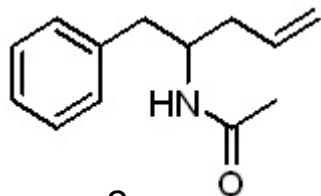
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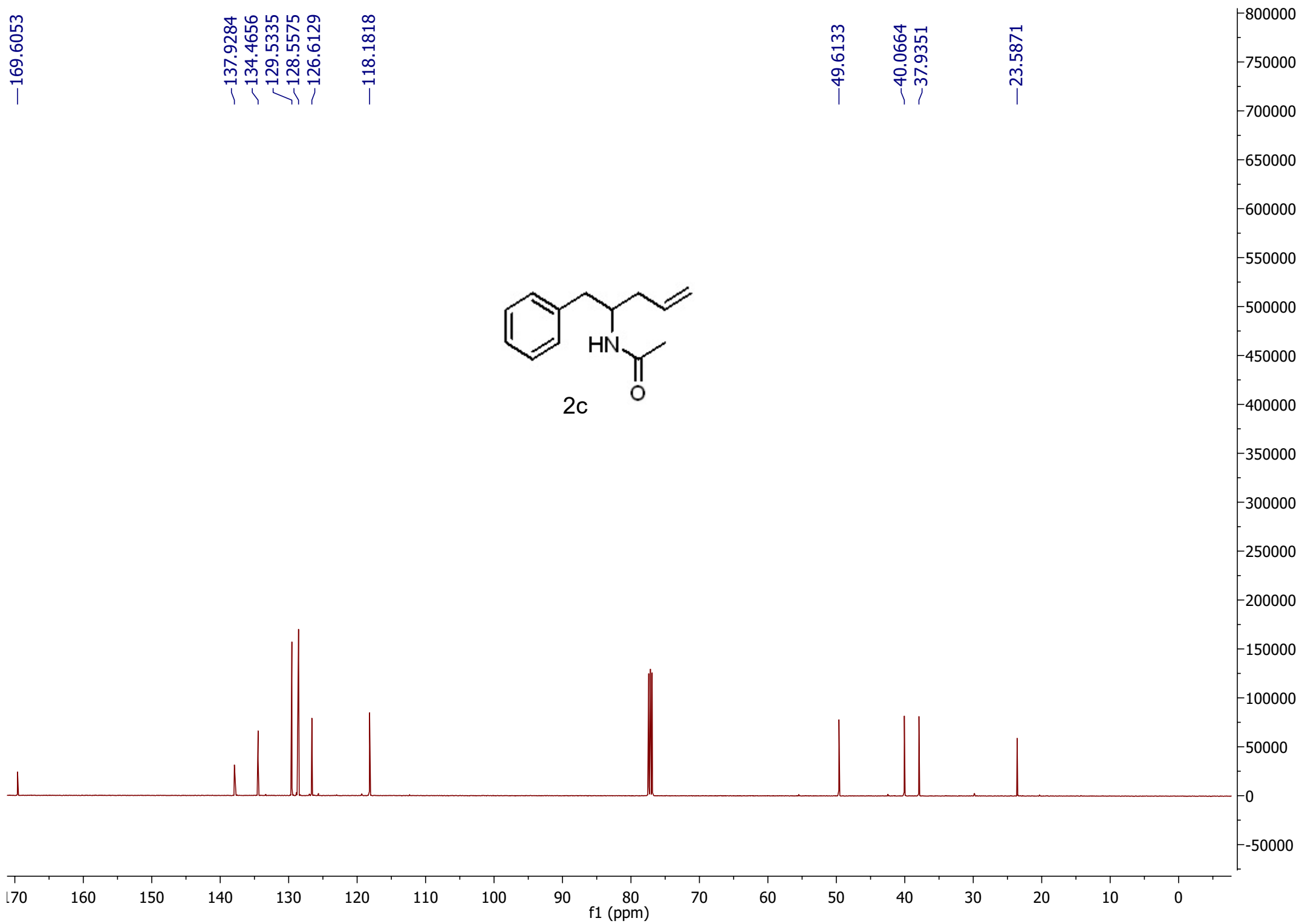
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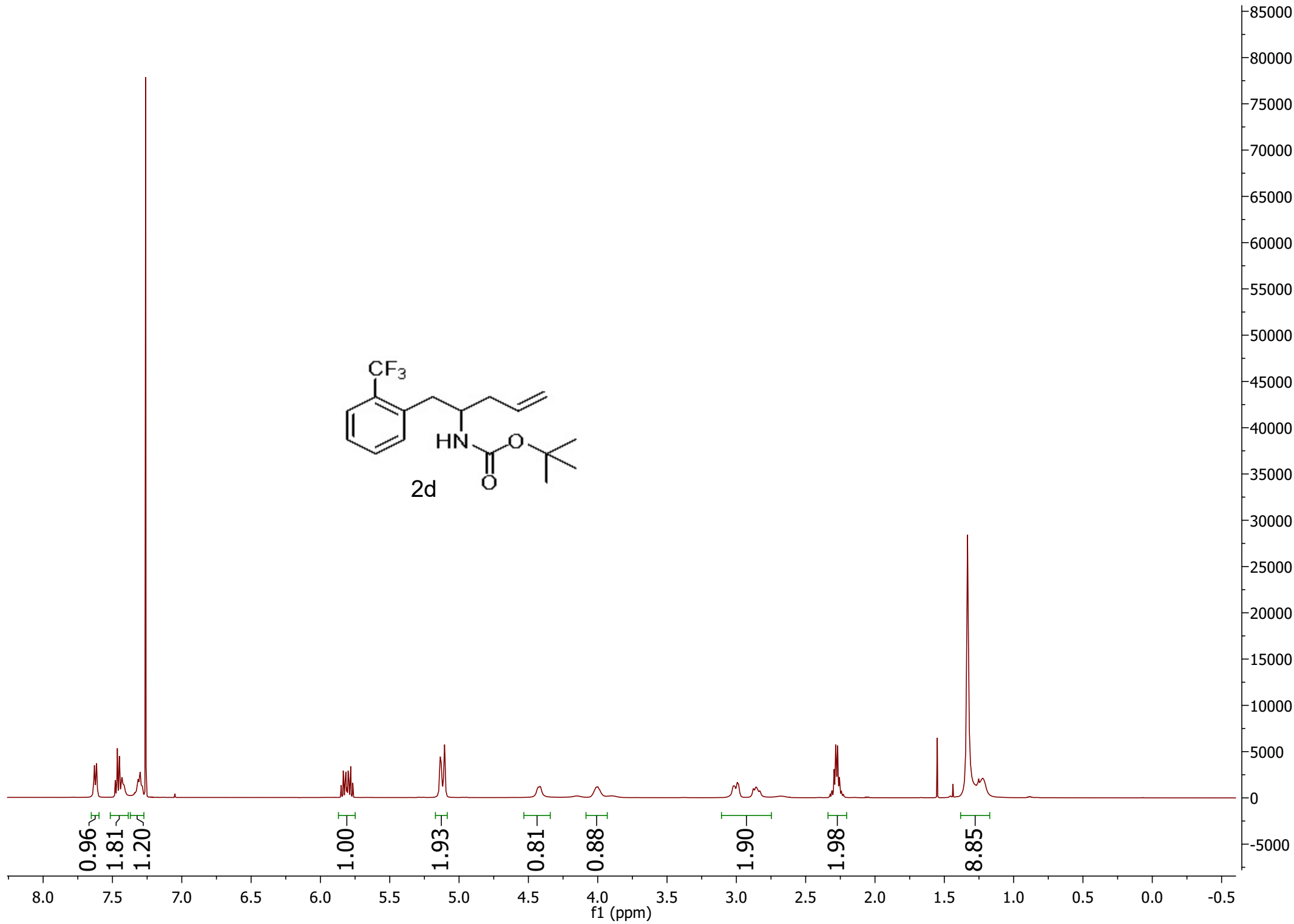
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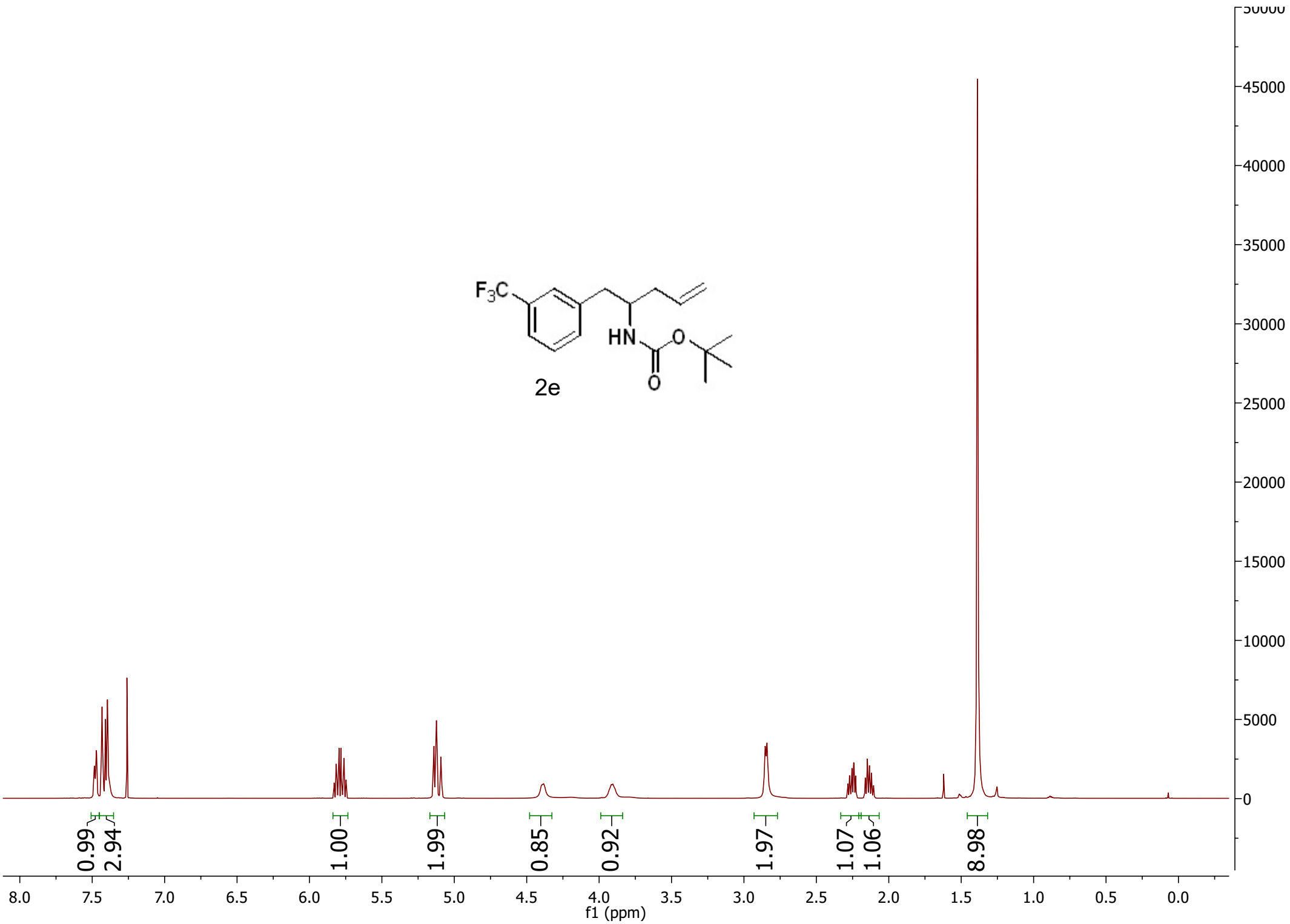
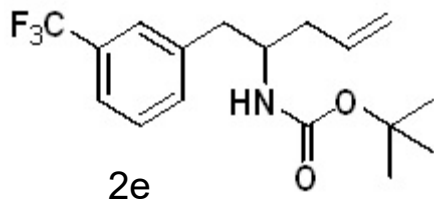
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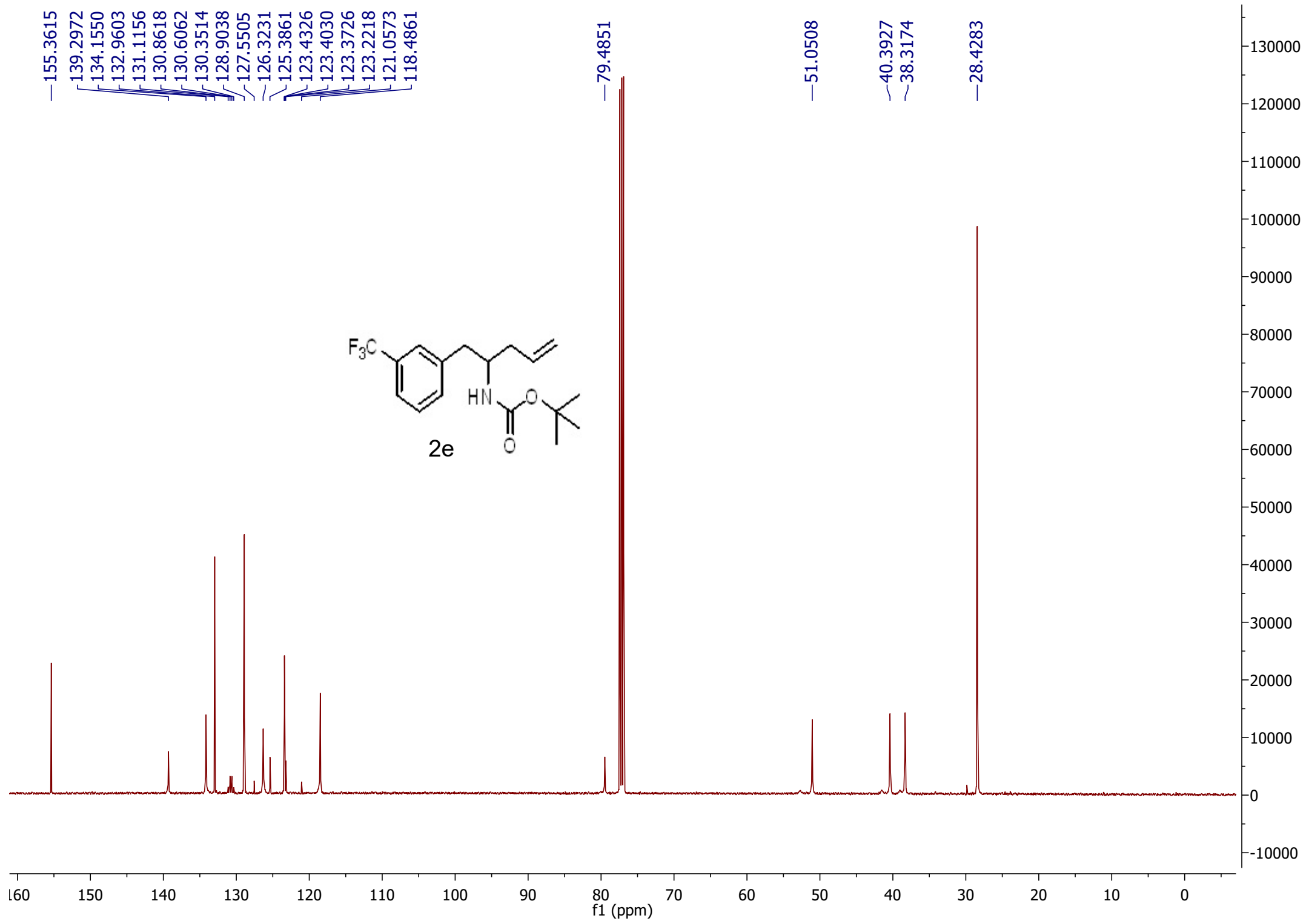


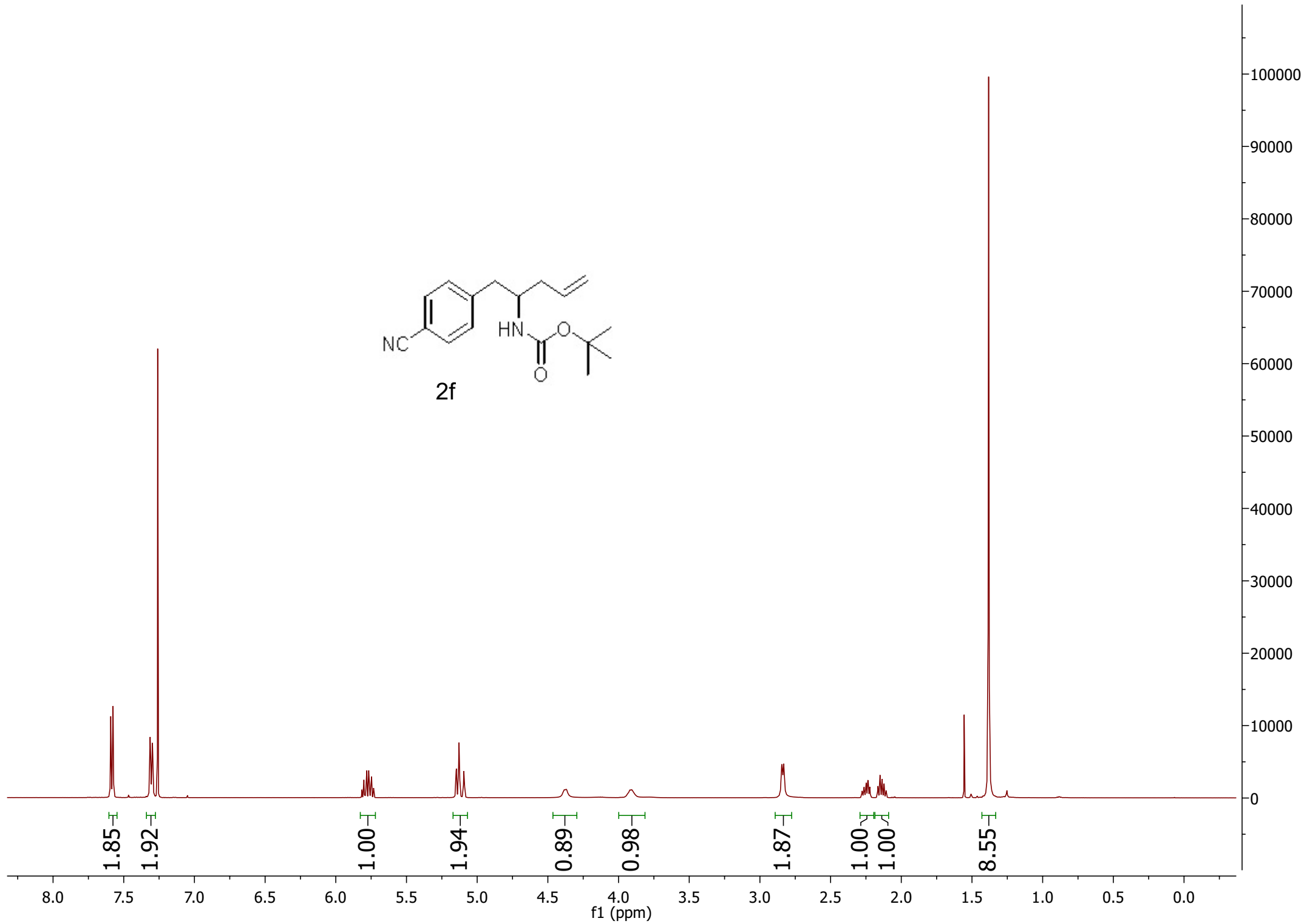
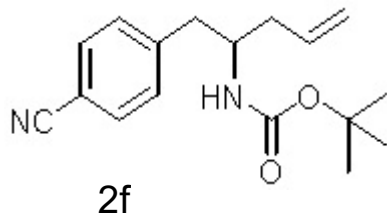
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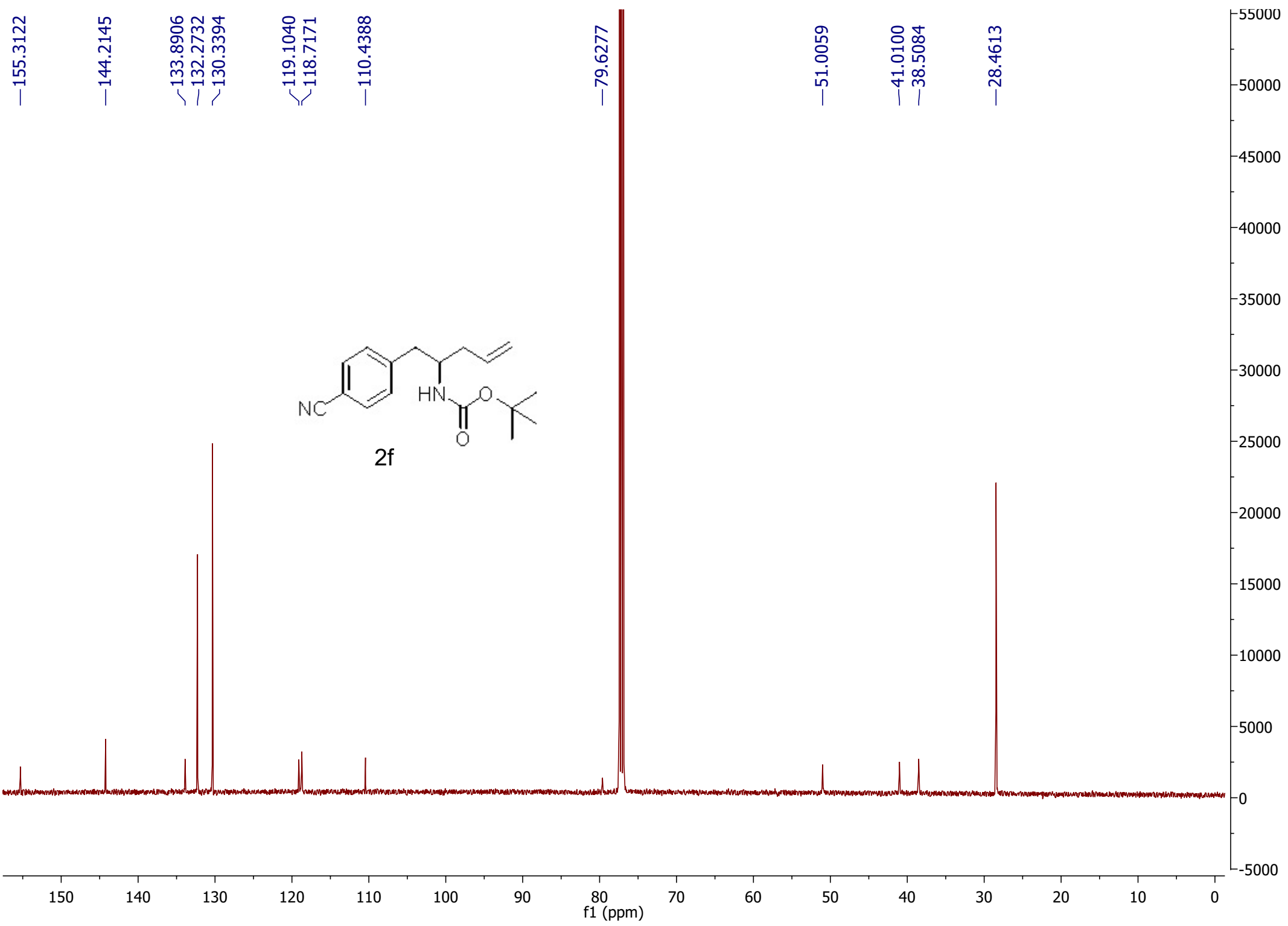


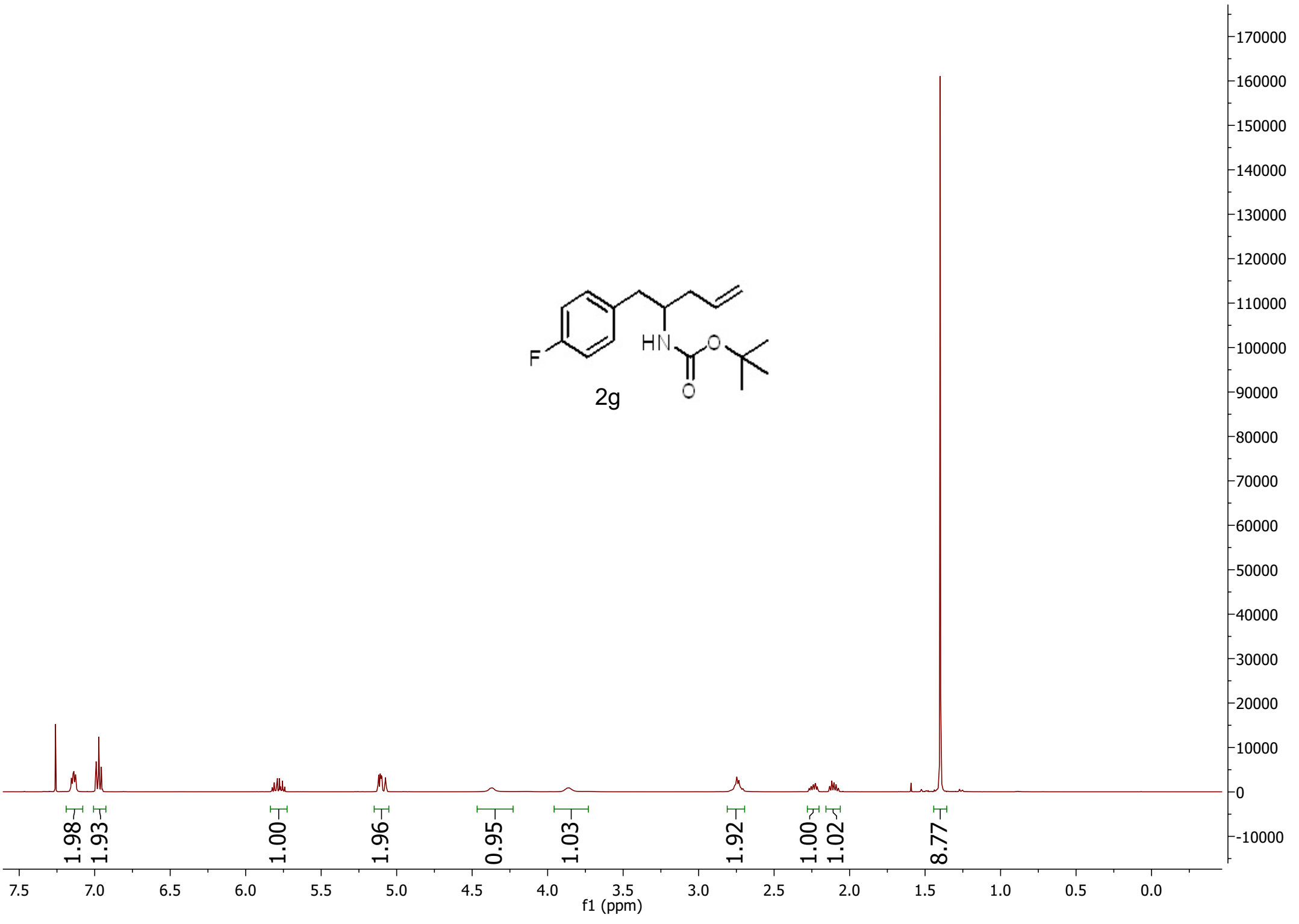
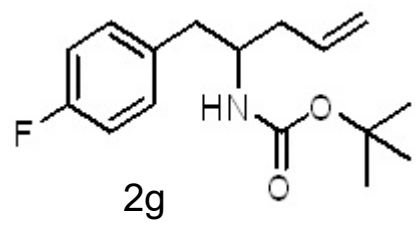












~162.7109
~160.7696
~155.4335

~134.3801
~133.9795
~133.9575
~130.9729
~130.9107

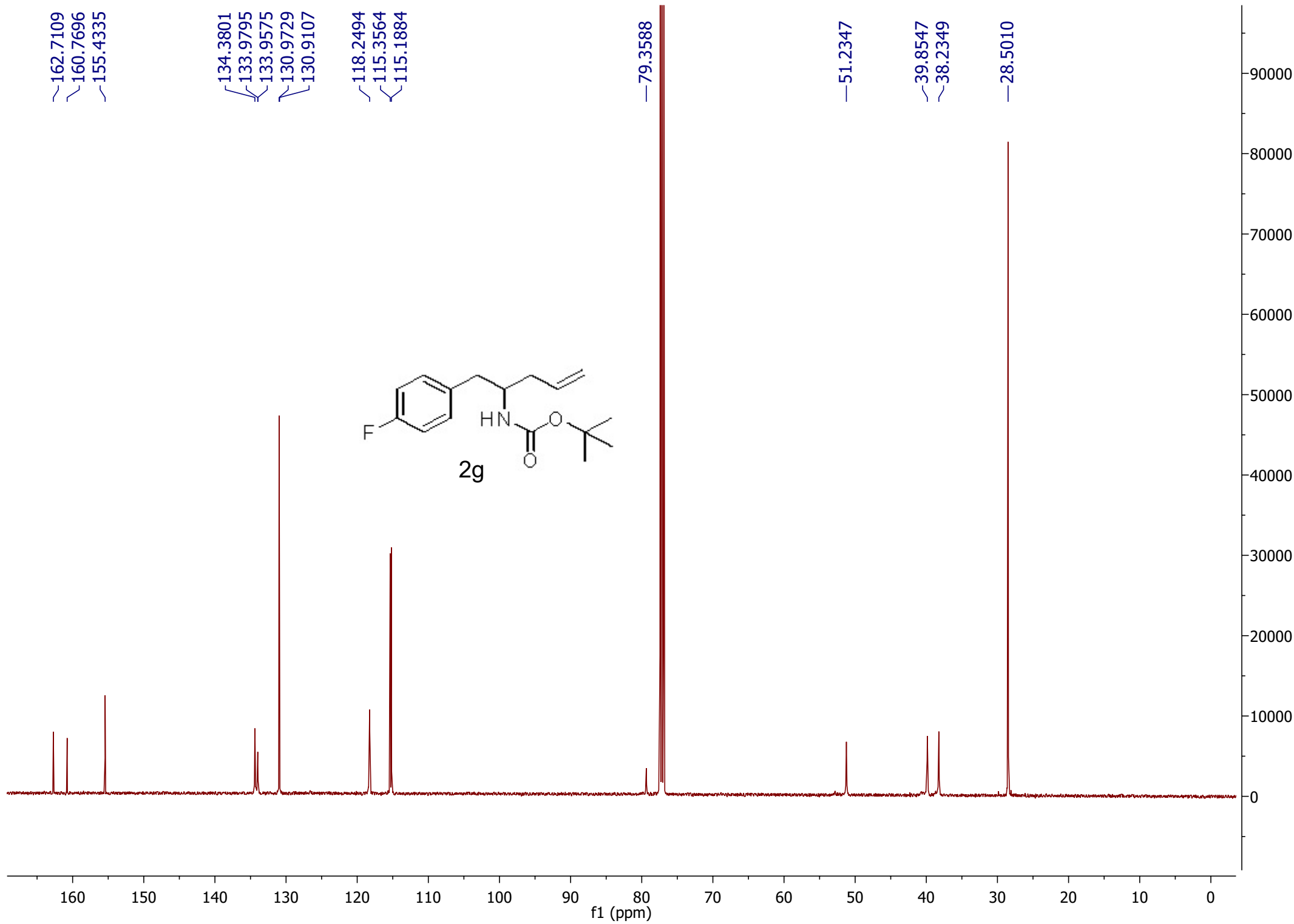
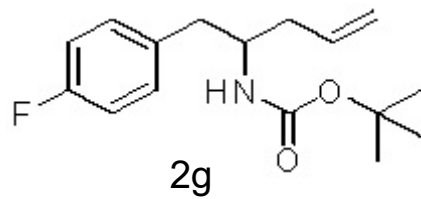
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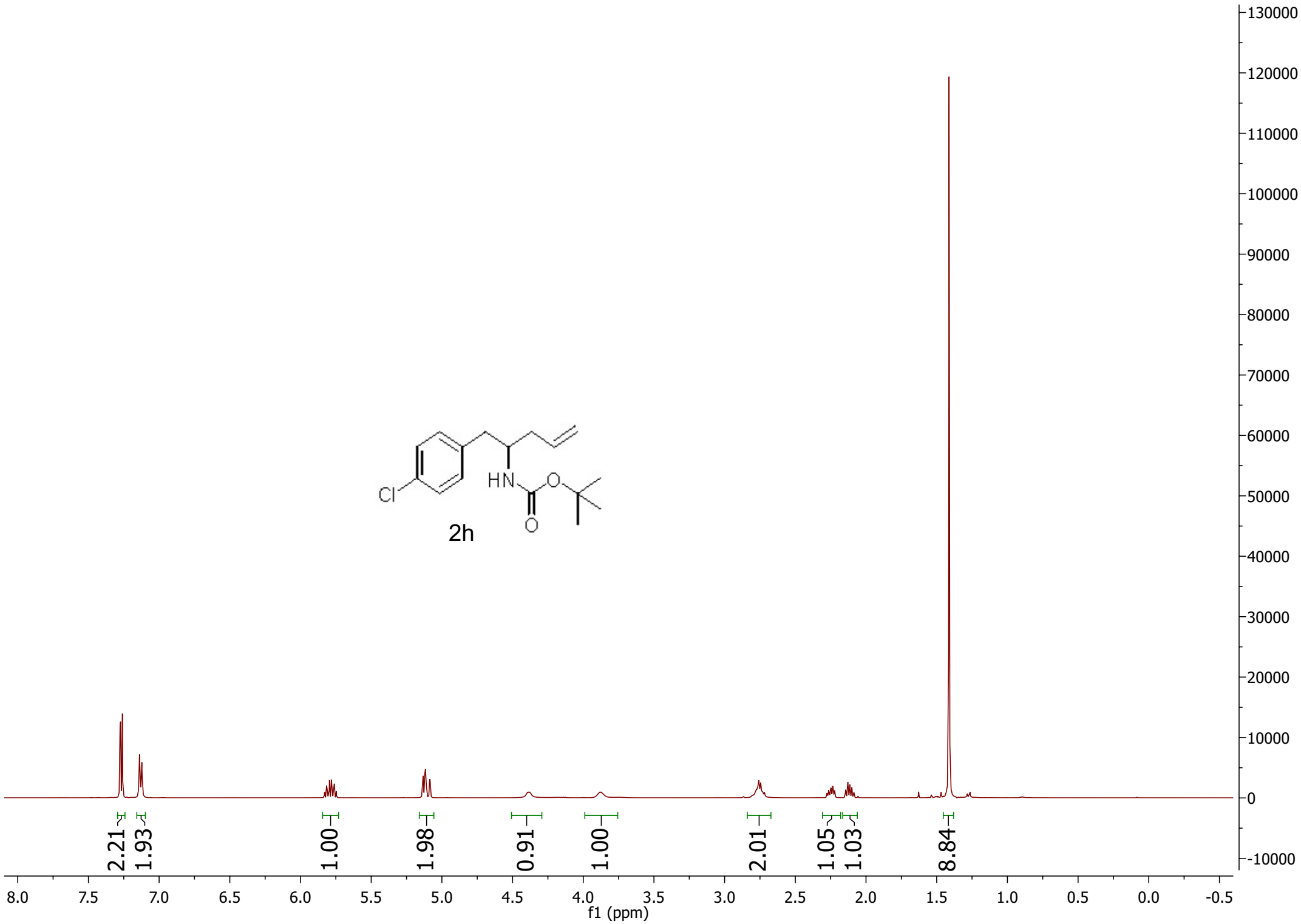
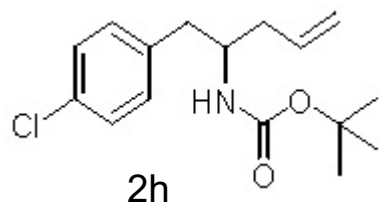
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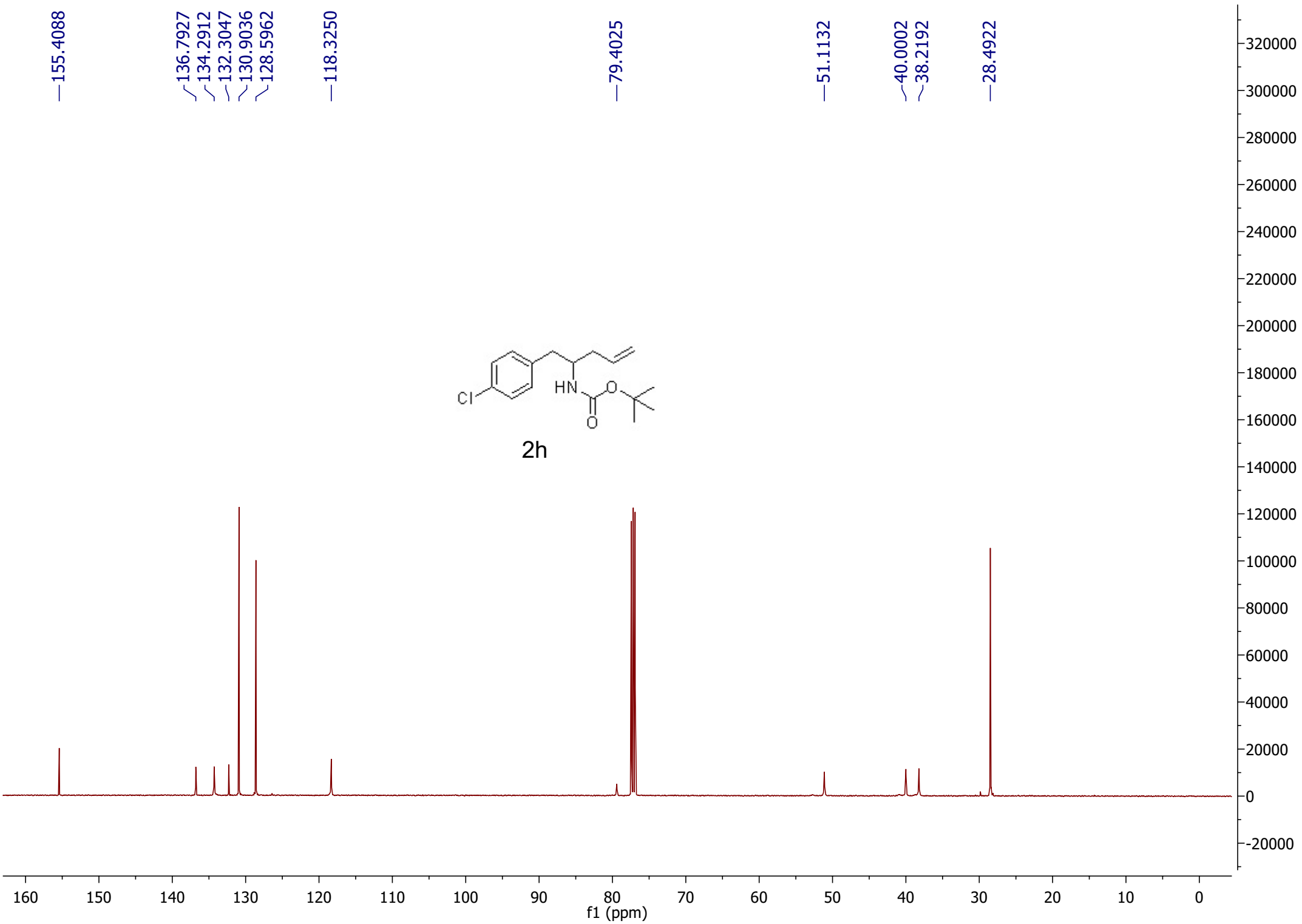
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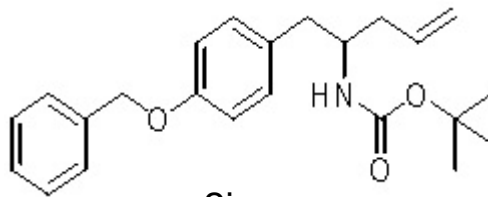
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~38.2349

~28.5010

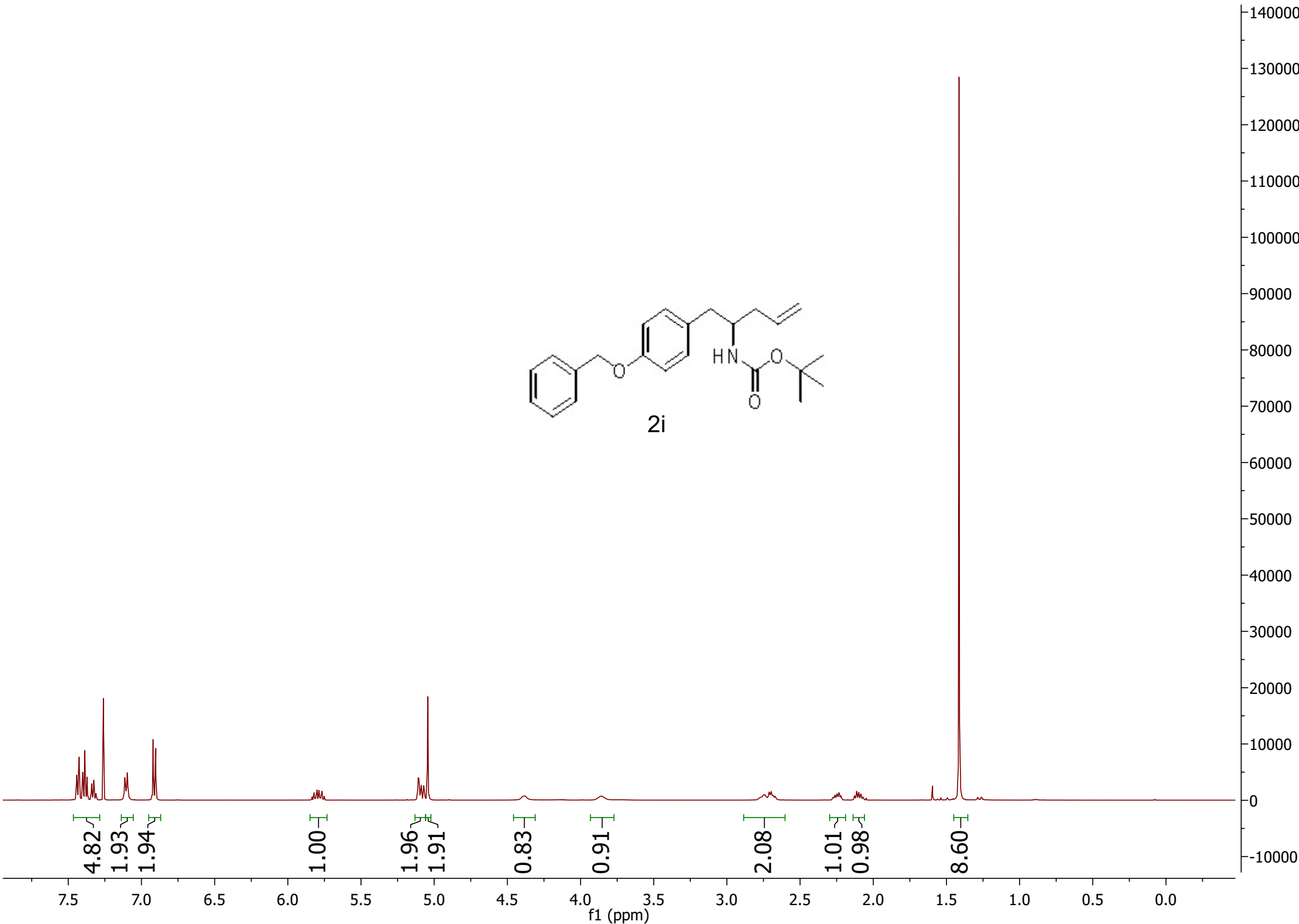


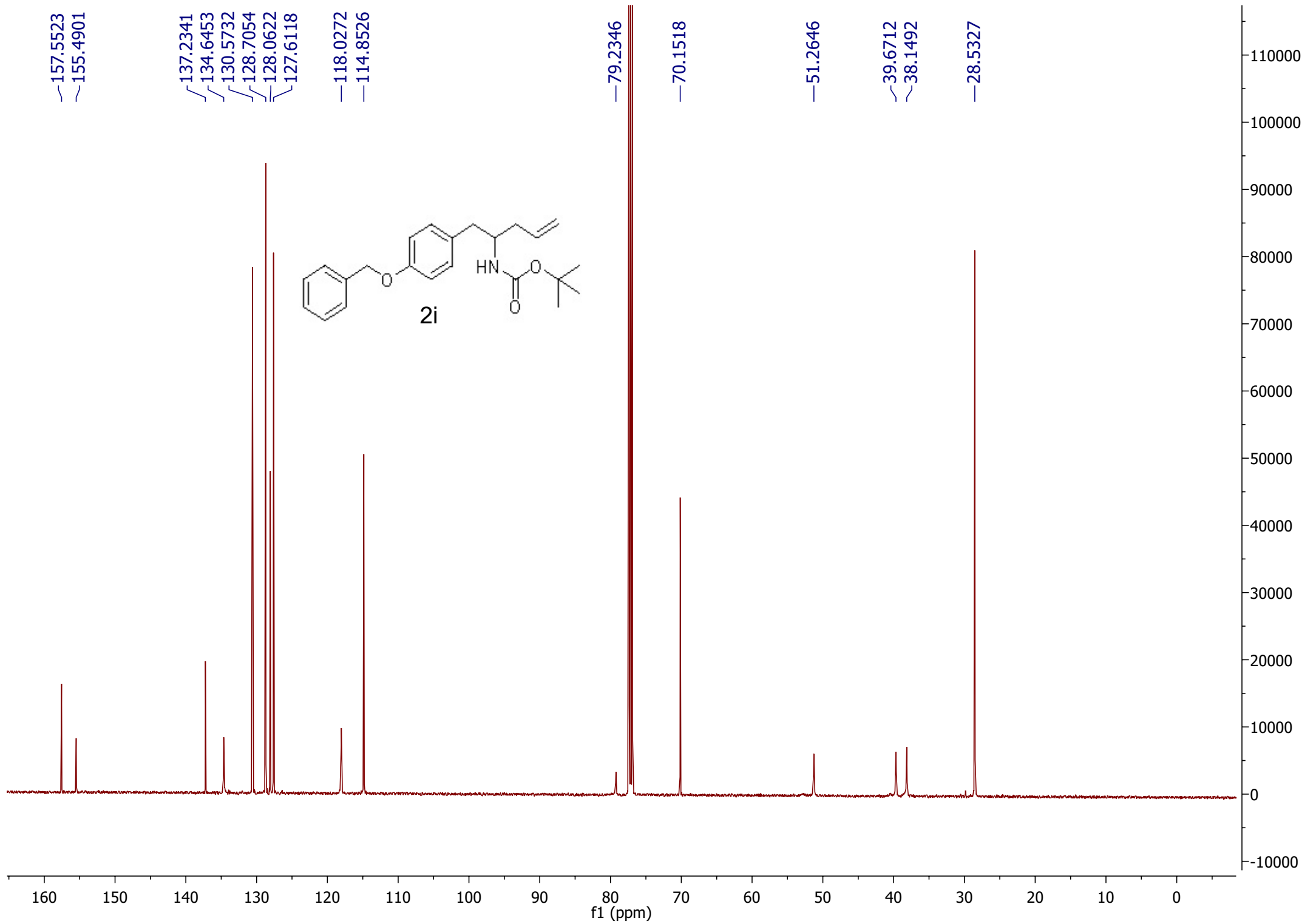


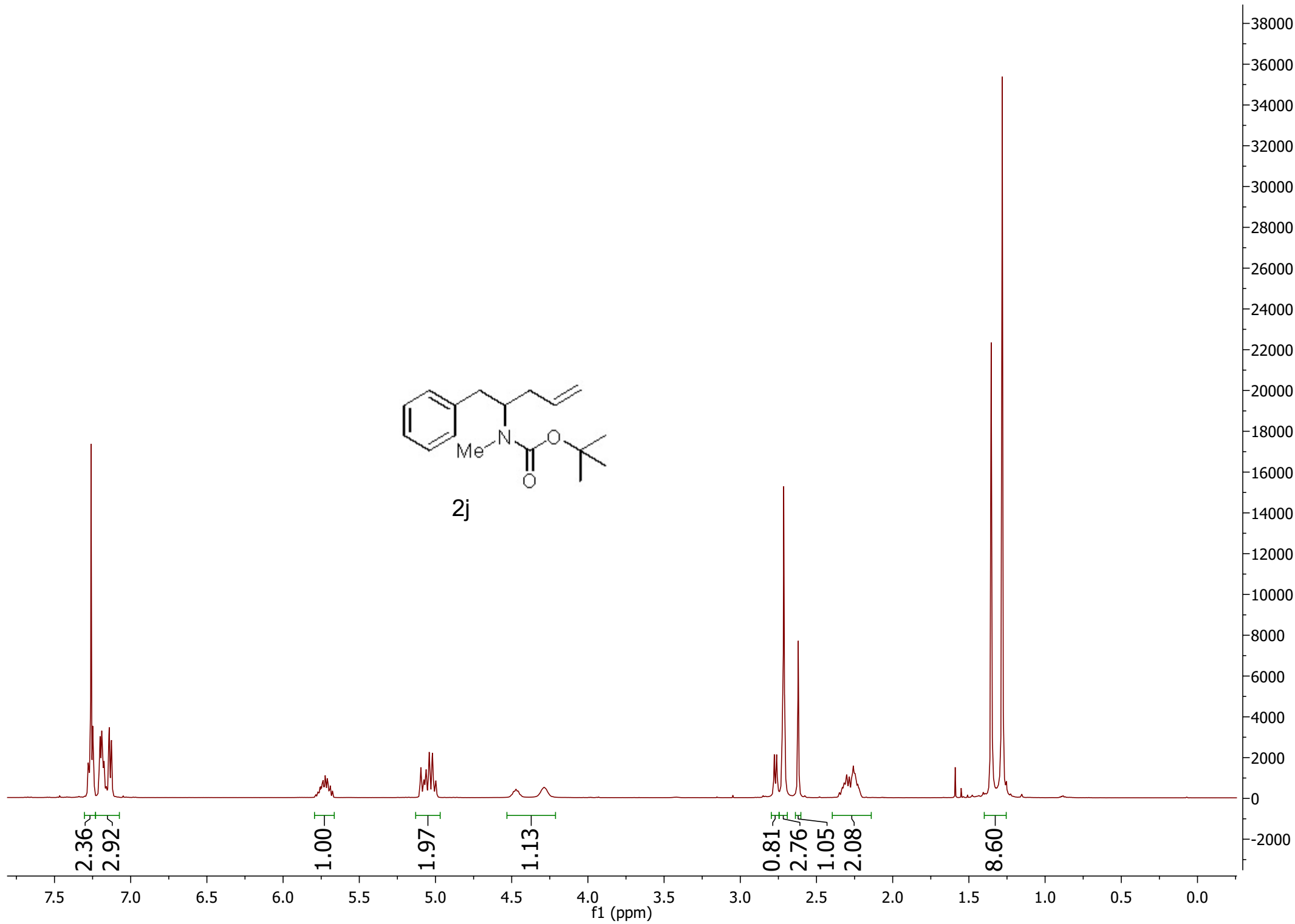


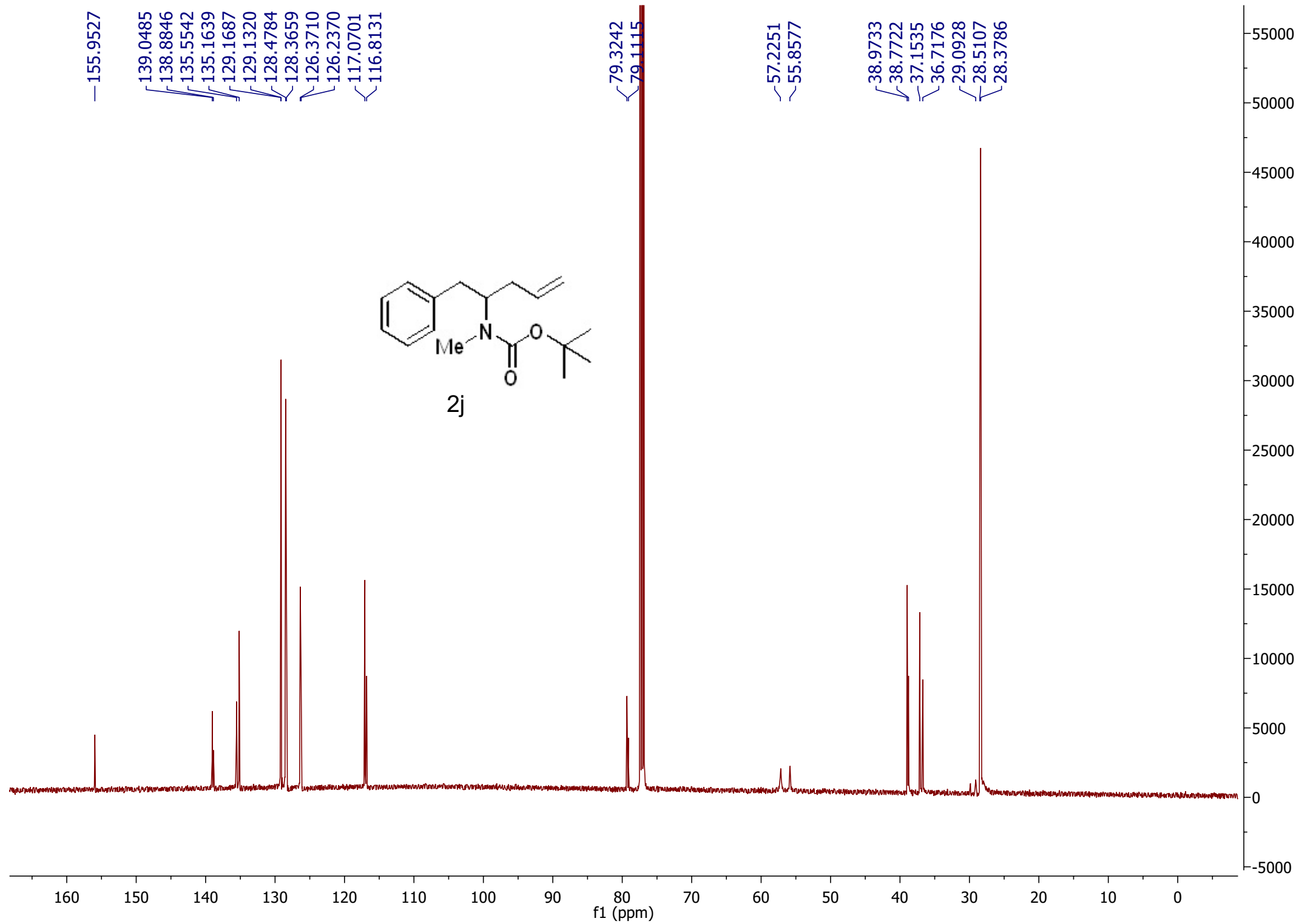


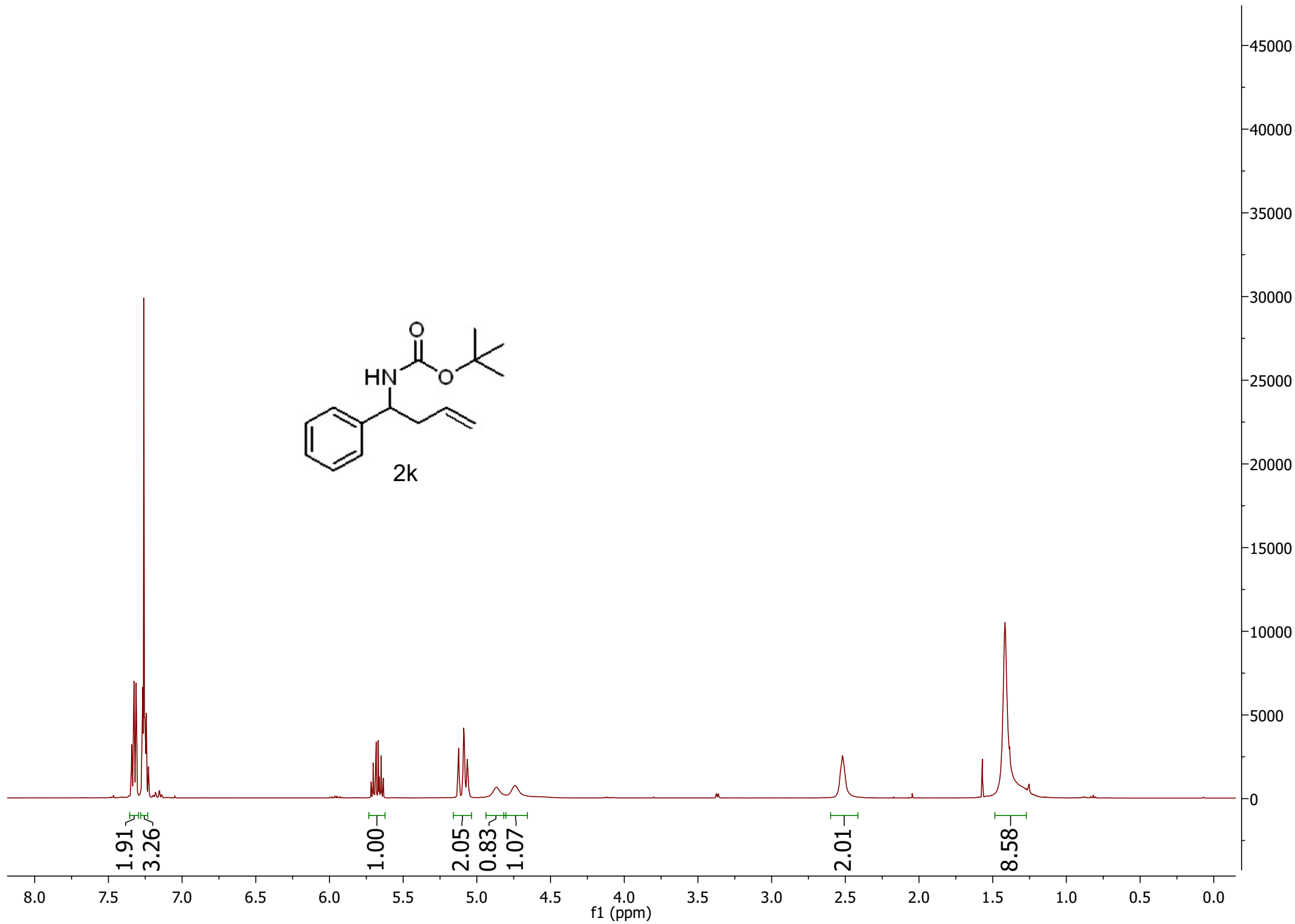
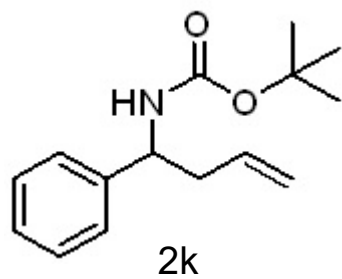
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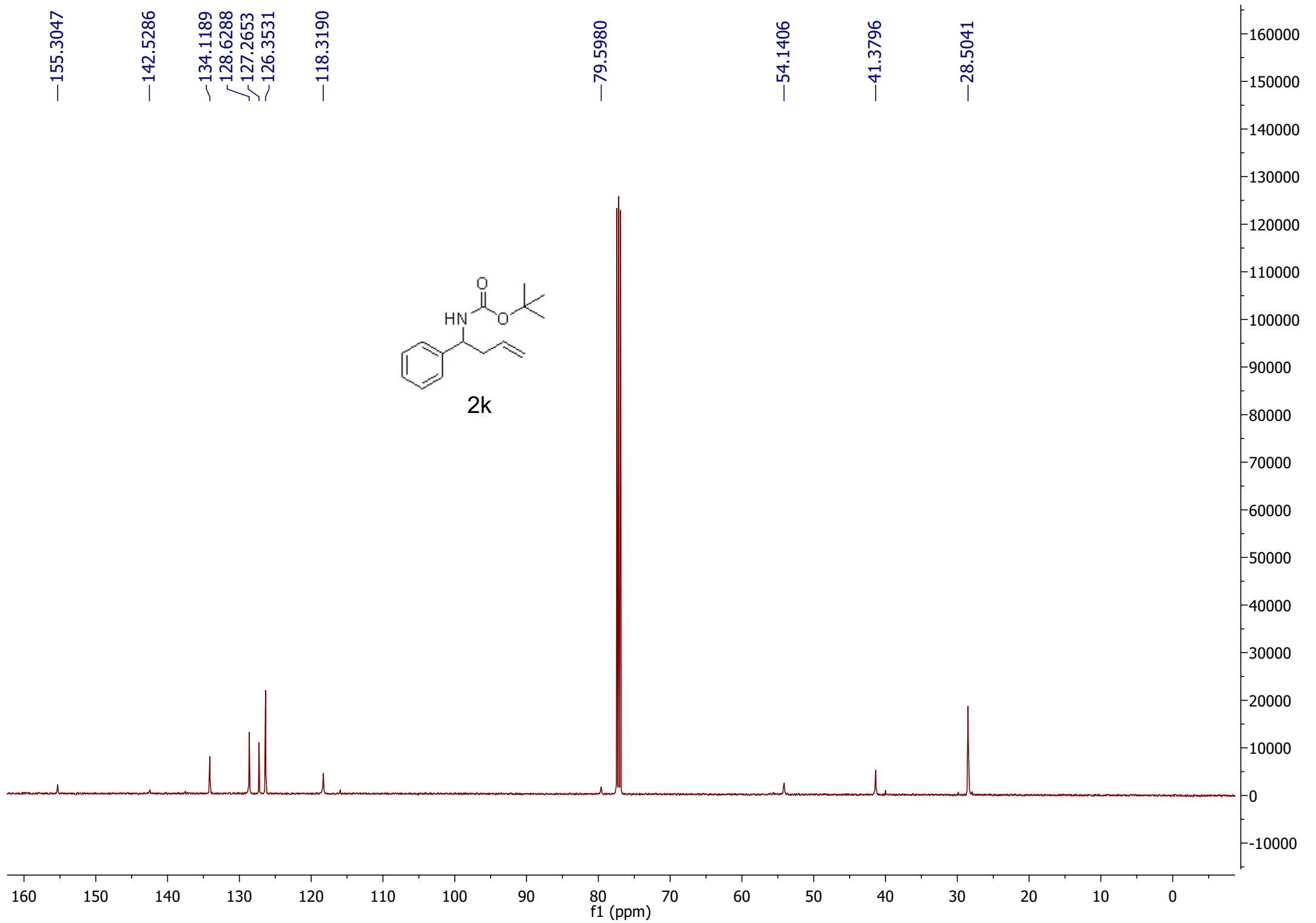


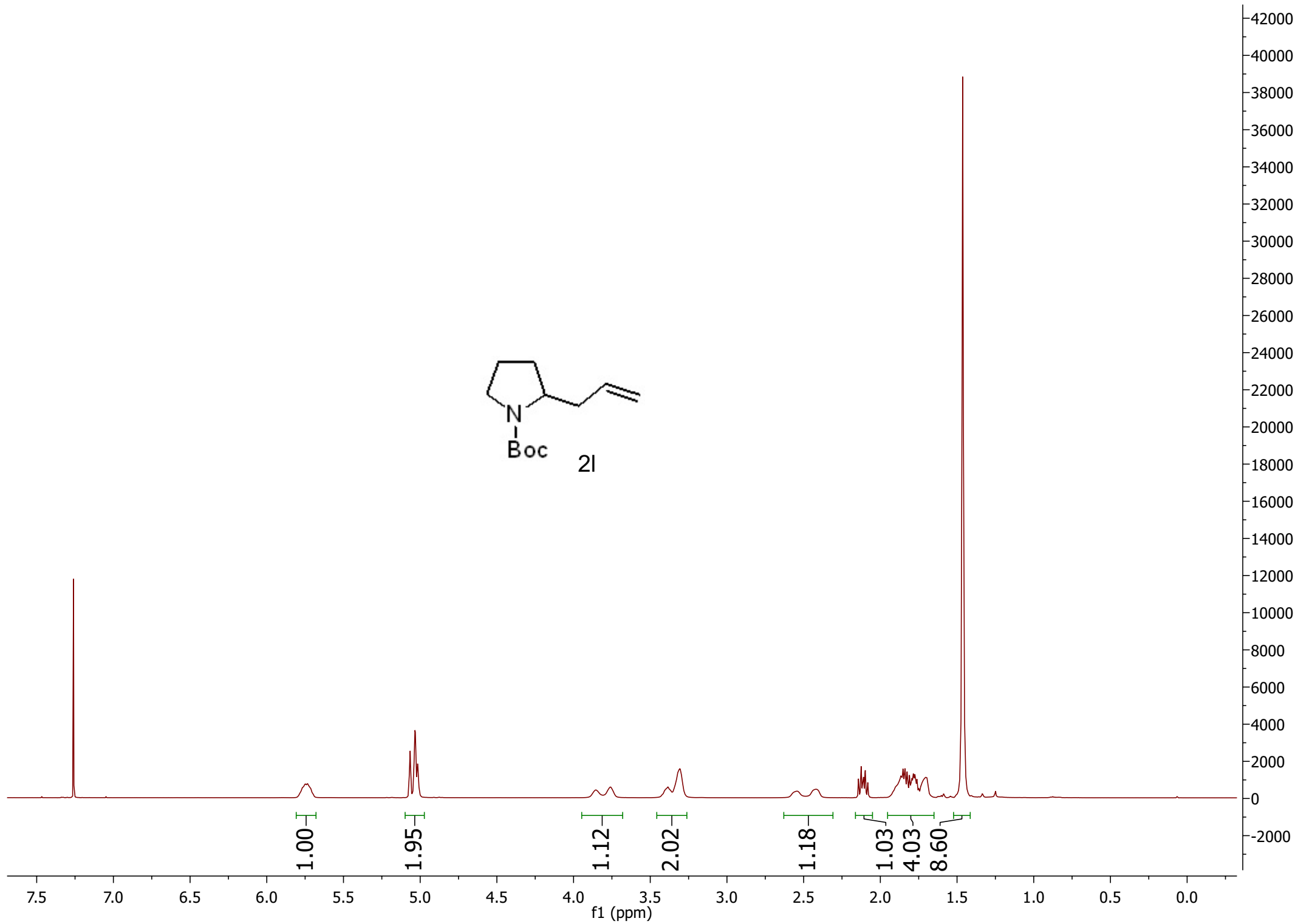
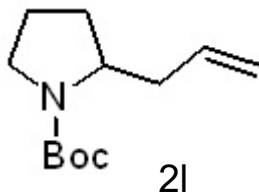


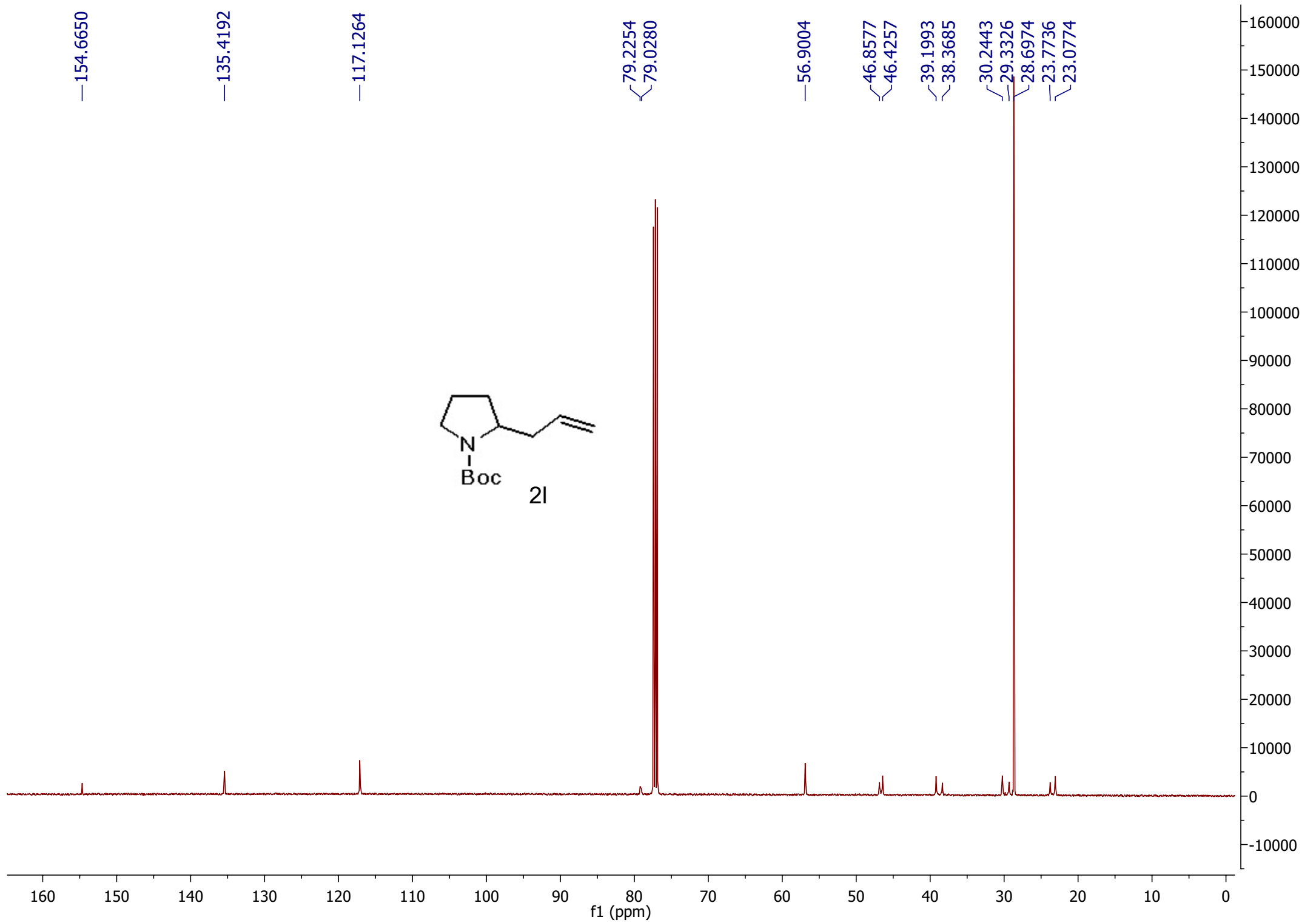


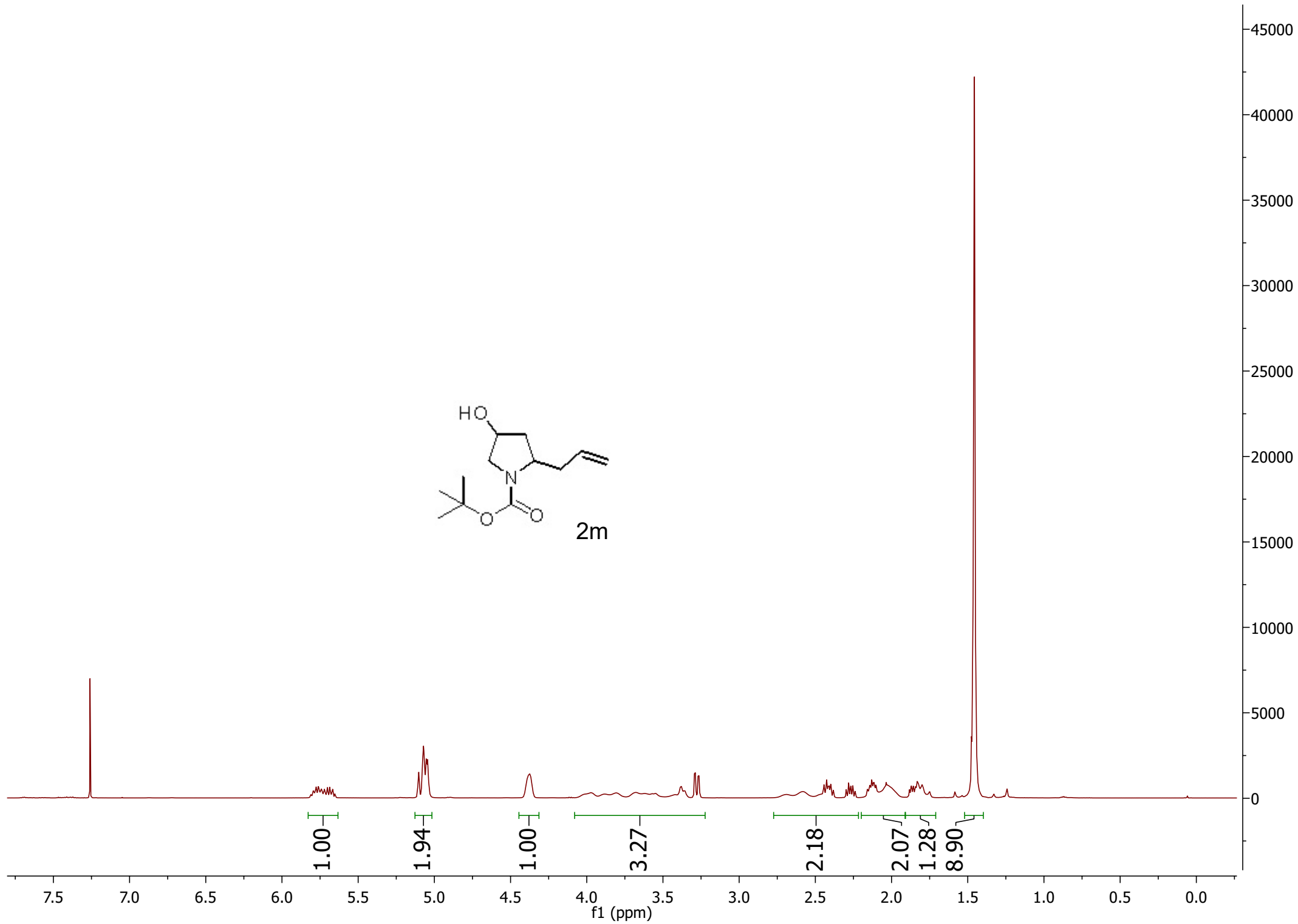


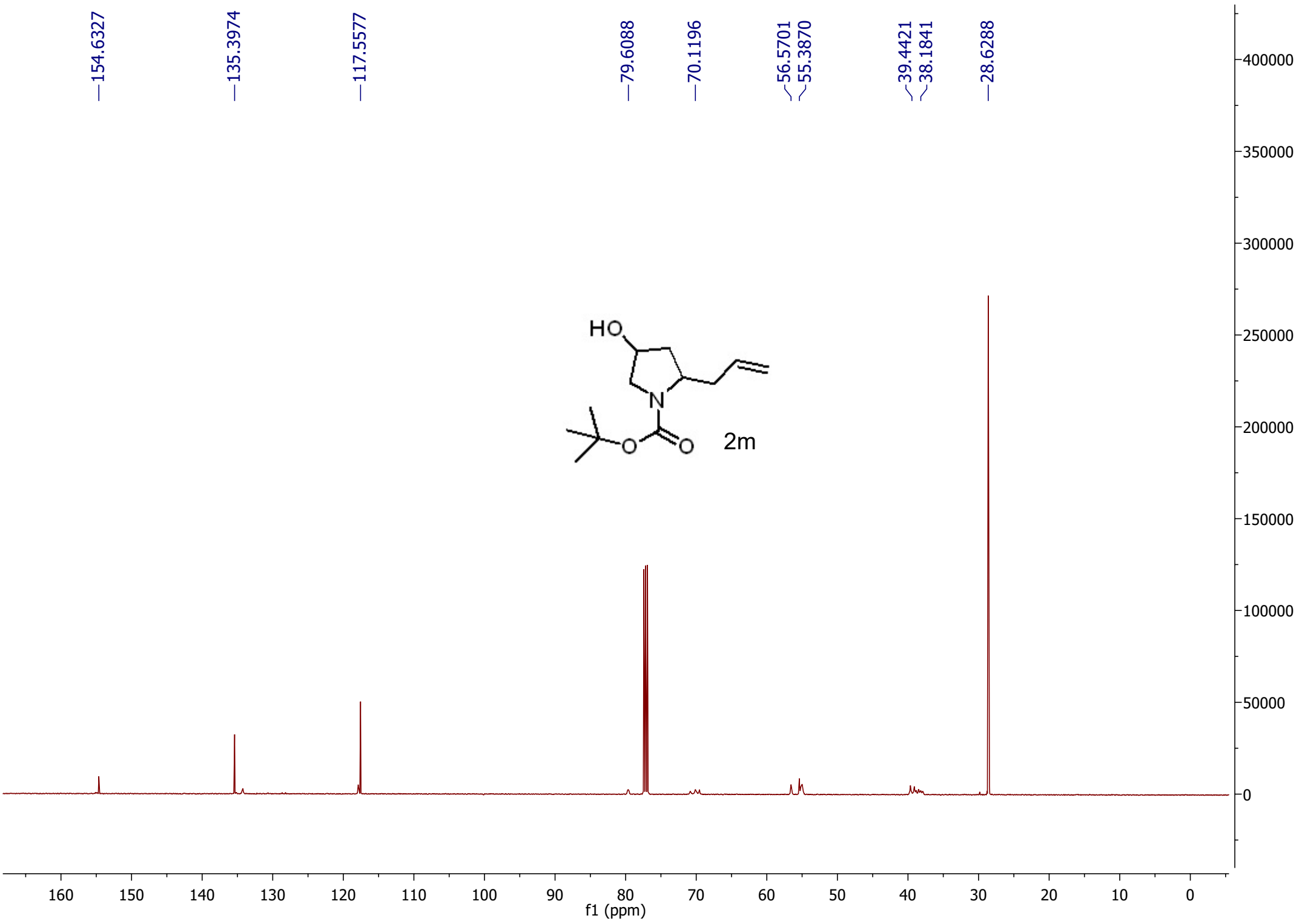
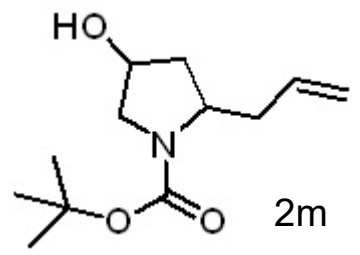


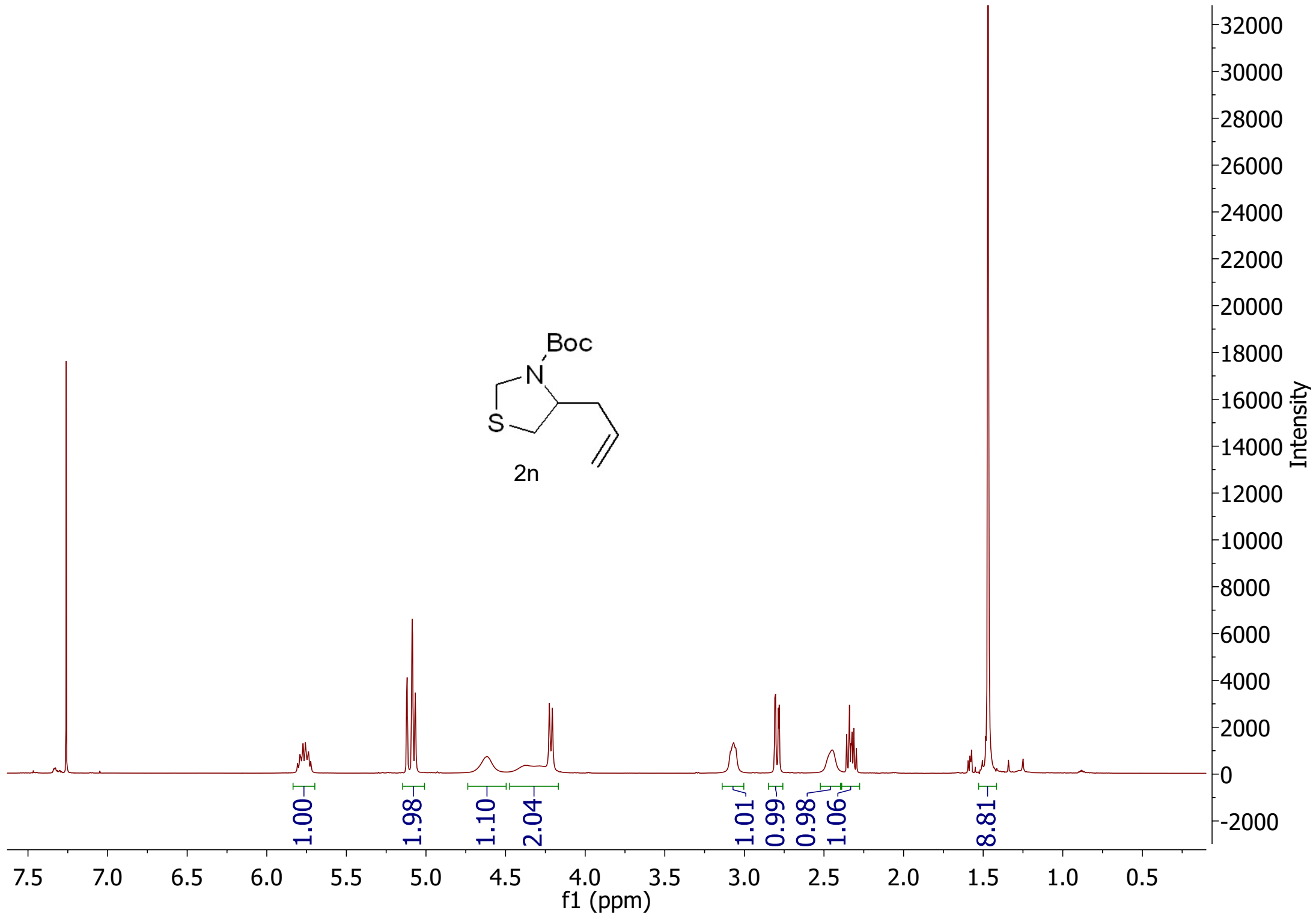


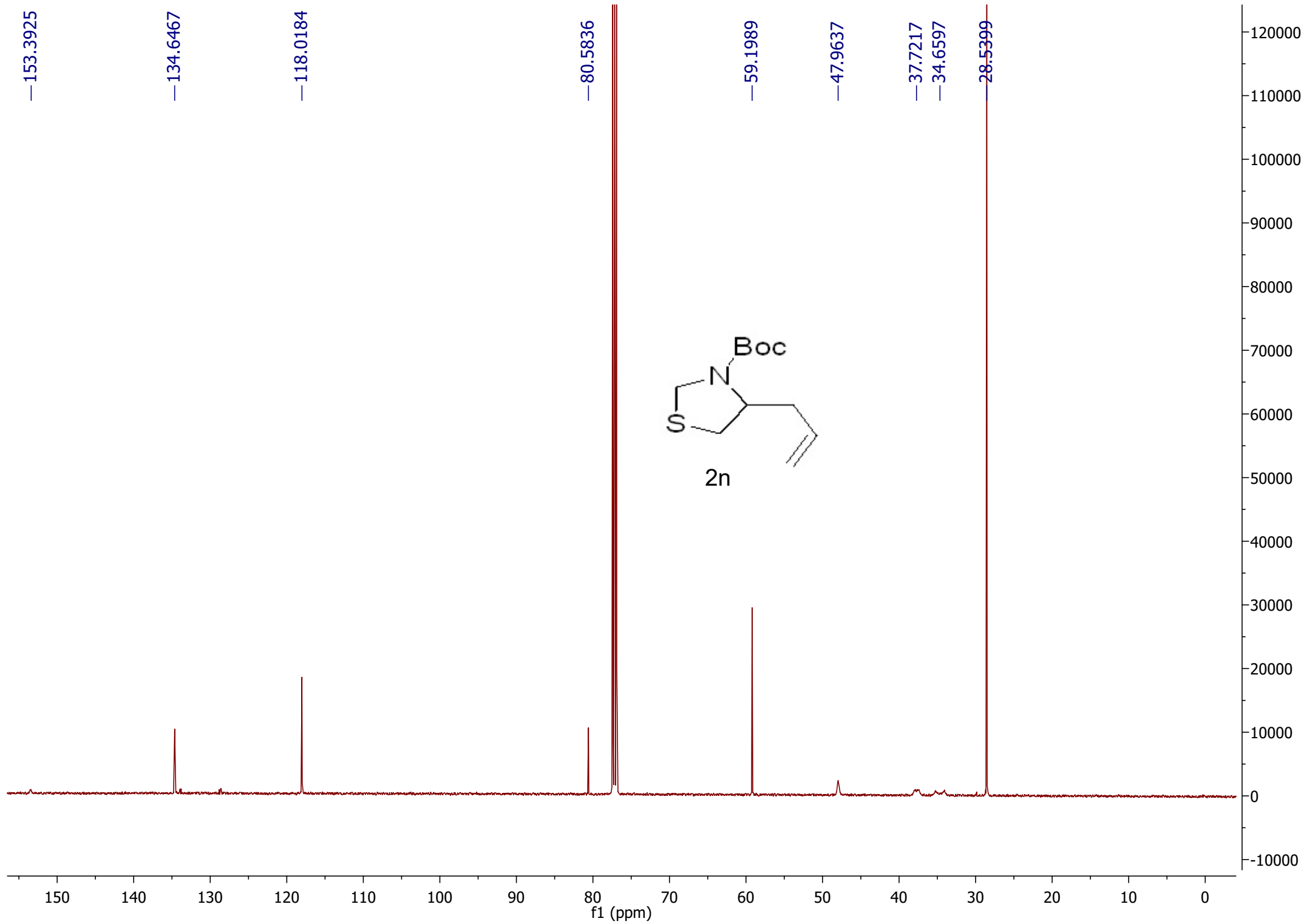


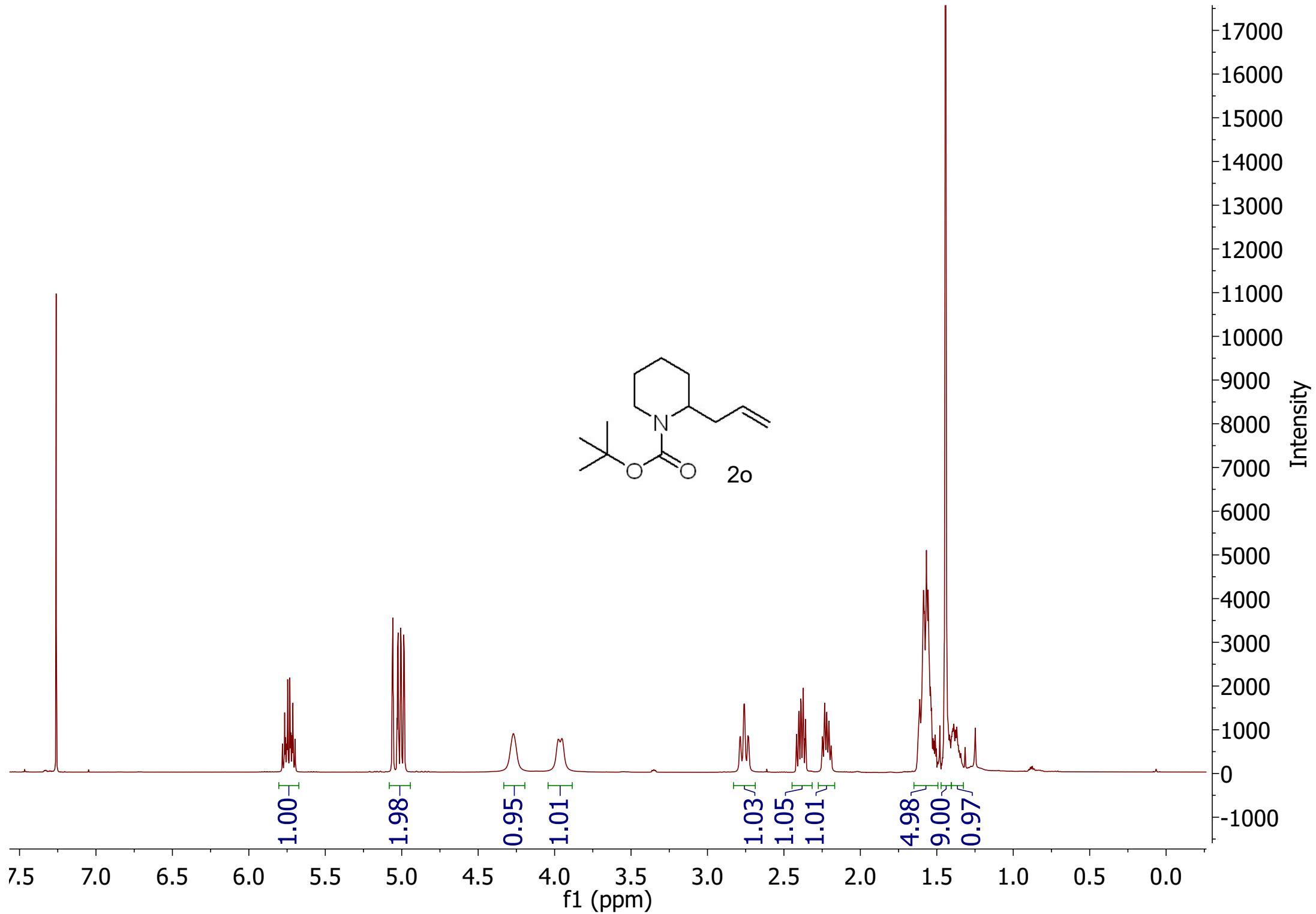


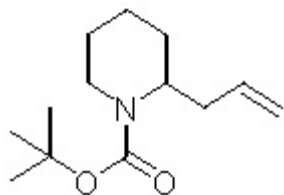




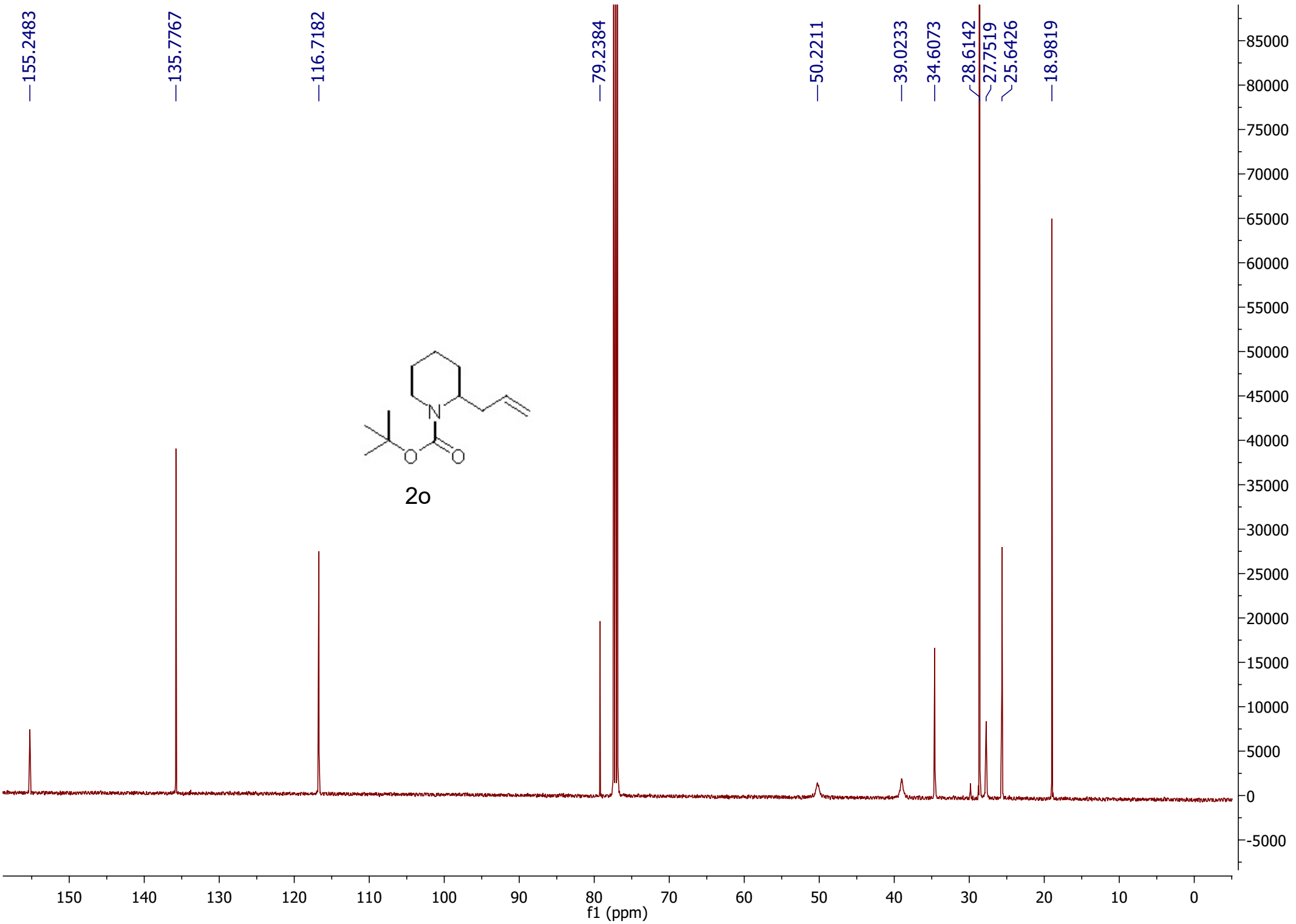


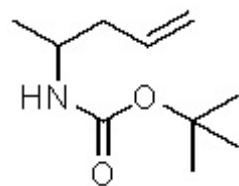




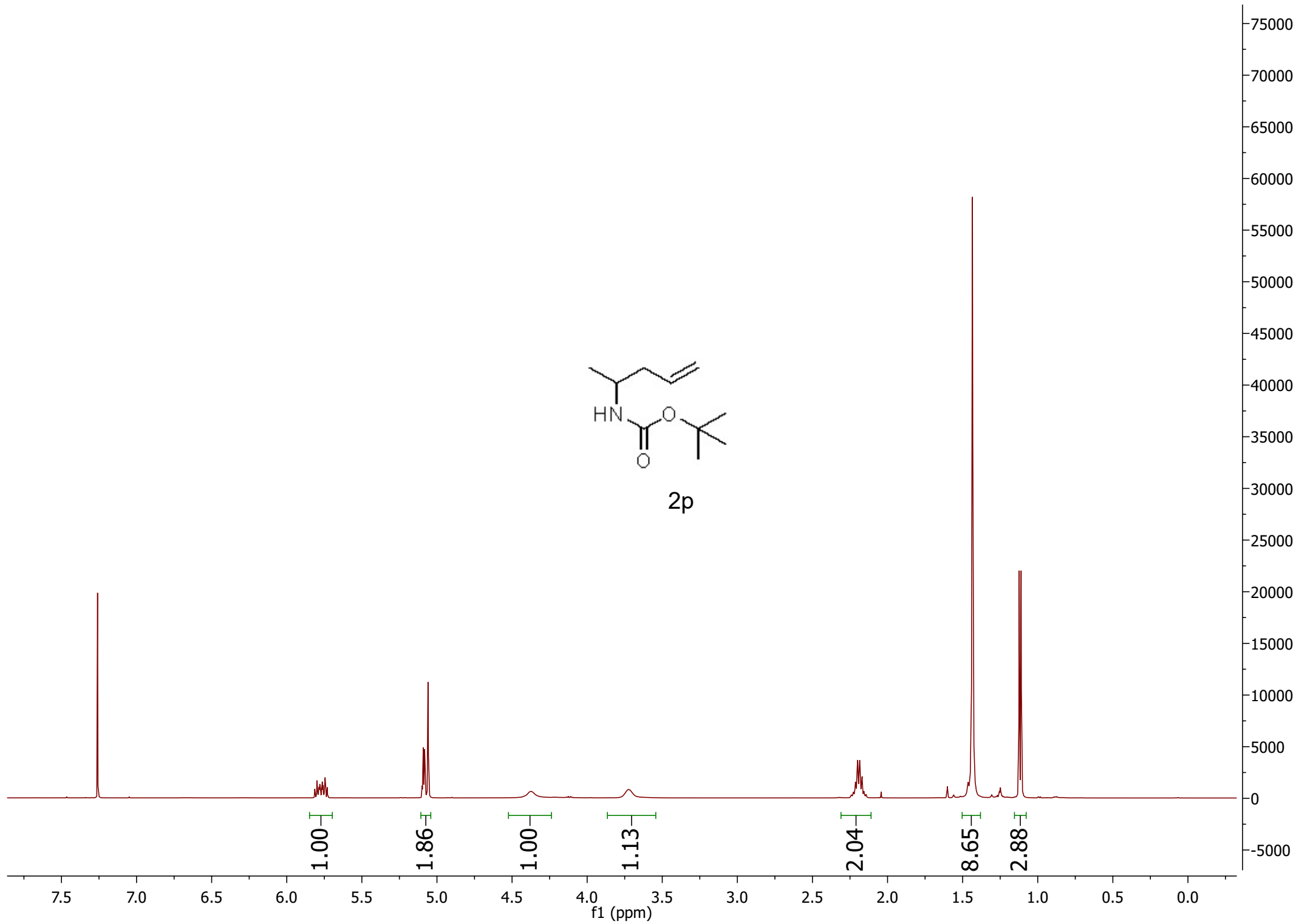


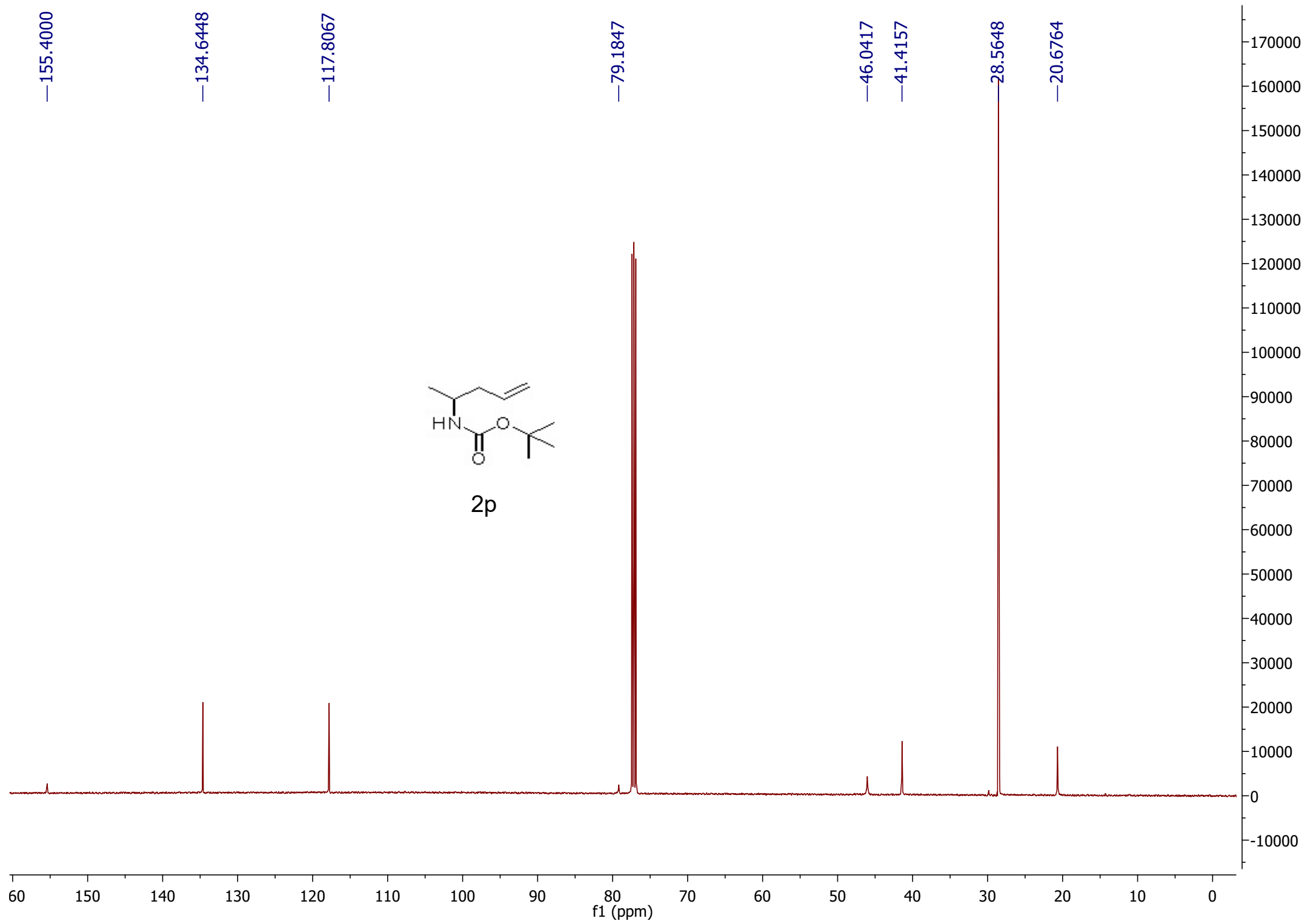
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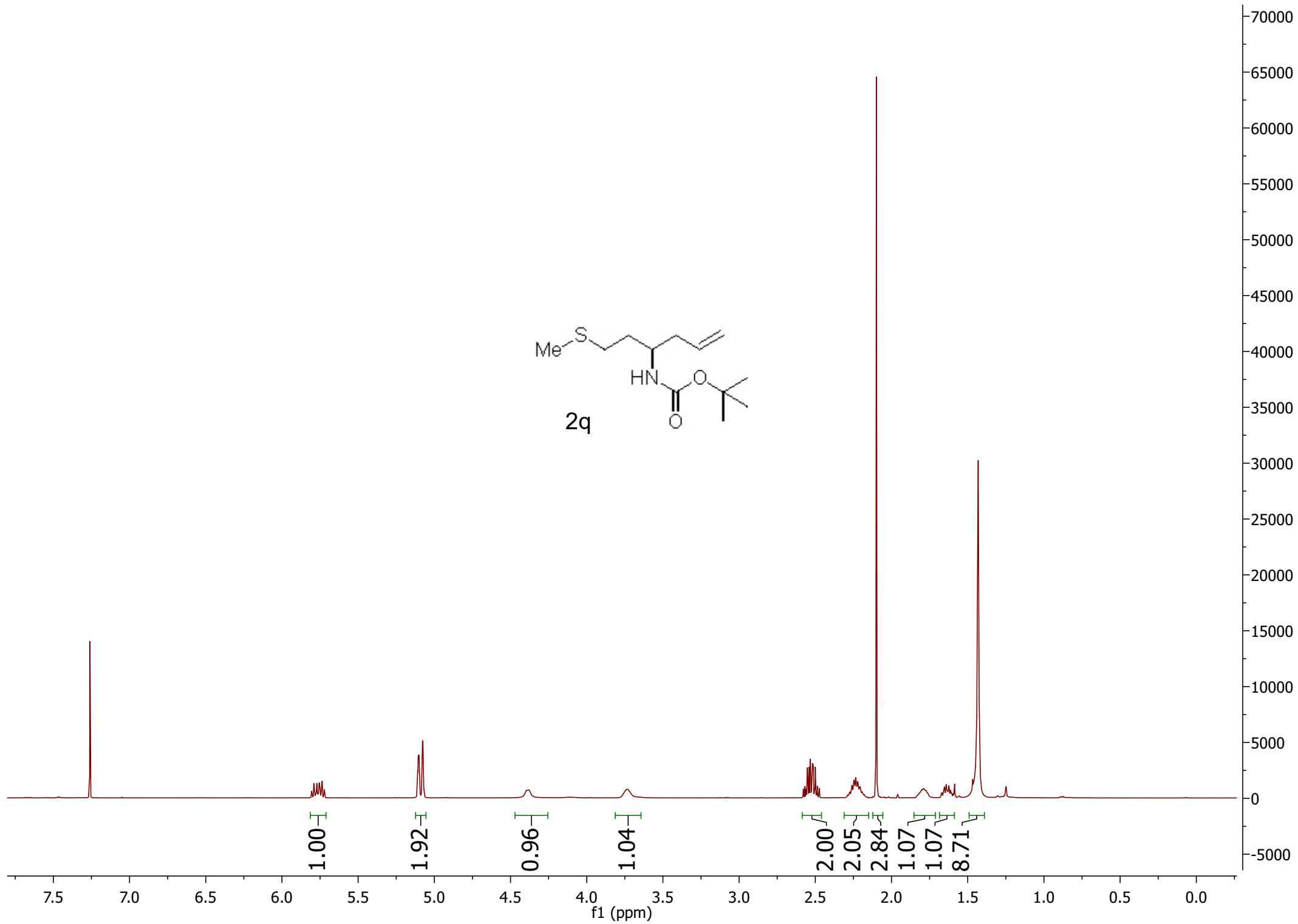


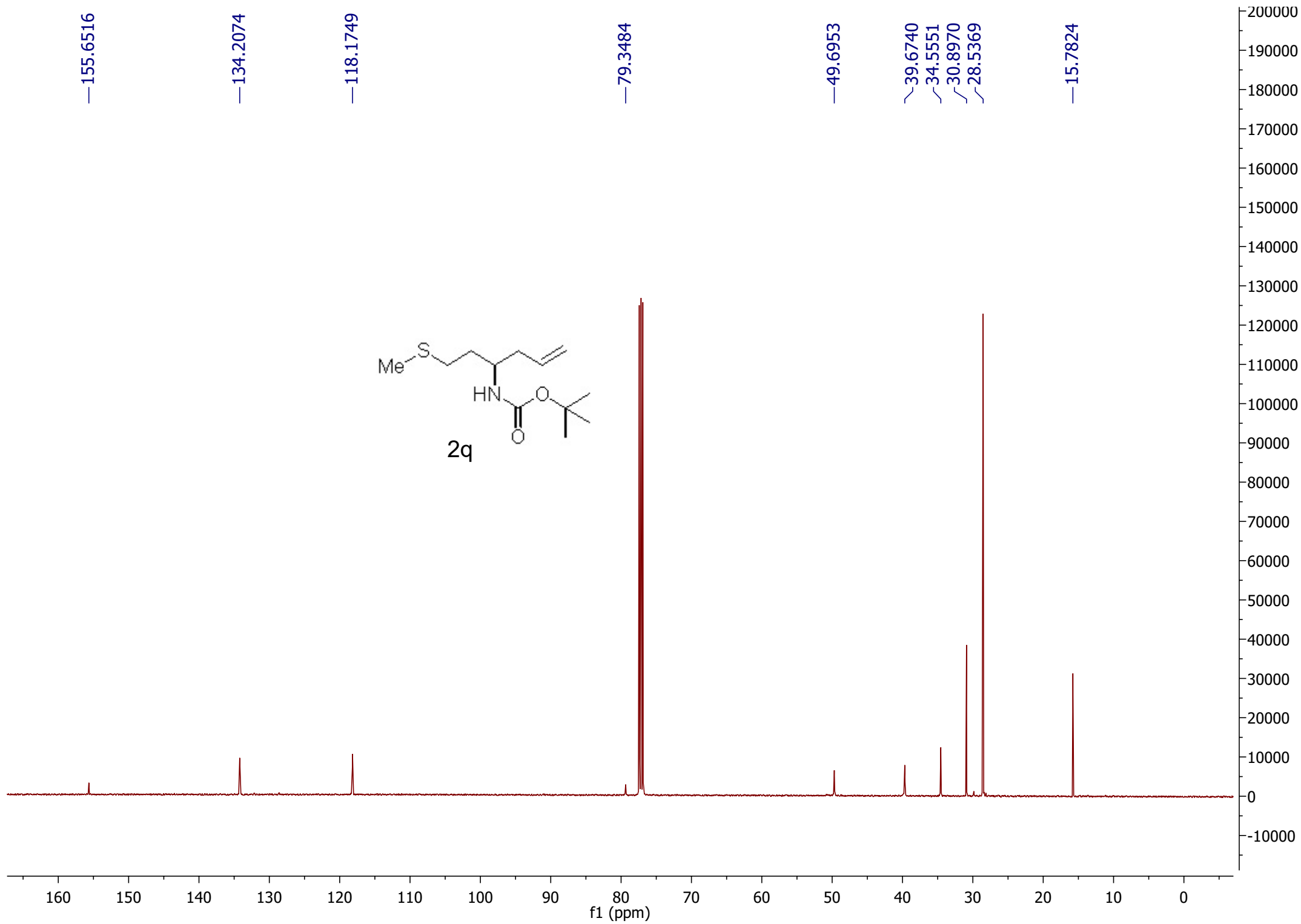


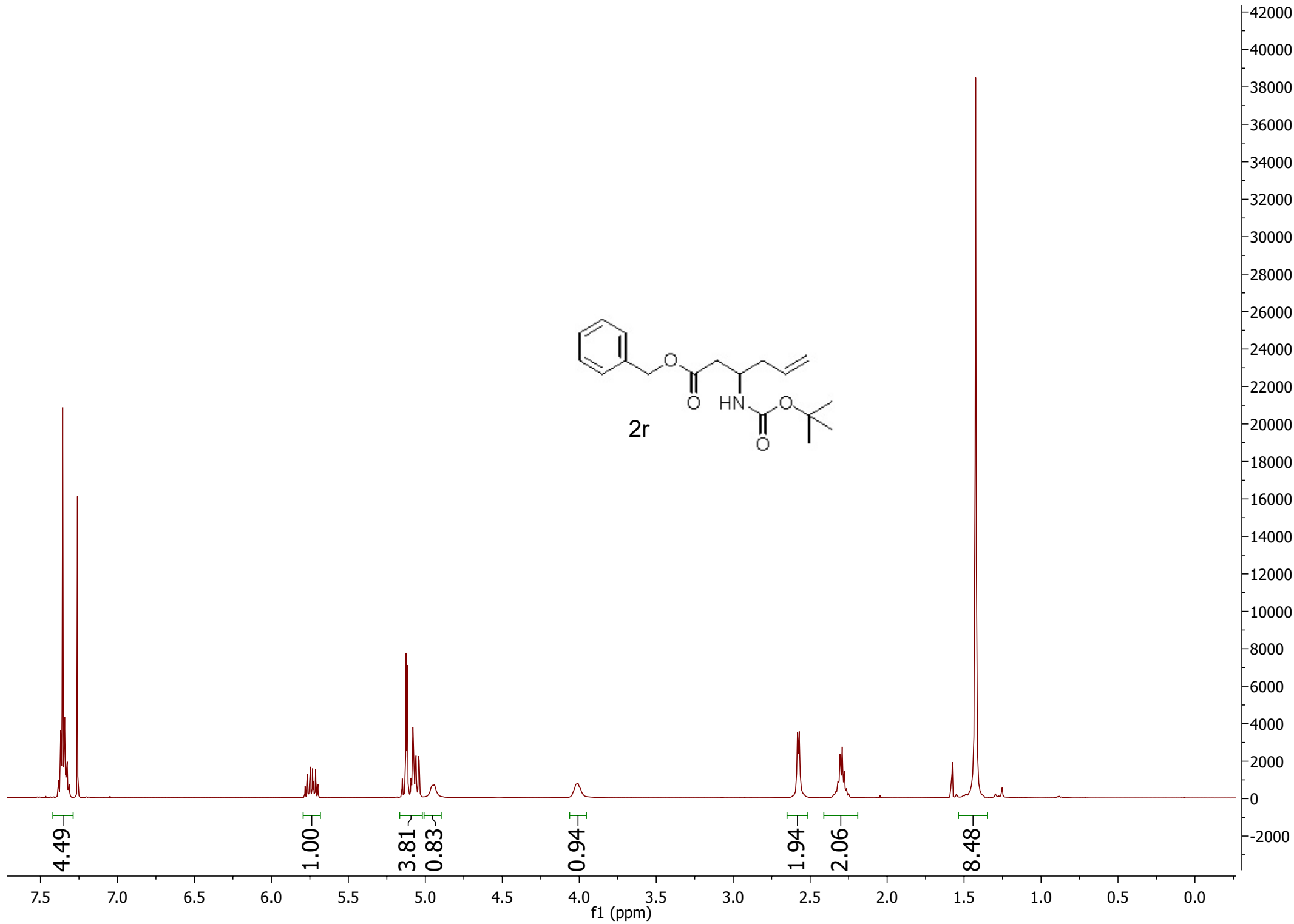
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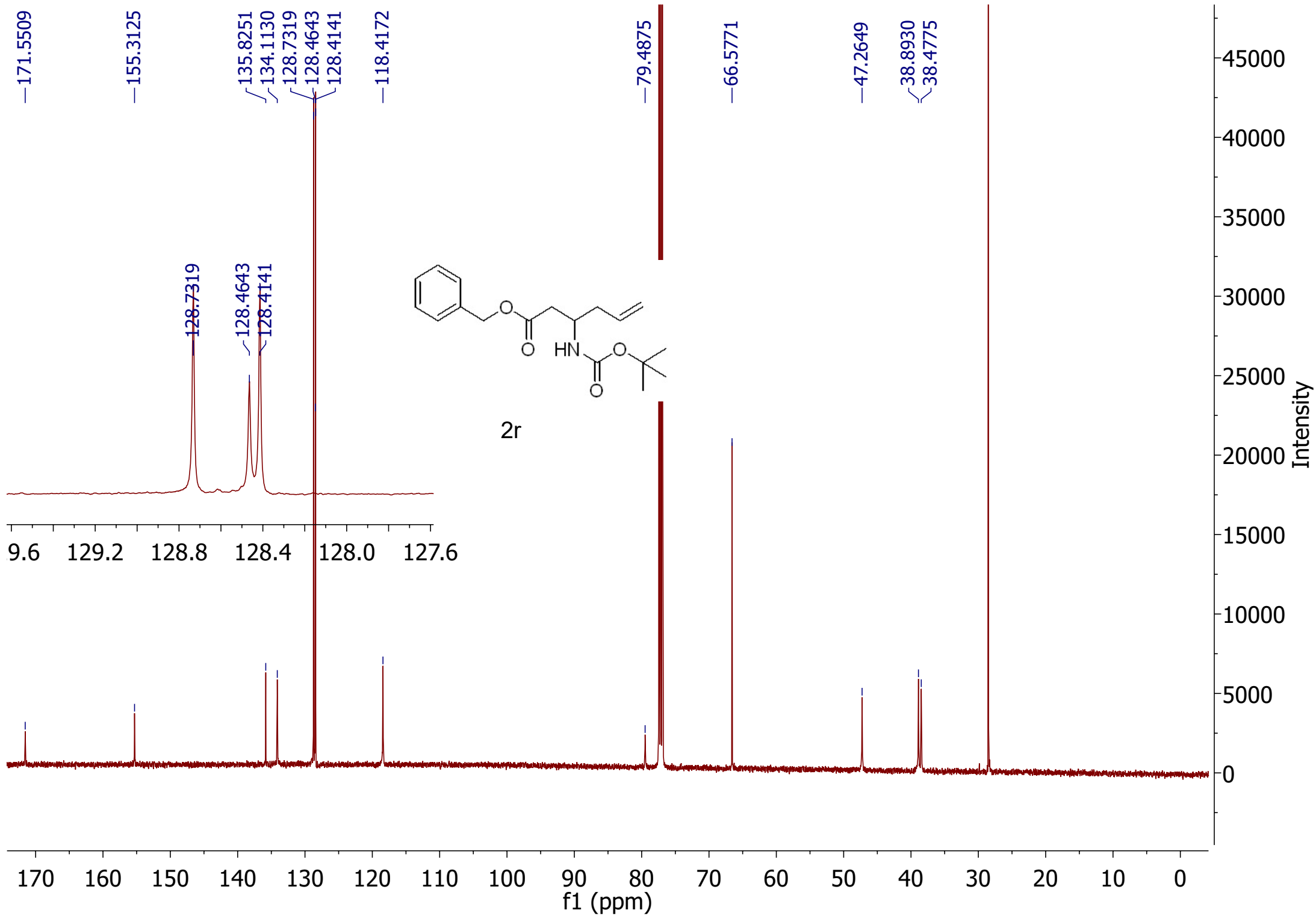


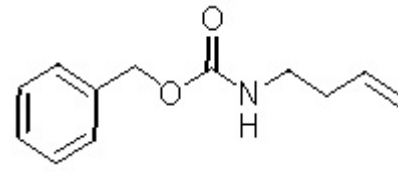




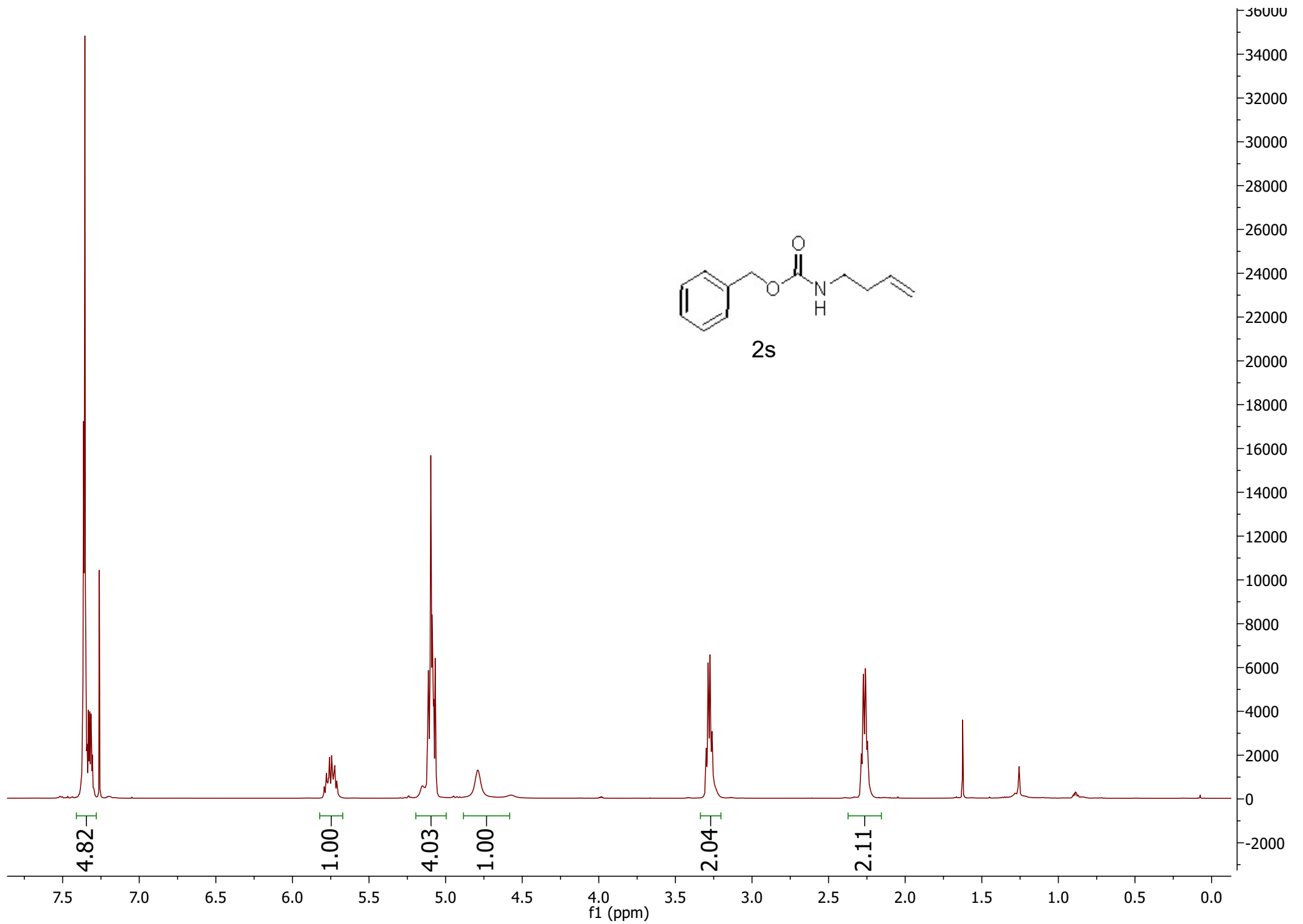


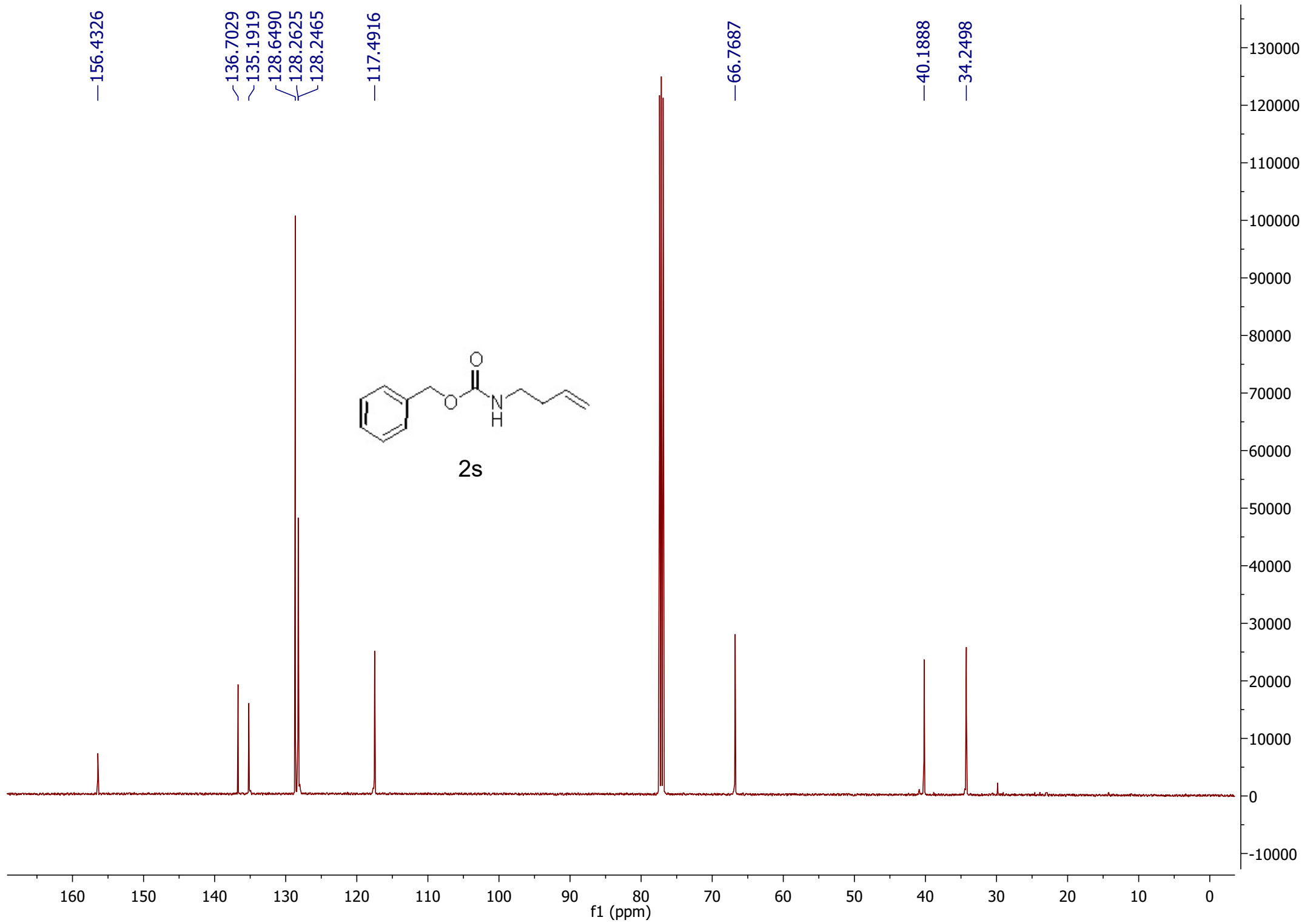


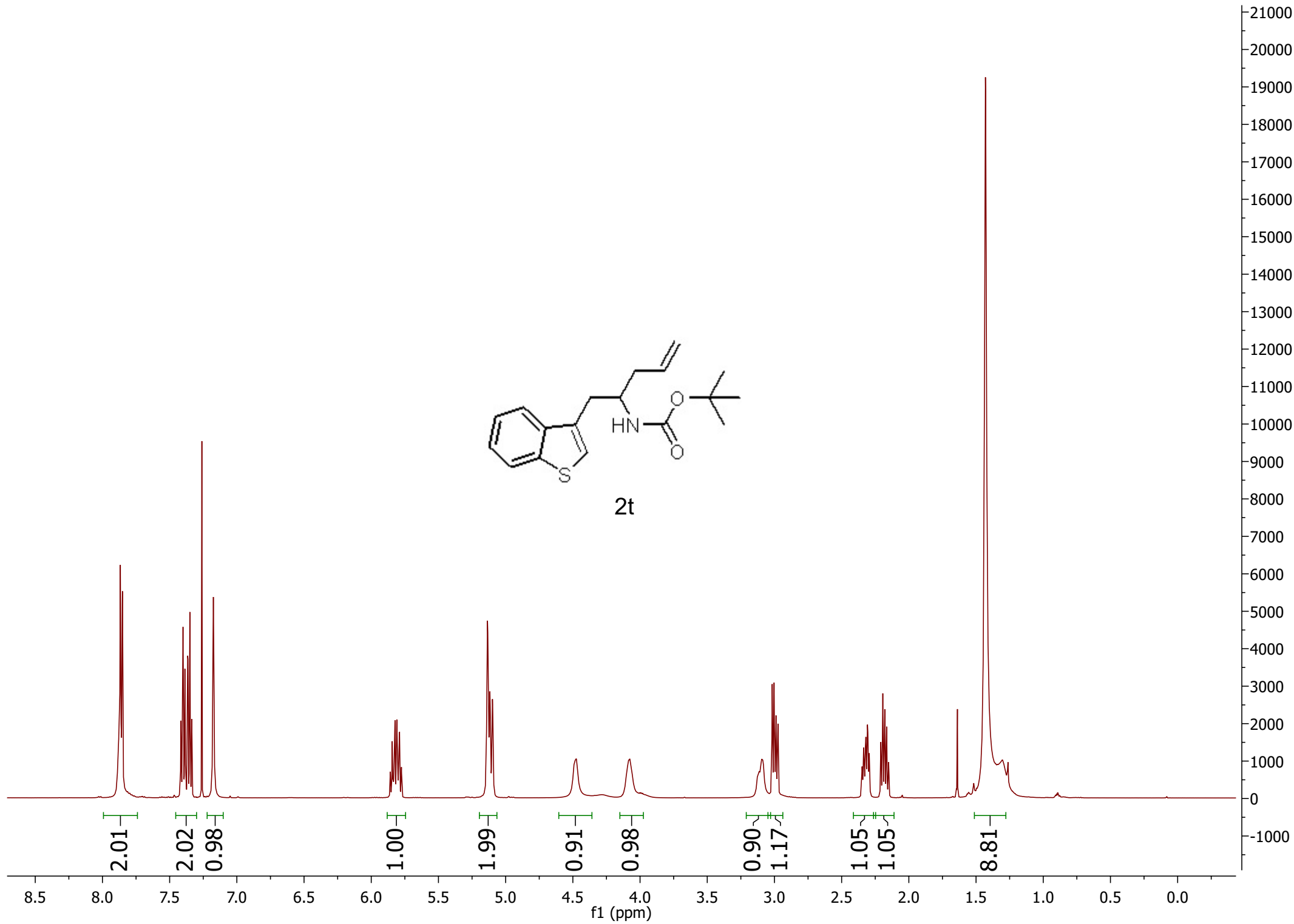


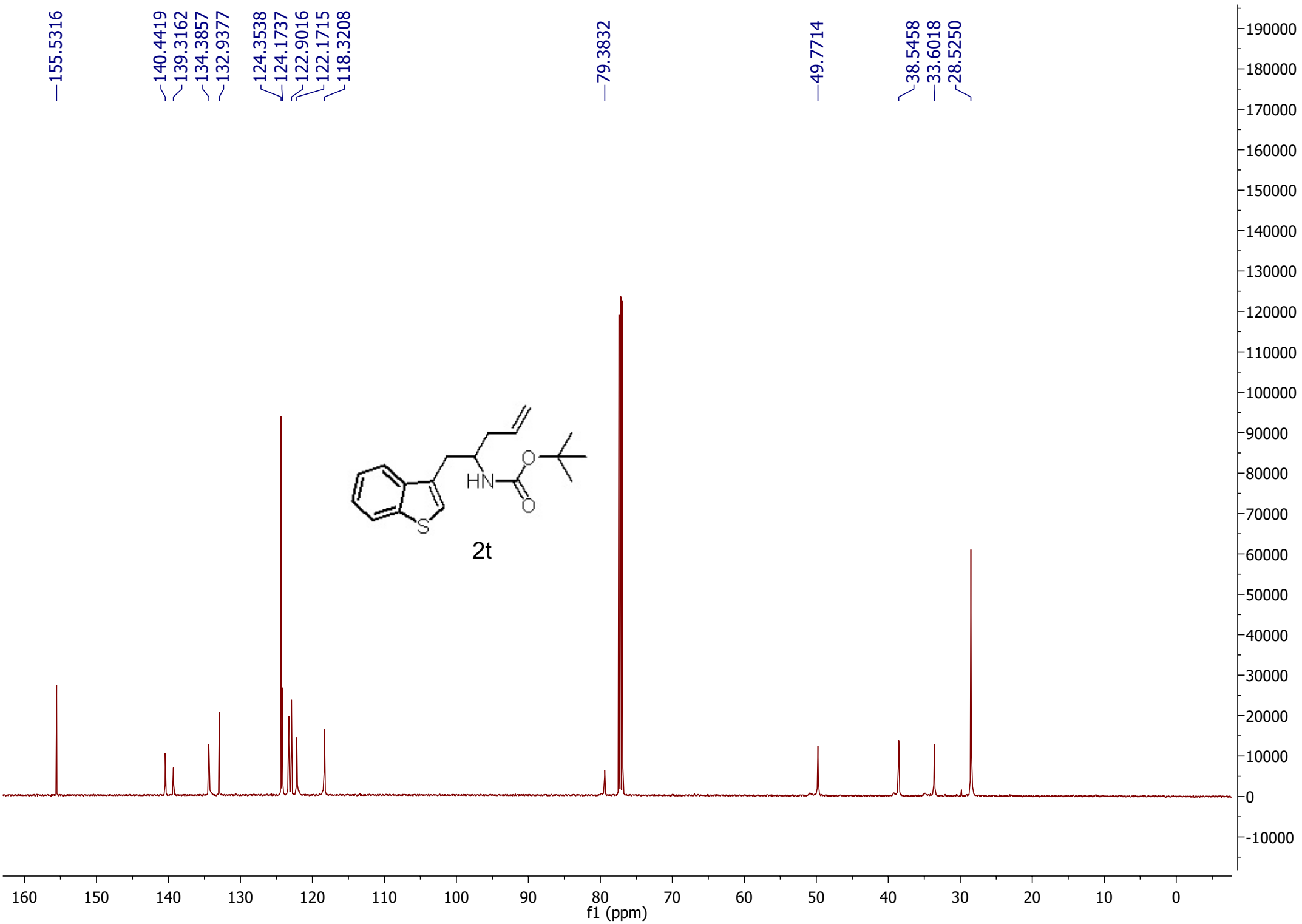


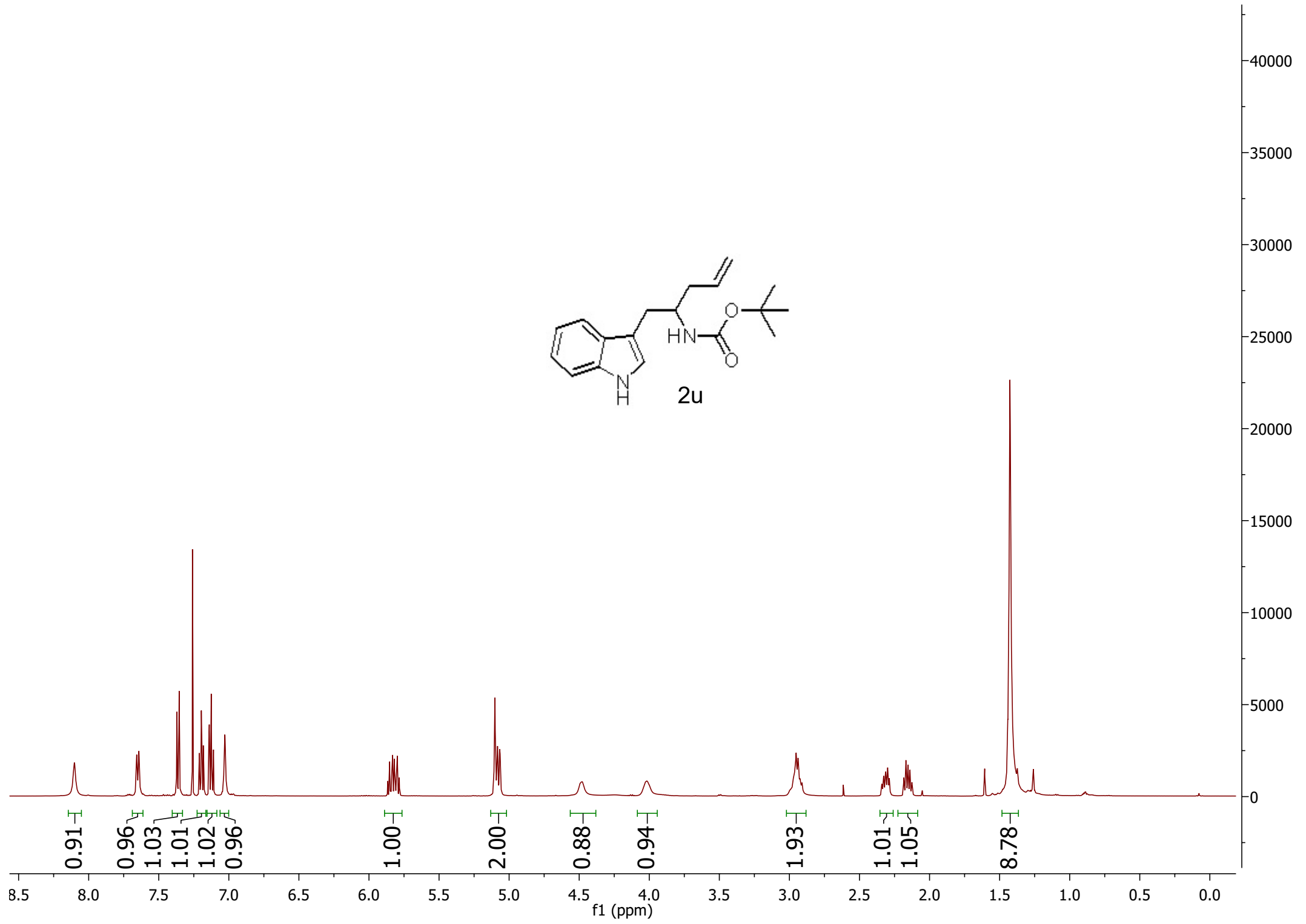
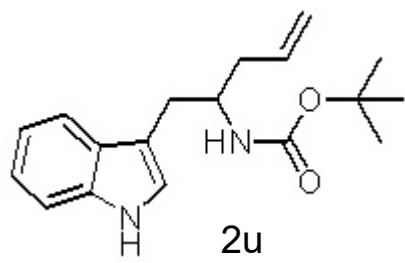
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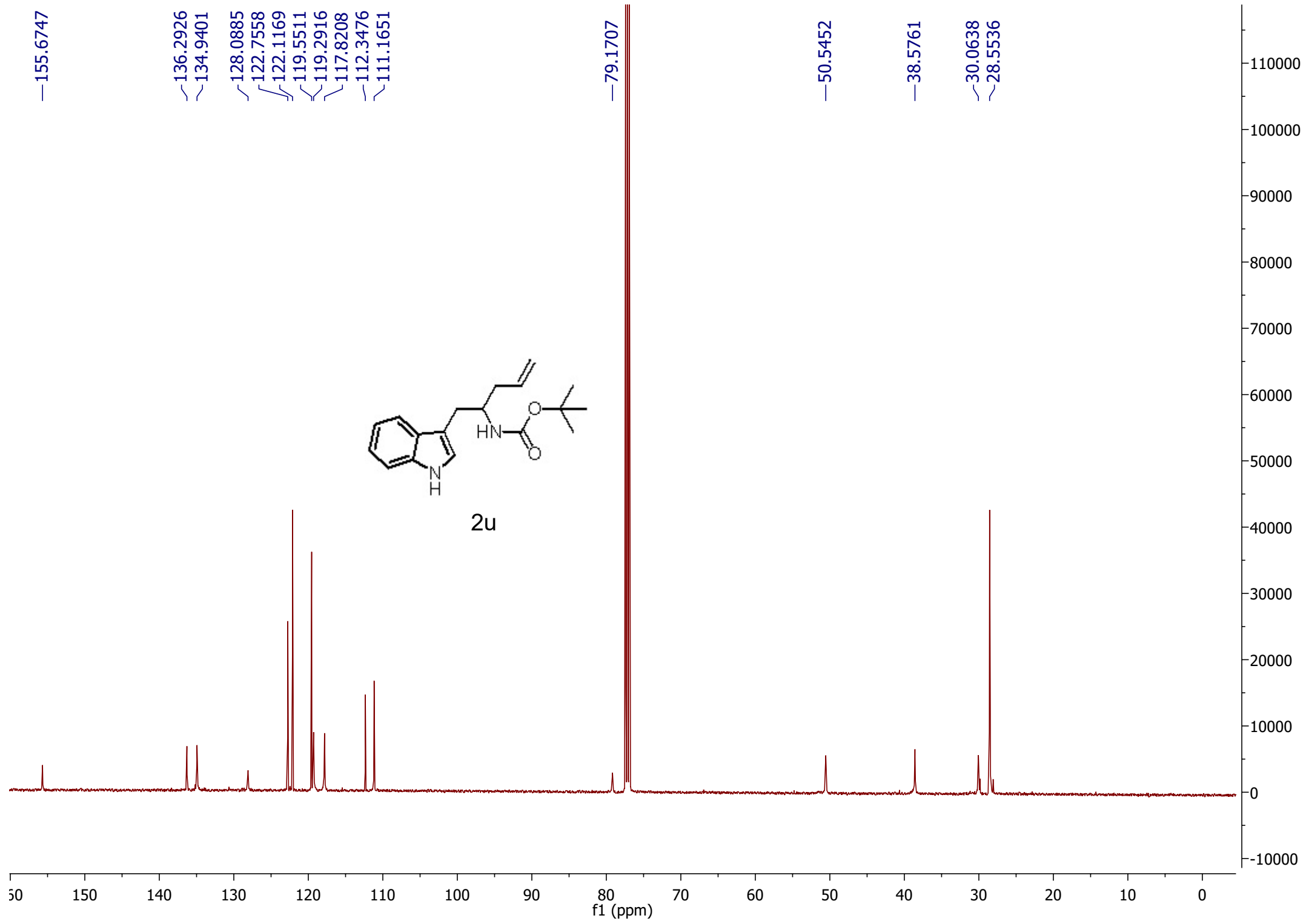


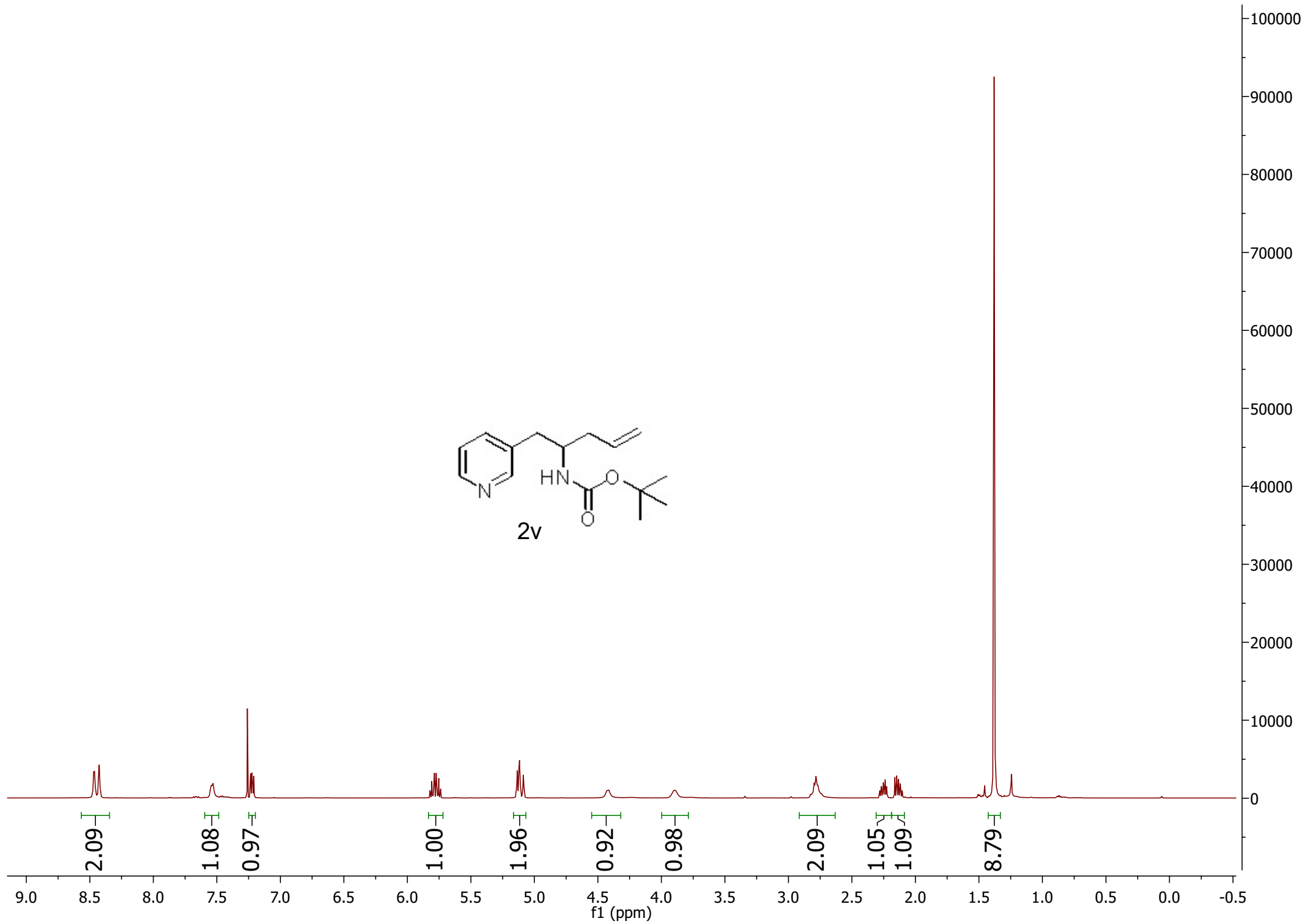
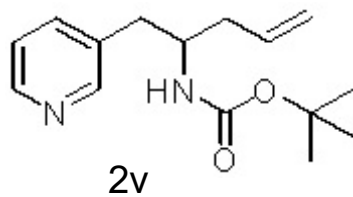


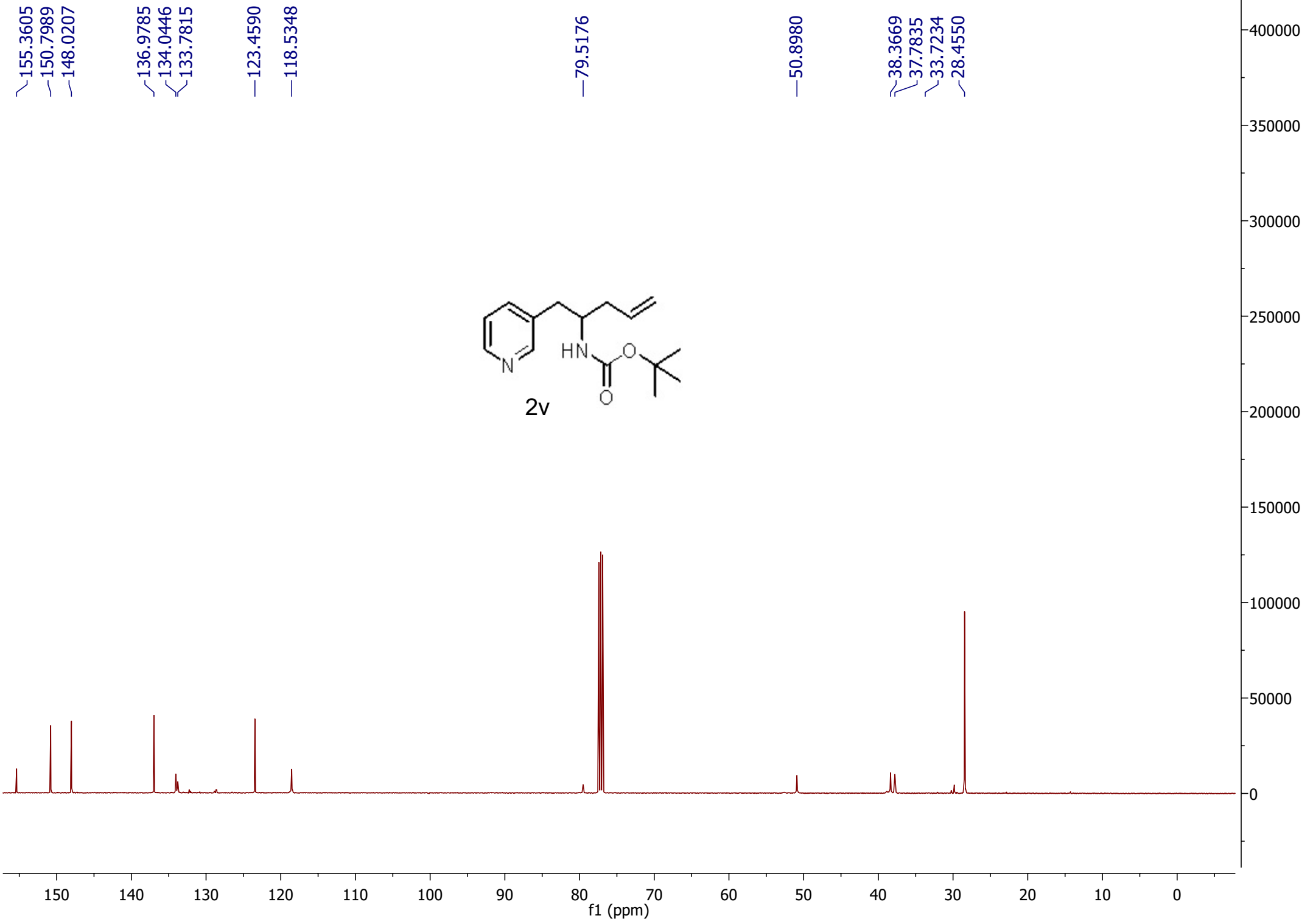


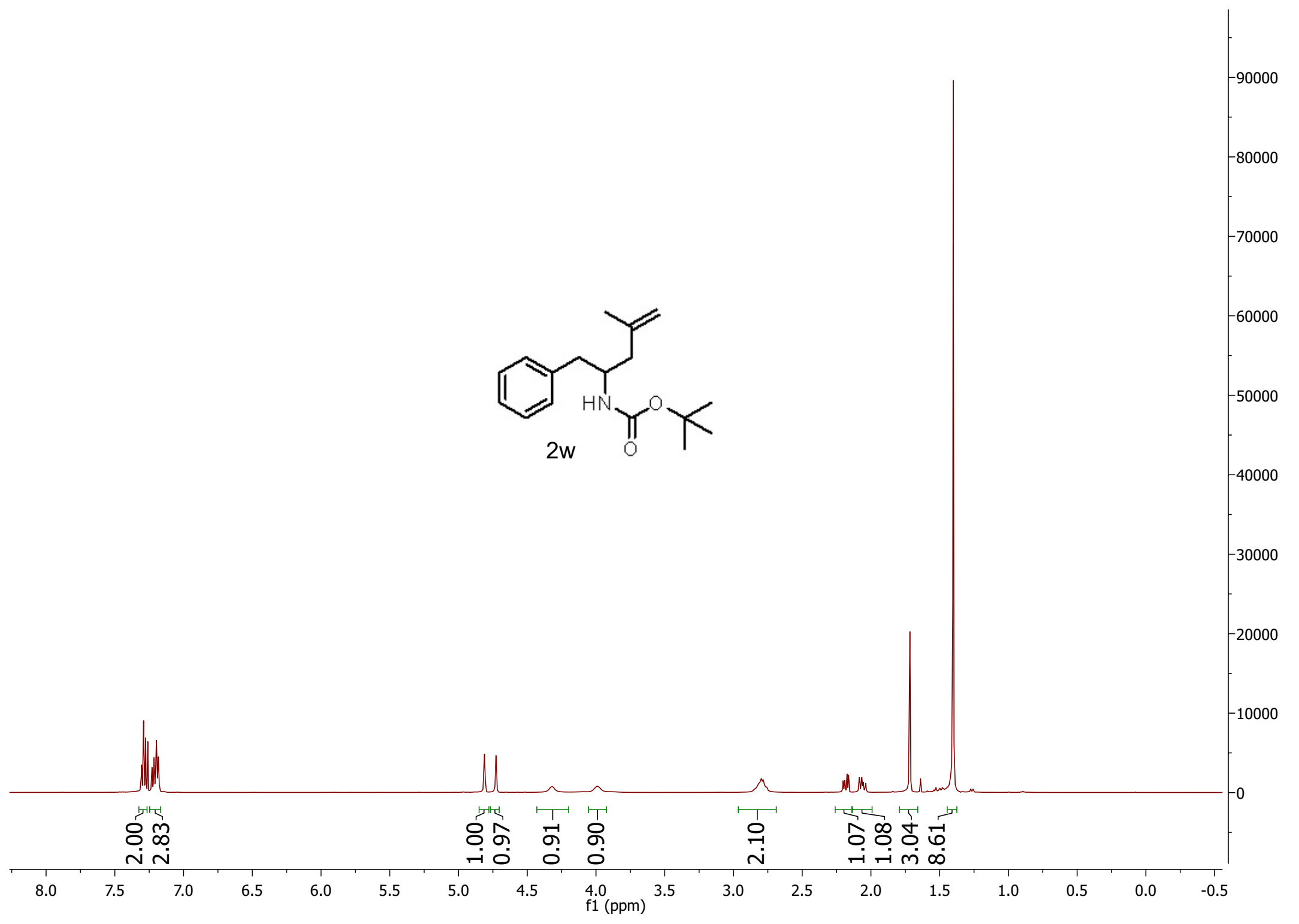
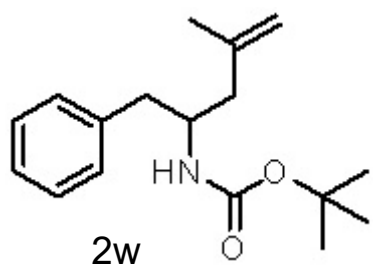


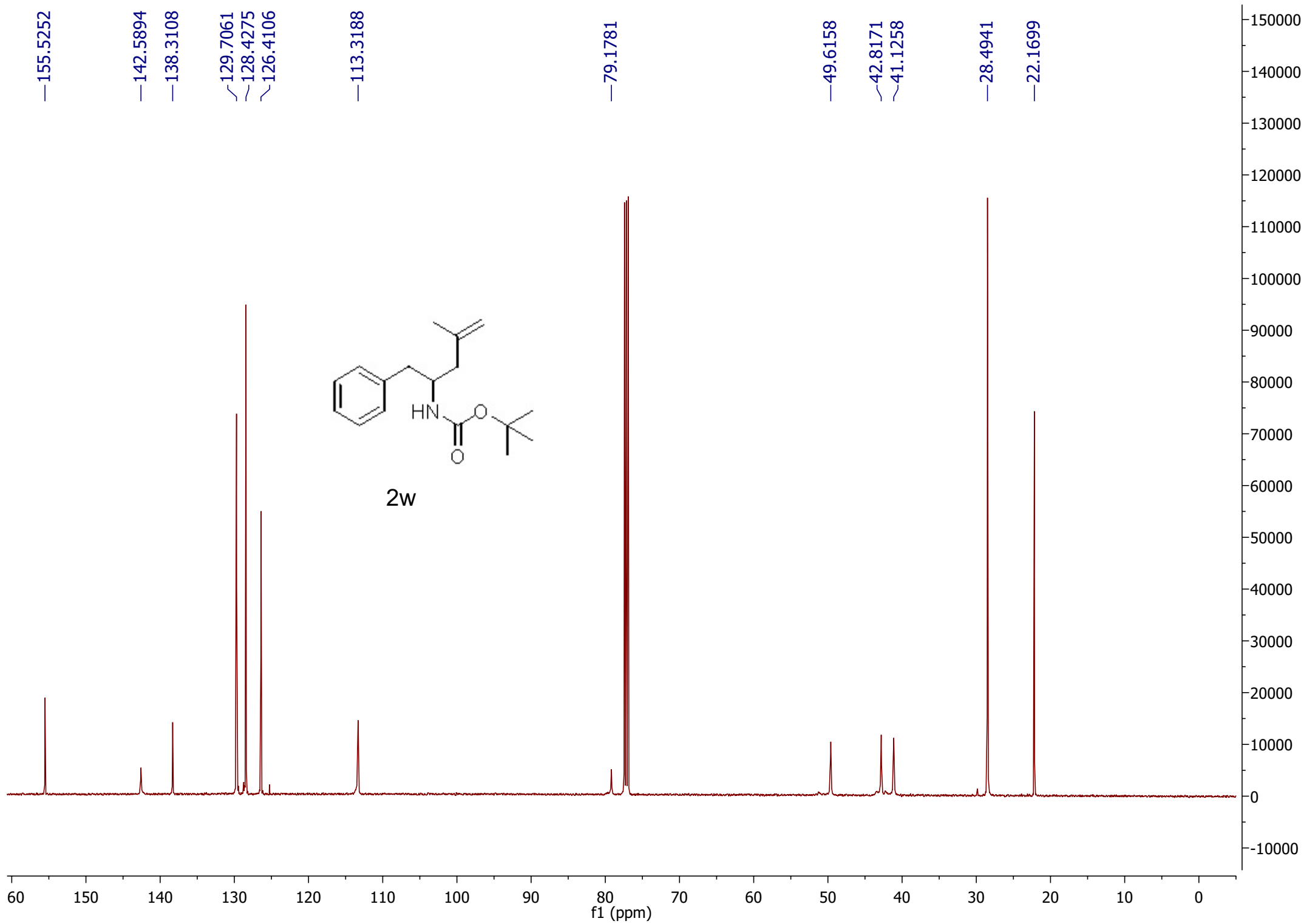


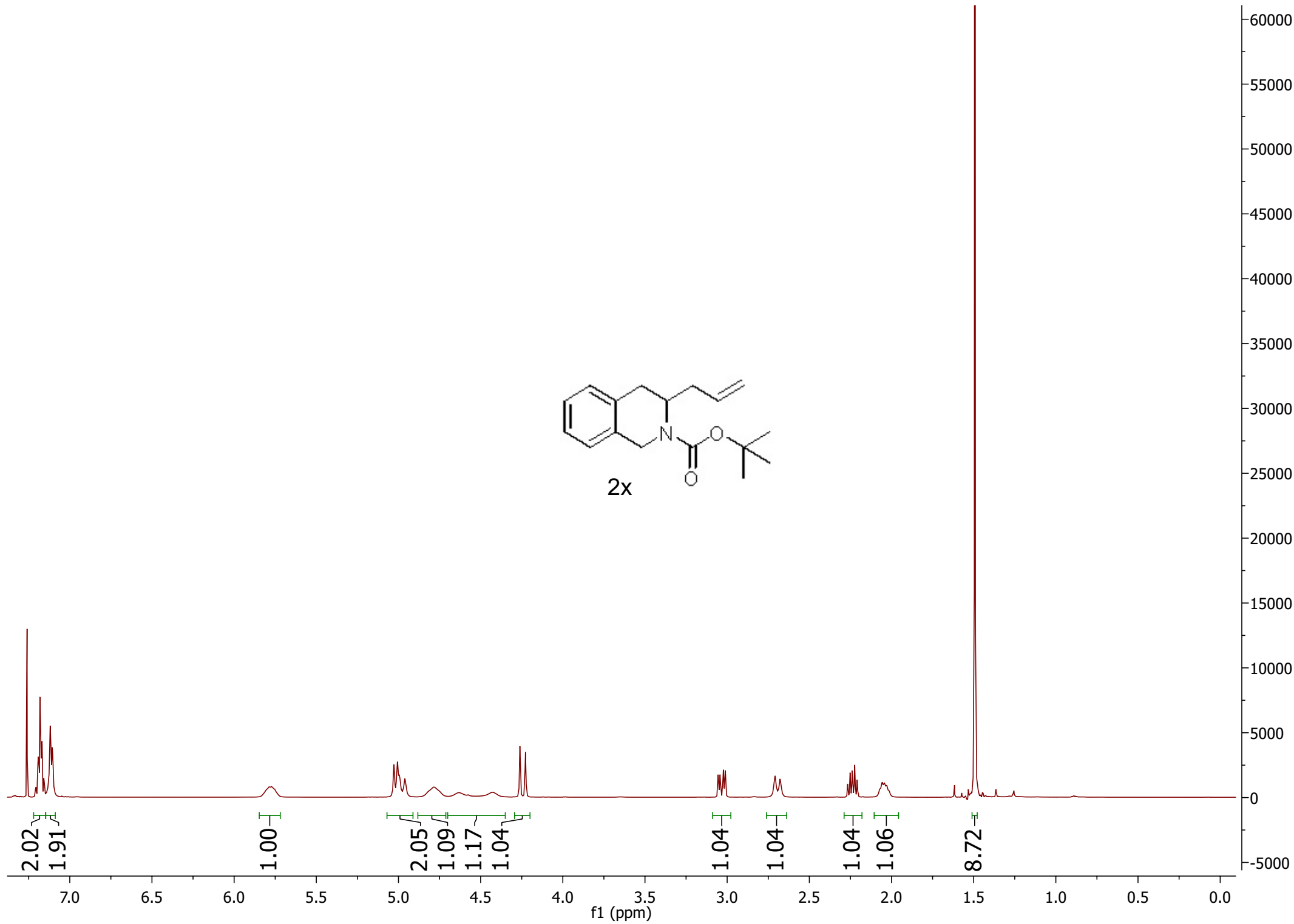


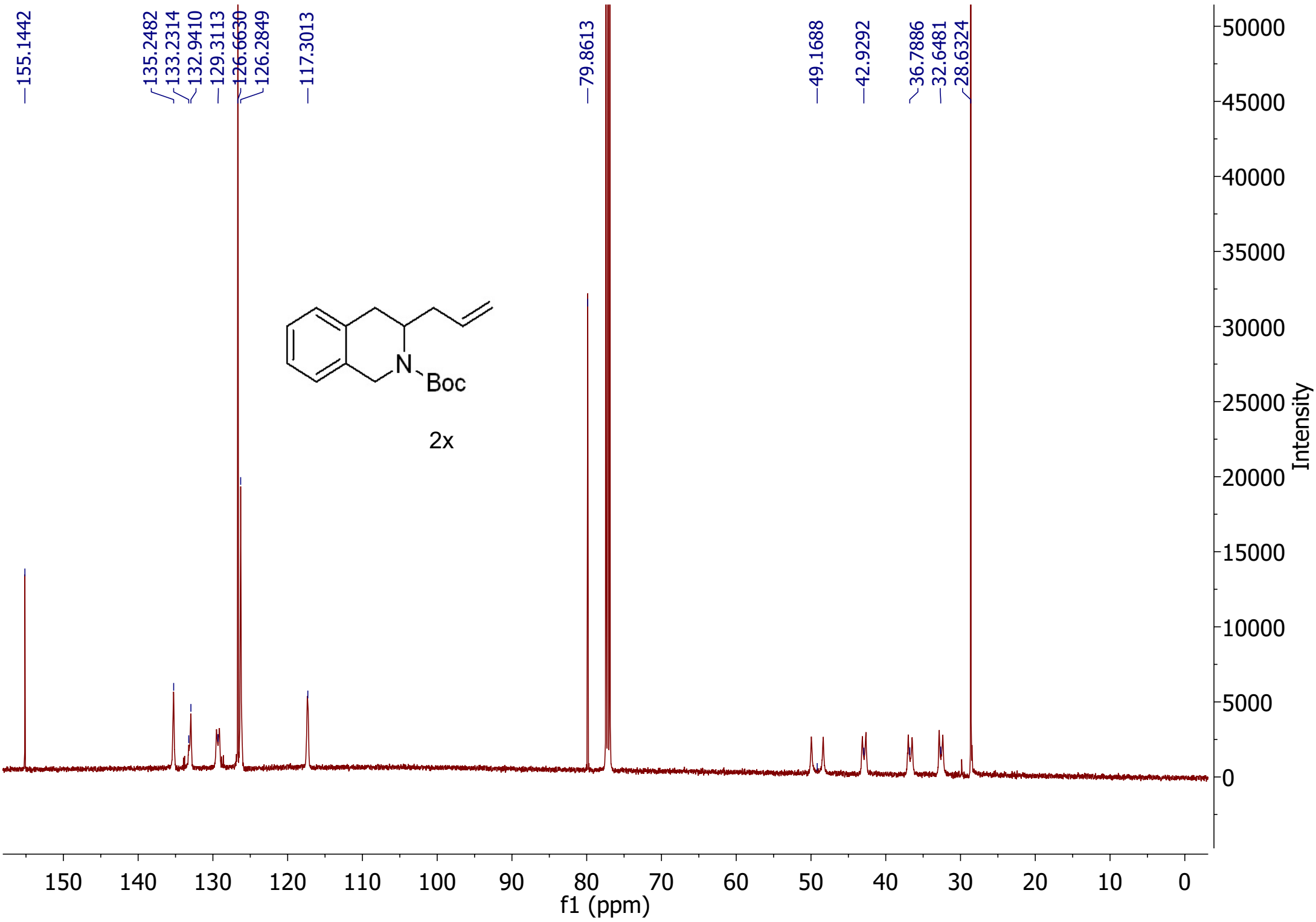


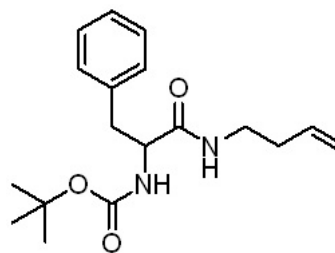












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