

The American Journal of Human Genetics

Supplemental Data

**Invariant TAD Boundaries Constrain Cell-Type-Specific  
Looping Interactions between Promoters  
and Distal Elements around the *CFTR* Locus**

Emily M. Smith, Bryan R. Lajoie, Gaurav Jain, and Job Dekker

**Table S1: 5C probe set.** This table contains all the probes used in the 5C experiment: their name, sequence, and genomic location.

**Table S2: Mapping Statistics.** This table contains all mapping statistics for the raw data of our 10 5C libraries.

**Table S3: 5C library read depth.** This table summarizes the 5C read depth for each replicate, indicating the number of reads in the raw data, cis-purged data, singleton- removed data, and final coverage corrected data.

	Raw Data		<i>cis</i> -purged Data		Singleton-removed Data		Coverage-corrected Data	
	#interactions	#reads	#interactions	#reads	#interactions	#reads	#interactions	#reads
Caco2-R1	50,986	12,930,158	24,492	9,240,611	24,449	7,265,532	20,351	7,354,017
Caco2-R2	50,970	12,724,492	27,090	9,783,411	27,046	9,453,039	22,278	8,248,685
Calu3-R1	66,029	18,308,618	36,812	12,073,332	36,768	11,970,277	32,886	8,222,536
Calu3-R2	148,785	25,380,481	39,883	15,507,757	39,908	15,378,124	36,733	8,522,545
Capan1-R1	126,462	18,101,240	38,046	6,041,896	38,002	5,760,920	35,337	8,853,211
Capan1-R2	161,499	27,547,883	39,308	8,722,529	39,264	8,380,681	36,799	8,897,244
GM12878-R1	88,198	24,408,480	37,880	9,907,568	37,836	9,687,772	34,869	10,734,618
GM12878-R2	190,199	28,011,778	40,190	15,253,503	40,146	14,800,666	39,392	9,711,216
HepG2-R1	161,365	31,681,378	39,568	24,630,673	39,524	24,508,802	35,515	11,181,660
HepG2-R2	147,911	26,426,516	39,909	16,945,637	39,865	16,718,446	35,701	9,771,604



**Table S5: Probes Removed in Probe Filtering Step.** This table shows the 34 probes removed from all datasets after our probe filtering step.

Trim Amount: 0.85

Flagged Probe	#datasets in which this probe is flagged
5C_2410_EMS03_FOR_102	10
5C_2410_EMS03_FOR_140	10
5C_2410_EMS03_FOR_349	10
5C_2410_EMS03_FOR_429	10
5C_2410_EMS03_FOR_773	10
5C_2410_EMS03_FOR_165	10
5C_2410_EMS03_FOR_2	10
5C_2410_EMS03_REV_10	9
5C_2410_EMS03_FOR_75	9
5C_2410_EMS03_FOR_74	9
5C_2410_EMS03_REV_13	8
5C_2410_EMS03_FOR_8	8
5C_2410_EMS03_FOR_51	8
5C_2410_EMS03_FOR_54	7
5C_2410_EMS03_FOR_197	7
5C_2410_EMS03_FOR_762	7
5C_2410_EMS03_REV_111	7
5C_2410_EMS03_FOR_228	7
5C_2410_EMS03_FOR_389	7
5C_2410_EMS03_FOR_350	7
5C_2410_EMS03_REV_523	6
5C_2410_EMS03_FOR_407	6
5C_2410_EMS03_FOR_1	6
5C_2410_EMS03_FOR_57	5
5C_2410_EMS03_FOR_129	5
5C_2410_EMS03_FOR_677	5
5C_2410_EMS03_FOR_117	5
5C_2410_EMS03_FOR_248	4
5C_2410_EMS03_FOR_246	4
5C_2410_EMS03_FOR_864	4
5C_2410_EMS03_FOR_283	4
5C_2410_EMS03_FOR_607	4
5C_2410_EMS03_FOR_658	4
5C_2410_EMS03_FOR_298	4

**Table S6: Individual Interactions Removed in Singleton Removal Step.** This table lists the 44 individual interactions removed in the singleton removal step.

z-score 12	#datasets in which this interaction is flagged
Probe-Probe interaction	
5C_2410_EMS03_FOR_124_5C_2410_EMS03_REV_17	10
5C_2410_EMS03_FOR_128_5C_2410_EMS03_REV_17	10
5C_2410_EMS03_FOR_464_5C_2410_EMS03_REV_405	10
5C_2410_EMS03_FOR_126_5C_2410_EMS03_REV_17	9
5C_2410_EMS03_FOR_128_5C_2410_EMS03_REV_203	9
5C_2410_EMS03_FOR_21_5C_2410_EMS03_REV_203	9
5C_2410_EMS03_FOR_339_5C_2410_EMS03_REV_898	9
5C_2410_EMS03_FOR_605_5C_2410_EMS03_REV_777	9
5C_2410_EMS03_FOR_7_5C_2410_EMS03_REV_405	9
5C_2410_EMS03_FOR_817_5C_2410_EMS03_REV_405	9
5C_2410_EMS03_FOR_90_5C_2410_EMS03_REV_613	9
5C_2410_EMS03_FOR_93_5C_2410_EMS03_REV_405	9
5C_2410_EMS03_FOR_467_5C_2410_EMS03_REV_719	8
5C_2410_EMS03_FOR_548_5C_2410_EMS03_REV_405	8
5C_2410_EMS03_FOR_579_5C_2410_EMS03_REV_72	8
5C_2410_EMS03_FOR_125_5C_2410_EMS03_REV_17	7
5C_2410_EMS03_FOR_19_5C_2410_EMS03_REV_203	7
5C_2410_EMS03_FOR_214_5C_2410_EMS03_REV_761	7
5C_2410_EMS03_FOR_71_5C_2410_EMS03_REV_761	7
5C_2410_EMS03_FOR_87_5C_2410_EMS03_REV_782	7
5C_2410_EMS03_FOR_105_5C_2410_EMS03_REV_405	6
5C_2410_EMS03_FOR_208_5C_2410_EMS03_REV_711	6
5C_2410_EMS03_FOR_503_5C_2410_EMS03_REV_420	6
5C_2410_EMS03_FOR_125_5C_2410_EMS03_REV_665	5
5C_2410_EMS03_FOR_132_5C_2410_EMS03_REV_17	5
5C_2410_EMS03_FOR_139_5C_2410_EMS03_REV_405	5
5C_2410_EMS03_FOR_154_5C_2410_EMS03_REV_405	5
5C_2410_EMS03_FOR_641_5C_2410_EMS03_REV_420	5
5C_2410_EMS03_FOR_68_5C_2410_EMS03_REV_594	5
5C_2410_EMS03_FOR_98_5C_2410_EMS03_REV_405	5
5C_2410_EMS03_FOR_12_5C_2410_EMS03_REV_203	4
5C_2410_EMS03_FOR_136_5C_2410_EMS03_REV_405	4
5C_2410_EMS03_FOR_237_5C_2410_EMS03_REV_665	4
5C_2410_EMS03_FOR_394_5C_2410_EMS03_REV_405	4
5C_2410_EMS03_FOR_5_5C_2410_EMS03_REV_808	4
5C_2410_EMS03_FOR_502_5C_2410_EMS03_REV_420	4
5C_2410_EMS03_FOR_503_5C_2410_EMS03_REV_421	4
5C_2410_EMS03_FOR_597_5C_2410_EMS03_REV_666	4

5C_2410_EMS03_FOR_626_5C_2410_EMS03_REV_420	4
5C_2410_EMS03_FOR_63_5C_2410_EMS03_REV_405	4
5C_2410_EMS03_FOR_641_5C_2410_EMS03_REV_421	4
5C_2410_EMS03_FOR_672_5C_2410_EMS03_REV_203	4
5C_2410_EMS03_FOR_751_5C_2410_EMS03_REV_665	4
5C_2410_EMS03_FOR_883_5C_2410_EMS03_REV_405	4



**Table S7: Significant Interactions.** This table lists all significant intra-TAD and inter - TAD interactions in the 5 cell types.

**Table S8: RT-PCR Primers.** Primers used in our qPCR experiment testing gene expression in the 5C region.

Genes in 5C Region - qPCR primers

<u>Primer Name</u>	<u>Primer Sequence</u>	<u>Exon #</u>	<u>Genome Coordinates (hg18)</u>
<b><u>TES:</u> <i>testin isoform 1</i></b>			
TES_F_A	GCCCCTTGTTTAAAATGCAA	2	115661854-115661873
TES_R_A	TGCTCAAGAGGACATCATGC	3	115676346-115676365
<b><u>CAV2:</u> <i>caveolin 2 isoform a and b</i></b>			
CAV2_F_A	GGCTCAACTCGCATCTCAAG	1	115927202-115927221
CAV2_R_A	CAGGAACACCGTCAGGAACT	2	115927656-115927675
<b><u>CAV1:</u> <i>caveolin 1</i></b>			
CAV1_F_A	GAGCTGAGCGAGAAGCAAGT	2	115953887-115953906
CAV1_R_A	CAAATGCCGTCAAACTGTG	3	115986275-115986294
<b><u>MET:</u> <i>met proto-oncogene isoform a precursor</i></b>			
MET_F_B	CCAATGACCTGCTGAAATTG	11	116197041-116197060
MET_R_B	CTTTTCCAAGGACGGTTGAA	12	116198805-116198824
<b><u>CAPZA2:</u> <i>capping protein (actin filament) muscle Z-line</i></b>			
CAPZA2_F_A	GAAGGAGGCAACTGATCCAA	5	116331553-116331572
CAPZA2_R_A	GCTTGGAAGTGTGGCTTTTC	6	116333601-116333620
<b><u>ST7:</u> <i>suppression of tumorigenicity 7 isoform b</i></b>			
ST7_F_A	TTCCAGTAACGGGGACTCAG	3	116546967-116546986
ST7_R_A	TGGATTTCCGCATACTTTGC	4	116557086-116557105
<b><u>WNT2:</u> <i>wingless-type MMTV integration site family</i></b>			
WNT2_F_B	GTGGATGCAAAGGAAAGGAA	3	116742416-116742435
WNT2_R_B	AGCCAGCATGTCCTGAGAGT	4	116725090-116725109
<b><u>ASZ1:</u> <i>ankyrin repeat, SAM and basic leucine zipper</i></b>			
ASZ1_F_B	CACGTCAGGGTCATAAA	6	116812115-116812137
ASZ1_R_B	GCTGTTGAAGTTTTCTTCCA	7	116810346-116810366
<b><u>CFTR:</u> <i>cystic fibrosis transmembrane conductance regulator</i></b>			
CFTR2-3F	CCCTTCTGTTGATTCTGCTG	2	116931609-116931628
CFTR2-3R	AAGGGCATTAAATGAGTTTAGGA	3	116936357-116936378
<b><u>CTTNBP2:</u> <i>cortactin binding protein 2</i></b>			
CTTNBP2_F_C	AAAATGGCTTCACACCCTTG	6	117210181-117210200
CTTNBP2_R_C	TGTCTGTCCATCAGCAG	7	117207818-117207837
<b><u>LSM8:</u> <i>U6 snRNA-associated Sm-like protein LSM8</i></b>			
LSM8_F_A	CAGCTCTTCACAGGGGGTAG	3	117615633-117615652
LSM8_R_A	CTGCTCGAATATTCGCCAAA	4	117619247-117619000
<b><u>ANKRD7:</u> <i>ankyrin repeat domain 7 isoform b</i></b>			
ANKRD7_F_A	ACTTTTGCACCTAGCCTGTG	2	117661724-117661743

ANKRD7_R_A	ATCTGGGTCTGCACCAAAGT	3	117662034-117662053
<b>HPRT1:</b> <i>Chromosome X: hypoxanthine phosphoribosyltransferase 1</i>			
HPRT2-3F	TGAGGATTTGGAAAGGGTGT	2	133435114-133435133
HPRT2-3R	TAATCCAGCAGGTCAGCAA	3	133436965-133436984

**Figures S1-S10: Correction pipeline for each replicate.** A) Raw data. B) Data after cis-purge. Grey stripes represent primers that were removed. C) Data after singleton removal. Grey stripes are the primers removed in the previous step, grey pixels are the individual reactions that were removed in this step. D) Final coverage corrected data. Grey lines have been removed and are now represented in white. E) Binned raw data (100kb, 10kb step). F) Binned coverage corrected data (100kb, 10kb step).

Caco2 R1

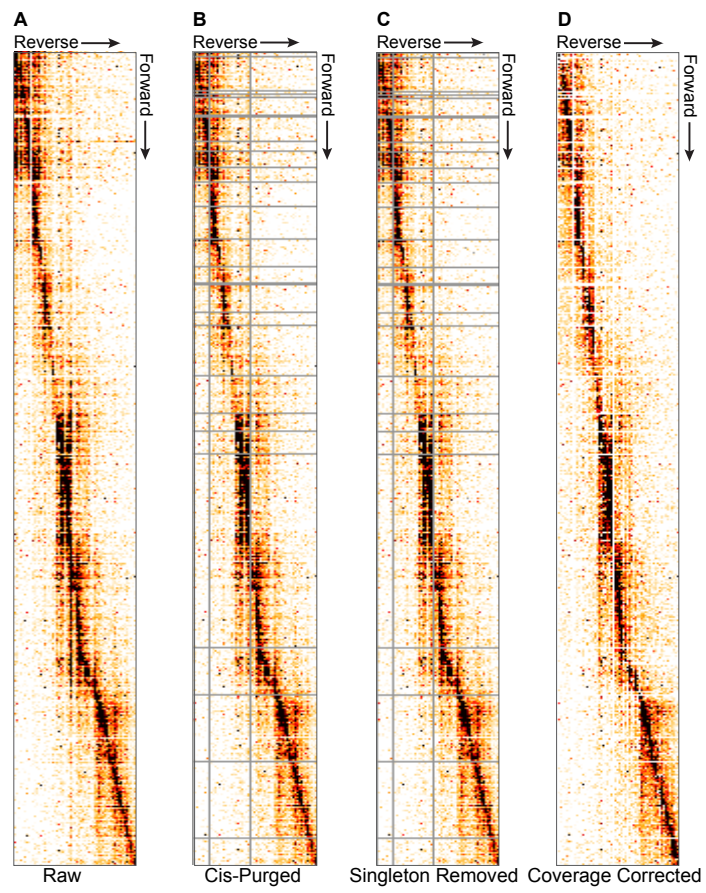
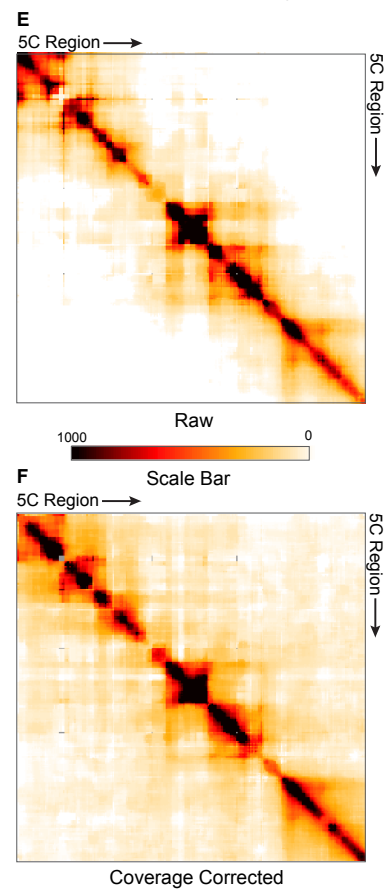
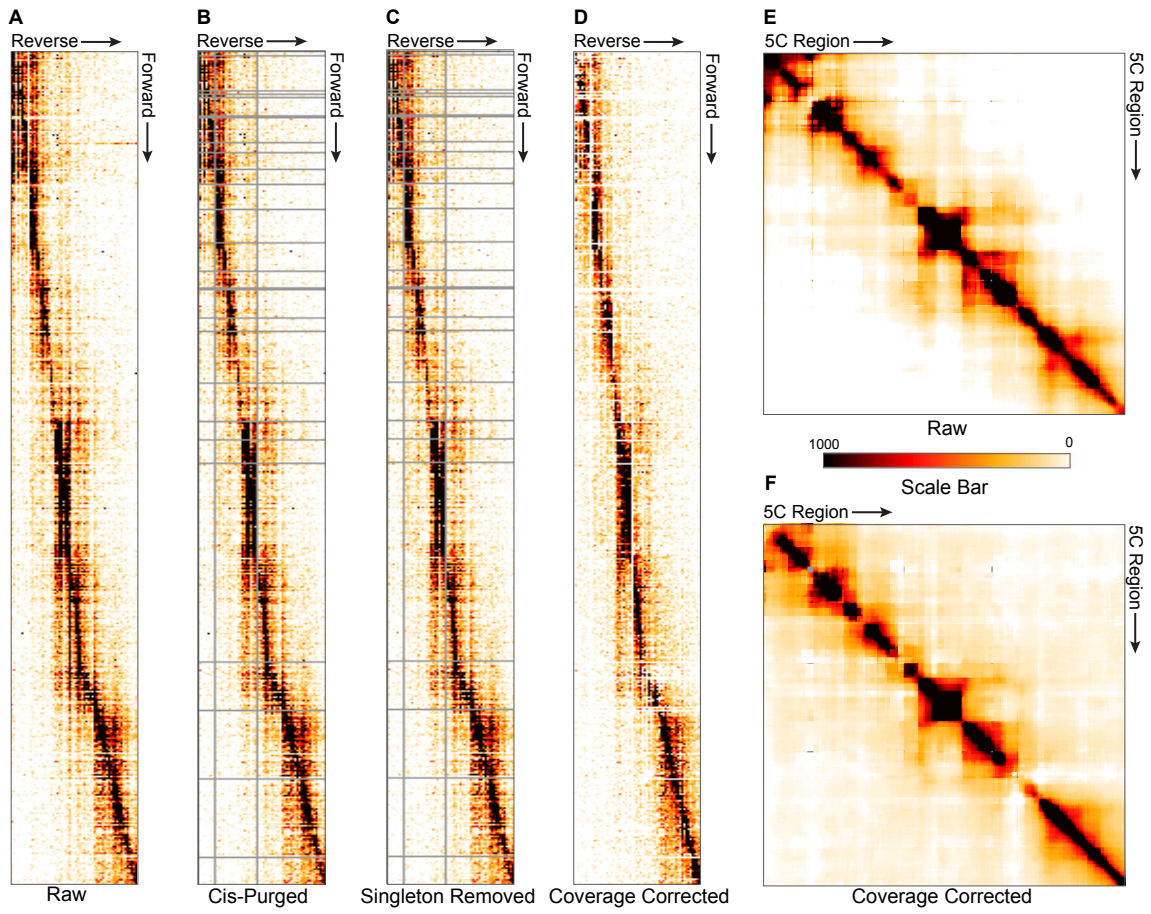


Figure S1



Caco2 R2

Figure S2



Calu3 R1

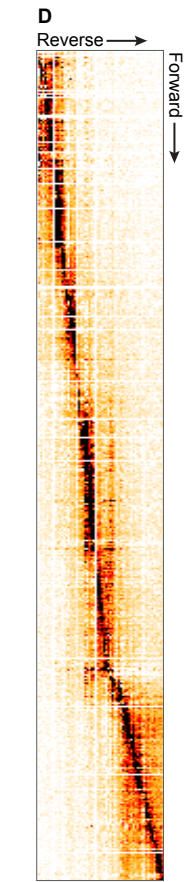
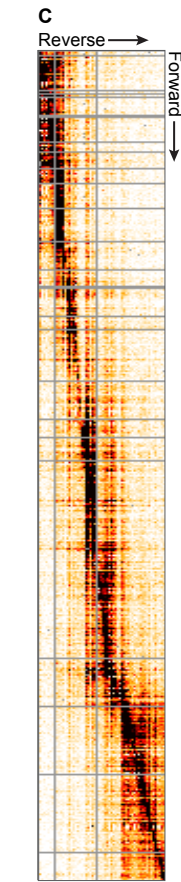
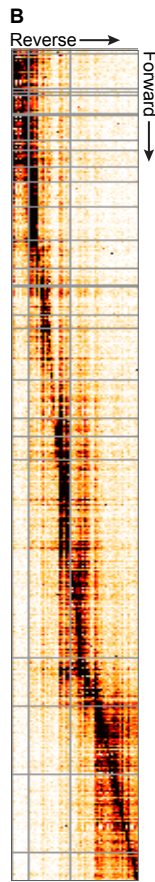
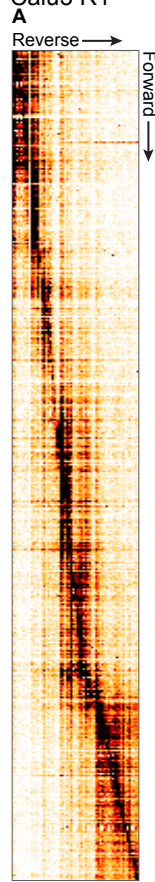
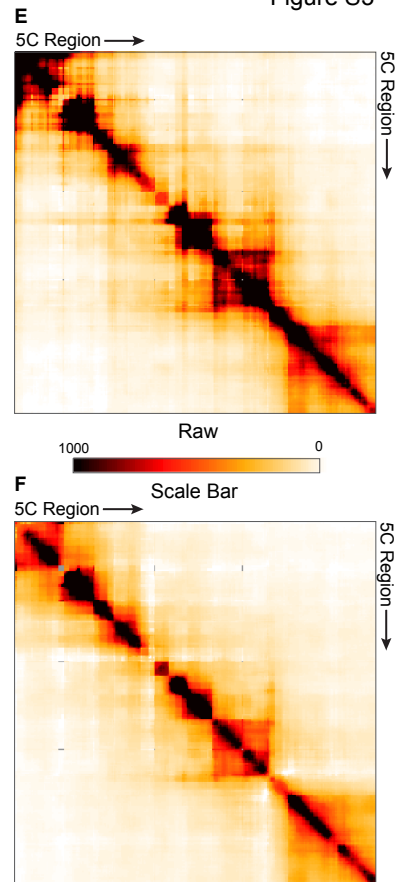
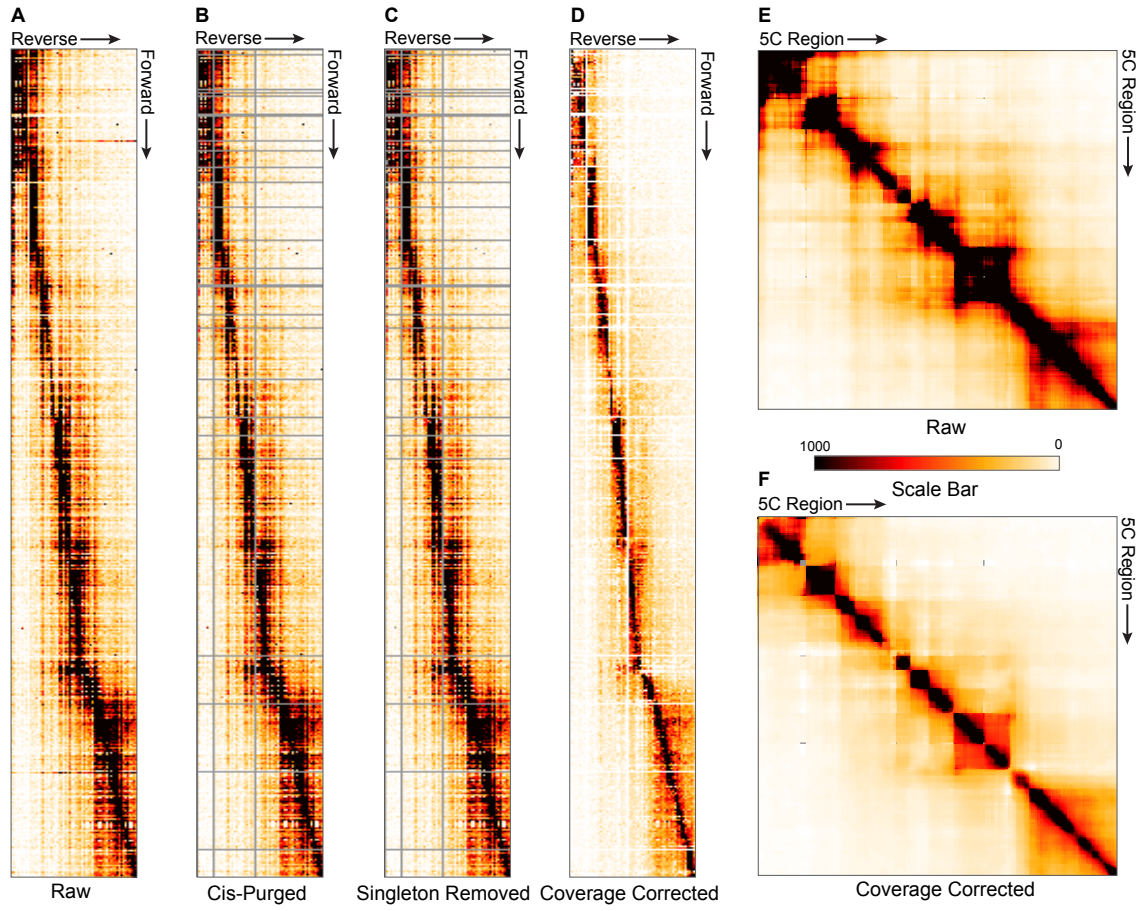


Figure S3



Calu3 R2

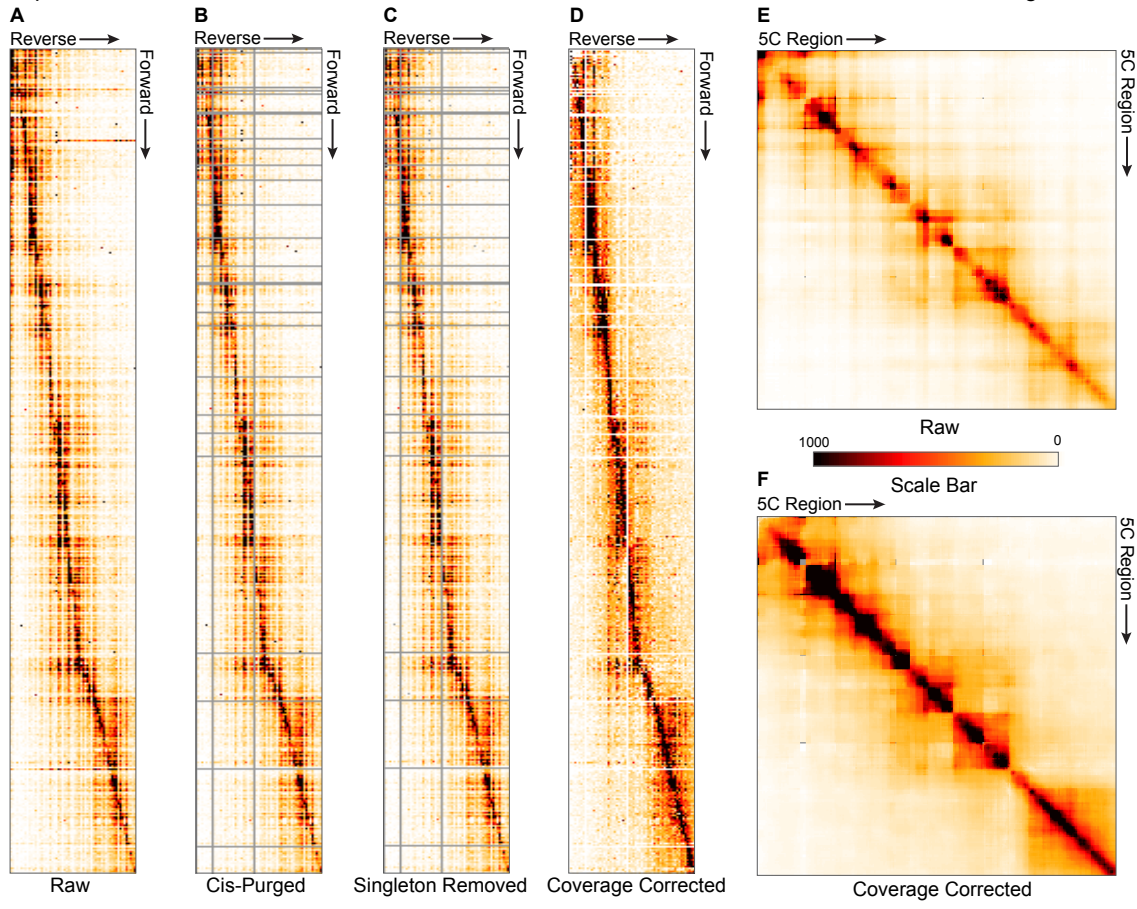
Figure S4





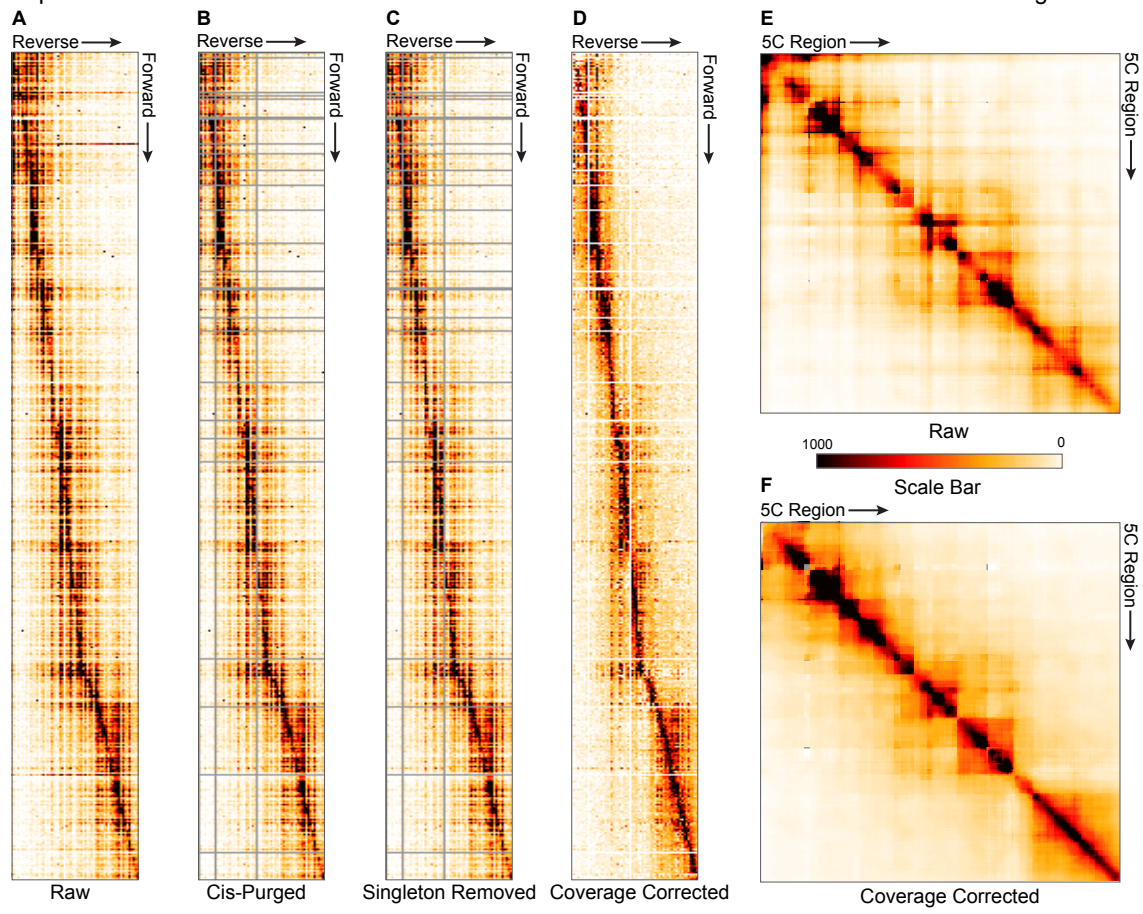
Capan1 R1

Figure S5



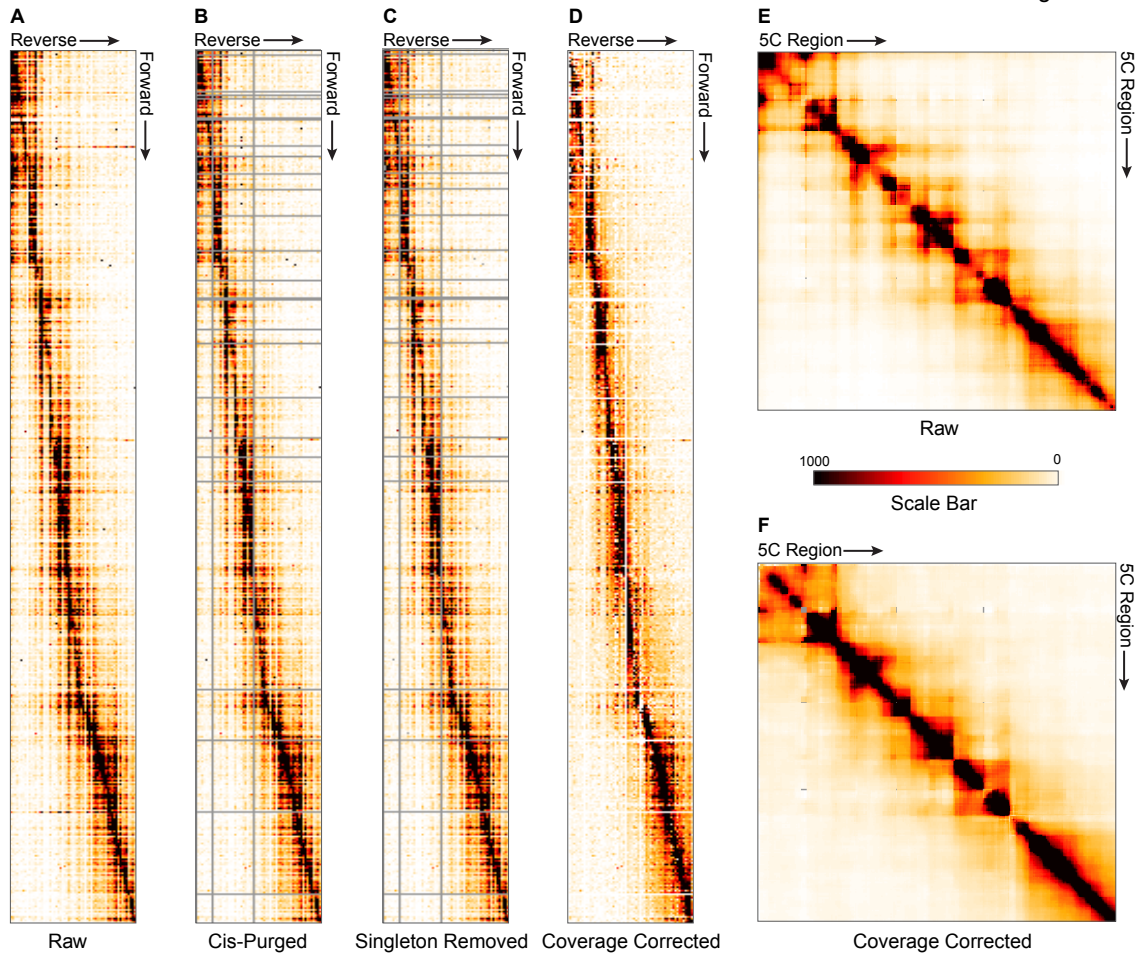
Capan1 R2

Figure S6



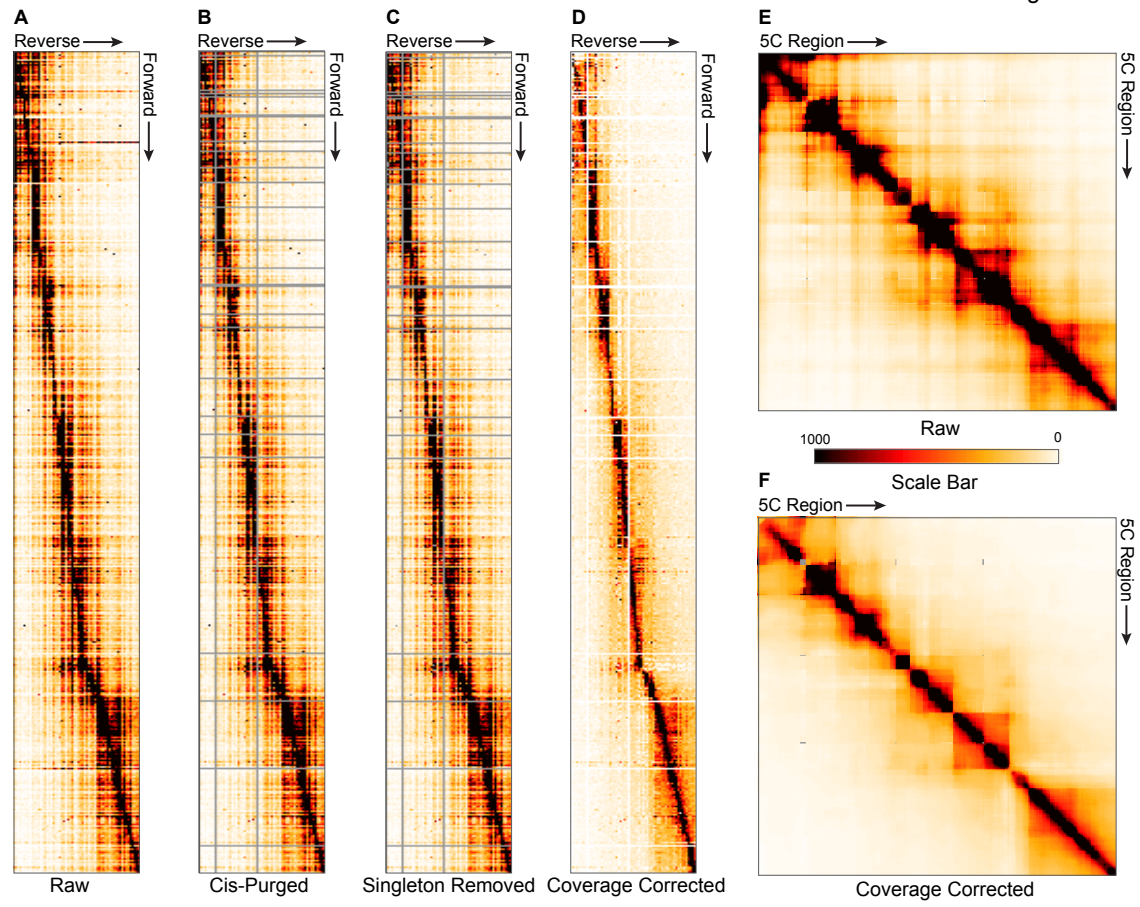
GM12878 R1

Figure S7



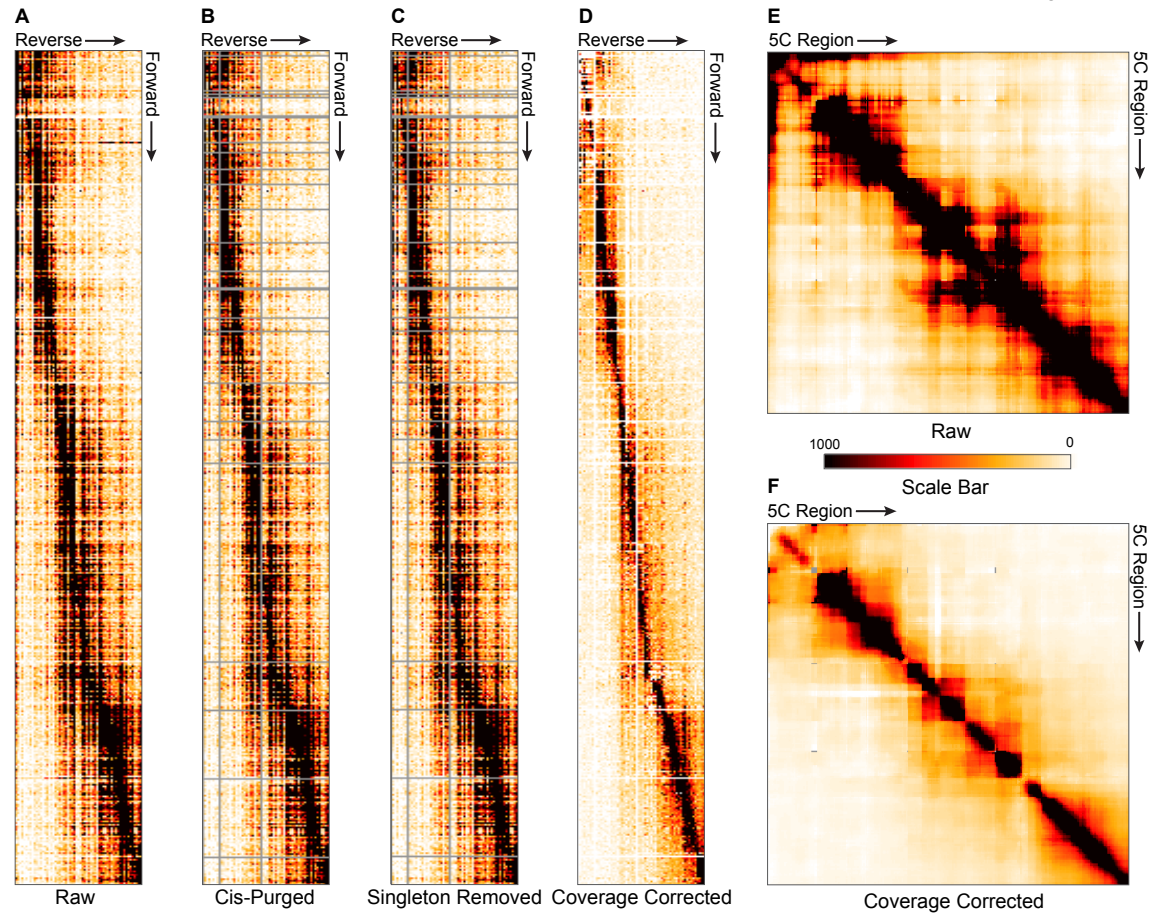
GM12878 R2

Figure S8



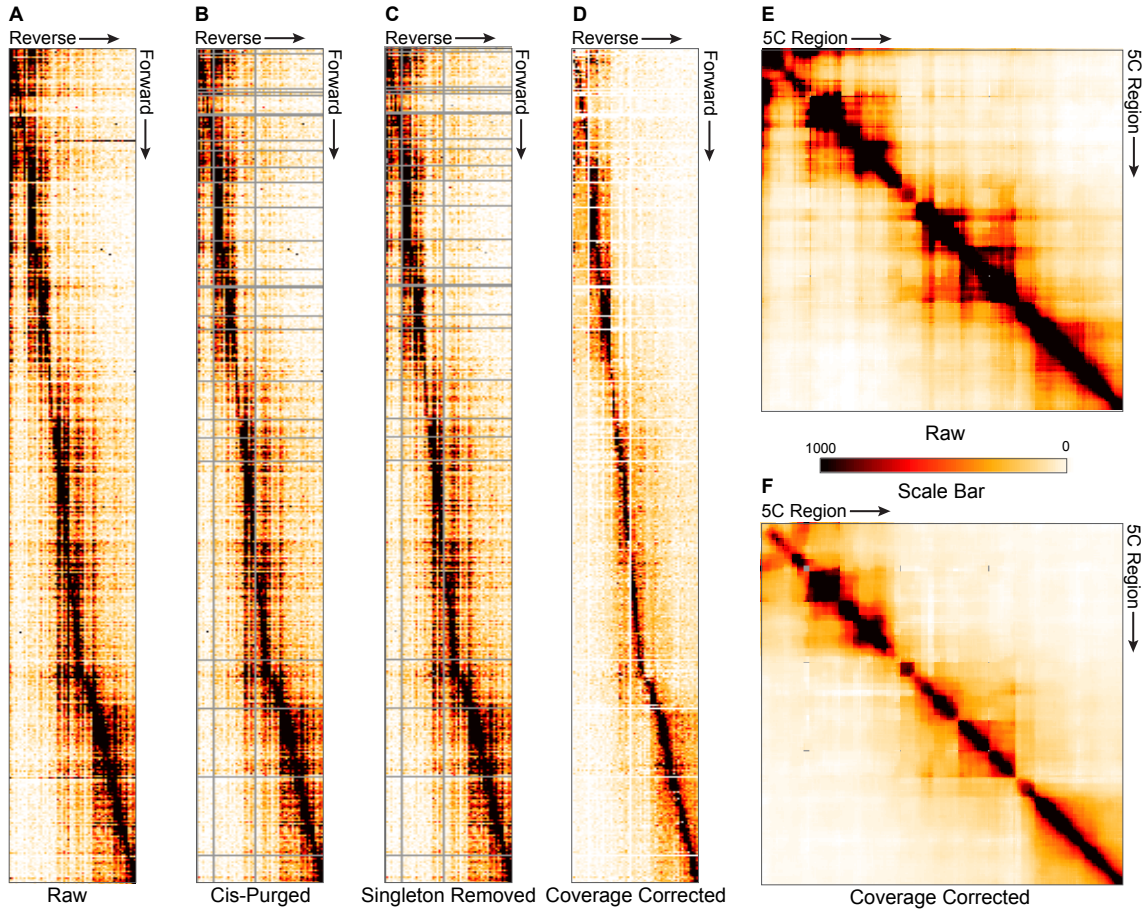
HepG2 R1

Figure S9



HepG2 R2

Figure S10

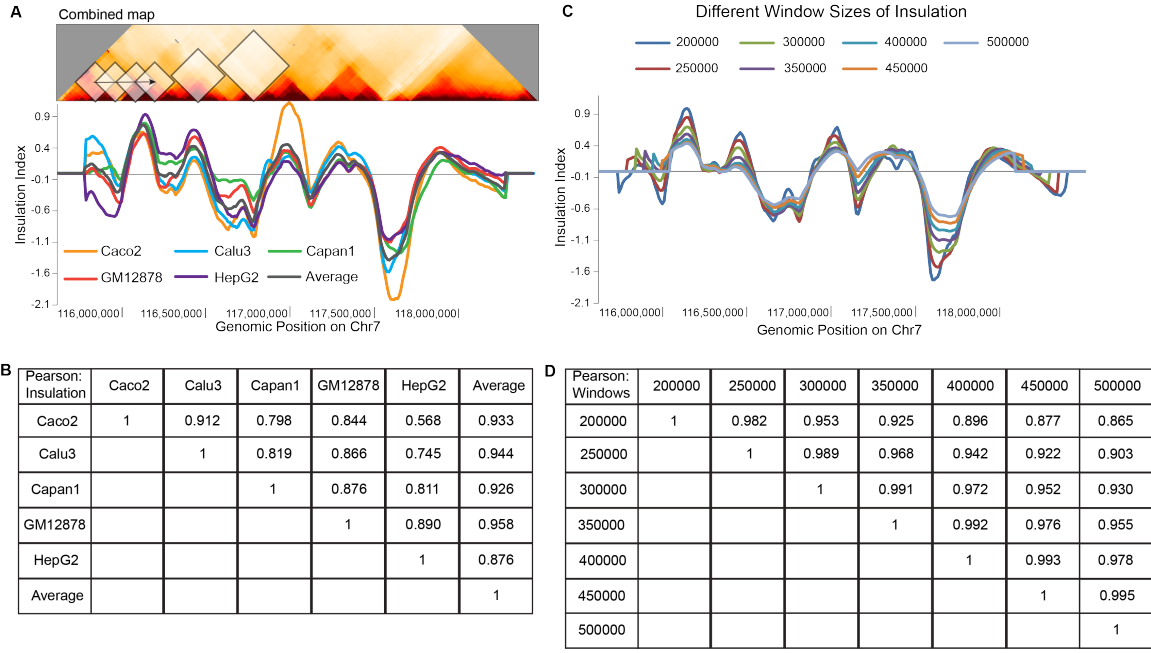


**Figure S11: Insulation Index of all cell lines and different window search spaces, and correlation with CTCF binding**

A) The insulation index method is illustrated by sliding a diamond along the bottom of the triangular heatmap. Interactions within the diamond are summed and divided by the average sum for the region to calculate the insulation score of the bin at the bottom corner of the diamond. Below the heatmap is plotted the insulation index for all cell lines and the average, as in **Figure 2**. The size of the window over which can be changed, but does not affect the insulation profile (panel C).

B) A) Insulation profiles run on the combined heatmap (all 10 datasets) using different diamond (window) sizes. As the size increases the index smoothens out but maintains the same peaks and minima. B). Browser shot (hg18) of the 2.8 Mb region studied here with the positions of the 7 TAD indicated (Top track, in black bars). Below are CTCF ChIP seq tracks (CTCF P = peaks, CTCF s= raw signal) for GM12878 and HepG2 cells. CTCF sites are within 10-20 Kb of TAD boundaries but are also found inside TADs. Also indicated are positions of all genes (UCSC gene track). CTCF ChIP- seq data were generated at the Broad Institute and in the lab of Dr. Bradley Bernstein at Massachusetts General Hospital/Harvard Medical School. This data is publicly available as part of the ENCODE dataset. B) Pearson correlations between the insulation profiles for the different cell types studies here. C) Pearson correlation of insulation profiles for the combined dataset calculated with the different diamond sizes.

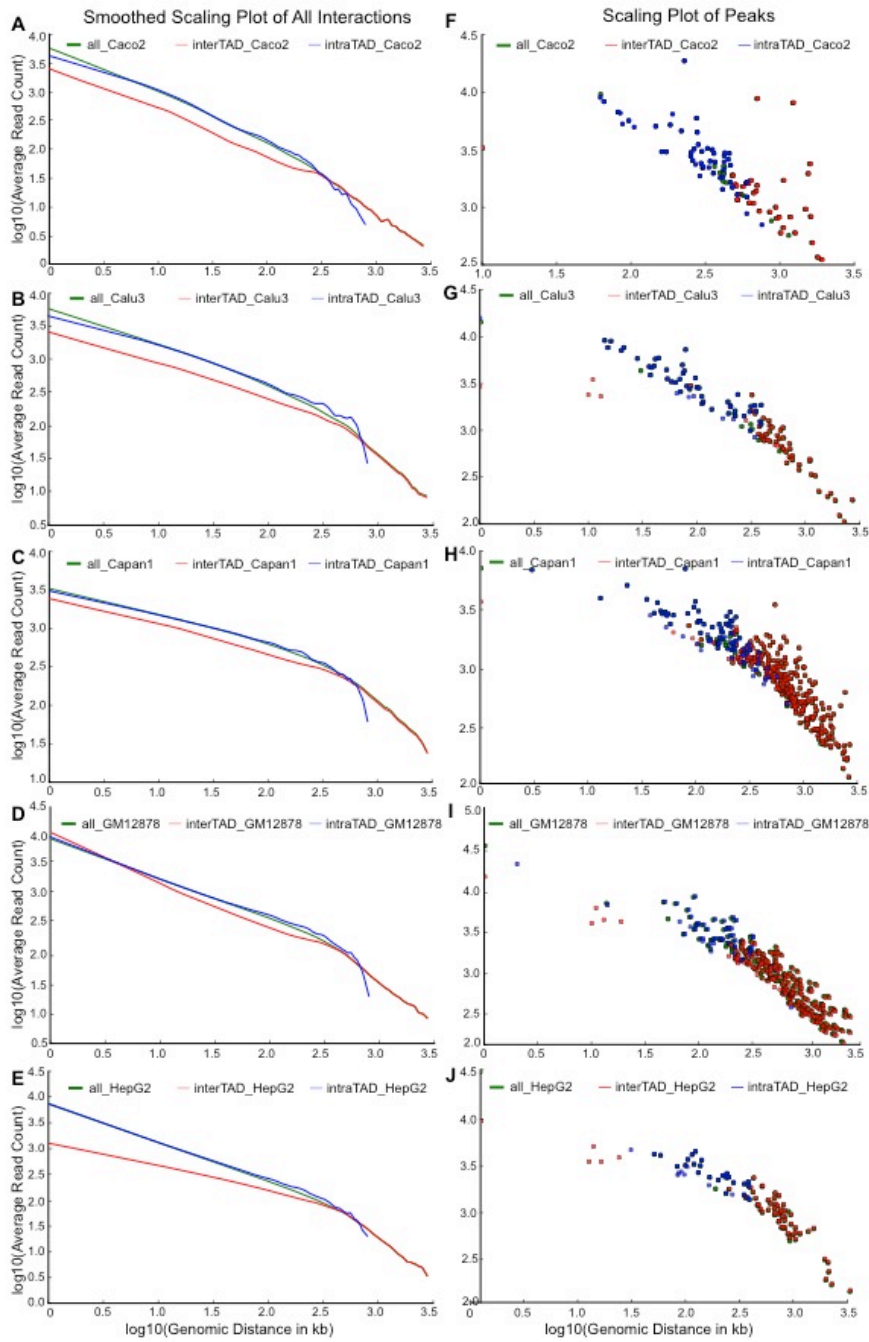
Figure S11





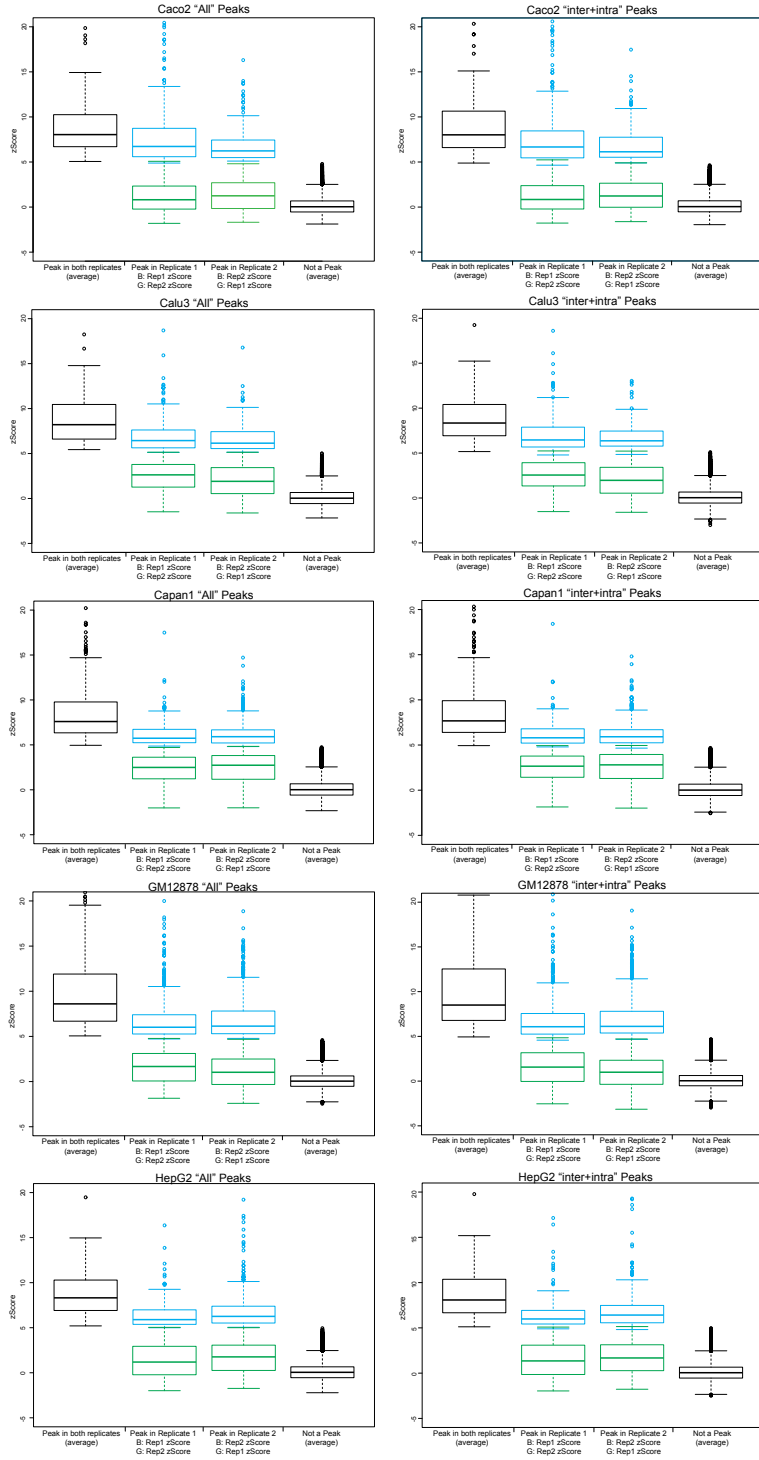
**Figure S12: Scaling plots for all cell lines, interactions and peaks.** For all plots: red represents inter-TAD interactions, blue intra-TAD interactions, and green all interactions. A) Scaling plot of all interactions for Caco2 cells. B) Scaling plot of all interactions for Calu3 cells C) Scaling plot of all interactions for Capan1 cells. D) C) Scaling plot of all interactions for GM12878 cells. E) D) Scaling plot of all interactions for HepG2 cells. F) E) Scaling plot of peaks significant interactions in Caco2 cells. G) F) Scaling plot of significant interactions in Capan1 cells. H) G) Scaling plot of significant interactions in Calu3 cells. H). I) Scaling plots of significant interactions in GM12878 cells. J) H) Scaling plots of significant interactions in HepG2 cells.

Figure S12



**Figure S13: Interactions deemed significant in only one replicate have higher signal in the other replicate as compared to interactions deemed not significant in both replicates.** For all plots: The box plot on the far left represents z-scores for interactions called significant in both replicates. The blue box plot of peaks in replicate 1 represents those interactions in replicate 1 that were deemed significant, but were not significant in replicate 2. The green box shows the z-scores of those same interactions in the second replicate, where they were not counted significant. This coloring is reversed when we look at replicate 2. The blue box now indicates the z-score of interactions deemed significant in replicate 2 but not in replicate 1. The green box shows the z-scores of those same interactions in replicate 1, where they are not significant. The box plot on the far right shows the z-scores of all interactions in both replicates that were not significant. This is the same for all graphs.

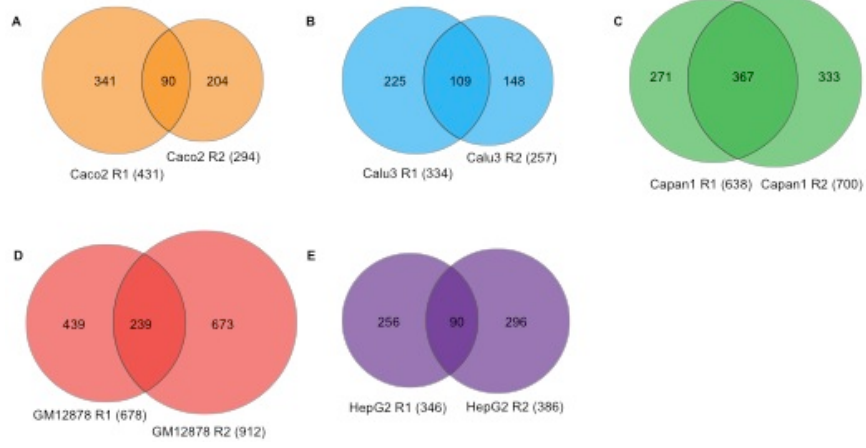
Figure S13



**Figure S14: Overlap of peak calling between replicates on the whole data set.**

Overlap of significant interactions in Caco2 replicates (A), Calu3 replicates (B), Capan1 replicates (C), GM12878 replicates (D) and HepG2 replicates (E).

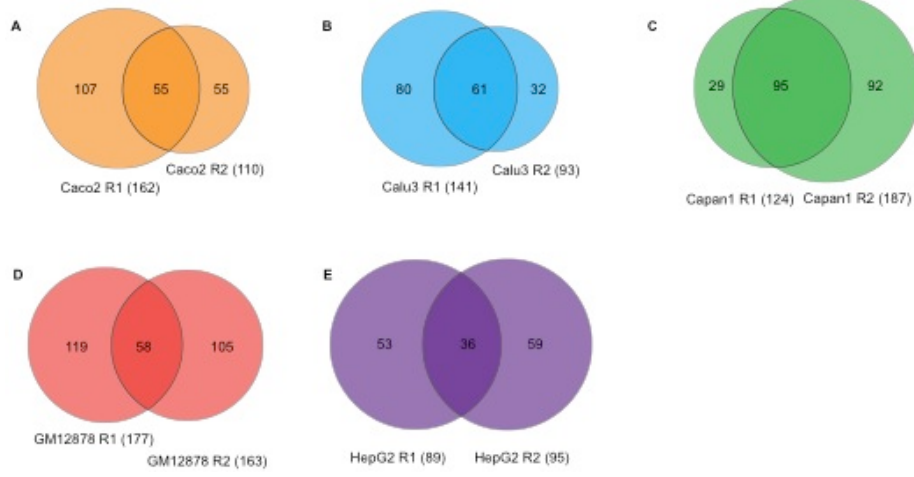
Figure S14



**Figure S15: Overlap of peak calling between replicates on the intraTAD dataset.**

Overlap of significant interactions in Caco2 replicates (A), Calu3 replicates (B), Capan1 replicates (C), GM12878 replicates (D) and HepG2 replicates (E).

Figure S15

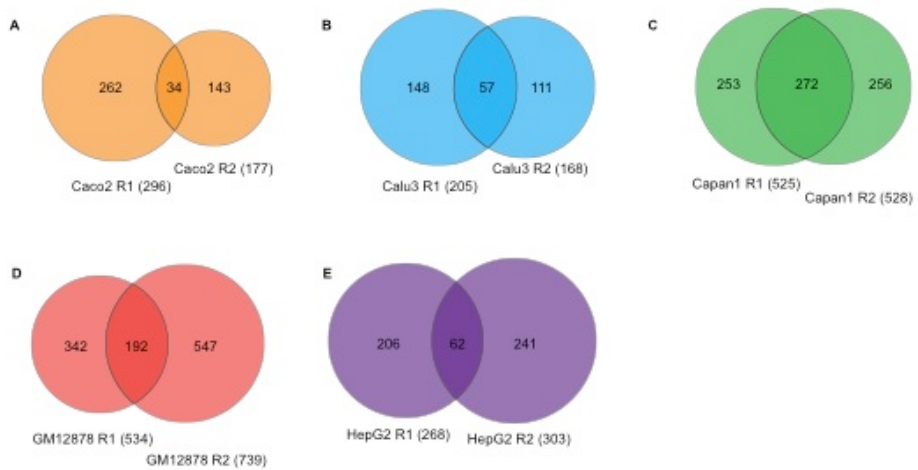




**Figure S16: Overlap of peak calling between replicates on the interTAD dataset.**

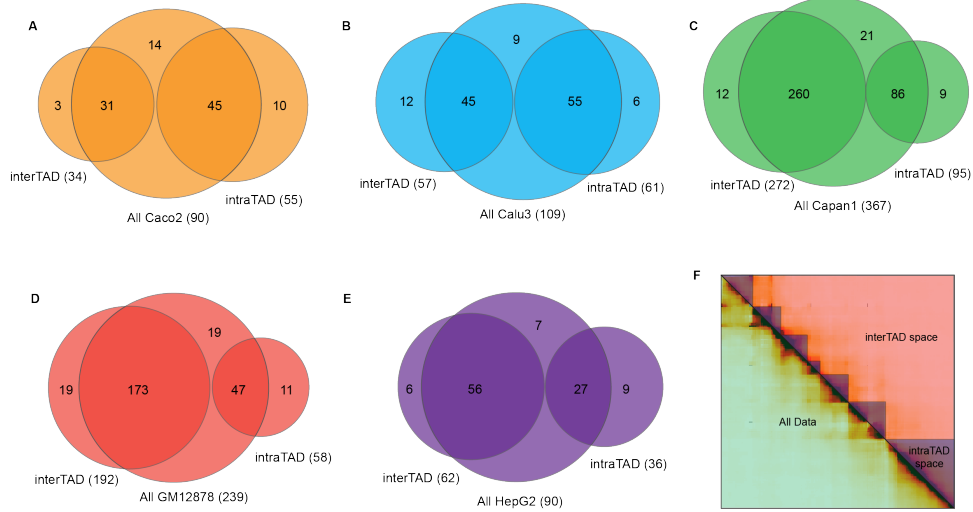
Overlap of significant interactions in Caco2 replicates (A), Calu3 replicates (B), Capan1 replicates (C), GM12878 replicates (D) and HepG2 replicates (E).

Figure S16



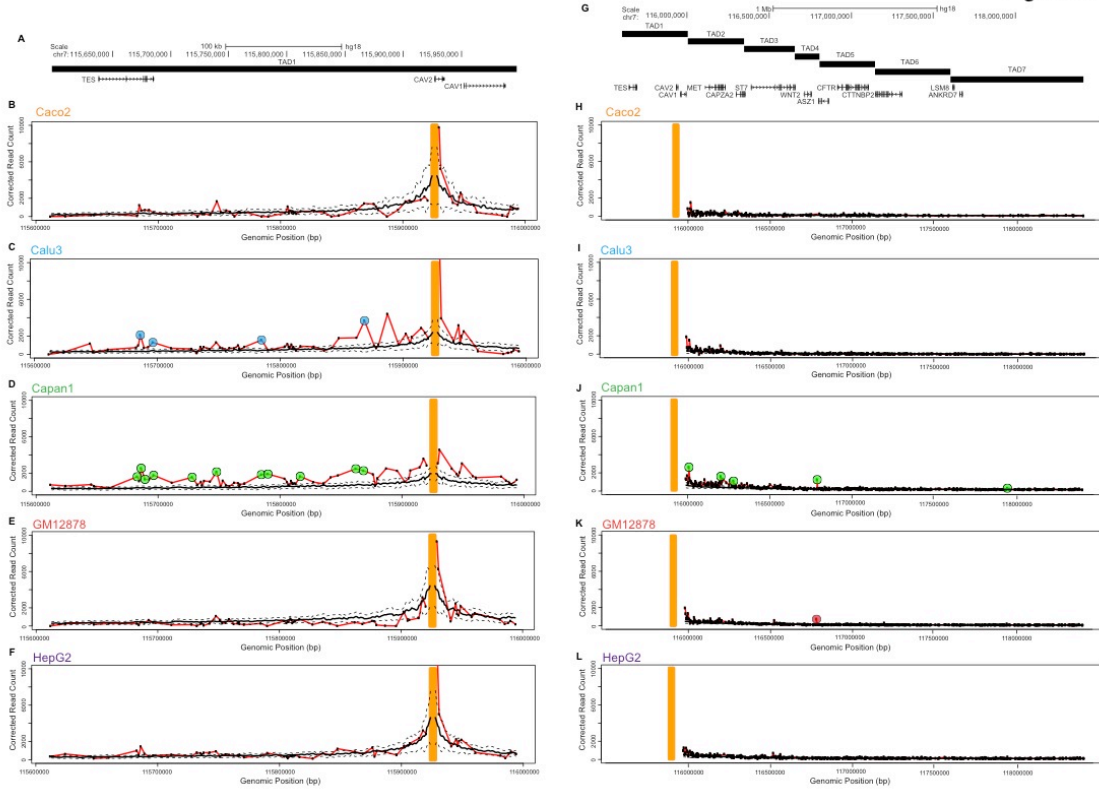
**Figure S17: Overlap of peaks called using the different spaces: whole, intra-TAD and inter-TAD, for all cell lines.** A) Overlap of peak calling methods in Caco2 cells. B) Overlap of peak calling methods in Calu3 cells. C) Overlap of peak calling methods in Capan1 cells. D) Overlap of peak calling methods in GM12878 cells. E) Overlap of peak calling methods in HepG2 cells. F) Diagram of peak calling space (same as Figure 4A).

Figure S17



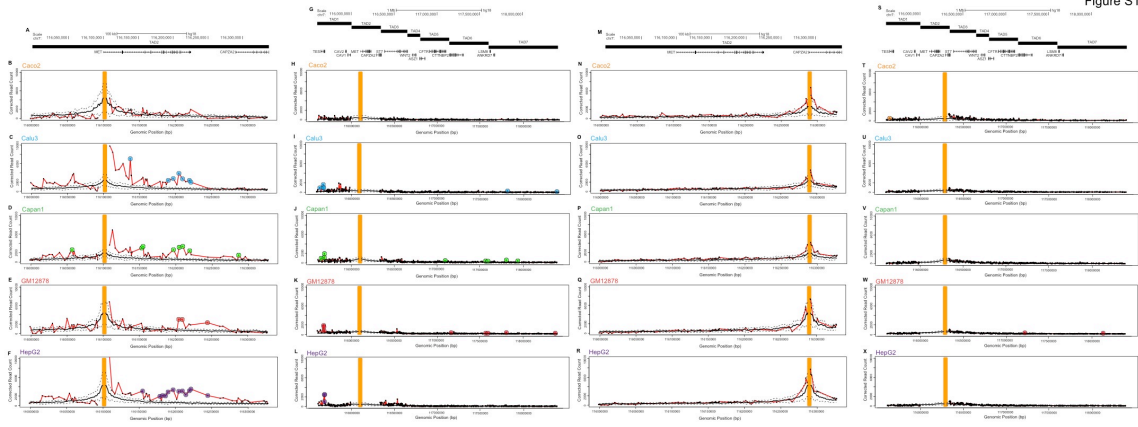
**Figure S18: Significant interactions of the *CAV2* promoter.** A genome browser snapshot of the region is above each profile view. In each anchor plot: Corrected read count is plotted versus genomic distance. The yellow bar represents the location of the anchoring interaction. The dotted black lines represent the loess standard deviation, and the thick black line represents the loess average. The red line represents the interactions of the genomic fragment indicated by the yellow line with all other interrogated fragments in the region. Significant interactions are indicated by large circles.

Figure S18



**Figure S19: Significant interactions of *MET* and *CAPZA2* promoters.** A genome browser snapshot of the region is above each profile view. In each anchor plot: Corrected read count is plotted versus genomic distance. The yellow bar represents the location of the anchoring interaction. The dotted black lines represent the loess standard deviation, and the thick black line represents the loess average. The red line represents the interactions of the genomic fragment indicated by the yellow line with all other interrogated fragments in the region. Significant interactions are indicated by large circles.

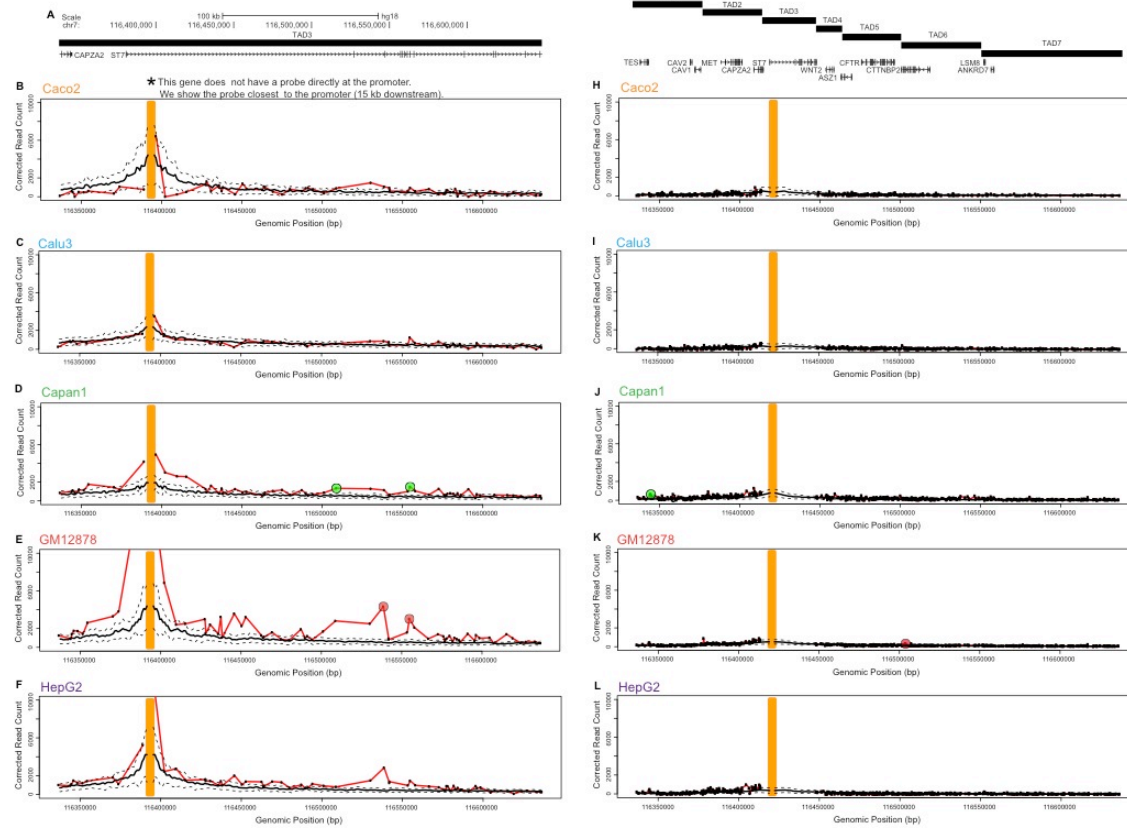
Figure S19





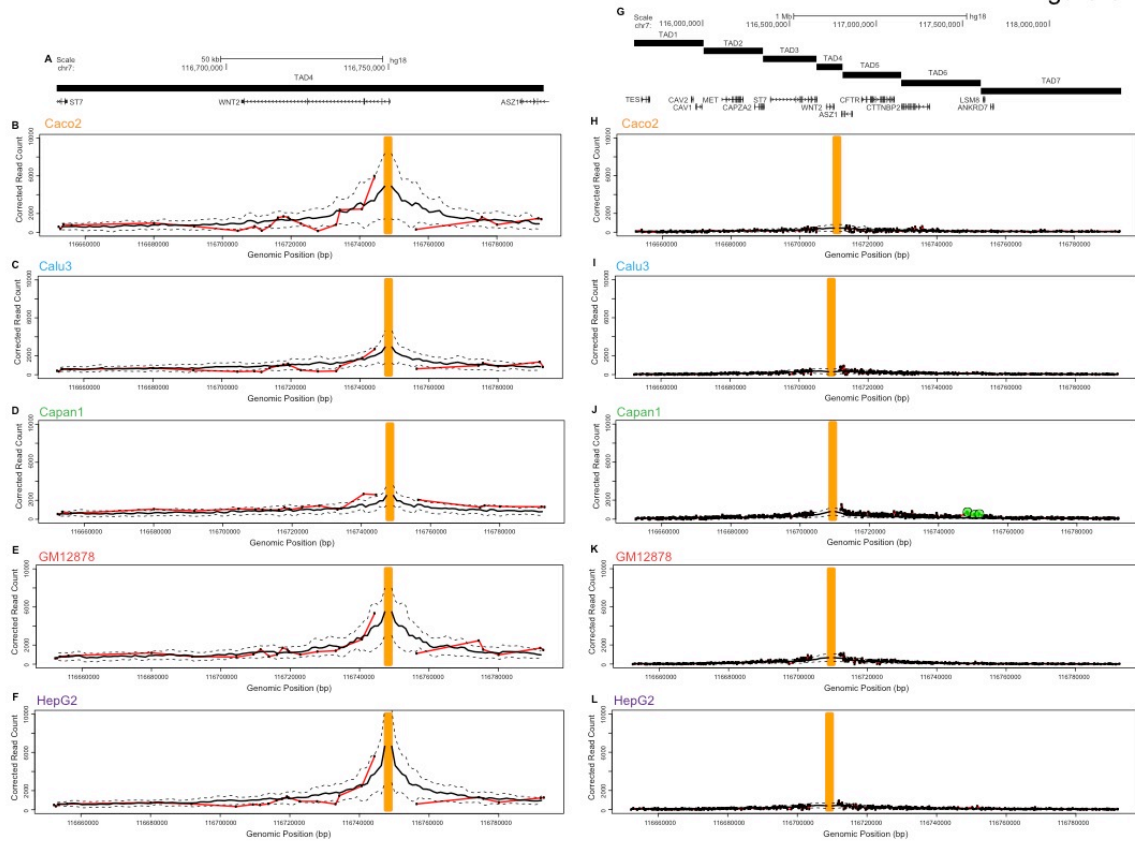
**Figure S20: Significant interactions of the *ST7* promoter.** A genome browser snapshot of the region is above each profile view. In each anchor plot: Corrected read count is plotted versus genomic distance. The yellow bar represents the location of the anchoring interaction. The dotted black lines represent the loess standard deviation, and the thick black line represents the loess average. The red line represents the interactions of the genomic fragment indicated by the yellow line with all other interrogated fragments in the region. Significant interactions are indicated by large circles.

Figure S20



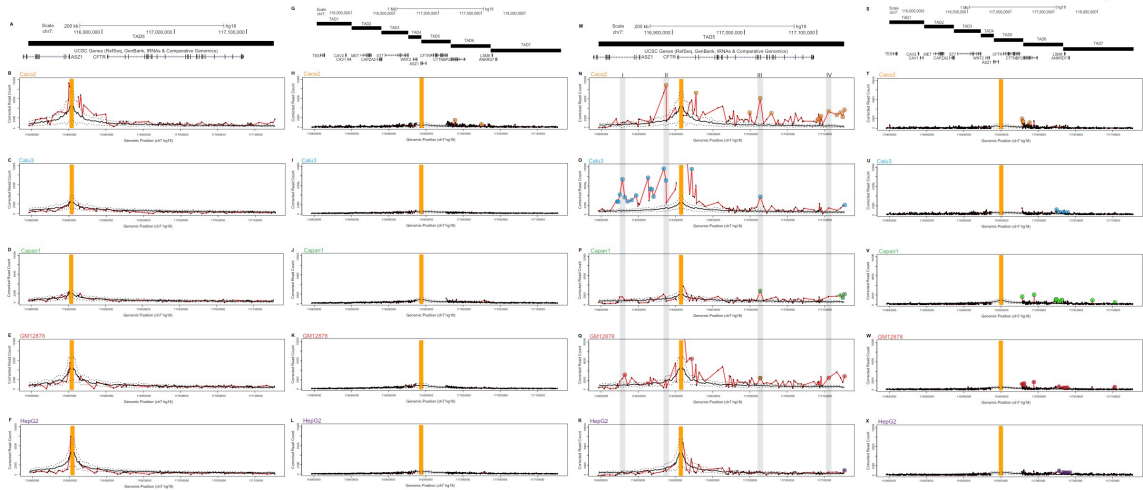
**Figure S21: Significant interactions of the *WNT2* promoter.** A genome browser snapshot of the region is above each profile view. In each anchor plot: Corrected read count is plotted versus genomic distance. The yellow bar represents the location of the anchoring interaction. The dotted black lines represent the loess standard deviation, and the thick black line represents the loess average. The red line represents the interactions of the genomic fragment indicated by the yellow line with all other interrogated fragments in the region. Significant interactions are indicated by large circles.

Figure S21



**Figure S22: Significant interactions of *ASZ1* and *CFTR* promoters.** A genome browser snapshot of the region is above each profile view. In each anchor plot: Corrected read count is plotted versus genomic distance. The yellow bar represents the location of the anchoring interaction. The dotted black lines represent the loess standard deviation, and the thick black line represents the loess average. The red line represents the interactions of the genomic fragment indicated by the yellow line with all other interrogated fragments in the region. Significant interactions are indicated by large circles.

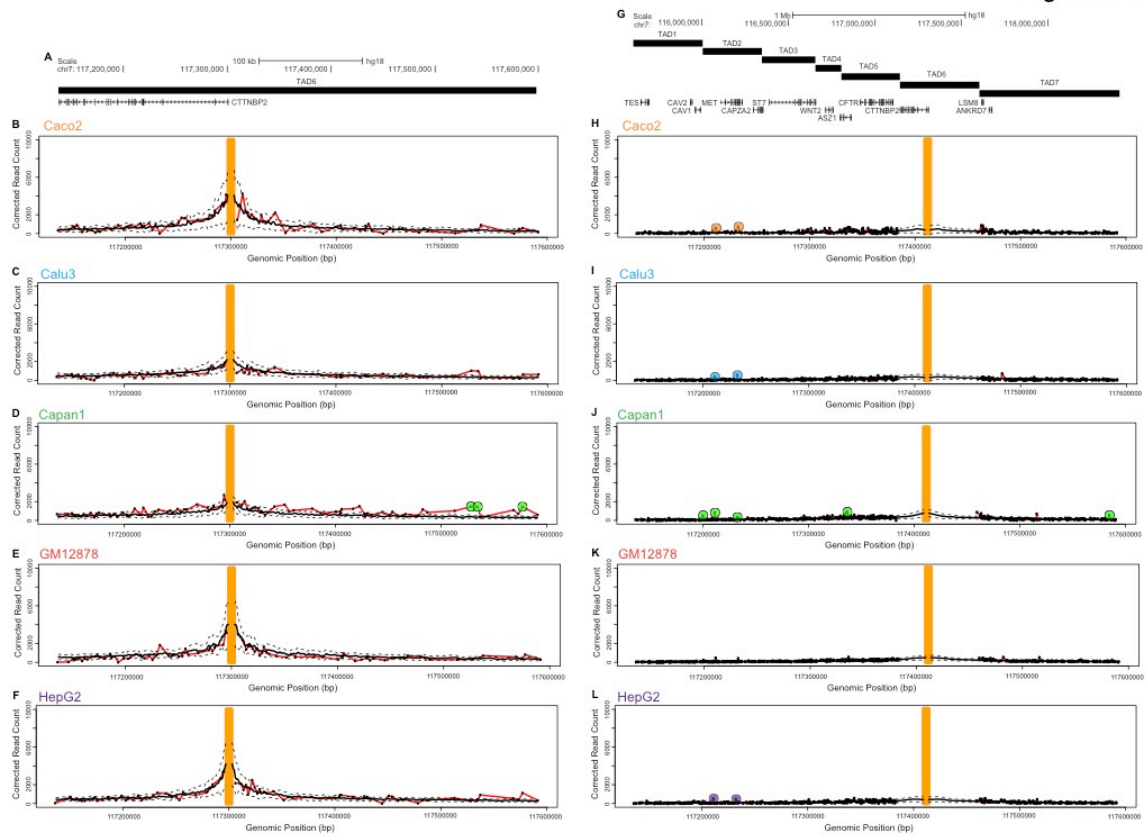
Figure S22



**Figure S23: Significant interactions of the *CTTNBP2* promoter within TAD6.**

A genome browser snapshot of the region is above each profile view. In each anchor plot: Corrected read count is plotted versus genomic distance. The yellow bar represents the location of the anchoring interaction. The dotted black lines represent the loess standard deviation, and the thick black line represents the loess average. The red line represents the interactions of the genomic fragment indicated by the yellow line with all other interrogated fragments in the region. Significant interactions are indicated by large circles.

Figure S23





**Figure S24: Significant interactions of the *LSM8* and Ankyrin Repeat Domain- containing Protein 7 (*ANKRD7* [MIM 610731]) promoters.** A genome browser snapshot of the region is above each profile view. In each anchor plot: Corrected read count is plotted versus genomic distance. The yellow bar represents the location of the anchoring interaction. The dotted black lines represent the loess standard deviation, and the thick black line represents the loess average. The red line represents the interactions of the genomic fragment indicated by the yellow line with all other interrogated fragments in the region. Significant interactions are indicated by large circles.

Figure S24

