

# Three new sesquiterpene glycosides from the roots of *Codonopsis pilosula*

Yueping Jiang<sup>a,b</sup>, Yufeng Liu<sup>a</sup>, Qinglan Guo<sup>a</sup>, Chengbo Xu<sup>a</sup>, Chenggen Zhu<sup>a</sup>, and Jiangong Shi<sup>a\*</sup>

<sup>a</sup> *State Key Laboratory of Bioactive Substance and Function of Natural Medicines, Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100050, China*

<sup>b</sup> *Department of Pharmacy, Xiangya Hospital, Central South University, Changsha, 410008, China*

## Supporting Information

---

\*Corresponding author. E-mail: shijg@imm.ac.cn.

No.	Content	Page
1	<b>Detailed ECD Calculation of 1–3, their aglycones, the model compounds, and (–)-<math>\alpha</math>-barbatenal-reduction product</b>	S4
2	<b>Figure S1.</b> The re-optimized conformer of <b>1</b> .	S4
3	<b>Figure S2.</b> The re-optimized fourteen conformers of the aglycone of <b>1</b> and their equilibrium populations.	S5
4	<b>Figure S3.</b> The re-optimized sixteen conformers of the <b>model</b> of <b>1</b> and their equilibrium populations.	S6
5	<b>Figure S4.</b> The experimental CD spectrum of <b>1</b> (black) and the calculated ECD spectra of <b>1</b> (dash dotted red), its aglycone (dashed green), and the <b>model</b> (dotted blue) in MeOH.	S7
6	<b>Figure S5.</b> The re-optimized conformer of <b>2</b> .	S7
7	<b>Figure S6.</b> The re-optimized five conformers of the aglycone of <b>2</b> and their equilibrium populations.	S7
8	<b>Figure S7.</b> The re-optimized ten conformers of the <b>model</b> of <b>2</b> and their equilibrium populations.	S8
9	<b>Figure S8.</b> The experimental CD spectrum of <b>2</b> (black) and the calculated ECD spectra of <b>2</b> (dash dotted red), its aglycone (dashed green), and the <b>model</b> (dotted blue) in MeOH.	S8
10	<b>Figure S9.</b> The optimized conformer of <b>3</b> .	S9
11	<b>Figure S10.</b> The optimized conformer of the aglycone of <b>3</b> .	S9
12	<b>Figure S11.</b> The experimental CD (black and blue) and calculated ECD (dash dotted red and dashed green) spectra of <b>3</b> and its aglycone in MeOH.	S9
13	<b>Figure S12.</b> The reoptimized six conformers of (–)- $\alpha$ -barbatenal-reduction product	S9
14	<b>Figure S13.</b> The calculated ECD spectrum of (–)- $\alpha$ -barbatenal-reduction product.	S10
15	<b>Figure S12.</b> The HPLC chromatograms of the sugar derivatives, from top to bottom, for D-glucose, L-glucose, D-apiose, L-apiose, and sugars from hydrolysates <b>1</b> and <b>2</b> .	S10
16	<b>Figure S13.</b> The UV spectrum of compound <b>1</b> in MeOH.	S11
17	<b>Figure S14.</b> The CD spectrum of compound <b>1</b> in MeOH.	S12
18	<b>Figure S15.</b> The IR spectrum of compound <b>1</b> .	S13
19	<b>Figure S16.</b> The ESI mass spectrum of compound <b>1</b> .	S14
20	<b>Figure S17.</b> The (+)-HR-ESI-MS report of compound <b>1</b> , page 1.	S15
21	<b>Figure S18.</b> The (+)-HR-ESI-MS report of compound <b>1</b> , page 2.	S16
22	<b>Figure S19.</b> The (+)-HR-ESI-MS report of compound <b>1</b> , page 3.	S17
23	<b>Figure S20.</b> The $^1\text{H}$ NMR spectrum of compound <b>1</b> in DMSO- $d_6$ (600 MHz).	S18
24	<b>Figure S21.</b> The $^{13}\text{C}$ NMR spectrum of compound <b>1</b> in DMSO- $d_6$ (150 MHz).	S19
25	<b>Figure S22.</b> The DEPT spectrum of compound <b>1</b> in DMSO- $d_6$ (150 MHz).	S20
26	<b>Figure S23.</b> The $^1\text{H}$ - $^1\text{H}$ COSY spectrum of compound <b>1</b> in DMSO- $d_6$ (600 MHz).	S21
27	<b>Figure S24.</b> The HSQC spectrum of compound <b>1</b> in DMSO- $d_6$ (600 MHz for $^1\text{H}$ ).	S22
28	<b>Figure S25.</b> The HMBC spectrum of compound <b>1</b> in DMSO- $d_6$ (600 MHz for $^1\text{H}$ ).	S23
29	<b>Figure S26.</b> The NOE difference spectrum of <b>1</b> in DMSO- $d_6$ (600 MHz).	S24
30	<b>Figure S27.</b> The UV spectrum of compound <b>2</b> in MeOH.	S25
31	<b>Figure S28.</b> The CD spectrum of compound <b>2</b> in MeOH.	S26
32	<b>Figure S29.</b> The IR spectrum of compound <b>2</b> .	S27
33	<b>Figure S30.</b> The ESI mass spectrum of compound <b>2</b> .	S28
34	<b>Figure S31.</b> The (+)-HR-ESI-MS report of compound <b>2</b> , page 1.	S29
35	<b>Figure S32.</b> The (+)-HR-ESI-MS report of compound <b>2</b> , page 2.	S30
36	<b>Figure S33.</b> The (+)-HR-ESI-MS report of compound <b>2</b> , page 3.	S31
37	<b>Figure S34.</b> The $^1\text{H}$ NMR spectrum of compound <b>2</b> in MeOH- $d_4$ (600 MHz).	S32
38	<b>Figure S35.</b> The $^{13}\text{C}$ NMR spectrum of compound <b>2</b> in MeOH- $d_4$ (150MHz).	S33
39	<b>Figure S36.</b> The DEPT spectrum of compound <b>2</b> in MeOH- $d_4$ (150 MHz).	S34
40	<b>Figure S37.</b> The $^1\text{H}$ - $^1\text{H}$ COSY spectrum of compound <b>2</b> in MeOH- $d_4$ (600 MHz).	S35
41	<b>Figure S38.</b> The HSQC spectrum of compound <b>2</b> in MeOH- $d_4$ (600 MHz for $^1\text{H}$ ).	S36

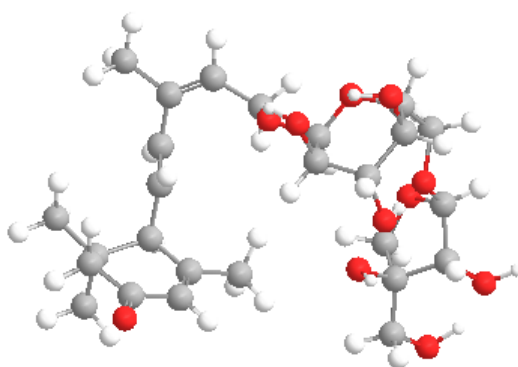
---

42	<b>Figure S39.</b> The HMBC spectrum of compound <b>2</b> in MeOH- <i>d</i> <sub>4</sub> (600 MHz for <sup>1</sup> H).	S37
43	<b>Figure S40.</b> The NOE difference spectrum of <b>2</b> in MeOH- <i>d</i> <sub>4</sub> (600 MHz).	S38
44	<b>Figure S41.</b> The <sup>1</sup> H NMR spectrum of compound <b>2a</b> in MeOH- <i>d</i> <sub>4</sub> (600 MHz).	S39
45	<b>Figure S42.</b> The UV spectrum of compound <b>3</b> in MeOH.	S40
46	<b>Figure S43.</b> The CD spectrum of compound <b>3</b> in MeOH.	S41
47	<b>Figure S44.</b> The IR spectrum of compound <b>3</b> .	S42
48	<b>Figure S45.</b> The ESI mass spectrum of compound <b>3</b> .	S43
49	<b>Figure S46.</b> The (+)-HR-ESI-MS report of compound <b>3</b> , page 1.	S44
50	<b>Figure S47.</b> The (+)-HR-ESI-MS report of compound <b>3</b> , page 2.	S45
51	<b>Figure S48.</b> The (+)-HR-ESI-MS report of compound <b>3</b> , page 3.	S46
52	<b>Figure S49.</b> The <sup>1</sup> H NMR spectra of compound <b>3</b> in acetone- <i>d</i> <sub>6</sub> (600 M).	S47
53	<b>Figure S50.</b> The <sup>13</sup> C NMR spectra of compound <b>3</b> in acetone- <i>d</i> <sub>6</sub> (150 M).	S48
54	<b>Figure S51.</b> The DEPT spectra of compound <b>3</b> in acetone- <i>d</i> <sub>6</sub> (150 M).	S49
55	<b>Figure S52.</b> The <sup>1</sup> H- <sup>1</sup> HgCOSY spectrum of compound <b>3</b> in acetone- <i>d</i> <sub>6</sub> (600 MHz).	S50
56	<b>Figure S53.</b> The gHSQC spectrum of compound <b>3</b> in acetone- <i>d</i> <sub>6</sub> (600 MHz for <sup>1</sup> H).	S51
57	<b>Figure S54.</b> The gHMBC spectrum of compound <b>3</b> in acetone- <i>d</i> <sub>6</sub> (600 MHz for <sup>1</sup> H).	S52
58	<b>Figure S55.</b> The NOE difference spectrum of <b>3</b> in acetone- <i>d</i> <sub>6</sub> (600 MHz).	S53
59	<b>Figure S56.</b> The <sup>1</sup> H NMR spectra of compound <b>3a</b> in acetone- <i>d</i> <sub>6</sub> (600 M).	S54
60	<b>Figure S57.</b> The CD spectrum of compound <b>3a</b> in MeOH.	S55
61	<b>Figure S58.</b> The <sup>1</sup> H NMR spectra of D-glucose hydrolyzed from <b>3</b> in D <sub>2</sub> O (600 M).	S56
62	<b>Figure S59.</b> The <sup>1</sup> H NMR spectra of the authentic D-glucose in D <sub>2</sub> O (600 M).	S57

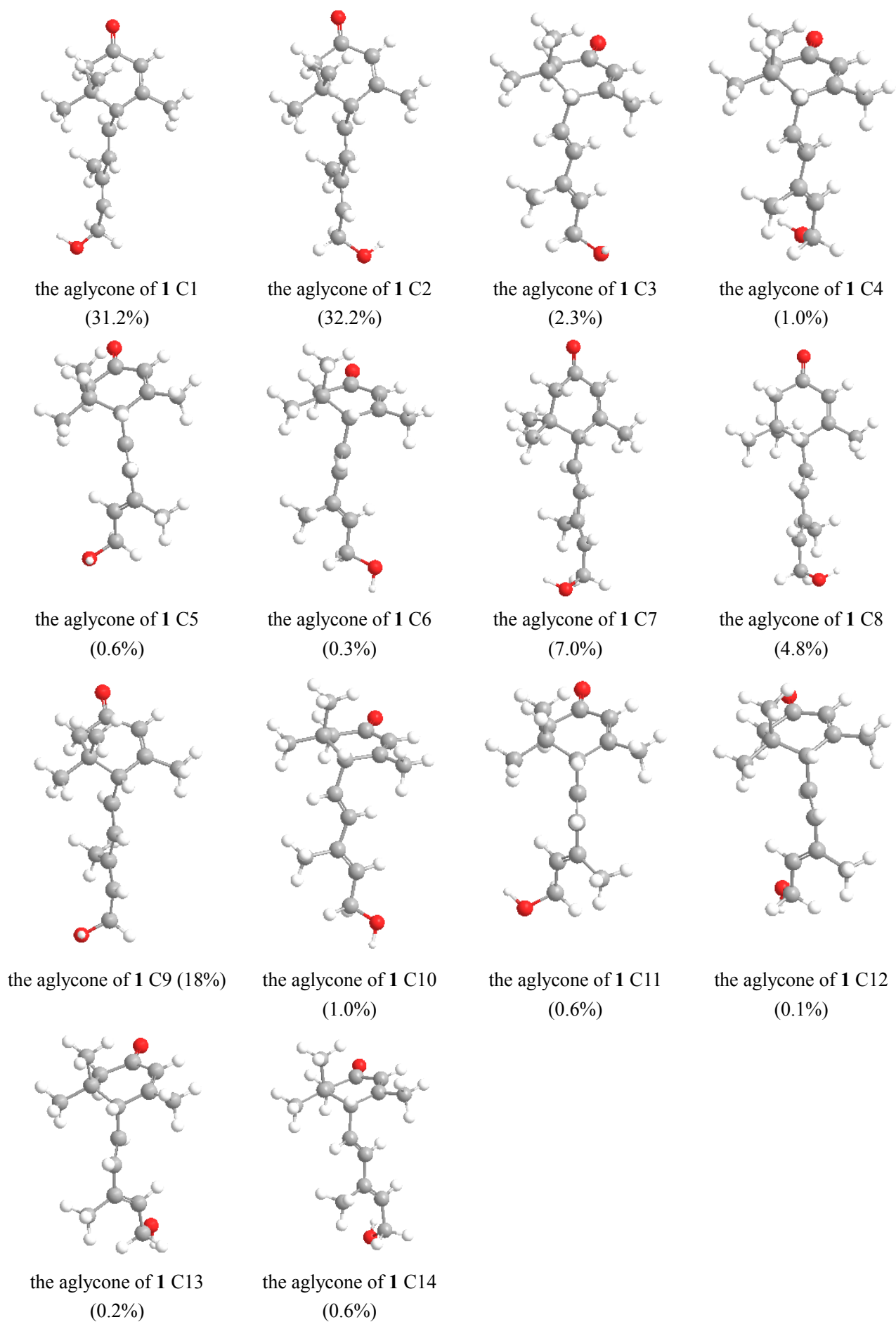
---

**Detailed ECD Calculation of 1–3, their aglycones, the model compounds, and (–)- $\alpha$ -barbatenal-reduction product.** Conformational analysis was performed by using the MMFF94 molecular mechanics force field calculation using the MOE (Molecular Operating Environment) software package.

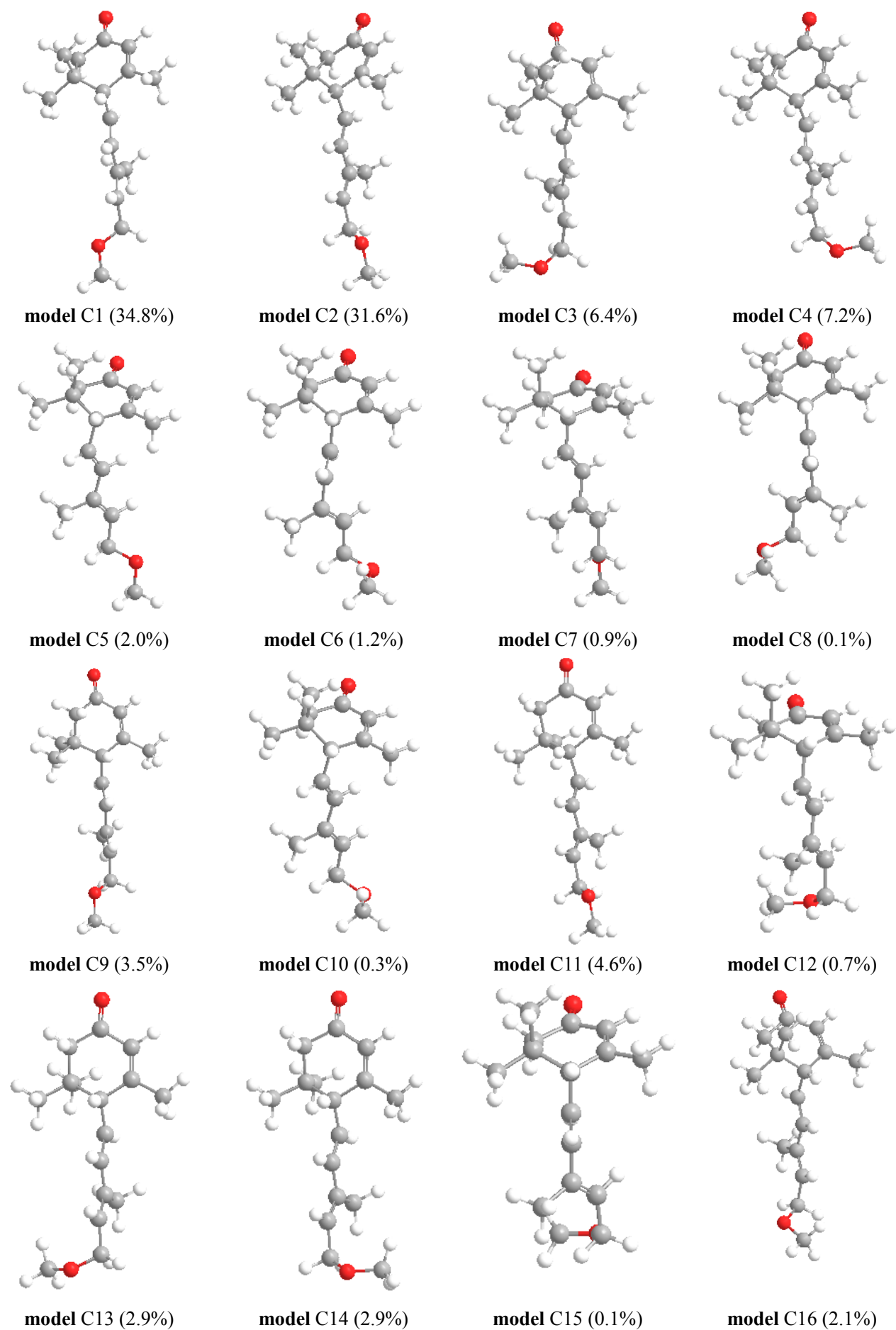
Conformational analysis showed conformer(s) for **1–3**, their aglycones, the model compounds, and (–)- $\alpha$ -barbatenal-reduction product having relative energy within 2 kcal/mol, which were considered for further DFT calculations. Subsequently, using Gaussian 09 program package, the conformers were re-optimized using DFT at the B3LYP/6-31+G(d) level, and conductor-like polarizable continuum model (CPCM) was adopted to consider solvent effects using the dielectric constant of MeOH ( $\epsilon = 32.6$ ). The B3LYP/6-31G+(d) harmonic vibrational frequencies were calculated to confirm their stability. The energies, oscillator strengths, and rotational strengths of the first 30 electronic excitations of the conformers were calculated using the TDDFT methodology at the B3LYP/6-311++G (2d,2p) level in gas phase, and conformer(s) for **1–3**, the aglycones, the model compounds, and (–)- $\alpha$ -barbatenal-reduction product (Figures S1–S3, S5–S7, S9, S10, and S12) showed relative Gibbs free energies ( $\Delta G$ ) under 2 kcal/mol. The ECD spectra of the conformers were then simulated using the Gaussian function ( $\sigma = 0.28$  eV), respectively. The final spectra of the compounds (Figures S4, S8, and S11) were obtained by averaging the simulated spectra of the lowest energy conformers according to the Boltzmann distribution theory, in which their Gibbs free energy (G) were adopted.



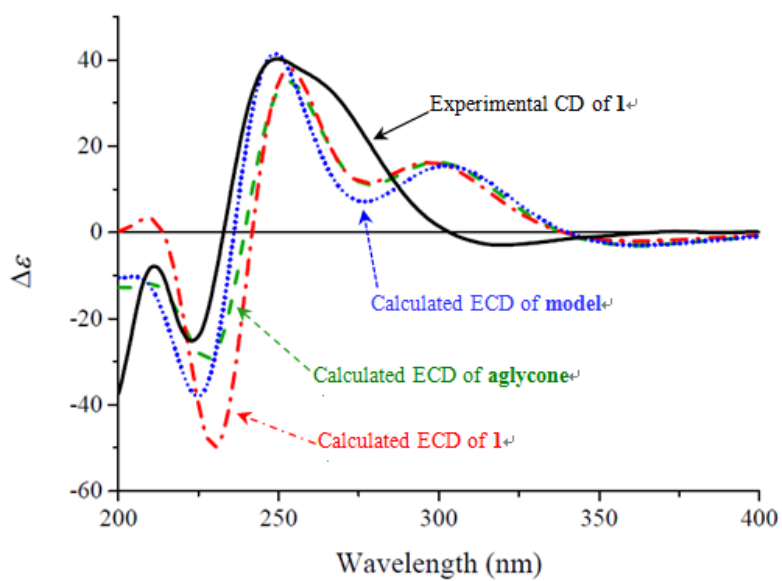
**Figure S1.** The re-optimized conformer of **1**.



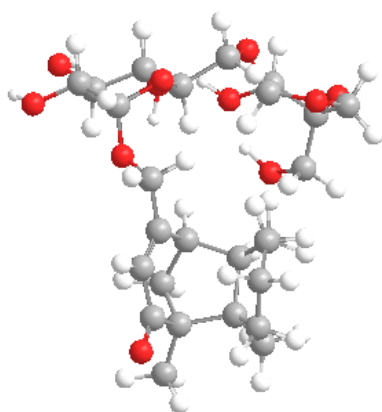
**Figure S2.** The re-optimized fourteen conformers of the aglycone of **1** and their equilibrium populations.



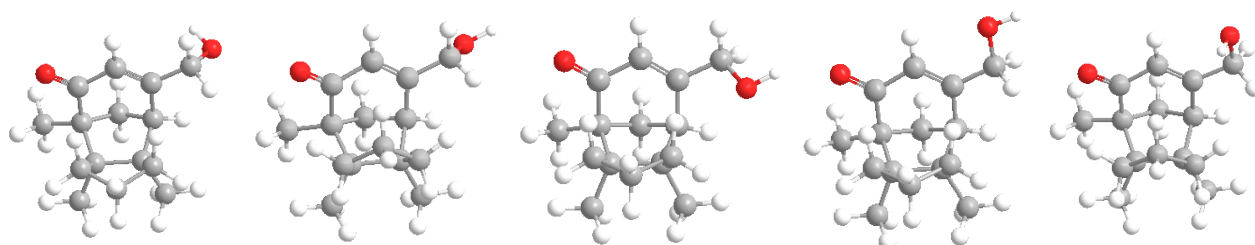
**Figure S3.** The re-optimized sixteen conformers of the **model** and their equilibrium populations.



**Figure 4.** The experimental CD spectrum of **1** (black) and the calculated ECD spectra of **1** (dash dotted red), its aglycone (dashed green), and the **model** (dotted blue) in MeOH.

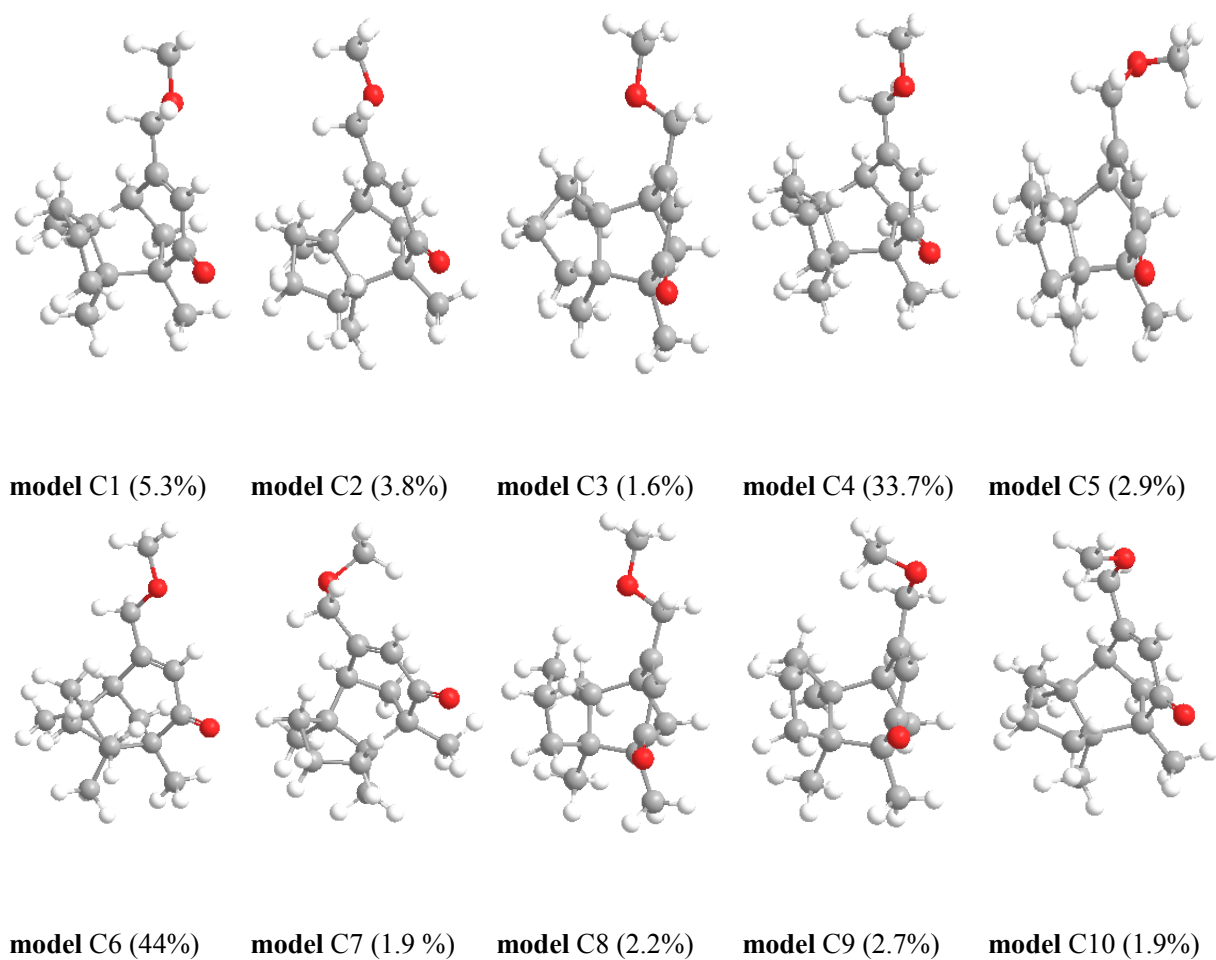


**Figure S5.** The re-optimized conformer of **2**.

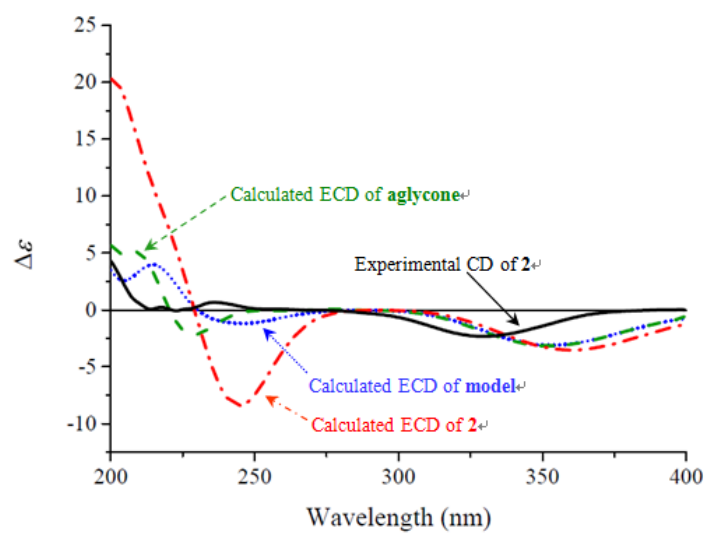


the aglycone of **2** C1 (8.8%)    the aglycone of **2** C2 (4.5%)    the aglycone of **2** C3 (2.0%)    the aglycone of **2** C4 (30.6%)    the aglycone of **2** C5 (54.0%)

**Figure S6.** The re-optimized five conformers of the aglycone of **2** and their equilibrium populations.

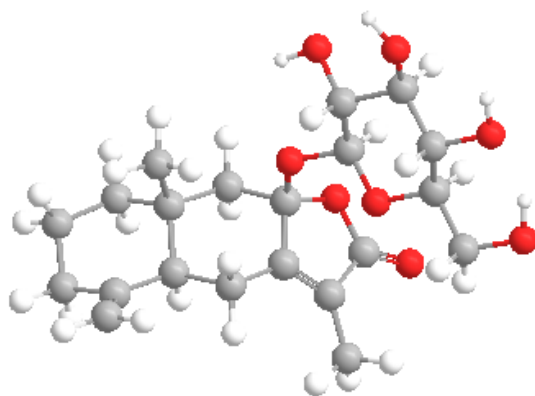


**Figure S7.** The re-optimized ten conformers of the **model** of **2** and their equilibrium populations.

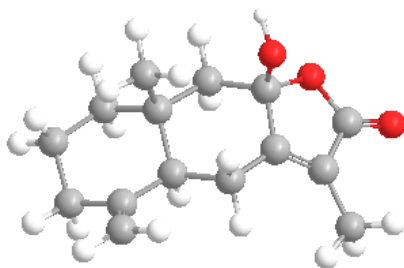


**Figure 8.** The experimental CD spectrum of **2** (black) and the calculated ECD spectra of **2** (dash dotted red), its aglycone (dashed green), and the **model** (dotted blue) in MeOH.

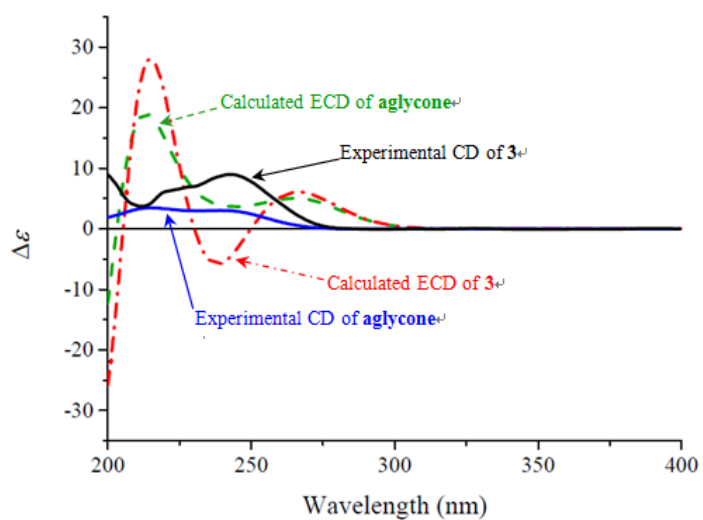




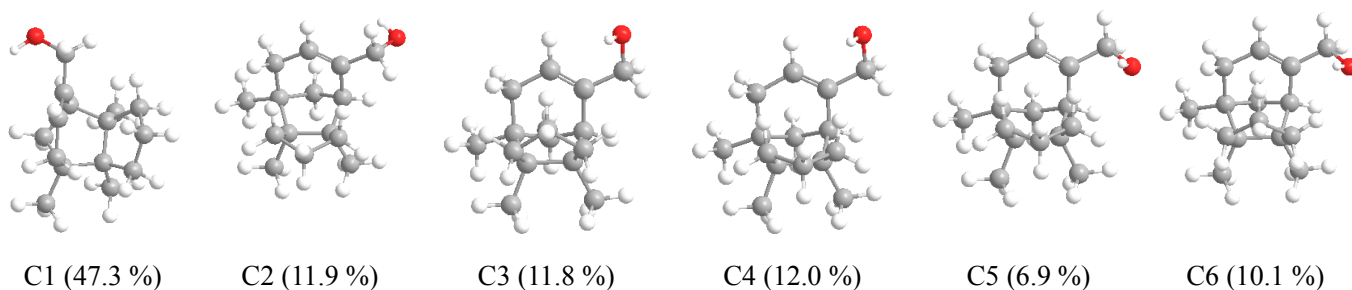
**Figure S9.** The re-optimized conformer of **3**.



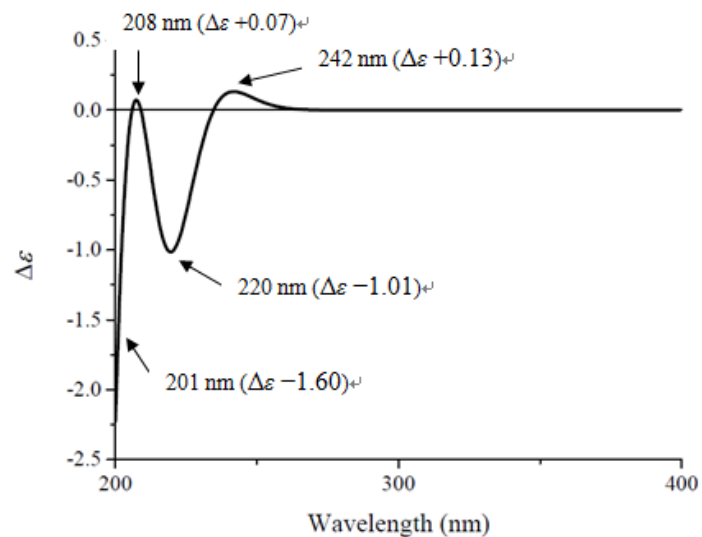
**Figure S10.** The re-optimized conformer of the aglycone of **3**.



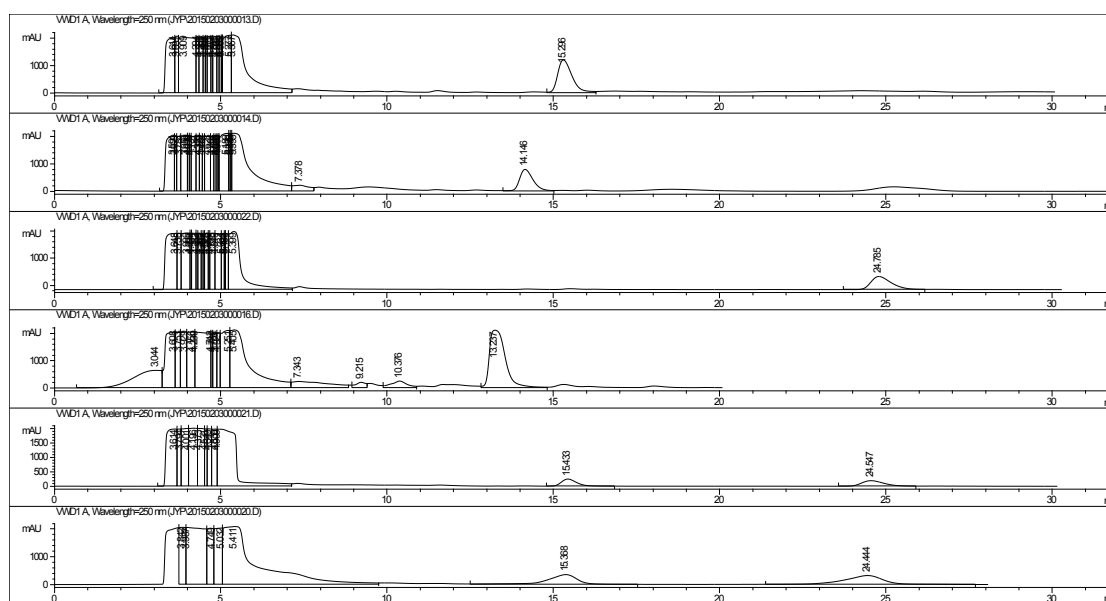
**Figure S11.** The experimental CD (black and blue) and calculated ECD (dashed and dotted red and green) spectra of **3** and its aglycone in MeOH.



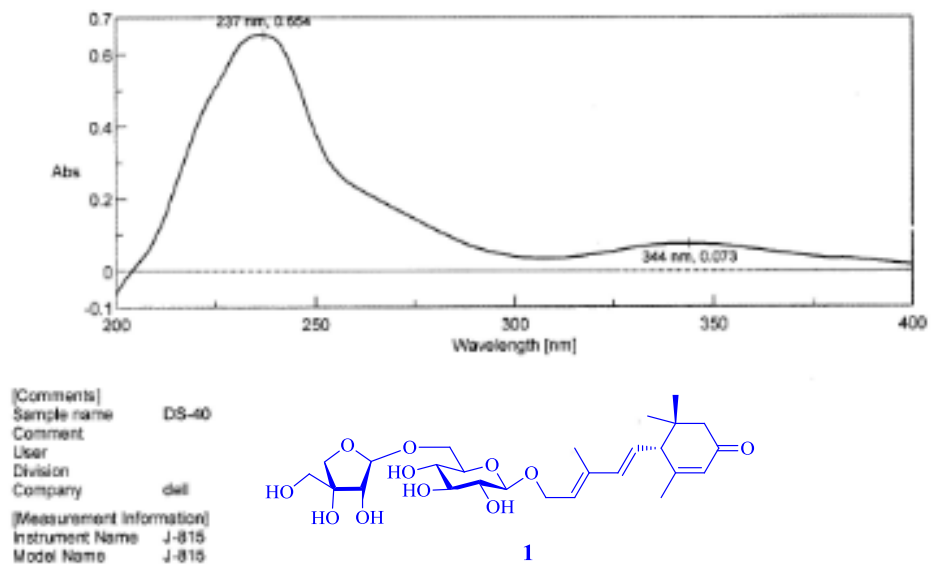
**Figure S12.** The reoptimized six conformers of (-)- $\alpha$ -barbatenol-reduction product.



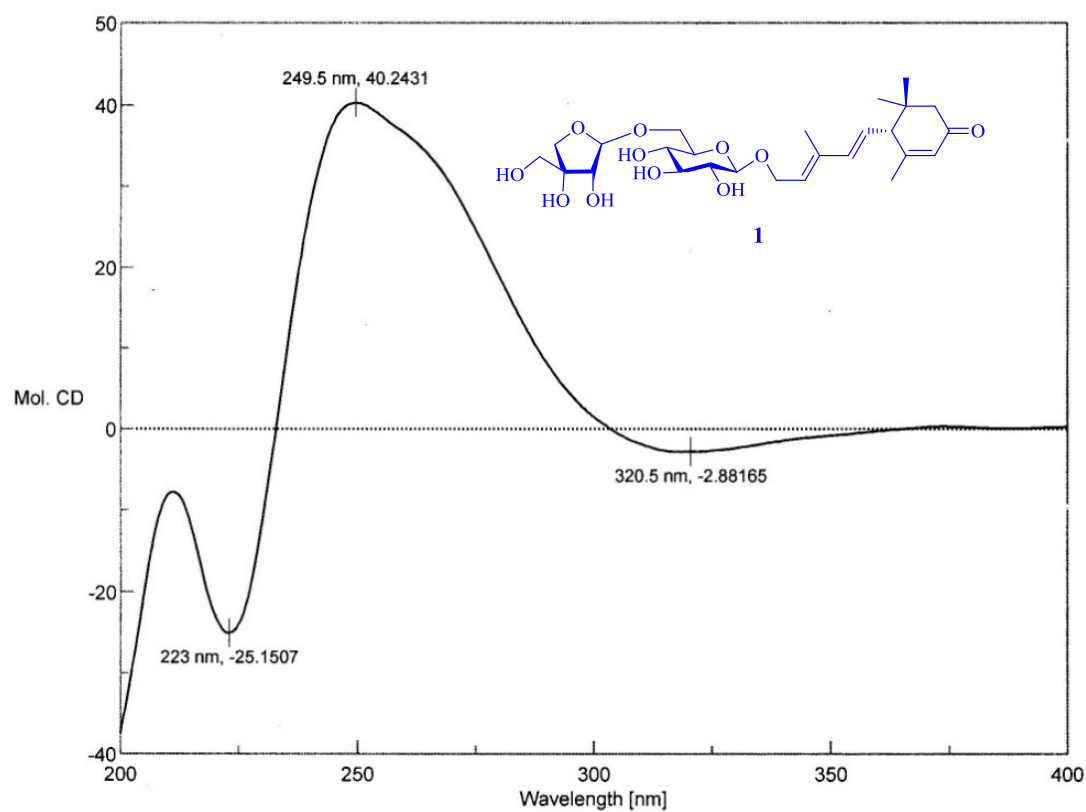
**Figure S13.** The calculated ECD spectrum of (-)- $\alpha$ -barbatenol-reduction product



**Figure S12.** The HPLC chromatograms of the sugar derivatives, from top to bottom, for D-glucose, L-glucose, D-apirose, L-apirose, and sugars from hydrolysates of **1** and **2**.



**Figure S13.** The UV spectrum of compound **1** in MeOH.



[Comments]  
 Sample name DS-40  
 Comment  
 User  
 Division  
 Company dell

[Measurement Information]  
 Instrument Name J-815  
 Model Name J-815  
 Serial No. A024461168

Accessory Standard  
 Accessory S/N A024461168  
 Cell Length 1 mm

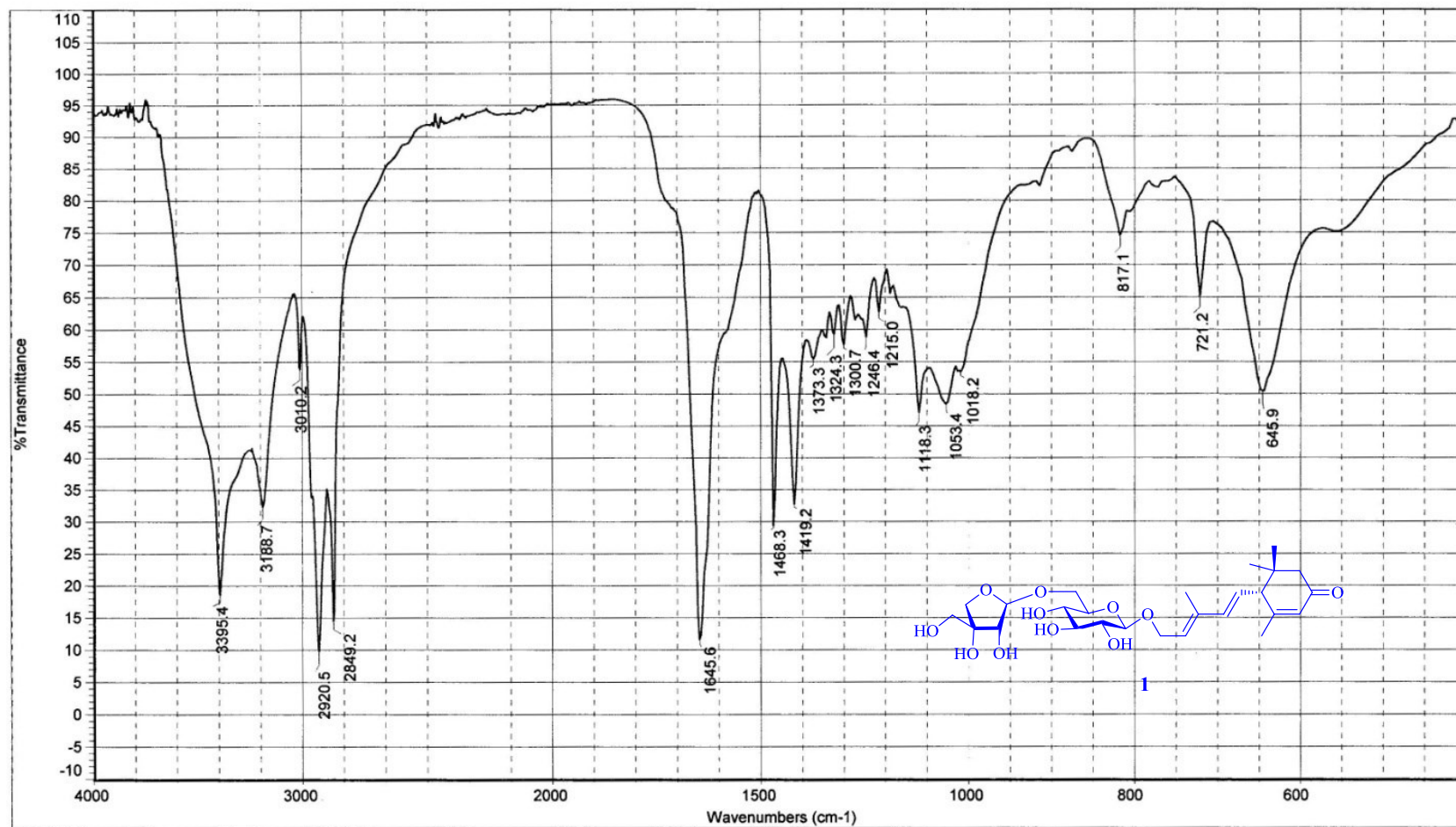
Photometric Mode CD, HT, Abs  
 Measure Range 400 - 200 nm  
 Data pitch 0.5 nm  
 Sensitivity Standard  
 D.I.T. 2 sec  
 Band width 2.00 nm  
 Start Mode Immediately  
 Scanning Speed 100 nm/min  
 Baseline Correction Baseline  
 Shutter Control Auto  
 PMT Voltage Auto  
 Accumulations 3  
 Solvent MEOH  
 Concentration 0.07 (w/v)%

[Detailed Information]

Creation date 2014-5-20 9:34

Data array type Linear data array \* 3  
 Horizontal axis Wavelength [nm]  
 Vertical axis(1) Mol. CD  
 Vertical axis(2) HT [V]  
 Vertical axis(3) Abs  
 Start 400 nm  
 End 200 nm  
 Data interval 0.5 nm  
 Data points 401

Figure S14. The CD spectrum of compound 1 in MeOH.



日期: 星期四 6月 05 12:11:44 2014 (GMT+08:00) Sample Name: DS - 40 (显微镜透射法FT- IR Microscope Transmission)

扫描次数: 100

傅里叶变换显微镜红外(FT-IR Microscope): Centaurus

分辨率: 8.000

美国热电公司(Thermo)傅里叶变换红外光谱仪:Nicolet 5700

Figure S15. The IR spectrum of compound 1.

# Single Mass Spectrum Deconvolution Report

**Analysis Name:** jngyp103.d    **Instrument:** LC-MSD-Trap-SL    **Print Date:** 12/23/2013 12:41:42 PM  
**Method:** TEST.MS    **Operator:** Operator    **Acq. Date:** 12/23/2013 11:42:49 AM  
**Sample Name:** DS-40  
**Analysis Info:**

## Acquisition Parameter:

Mass Range Mode	Std/Normal	Trap Drive	36.3	Scan Begin	150 m/z
Ion Polarity	Positive	Octopole RF Amplitude	171.0 Vpp	Scan End	700 m/z
Ion Source Type	ESI	Capillary Exit	106.0 Volt	Averages	5 Spectra
Dry Temp (Set)	330 °C	Skimmer	40.0 Volt	Max. Accu Time	200000 µs
Nebulizer (Set)	15.00 psi	Oct 1 DC	12.00 Volt	ICC Target	20000
Dry Gas (Set)	6.00 l/min	Oct 2 DC	1.70 Volt	Charge Control	on

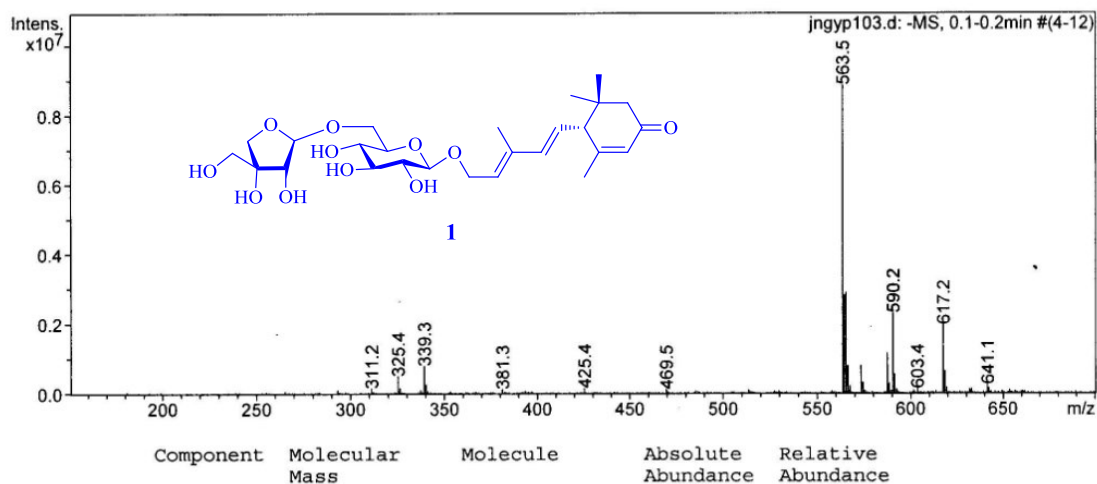
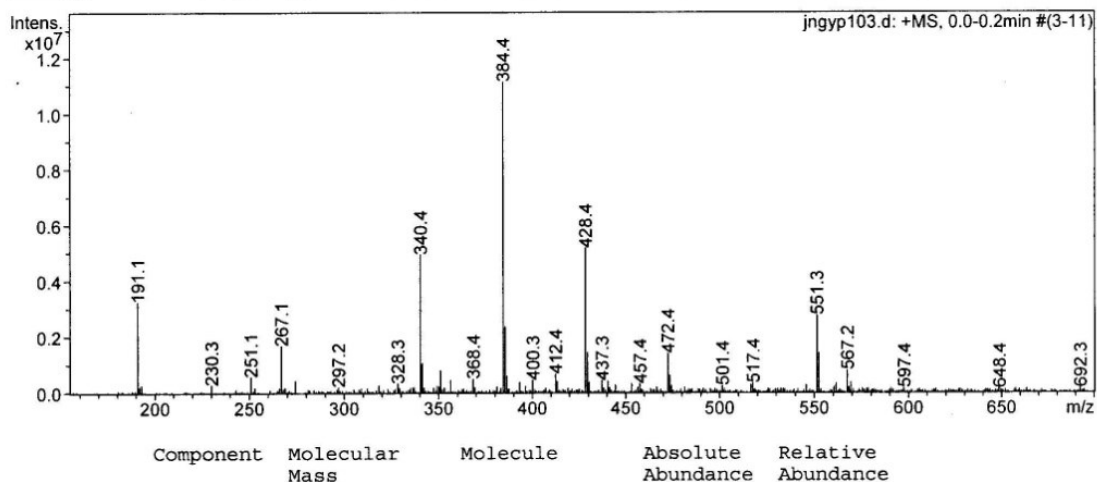


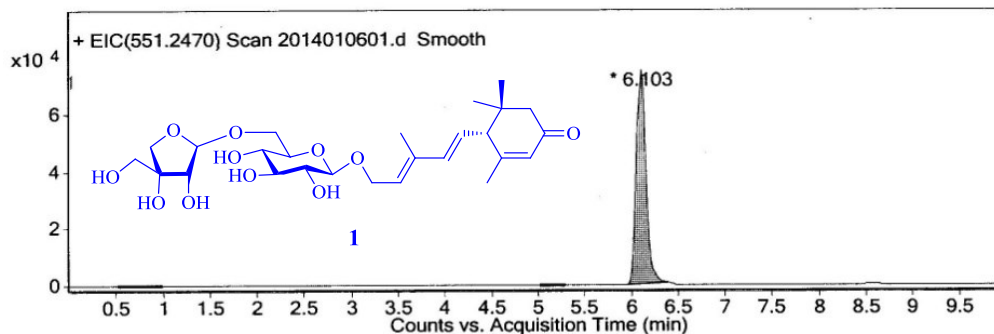
Figure S16. The ESI mass spectrum of compound 1.

# Qualitative Analysis Report

Data Filename	2014010601.d	Sample Name	DS-40
Sample Type	Sample	Position	P1-C3
Instrument Name	Instrument 1	User Name	
Acq Method		IRM Calibration Status	XXXXXXXXXX
DA Method	TEST LCMS.m	Comment	

## User Chromatograms

Fragmentor Voltage 135    Collision Energy 0    Ionization Mode ESI



### Integration Peak List

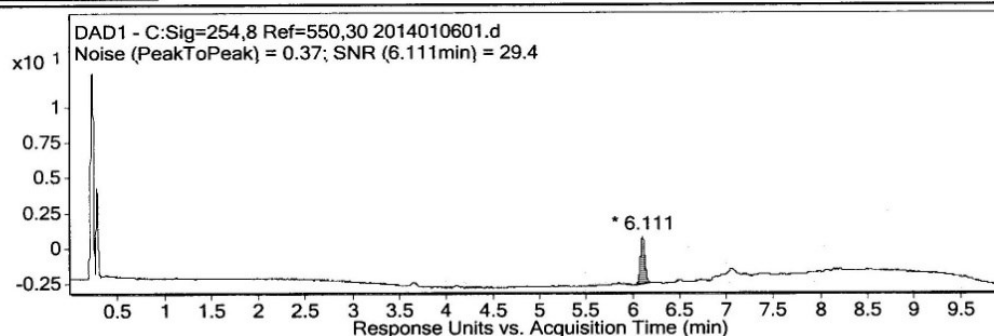
Peak	Start	RT	End	Height	Area	Area %	Signal To Noise
1	5.958	6.103	6.393	75115	534180	100	Infinity

### Noise Measurements

Noise Type	Signal Definition	Noise Multiplier	Noise Value
Peak-to-Peak	Area	1	0

### Noise Regions

Start	End
0.5	1
5	5.3
9.99	11



### Integration Peak List

Peak	Start	RT	End	Height	Area	Area %	Signal To Noise
1	6.016	6.111	6.189	3.35	10.84	100	29.4

### Noise Measurements

Noise Type	Signal Definition	Noise Multiplier	Noise Value
Peak-to-Peak	Area	1	0.36907196

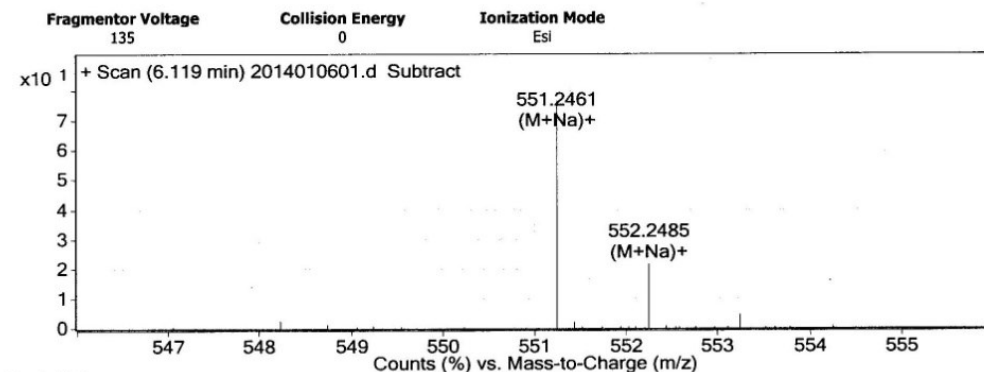
### Noise Regions

Start	End
0.5	1
5	5.3
9.99	11

## User Spectra

Figure S17. The (+)-HR-ESI-MS report of compound 1, page 1.

# Qualitative Analysis Report

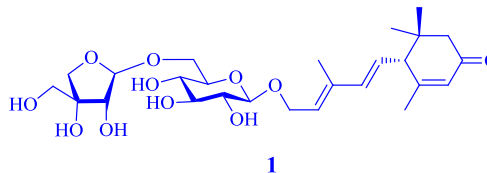


**Peak List**

m/z	z	Abund	Formula	Ion
101.0083		39257		
116.9859		43150		
133.0342	1	60415		
133.0966		39551		
277.128	1	90968		
284.1122	2	35480		
294.1545		45155		
299.1104	1	133236		
551.2461	1	102137	C26 H40 Na O11	(M+Na)+
597.2391	1	34096		

**Formula Calculator Element Limits**

Element	Min	Max
C	3	100
H	0	500
O	0	90
N	0	5
S	0	2
Cl	0	0
Br	0	0
Si	0	0
F	0	0
P	0	0



**Formula Calculator Results**

Formula	Best	Mass	Tgt Mass	Diff (ppm)	Ion Species	Score
C26 H40 O11	TRUE	528.2568	528.2571	0.43	C26 H40 Na O11	99.96
C27 H36 N4 O7		528.2568	528.2584	2.95	C27 H36 N4 Na O7	99.65
C30 H40 O6 S		528.2568	528.2546	-4.31	C30 H40 Na O6 S	98.51
C31 H36 N4 O2 S		528.2568	528.2559	-1.79	C31 H36 N4 Na O2 S	98.39
C27 H44 O6 S2		528.2568	528.2579	2.06	C27 H44 Na O6 S2	97.39
C28 H40 N4 O2 S2		528.2568	528.2593	4.58	C28 H40 N4 Na O2 S2	97.19
C22 H44 N2 O8 S2		528.2568	528.2539	-5.56	C22 H44 N2 Na O8 S2	96.83
C36 H36 N2 S		528.2568	528.2599	5.83	C36 H36 N2 Na S	96.57
C39 H32 N2		528.2568	528.2565	-0.55	C39 H32 N2 Na	96.3

--- End Of Report ---

Figure S18. The (+)-HR-ESI-MS report of compound 1, page 2.



MS Formula Results: + Scan (6.119 min) Sub (2014010601.d)

m/z	Ion	Formula	Abundance
551.2461	(M+Na) <sup>+</sup>	C26 H40 Na O11	102137

Best	Formula (M)	Ion Formula	Calc m/z	Score	Cross S	Mass	Calc Mass	Diff (ppm)	Abs Diff (ppm)	Abund Match	Spacing Mat	Mass Match	m/z	DBE
<input checked="" type="checkbox"/>	C26 H40 O11	C26 H40 Na O11	551.2463	99.96		528.2568	528.2571	0.43	0.43	99.93	99.91	99.99	551.2461	7
<input type="checkbox"/>	C27 H36 N4 O7	C27 H36 N4 Na O7	551.2476	99.65		528.2568	528.2584	2.95	2.95	99.26	99.96	99.72	551.2461	12
<input type="checkbox"/>	C30 H40 O6 S	C30 H40 Na O6 S	551.2438	98.51		528.2568	528.2546	-4.31	4.31	95.98	99.77	99.41	551.2461	11
<input type="checkbox"/>	C31 H36 N4 O2 S	C31 H36 N4 Na O2 S	551.2451	98.39		528.2568	528.2559	-1.79	1.79	94.69	99.8	99.9	551.2461	16
<input type="checkbox"/>	C27 H44 O6 S2	C27 H44 Na O6 S2	551.2472	97.39		528.2568	528.2579	2.06	2.06	91.51	99.52	98.86	551.2461	6
<input type="checkbox"/>	C28 H40 N4 O2 S2	C28 H40 N4 Na O2 S2	551.2485	97.19		528.2568	528.2593	4.58	4.58	91.67	99.54	99.33	551.2461	11
<input type="checkbox"/>	C22 H44 N2 O8 S2	C22 H44 N2 Na O8 S2	551.2431	96.83		528.2568	528.2539	-5.56	5.56	91.01	99.45	99.02	551.2461	2
<input type="checkbox"/>	C36 H36 N2 S	C36 H36 N2 Na S	551.2491	96.57		528.2568	528.2599	5.83	5.83	89.94	99.84	98.92	551.2461	20
<input type="checkbox"/>	C39 H32 N2	C39 H32 N2 Na	551.2458	96.3		528.2568	528.2565	-0.55	0.55	87.12	99.92	99.99	551.2461	25

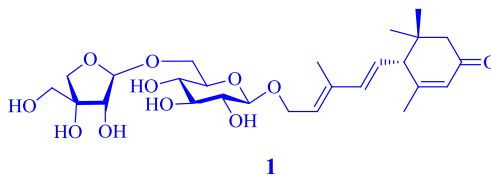


Figure S19. The (+)-HR-ESI-MS report of compound **1**, page 3.

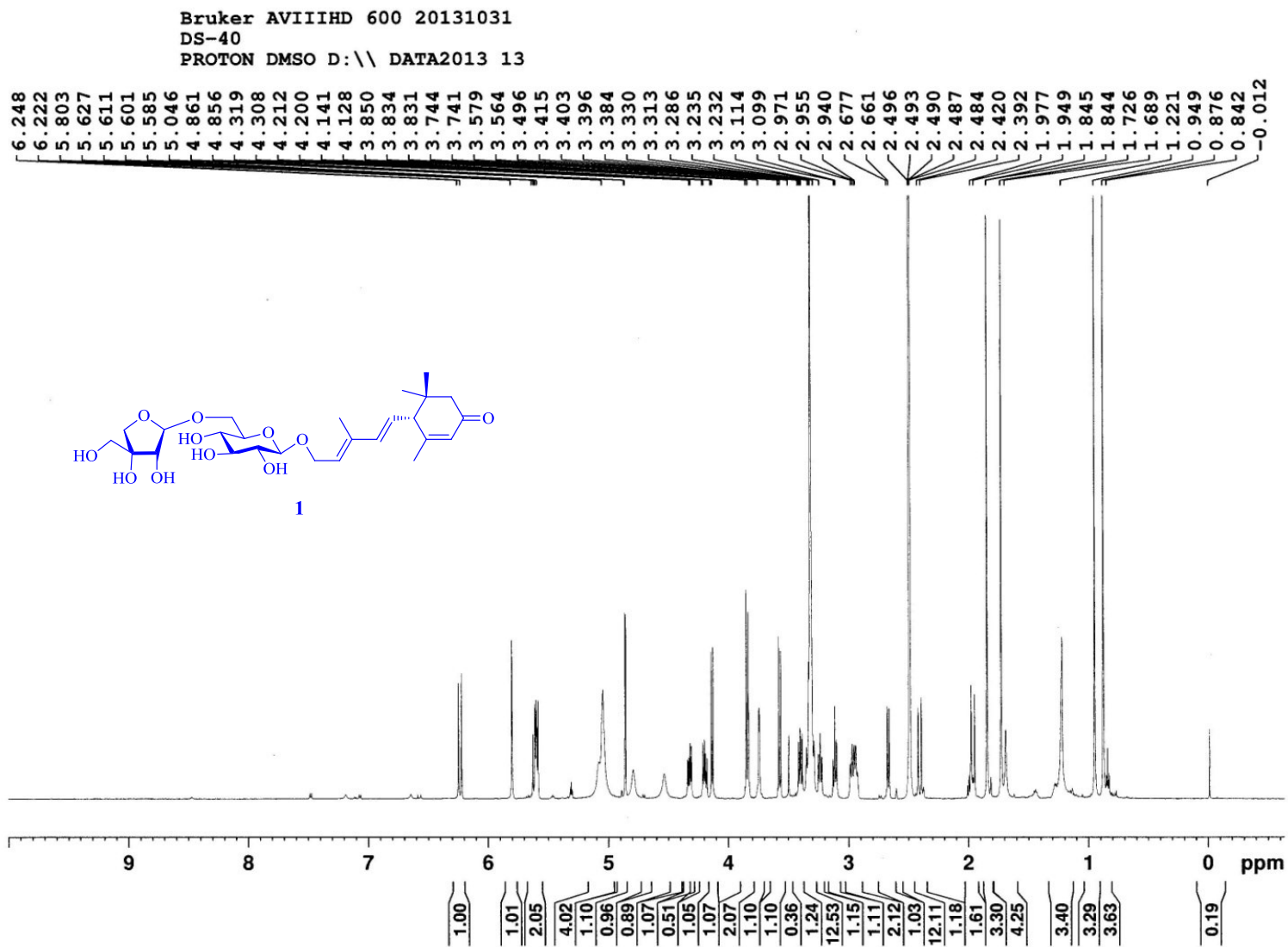


Figure S20. The  $^1\text{H}$  NMR spectrum of compound **1** in  $\text{DMSO-}d_6$  (600 MHz).

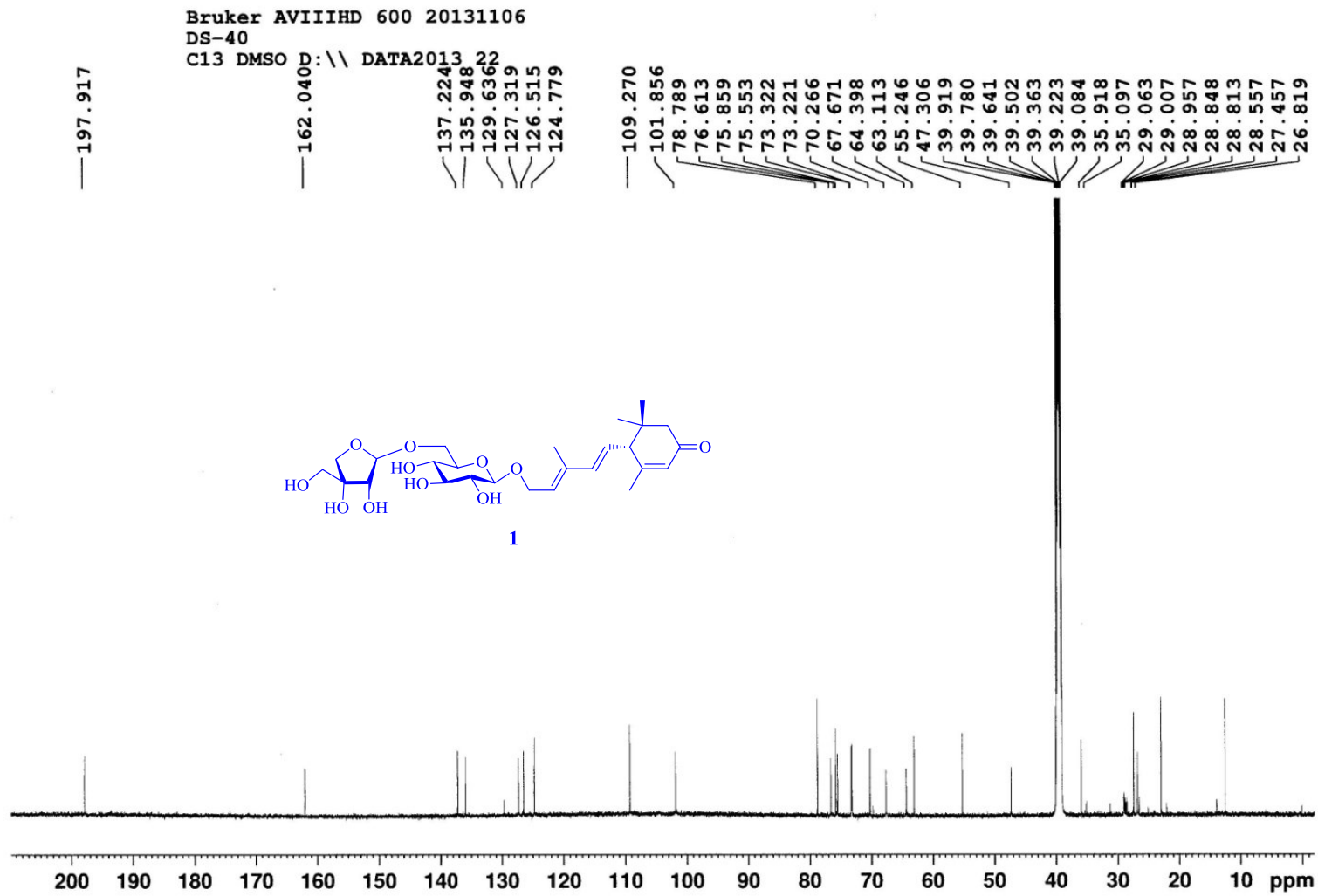


Figure S21. The  $^{13}\text{C}$  NMR spectrum of compound **1** in  $\text{DMSO-}d_6$  (150 MHz).



Bruker AVIIIHD 600 20131106  
DS-40 DEPT DMSO D:\ \ DATA2013 22

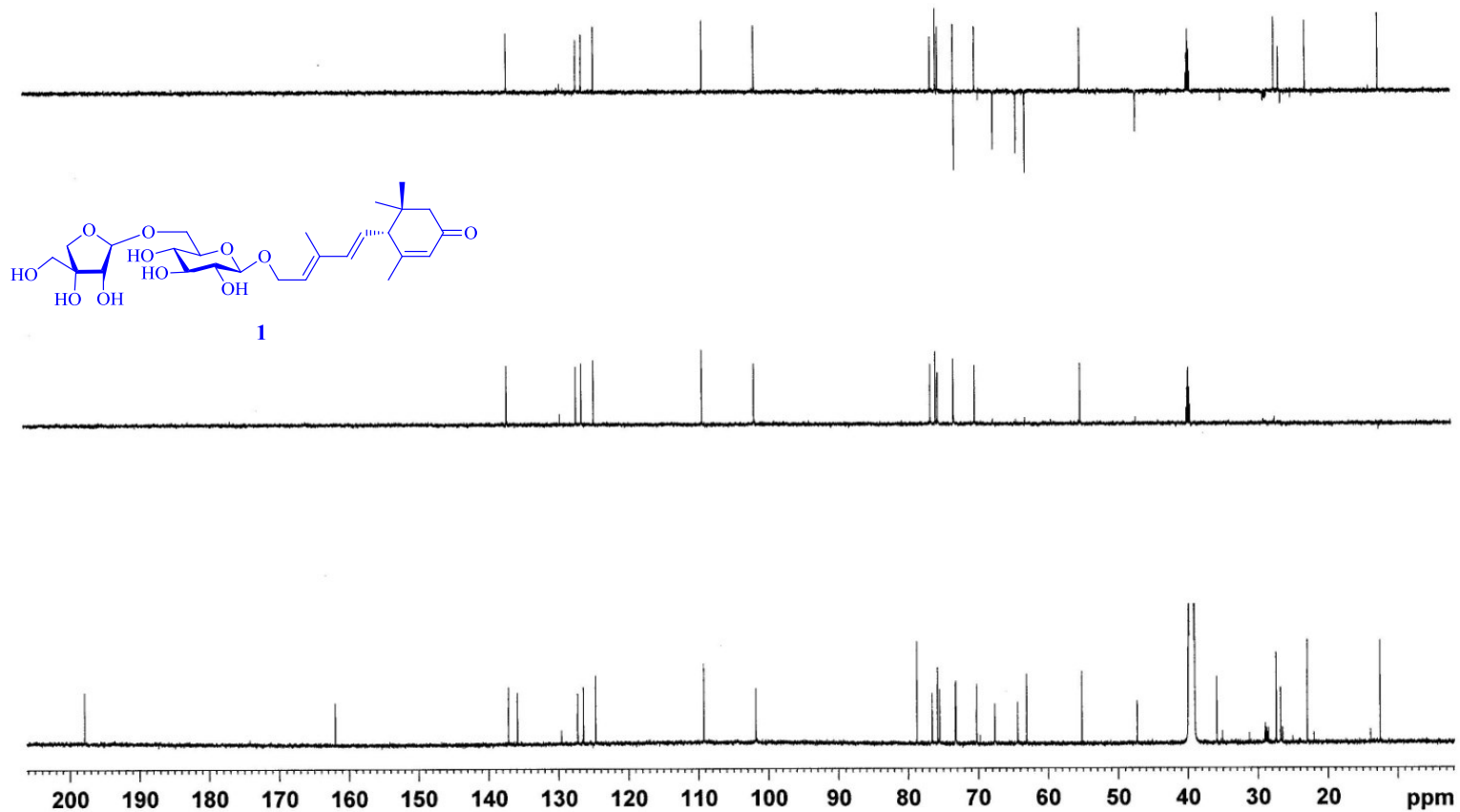


Figure S22. The DEPT spectrum of compound 1 in DMSO- $d_6$  (150 MHz).

Bruker AVIIIHD 600 20131121  
DS-40  
{H-H COSY} DMSO D:\ DATA2013 24

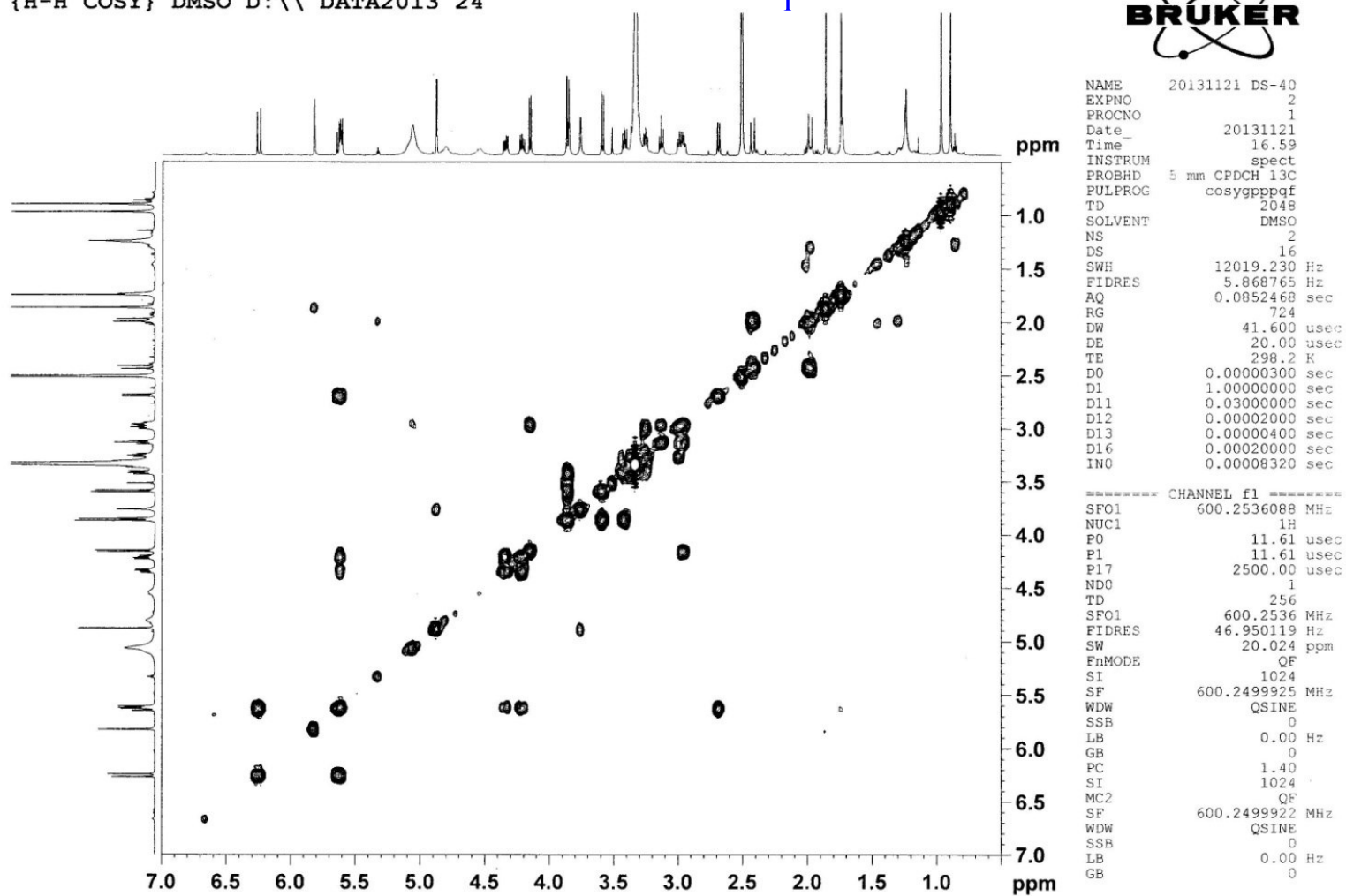
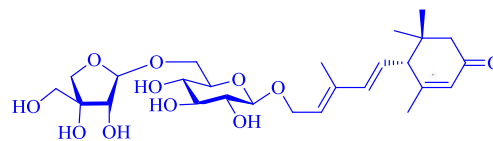


Figure S23. The  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of compound 1 in DMSO- $d_6$  (600 MHz).

Bruker AVIIIHD 600 20131121  
 DS-40  
 HSQC DMSO D:\ DATA2013 24

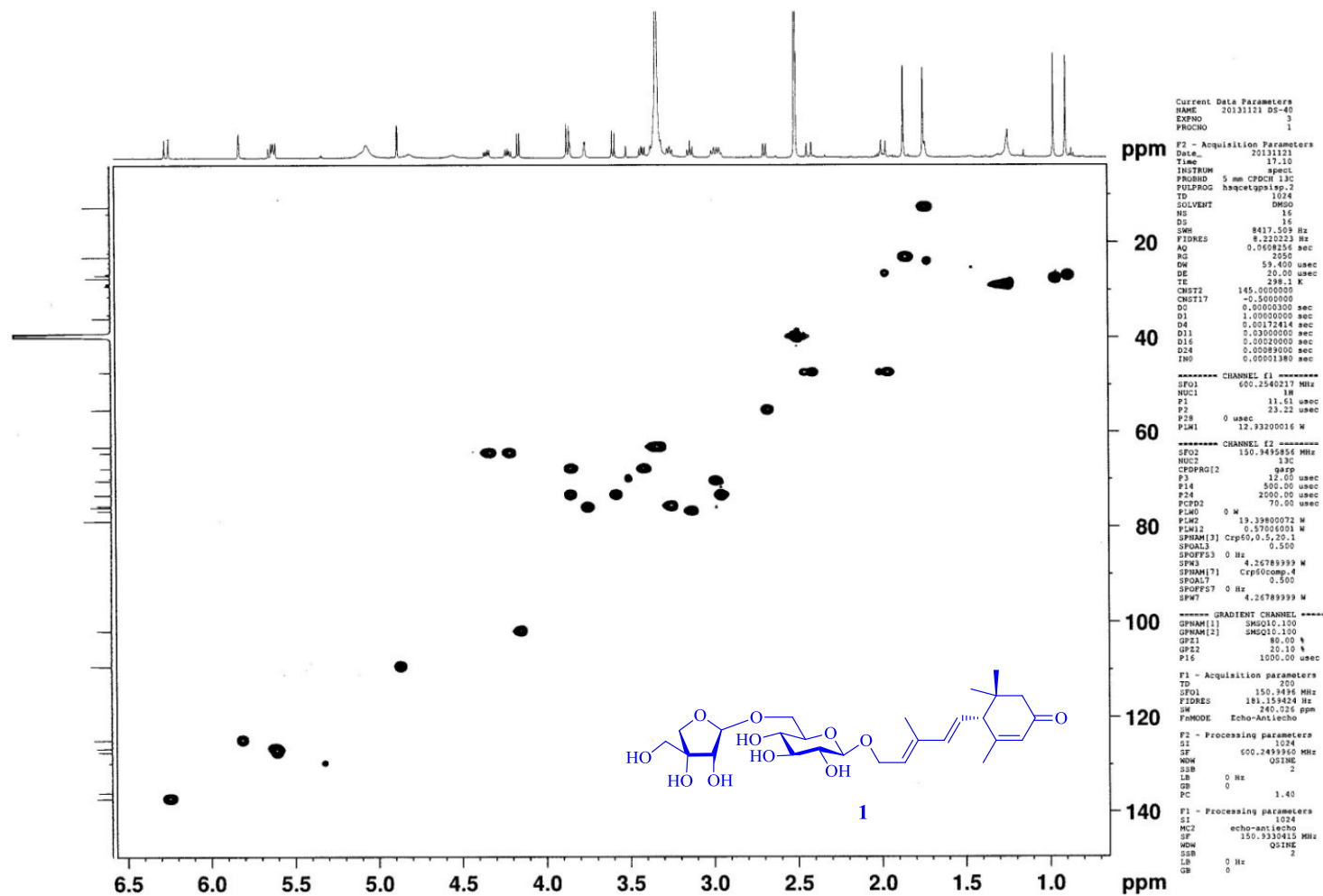


Figure S24. The HSQC spectrum of compound 1 in DMSO- $d_6$  (600 MHz for  $^1\text{H}$ ).

Bruker AVIIIHD 600 20131121  
DS-40  
HMBC DMSO D:\ DATA2013 24

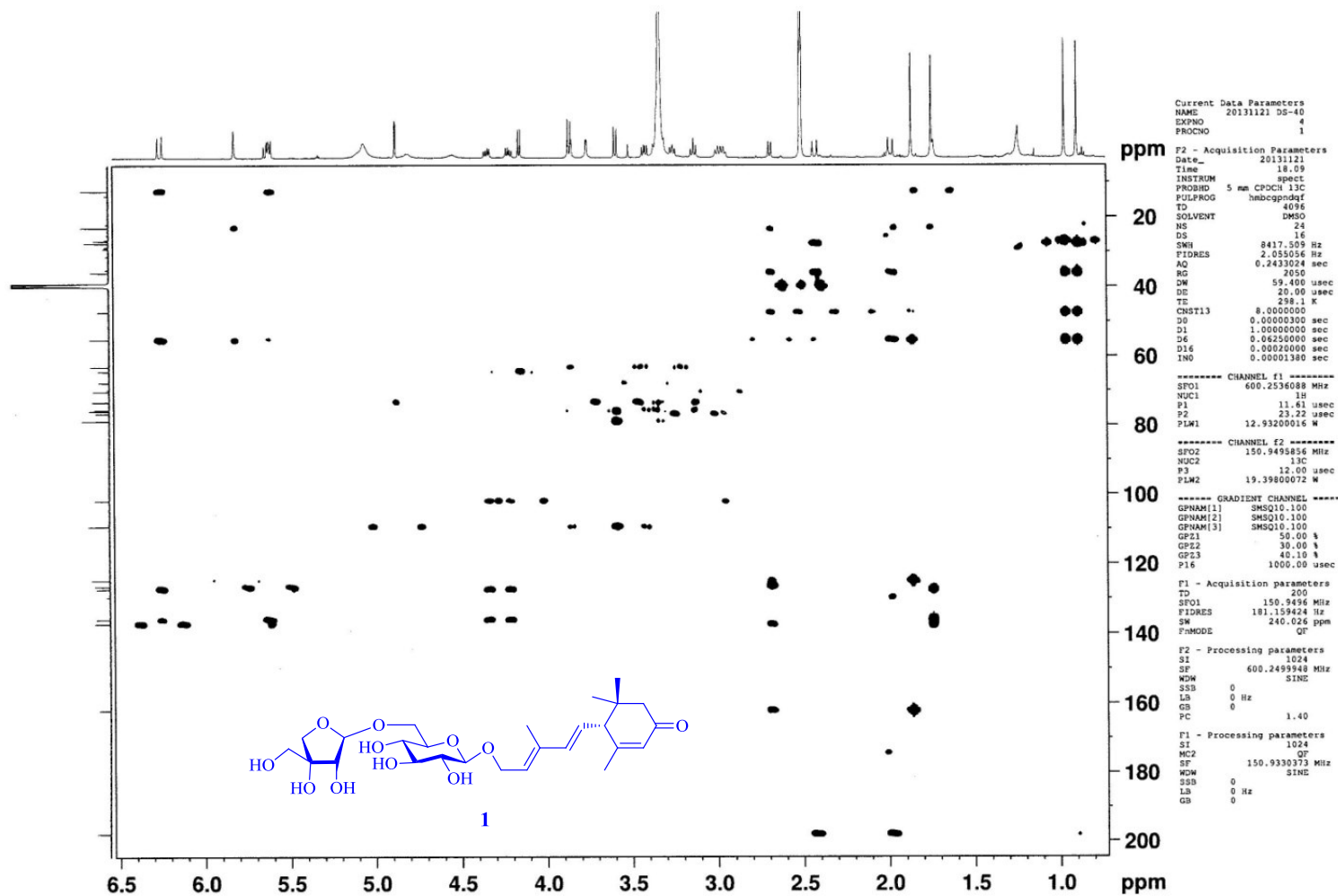
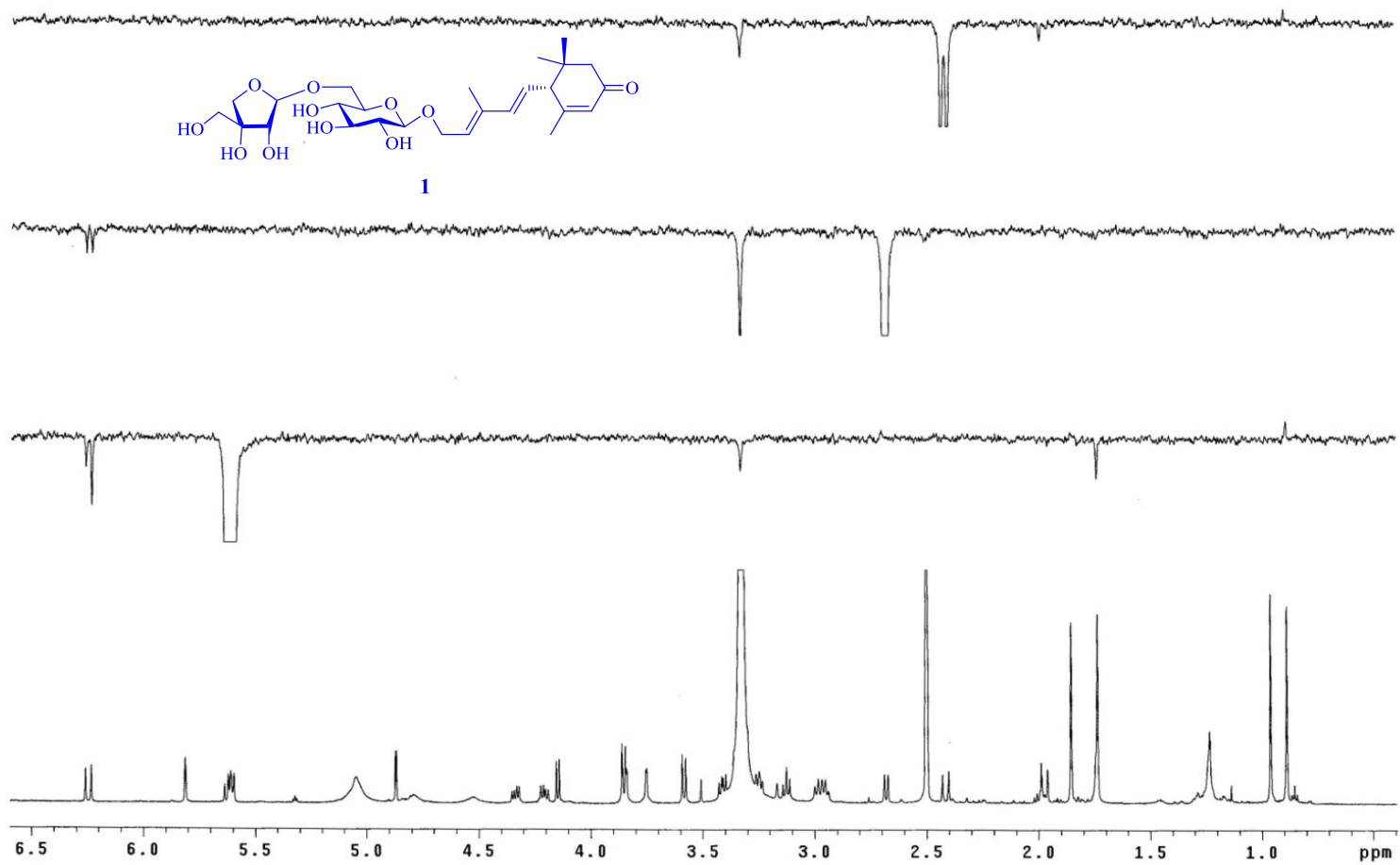
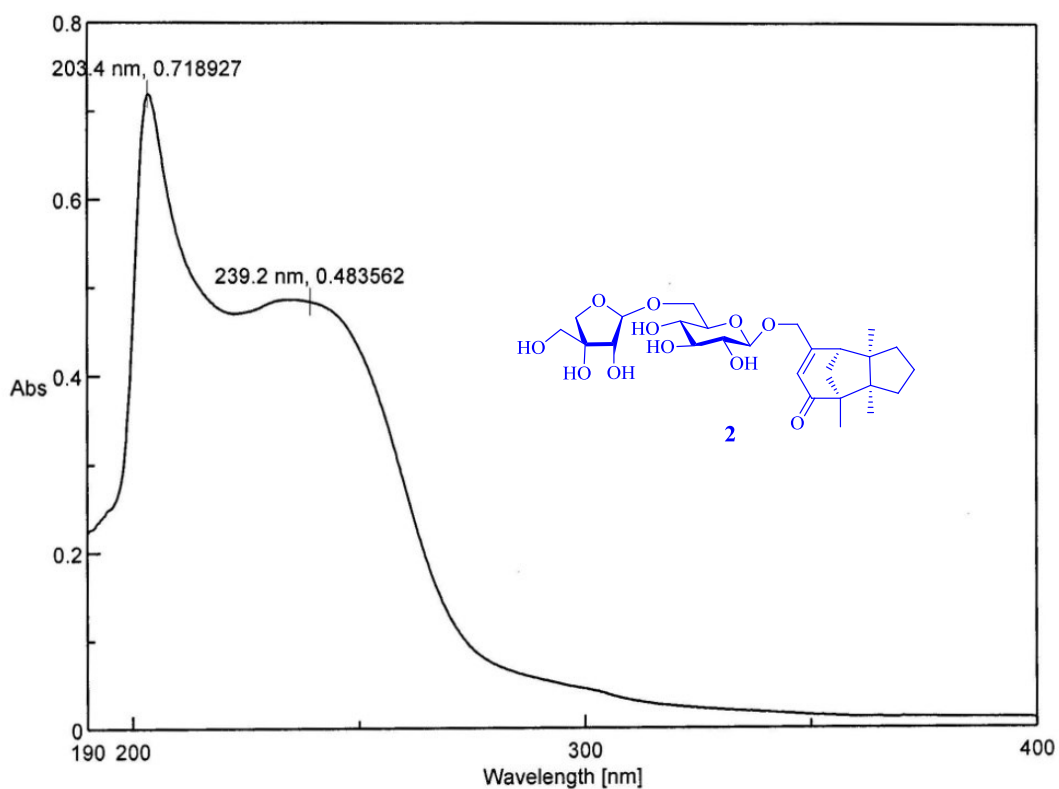


Figure S25. The HMBC spectrum of compound 1 in DMSO- $d_6$  (600 MHz for  $^1\text{H}$ ).



**Figure S26.** The NOE difference spectrum of **1** in DMSO-*d*<sub>6</sub> (600 MHz).





[Comment]  
 Sample Name ds-77b  
 Comment 0.02  
 User  
 Division UV  
 Company 324  
 [Measurement Information]  
 Instrument Name V-650  
 Model Name V-650  
 Serial No. A034461150

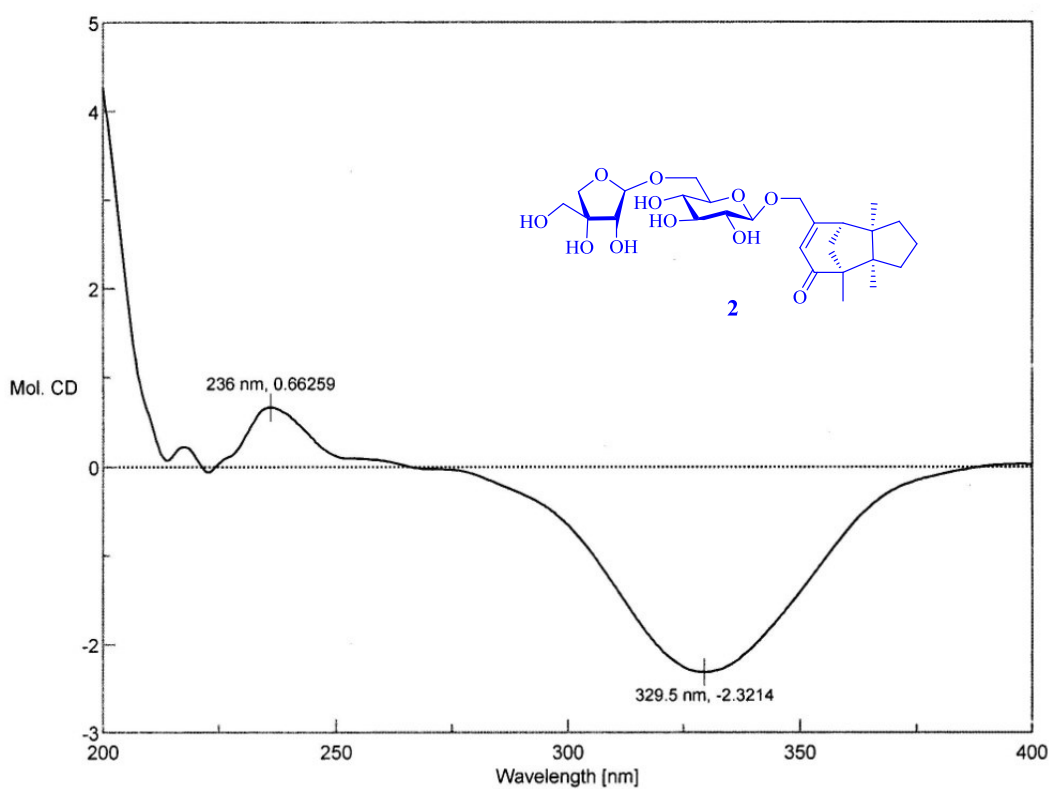
Accessory PSC-718  
 Accessory S/N A001761114  
 Position 1  
 Cell Length 10 mm  
 Temperature 19.97 C  
 Control Sensor Holder  
 Monitor Sensor Holder  
 Start Mode Start immediately

Photometric Mode Abs  
 Measurement range 400 - 190 nm  
 Data pitch 0.2 nm  
 Band width(UV/Vis) 2.0 nm  
 Response Medium  
 Scanning speed 200 nm/min  
 Source Change 340 nm  
 Light Source D2/WI  
 Filter Exchange Step  
 Correction Baseline

ds-77b

[Data Information]  
 Creation Date 2015-3-10 19:39  
 Data array type Linear data array  
 Horizontal Wavelength [nm]  
 Vertical Abs  
 Start 400 nm  
 End 190 nm  
 Data pitch 0.2 nm  
 Data points 1051

Figure S27. The UV spectrum of compound 2 in MeOH.



[Comments]  
 Sample name DS-77B  
 Comment  
 User  
 Division  
 Company dell

[Measurement Information]  
 Instrument Name J-815  
 Model Name J-815  
 Serial No. A024461168

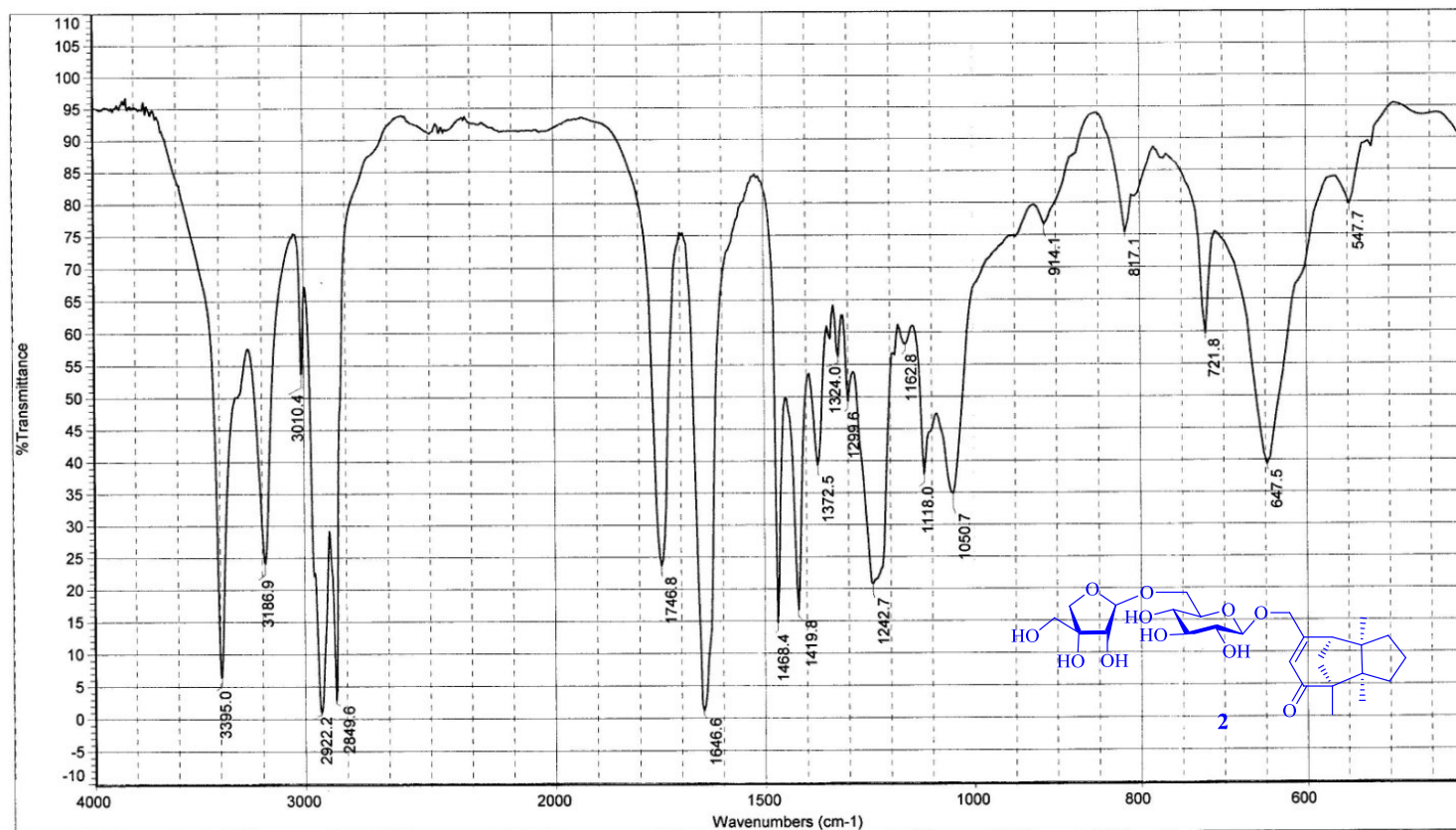
Accessory Standard  
 Accessory S/N A024461168  
 Cell Length 1 mm

Photometric Mode CD, HT, Abs  
 Measure Range 400 - 200 nm  
 Data pitch 0.5 nm  
 Sensitivity Standard  
 D.I.T. 2 sec  
 Band width 2.00 nm  
 Start Mode Immediately  
 Scanning Speed 100 nm/min  
 Baseline Correction Baseline  
 Shutter Control Auto  
 PMT Voltage Auto  
 Accumulations 3  
 Solvent MEOH  
 Concentration 0.48 (w/v)%

[Detailed Information]  
 Creation date 2014-5-20 10:25

Data array type Linear data array \* 3  
 Horizontal axis Wavelength [nm]  
 Vertical axis(1) Mol. CD  
 Vertical axis(2) HT [V]  
 Vertical axis(3) Abs  
 Start 400 nm  
 End 200 nm  
 Data interval 0.5 nm  
 Data points 401

Figure S28. The CD spectrum of compound 2 in MeOH.



日期: 星期四 6月 05 12:37:06 2014 (GMT+08:00) Sample Name: DS - 77b (显微镜透射法FT- IR Microscope Transmission)

扫描次数: 100

傅里叶变换显微镜红外(FT-IR Microscope): Centaurus

分辨率: 8.000

美国热电公司(Thermo)傅里叶变换红外光谱仪:Nicolet 5700

Figure S29. The IR spectrum of compound 2.

# Single Mass Spectrum Deconvolution Report

**Analysis Name:** jngyp161.d    **Instrument:** LC-MSD-Trap-SL    **Print Date:** 6/27/2014 3:10:18 PM  
**Method:** TEST.MS    **Operator:** Operator    **Acq. Date:** 6/27/2014 1:56:20 PM  
**Sample Name:** DS-77b  
**Analysis Info:**

## Acquisition Parameter:

Mass Range Mode	Std/Normal	Trap Drive	47.2	Scan Begin	300 m/z
Ion Polarity	Positive	Octopole RF Amplitude	194.5 Vpp	Scan End	700 m/z
Ion Source Type	ESI	Capillary Exit	132.3 Volt	Averages	5 Spectra
Dry Temp (Set)	330 °C	Skimmer	40.0 Volt	Max. Accu Time	200000 µs
Nebulizer (Set)	15.00 psi	Oct 1 DC	12.00 Volt	ICC Target	20000
Dry Gas (Set)	6.00 l/min	Oct 2 DC	1.70 Volt	Charge Control	on

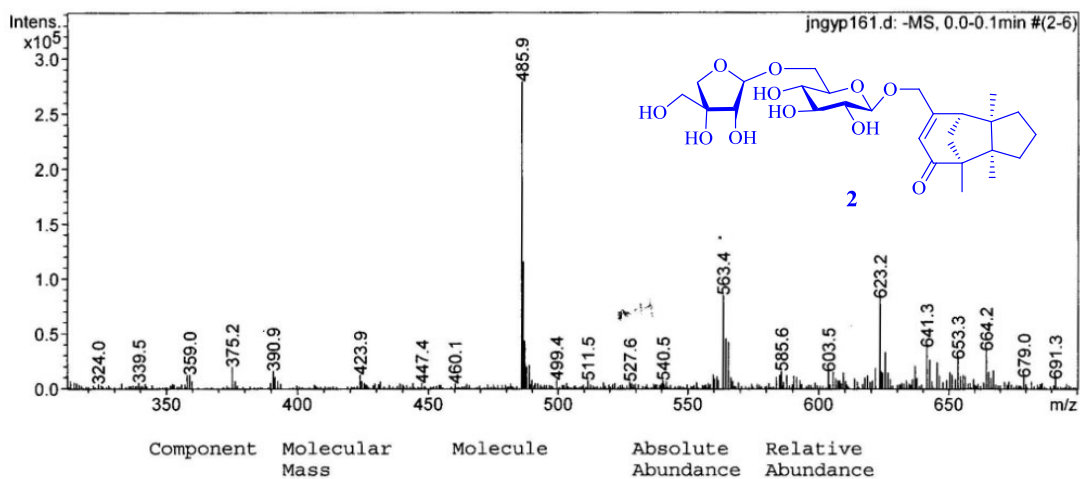
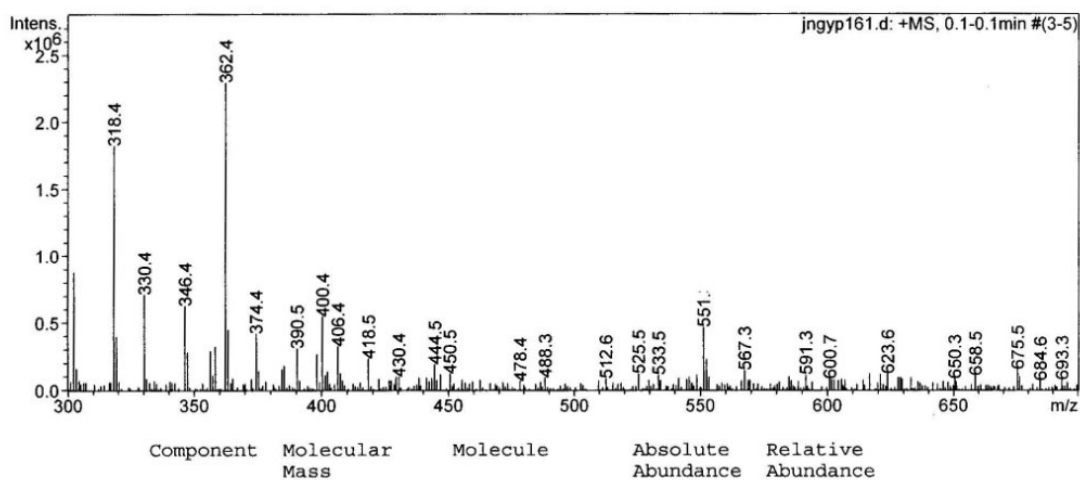


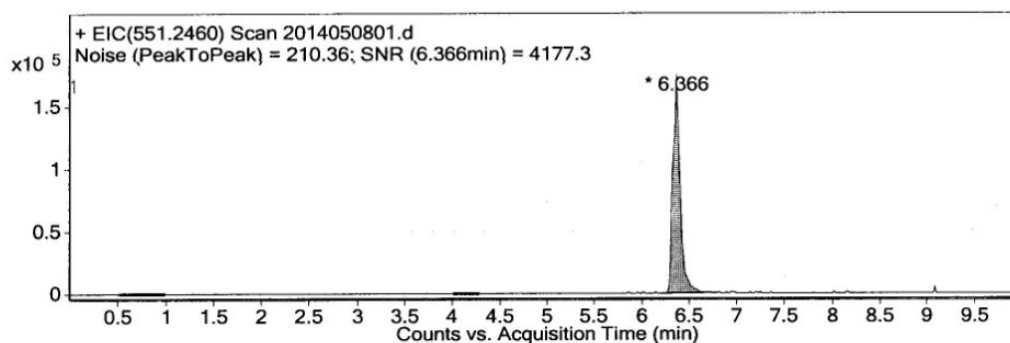
Figure S30. The ESI mass spectrum of compound 2.

# Qualitative Analysis Report

Data Filename	2014050801.d	Sample Name	DS-77b
Sample Type	Sample	Position	P1-C1
Instrument Name	Instrument 1	User Name	
Acq Method	TEST LCMS.m	IRM Calibration Status	XXXXXXXXXX
DA Method	TEST LCMS.m	Comment	

## User Chromatograms

Fragmentor Voltage 135    Collision Energy 0    Ionization Mode ESI



## Integration Peak List

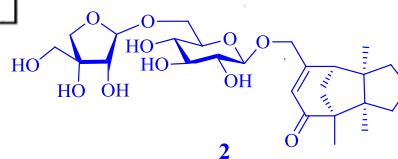
Peak	Start	RT	End	Height	Area	Area %	Signal To Noise
1	6.221	6.366	6.736	175229	878751	100	4177.3

## Noise Measurements

Noise Type	Signal Definition	Noise Multiplier	Noise Value
Peak-to-Peak	Area	1	210.3635864

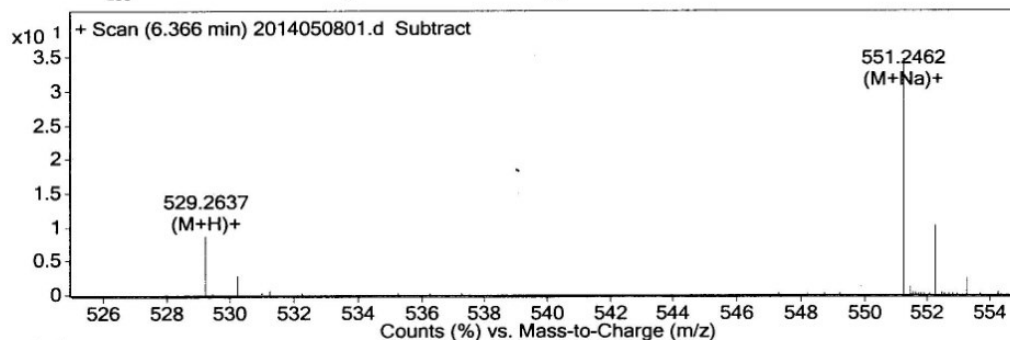
## Noise Regions

Start	End
0.5	1
4	4.3
9.99	11



## User Spectra

Fragmentor Voltage 135    Collision Energy 0    Ionization Mode ESI



## Peak List

m/z	z	Abund	Formula	Ion
199.1804	1	506216		
200.1827	1	71124		
214.0896	1	213460		
221.1609		56675		
231.1157		63052		
236.072		53527		
257.1678	1	254513		
397.2219	1	139477		

Figure S31. The (+)-HR-ESI-MS report of compound 2, page 1.

## Qualitative Analysis Report

<i>m/z</i>	<i>z</i>	Abund	Formula	Ion
551.2462	1	175385	C26 H40 Na O11	(M+Na)+
552.2494	1	51318	C26 H40 Na O11	(M+Na)+

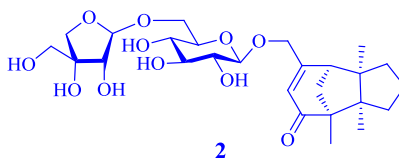
### Formula Calculator Element Limits

Element	Min	Max
C	3	100
H	0	500
O	0	90
N	0	5
S	0	0
Cl	0	0
Br	0	0
Si	0	0
F	0	0
P	0	0

### Formula Calculator Results

Formula	Best	Mass	Tgt Mass	Diff (ppm)	Ion Species	Score
C26 H40 O11	TRUE	528.257	528.2571	0.11	C26 H40 Na O11	99.98
C27 H36 N4 O7		528.257	528.2584	2.63	C27 H36 N4 Na O7	99.74
C27 H36 N4 O7		528.2565	528.2584	3.68	C27 H37 N4 O7	99.72
C26 H40 O11	TRUE	528.2564	528.2571	1.17	C26 H41 O11	99.67
C39 H32 N2		528.2565	528.2565	0.19	C39 H33 N2	98.2

--- End Of Report ---



MS Formula Results: + Scan (6.366 min) Sub (2014050801.d)

m/z	Ion	Formula	Abundance
529.2637	(M+H) <sup>+</sup>	C26 H41 O11	43614.1

Best	Formula (M)	Ion Formula	Calc m/z	Score	Cross S	Mass	Calc Mass	Diff (ppm)	Abs Diff (ppm)	Abund Match	Spacing Mat	Mass Match	m/z	DBE
☐	C27 H36 N4 O7	C27 H37 N4 O7	529.2657	99.72		528.2565	528.2584	3.68	3.68	99.83	99.96	99.53	529.2637	12
☐	C26 H40 O11	C26 H41 O11	529.2643	99.67		528.2564	528.2571	1.17	1.17	98.94	99.99	99.95	529.2637	7
☐	C39 H32 N2	C39 H33 N2	529.2638	98.2		528.2565	528.2565	0.19	0.19	93.72	99.98	100	529.2637	25

m/z	Ion	Formula	Abundance
551.2462	(M+Na) <sup>+</sup>	C26 H40 Na O11	175384.6

Best	Formula (M)	Ion Formula	Calc m/z	Score	Cross S	Mass	Calc Mass	Diff (ppm)	Abs Diff (ppm)	Abund Match	Spacing Mat	Mass Match	m/z	DBE
☐	C26 H40 O11	C26 H40 Na O11	551.2463	99.98		528.257	528.2571	0.11	0.11	99.95	99.99	100	551.2462	7
☐	C27 H36 N4 O7	C27 H36 N4 Na O7	551.2476	99.74		528.257	528.2584	2.63	2.63	99.47	100	99.78	551.2462	12

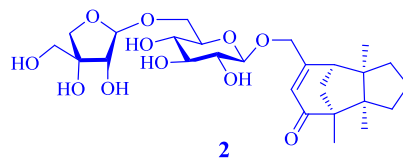


Figure S33. The (+)-HR-ESI-MS report of compound 2, page 3.

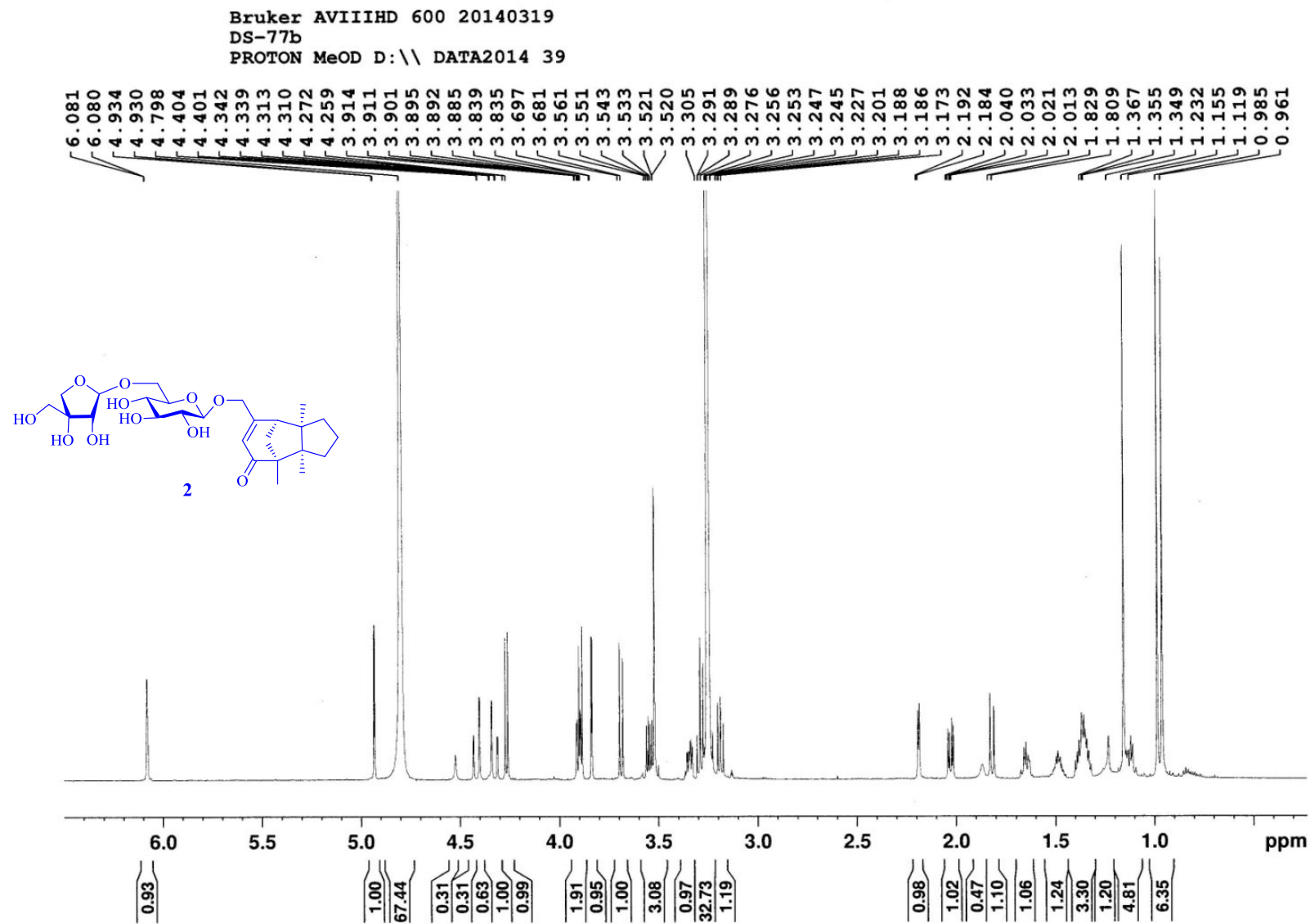


Figure S34. The  $^1\text{H}$  NMR spectrum of compound **2** in  $\text{MeOH-}d_4$  (600 MHz).



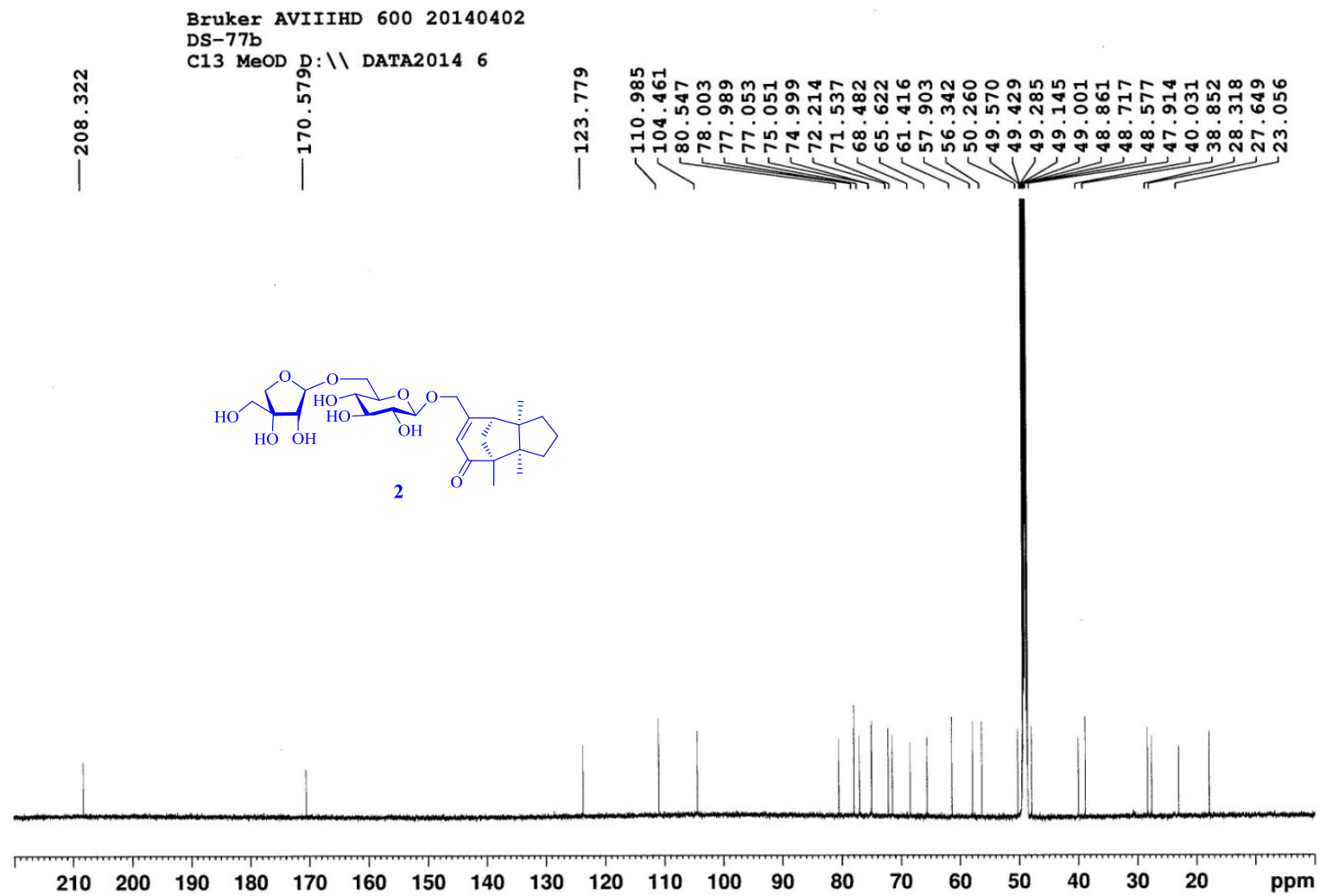


Figure S35. The  $^{13}\text{C}$  NMR spectrum of compound **2** in  $\text{MeOH-}d_4$  (150MHz).



Bruker AVIIIHD 600 20140402  
DS-77b DEPT MeOD D:\\ DATA2014 6

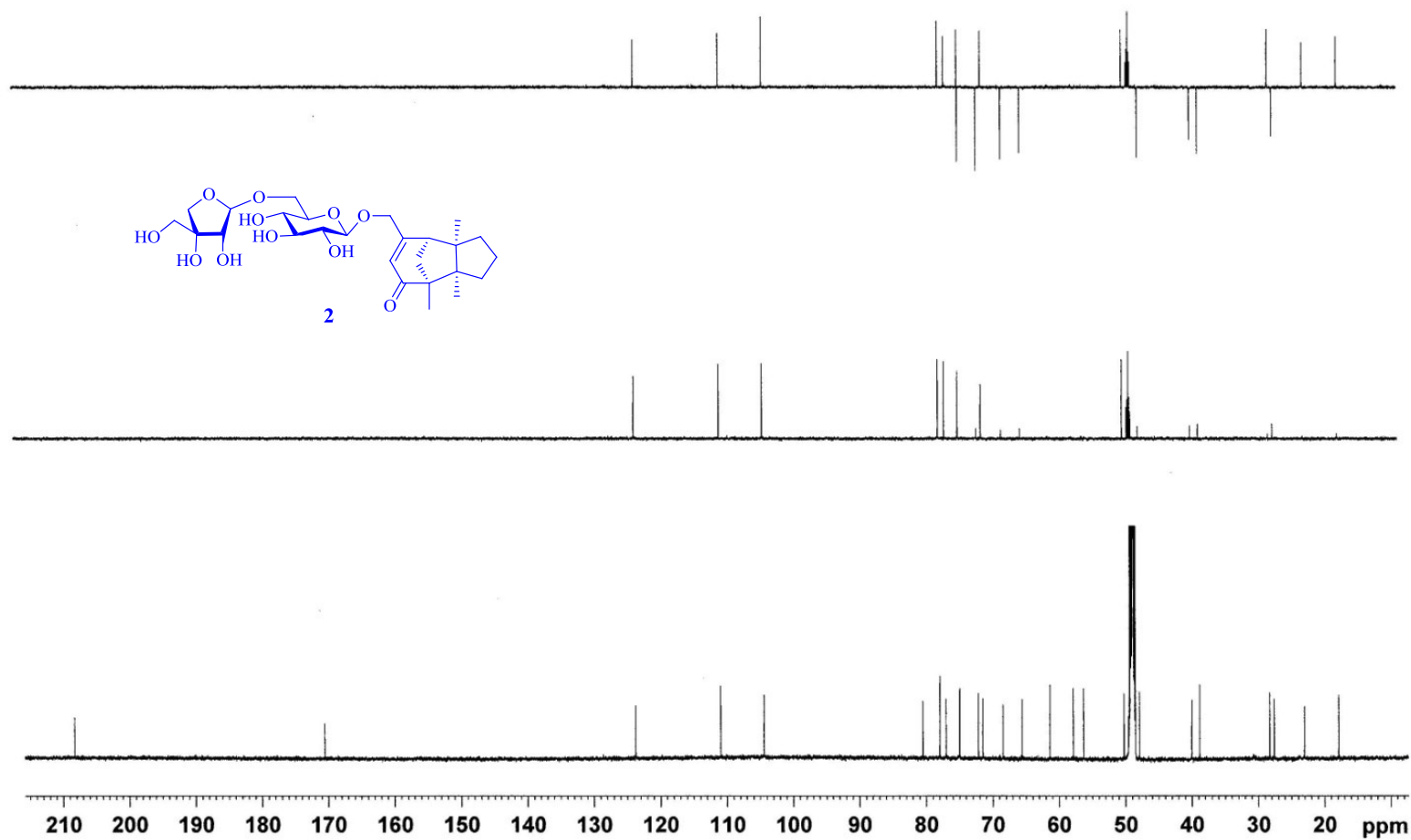


Figure S36. The DEPT spectrum of compound 2 in MeOH-*d*<sub>4</sub> (150 MHz).

Bruker AVIIIHD 600 20140404  
 DS-77b  
 {H-H COSY} MeOD D:\ DATA2014 10

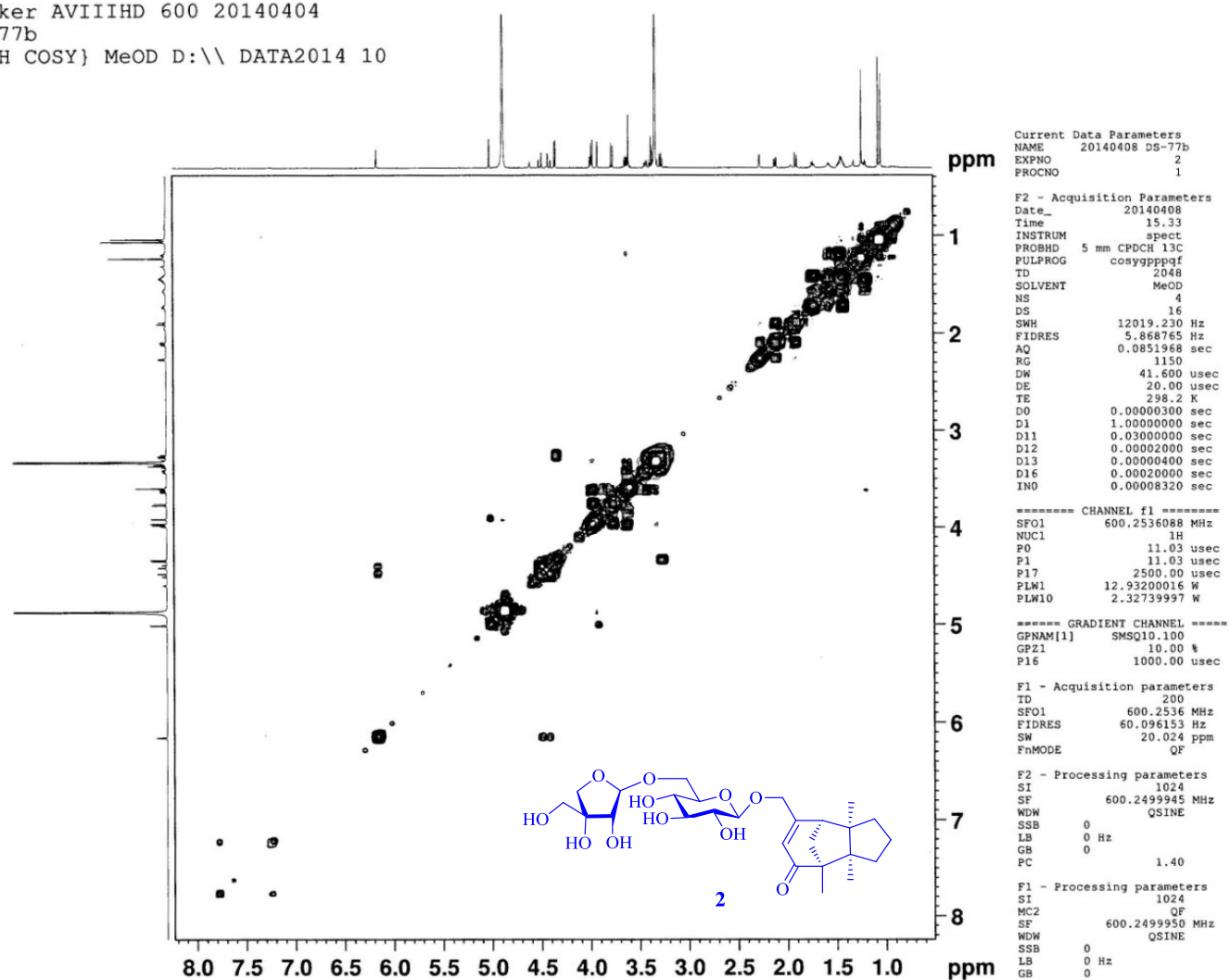


Figure S37. The <sup>1</sup>H-<sup>1</sup>H COSY spectrum of compound 2 in MeOH-d<sub>4</sub> (600 MHz).

Bruker AVIIIHD 600 20140404  
 DS-77b  
 HSQC MeOD D:\ DATA2014 10

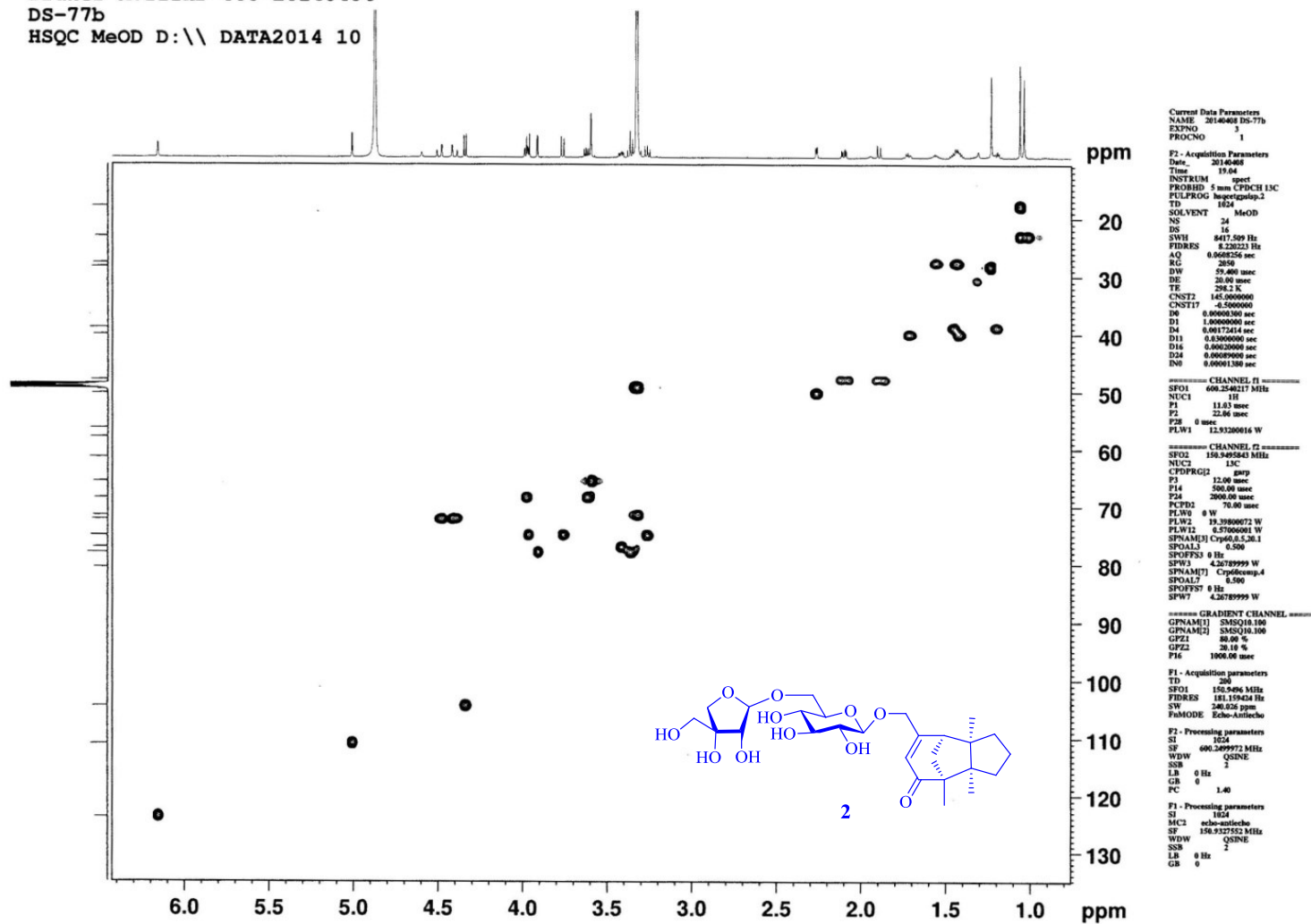
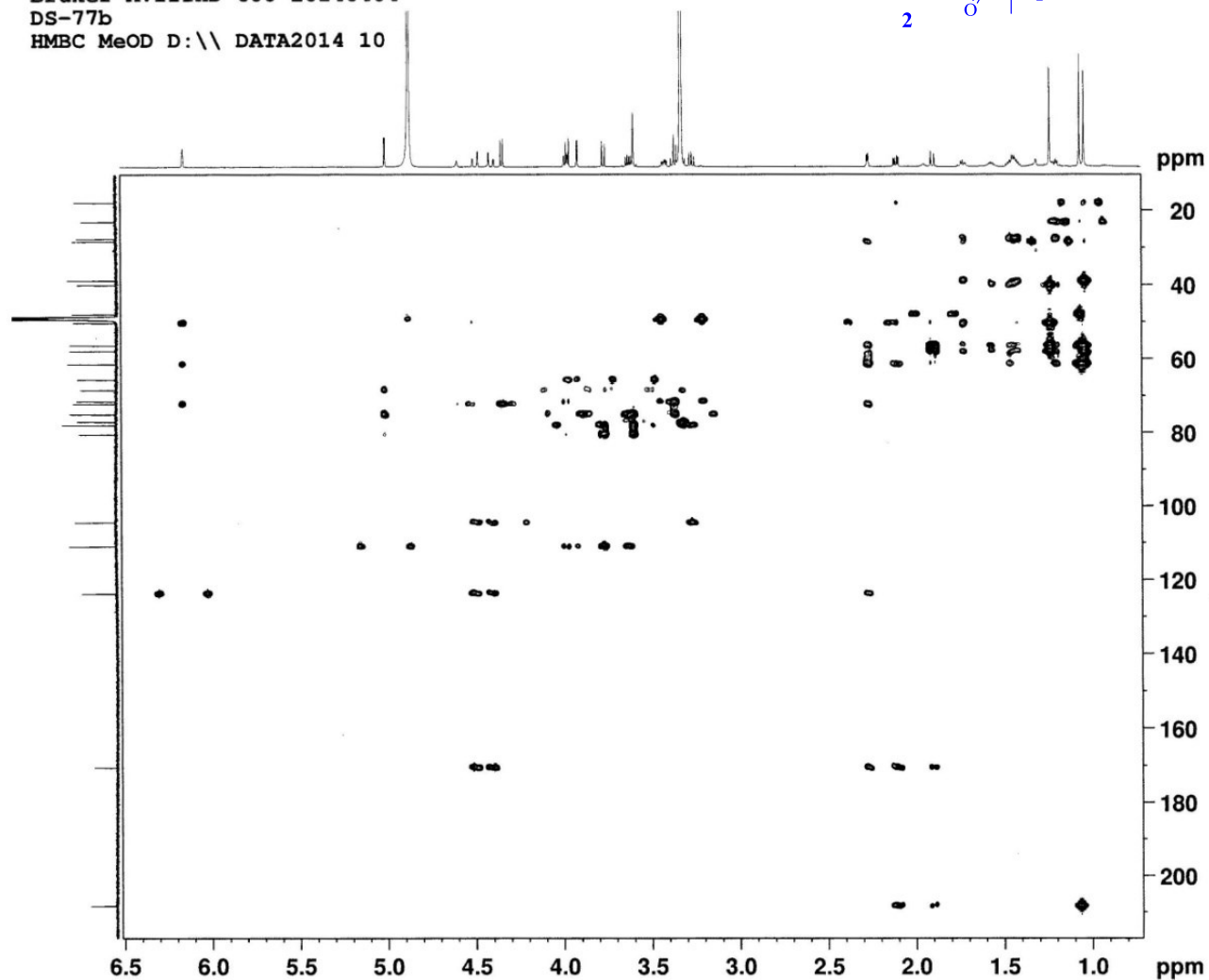
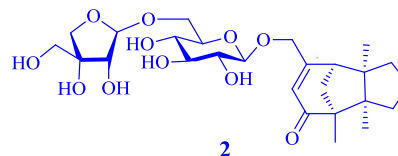


Figure S38. The HSQC spectrum of compound 2 in MeOH- $d_4$  (600 MHz for  $^1\text{H}$ ).

Bruker AVIIIHD 600 20140404  
 DS-77b  
 HMBC MeOD D:\ DATA2014 10



```

Current Data Parameters
NAME 20140408 DS-77b
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20140408
Time 20.32
INSTRUM spect
PROBHD 5 mm CPDCH 13C
PULPROG hmcgprsdqf
TD 4096
SOLVENT MeOD
NS 48
DS 16
SWH 8417.509 Hz
FIDRES 2.055056 Hz
AQ 0.2433024 sec
RG 2050
DW 59.400 usec
DE 20.00 usec
TE 298.2 K
CNST13 8.0000000
D0 0.00000300 sec
D1 1.00000000 sec
D6 0.06250000 sec
D16 0.00020000 sec
IN0 0.00001300 sec

===== CHANNEL f1 =====
SFO1 600.2536088 MHz
NUC1 1H
P1 11.03 usec
P2 22.06 usec
PLW1 12.93200016 W

===== CHANNEL f2 =====
SFO2 150.9495843 MHz
NUC2 13C
P3 12.00 usec
PLW2 19.39800072 W

===== GRADIENT CHANNEL =====
GPNAM[1] SMSQ10.100
GPNAM[2] SMSQ10.100
GPNAM[3] SMSQ10.100
GPZ1 50.00 %
GPZ2 30.00 %
GPZ3 40.10 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 200
SFO1 150.9496 MHz
FIDRES 181.159424 Hz
SW 240.026 ppm
PaMODE QF

F2 - Processing parameters
SI 1024
SF 600.2499981 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 150.937508 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
  
```

Figure S39. The HMBC spectrum of compound 2 in MeOH- $d_4$  (600 MHz for  $^1\text{H}$ ).

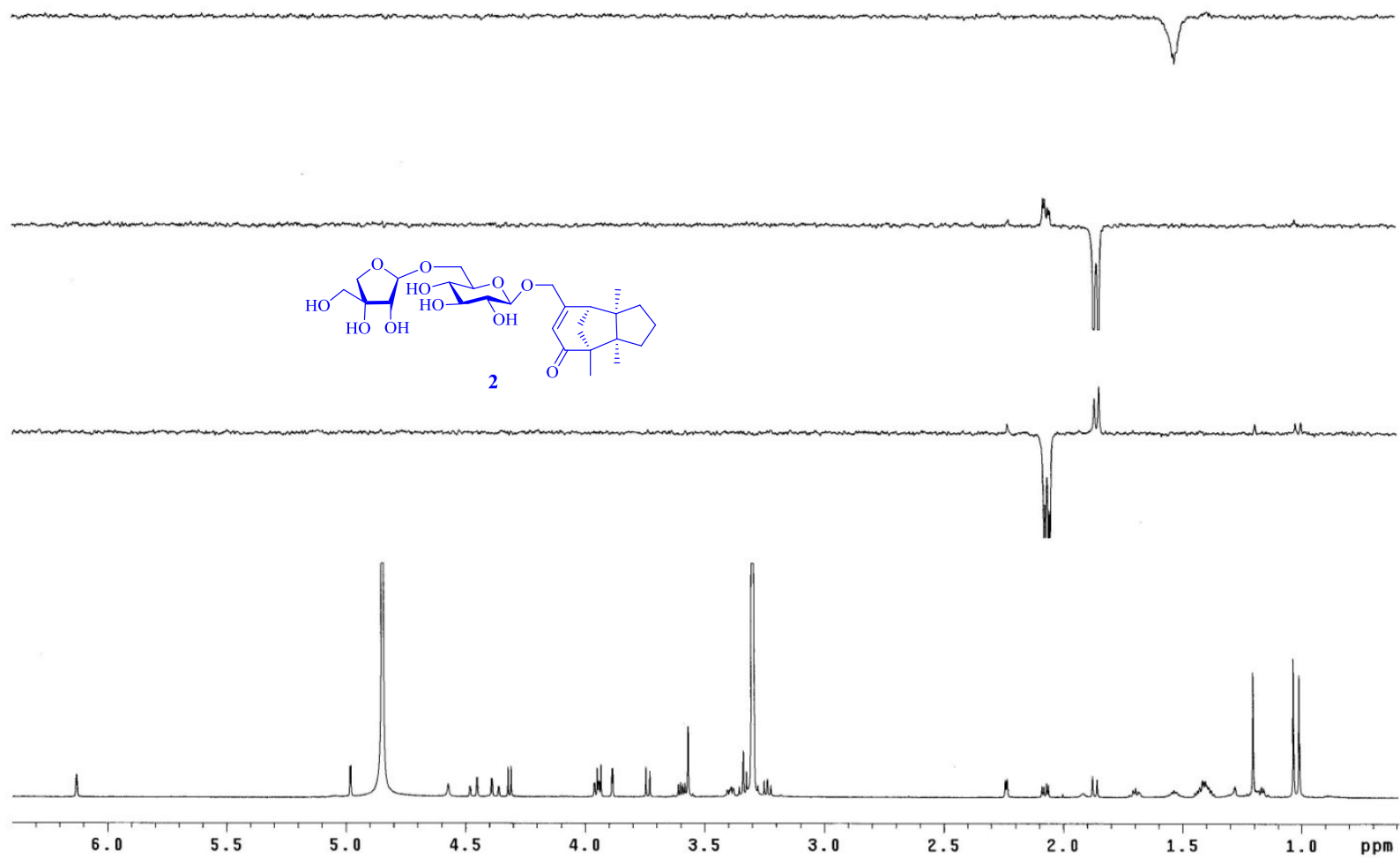
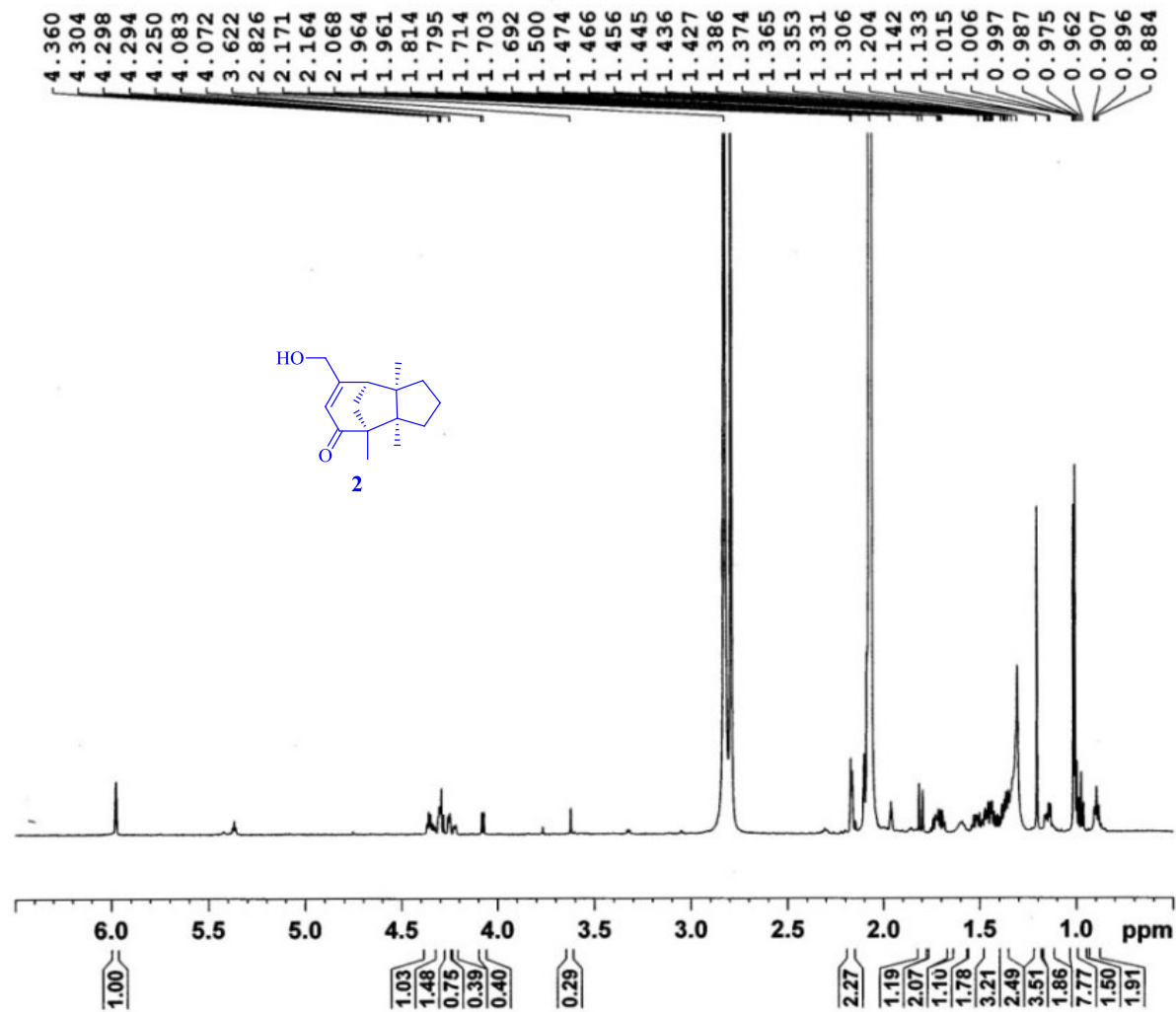


Figure S40. The NOE difference spectrum of 2 in MeOH- $d_4$  (600 MHz).

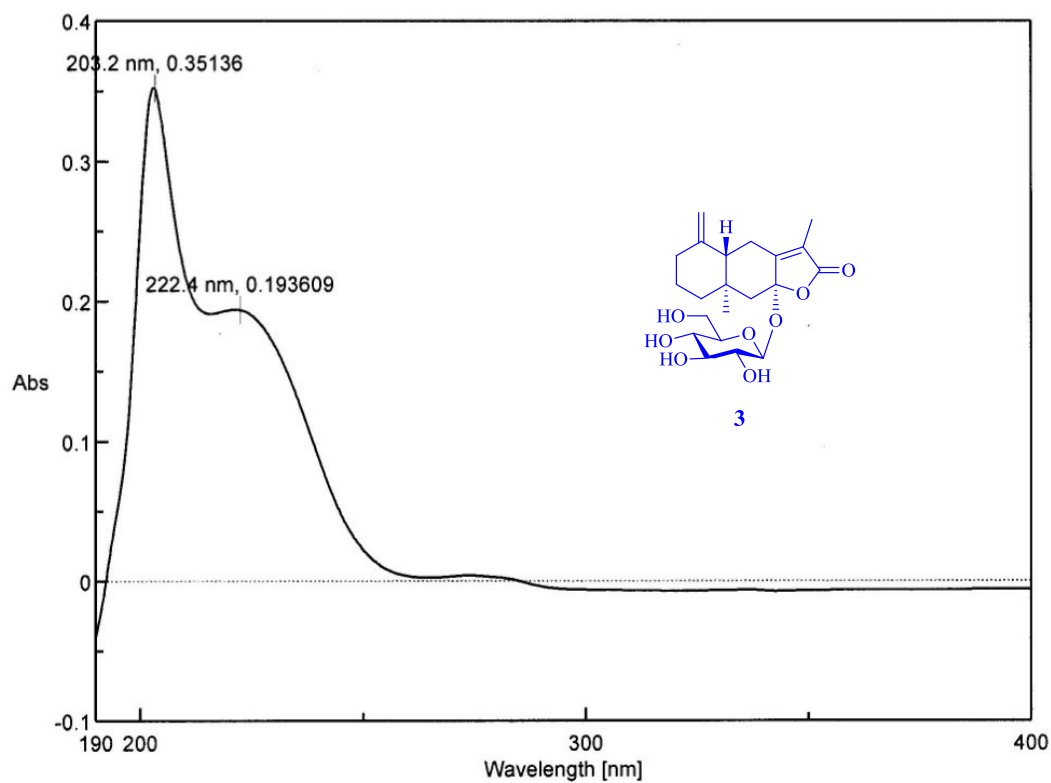
Bruker AVIIIHD 600 20141102  
 PROTON Acetone D:\\ DATA2014 7



NAME 20141103 DS-77a  
 EXPNO 2  
 PROCNO 1  
 Date\_ 20141103  
 Time\_ 9.59  
 INSTRUM spect  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zg30  
 TD 65536  
 SOLVENT Acetone  
 NS 32  
 DS 2  
 SWH 12019.230 Hz  
 FIDRES 0.183399 Hz  
 AQ 2.7263477 sec  
 RG 50.8  
 DW 41.600 usec  
 DE 20.00 usec  
 TE 298.2 K  
 D1 1.00000000 sec  
 TD0 2

----- CHANNEL f1 -----  
 SF01 600.2537068 MHz  
 NUC1 1H  
 P1 11.50 usec  
 SI 65536  
 SF 600.2500000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

Figure S41. The <sup>1</sup>H NMR spectrum of the aglycone of 2 in MeOH-*d*<sub>4</sub> (600 MHz).



[Comment]  
 Sample Name DBT  
 Comment 0.02  
 User  
 Division UV  
 324  
 [Measurement Information]  
 Instrument Name V-650  
 Model Name V-650  
 Serial No. A034461150

DS-135

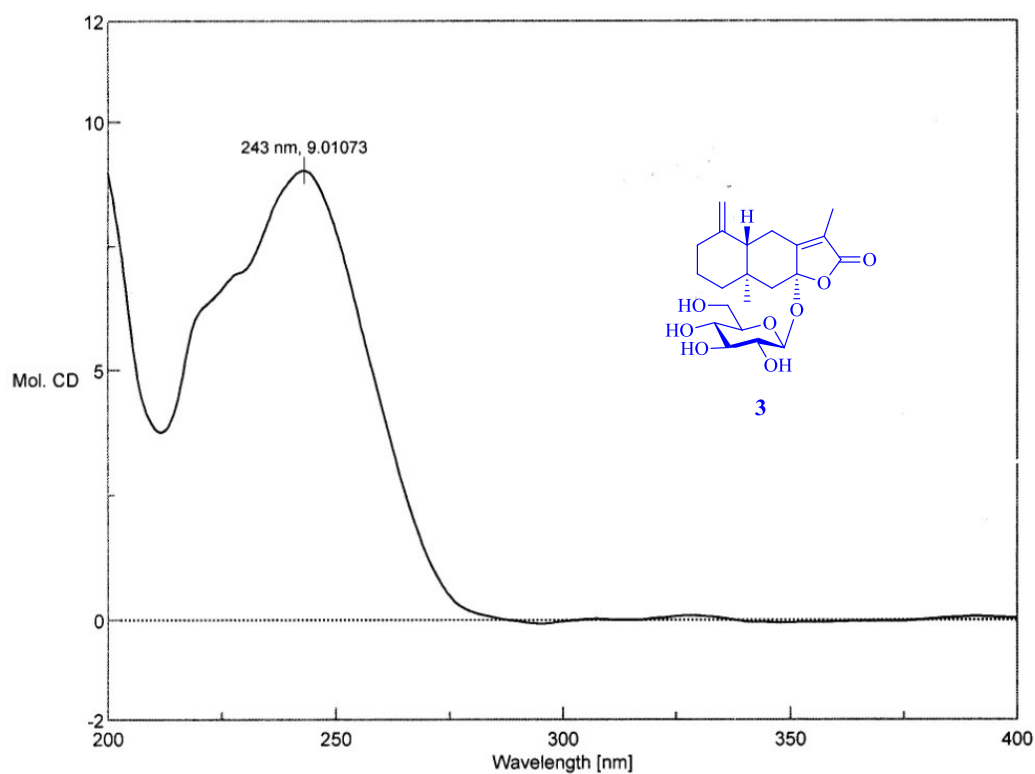
Accessory PSC-718  
 Accessory S/N A001761114  
 Position 1  
 Cell Length 10 mm  
 Temperature 19.94 C  
 Control Sensor Holder  
 Monitor Sensor Holder  
 Start Mode Start immediately

[Data Information]  
 Creation Date 2014-6-10 19:21  
 Data array type Linear data array  
 Horizontal Wavelength [nm]  
 Vertical Abs  
 Start 400 nm  
 End 190 nm  
 Data pitch 0.2 nm  
 Data points 1051

Photometric Mode Abs  
 Measurement range 400 - 190 nm  
 Data pitch 0.2 nm  
 Band width(UV/Vis) 2.0 nm  
 Response Medium  
 Scanning speed 200 nm/min  
 Source Change 340 nm  
 Light Source D2/WI  
 Filter Exchange Step  
 Correction Baseline

Figure S42. The UV spectrum of compound 3 in MeOH.





[Comments]  
 Sample name DS-135  
 Comment  
 User  
 Division  
 Company dell

[Measurement Information]  
 Instrument Name J-815  
 Model Name J-815  
 Serial No. A024461168

Accessory Standard  
 Accessory S/N A024461168  
 Cell Length 1 mm

Photometric Mode CD, HT, Abs  
 Measure Range 400 - 200 nm  
 Data pitch 0.5 nm  
 Sensitivity Standard  
 D.I.T. 2 sec  
 Band width 2.00 nm  
 Start Mode Immediately  
 Scanning Speed 100 nm/min  
 Baseline Correction Baseline  
 Shutter Control Auto  
 PMT Voltage Auto  
 Accumulations 3  
 Solvent MEOH  
 Concentration 0.2462 (w/v)%

[Detailed Information]  
 Creation date 2014-6-20 11:11

Data array type Linear data array \* 3  
 Horizontal axis Wavelength [nm]  
 Vertical axis(1) Mol. CD  
 Vertical axis(2) HT [V]  
 Vertical axis(3) Abs  
 Start 400 nm  
 End 200 nm  
 Data interval 0.5 nm  
 Data points 401

Figure S43. The CD spectrum of compound 3 in MeOH.

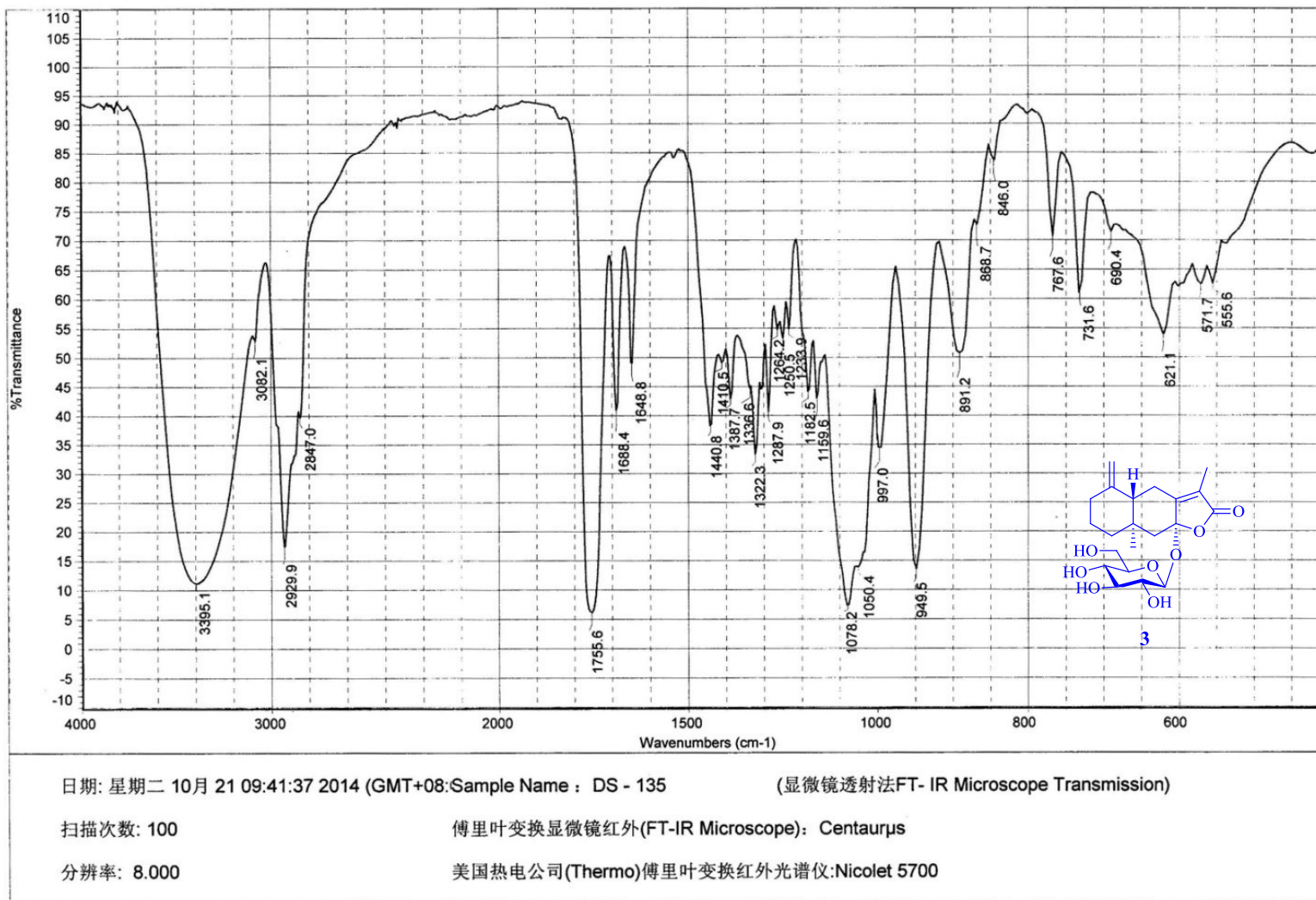


Figure S44. The IR spectrum of compound 3.

# Single Mass Spectrum Deconvolution Report

**Analysis Name:** jngyp173.d     **Instrument:** LC-MSD-Trap-SL     **Print Date:** 6/27/2014 3:31:53 PM  
**Method:** TEST.MS     **Operator:** Operator     **Acq. Date:** 6/27/2014 2:26:26 PM  
**Sample Name:** DS-135  
**Analysis Info:**

## Acquisition Parameter:

Mass Range Mode	Std/Normal	Trap Drive	36.3	Scan Begin	200 m/z
Ion Polarity	Positive	Octopole RF Amplitude	171.0 Vpp	Scan End	600 m/z
Ion Source Type	ESI	Capillary Exit	121.0 Volt	Averages	5 Spectra
Dry Temp (Set)	330 °C	Skimmer	40.0 Volt	Max. Accu Time	200000 µs
Nebulizer (Set)	15.00 psi	Oct 1 DC	12.00 Volt	ICC Target	20000
Dry Gas (Set)	6.00 l/min	Oct 2 DC	1.70 Volt	Charge Control	on

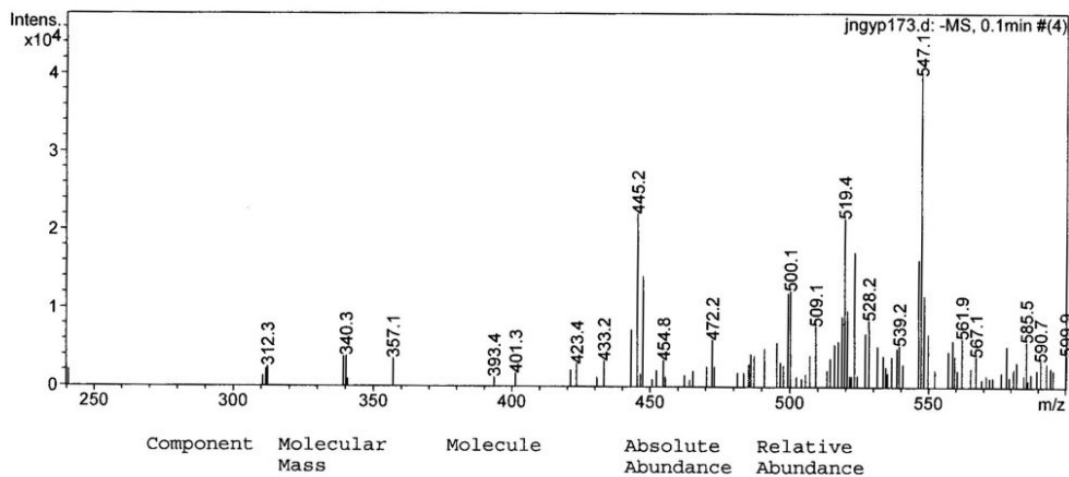
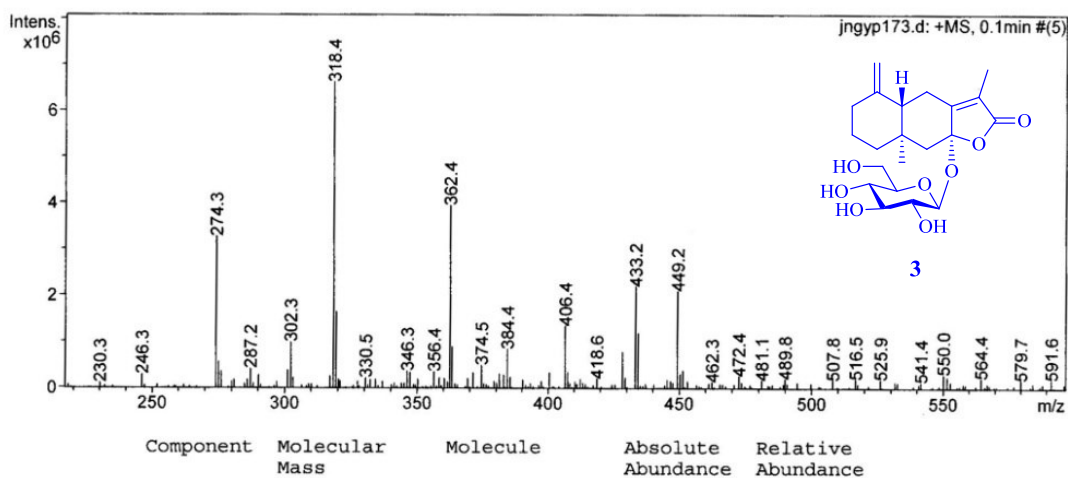
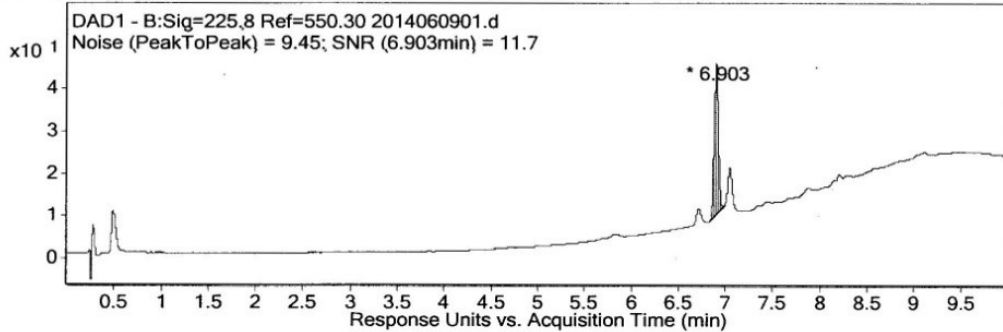


Figure S45. The ESI mass spectra of Compound 3.

# Qualitative Analysis Report

Data Filename	2014060901.d	Sample Name	DS-135
Sample Type	Sample	Position	P1-C2
Instrument Name	Instrument 1	User Name	
Acq Method		IRM Calibration Status	Success
DA Method	TEST LCMS.m	Comment	

## User Chromatograms



### Integration Peak List

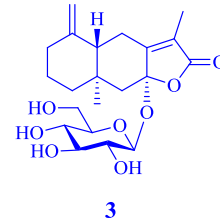
Peak	Start	RT	End	Height	Area	Area %	Signal To Noise
1	6.833	6.903	6.992	35.38	110.59	100	11.7

### Noise Measurements

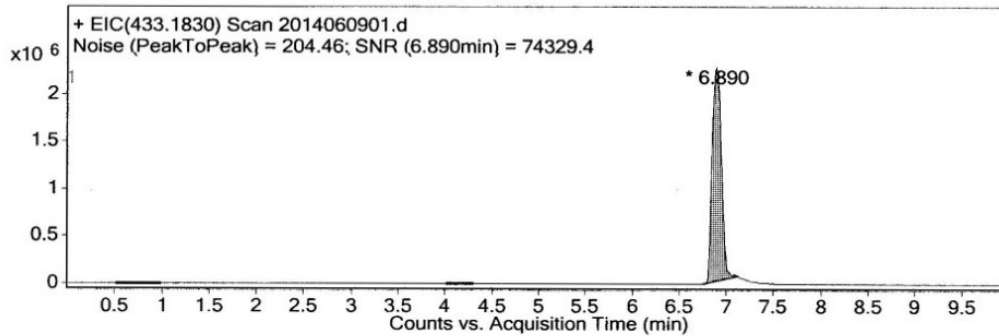
Noise Type	Signal Definition	Noise Multiplier	Noise Value
Peak-to-Peak	Area	1	9.44852829

### Noise Regions

Start	End
0.5	1
4	4.3
9.99	11



Fragmentor Voltage 135    Collision Energy 0    Ionization Mode ESI



### Integration Peak List

Peak	Start	RT	End	Height	Area	Area %	Signal To Noise
1	6.745	6.89	7.132	2258026	15197083	100	74329.4

### Noise Measurements

Noise Type	Signal Definition	Noise Multiplier	Noise Value
Peak-to-Peak	Area	1	204.4558563

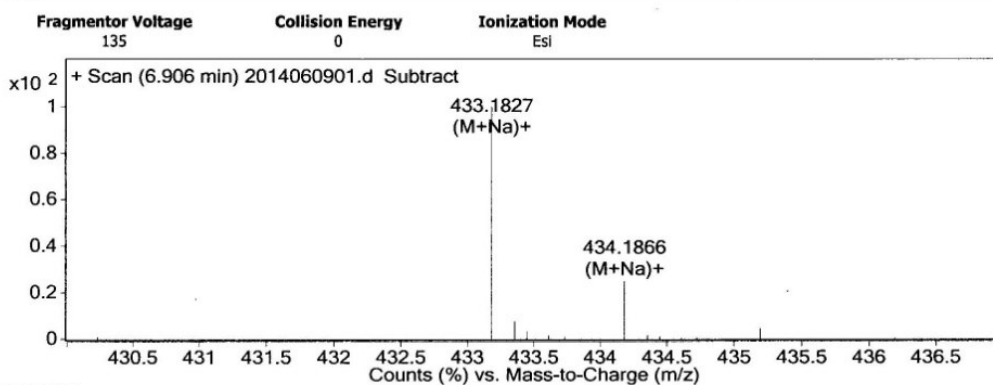
### Noise Regions

Start	End
0.5	1
4	4.3
9.99	11

## User Spectra

Figure S46. The (+)-HR-ESI-MS report of compound 3, page 1.

# Qualitative Analysis Report

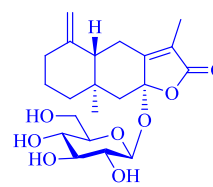


**Peak List**

m/z	z	Abund	Formula	Ion
249.1479		119082		
428.2276		405647		
433.1827	1	2145478	C21 H30 Na O8	(M+Na)+
433.3571		157937		
434.1866	1	532157	C21 H30 Na O8	(M+Na)+
843.3769	1	387325		
844.3787	1	175409		

**Formula Calculator Element Limits**

Element	Min	Max
C	3	100
H	0	500
O	0	90
N	0	5
S	0	2
Cl	0	0
Br	0	0
Si	0	0
F	0	0
P	0	0



3

**Formula Calculator Results**

Formula	Best	Mass	Tgt Mass	Diff (ppm)	Ion Species	Score
C21 H30 O8	TRUE	410.1935	410.1941	1.39	C21 H30 Na O8	99.91
C22 H26 N4 O4		410.1935	410.1954	4.64	C22 H26 N4 Na O4	99.62
C25 H30 O3 S		410.1935	410.1916	-4.72	C25 H30 Na O3 S	98.83
C22 H34 O3 S2		410.1935	410.1949	3.5	C22 H34 Na O3 S2	97.38

--- End Of Report ---

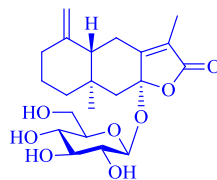
Figure S47. The (+)-HR-ESI-MS report of compound 3, page 2.

MS Formula Results: + Scan (6.906 min) Sub (2014060901.d)

m/z	Ion	Formula	Abundance
433.1827	(M+Na) <sup>+</sup>	C21 H30 Na O8	2145477.5

Best	Formula (M)	Ion Formula	Calc m/z	Score	Cross S	Mass	Calc Mass	Diff (ppm)	Abs Diff (ppm)	Abund Match	Spacing Mat	Mass Match	m/z	DBE
<input checked="" type="checkbox"/>	C21 H30 O8	C21 H30 Na O8	433.1833	99.91		410.1935	410.1941	1.39	1.39	99.8	99.96	99.94	433.1827	7
<input type="checkbox"/>	C22 H26 N4 O4	C22 H26 N4 Na O4	433.1846	99.62		410.1935	410.1954	4.64	4.64	99.82	99.89	99.36	433.1827	12
<input type="checkbox"/>	C25 H30 O3 S	C25 H30 Na O3 S	433.1808	98.83		410.1935	410.1916	-4.72	4.72	97.42	99.54	99.33	433.1827	11
<input type="checkbox"/>	C22 H34 O3 S2	C22 H34 Na O3 S2	433.1842	97.38		410.1935	410.1949	3.5	3.5	92.17	99.13	99.63	433.1827	6



3

Figure S48. The (+)-HR-ESI-MS report of compound 3, page 3.

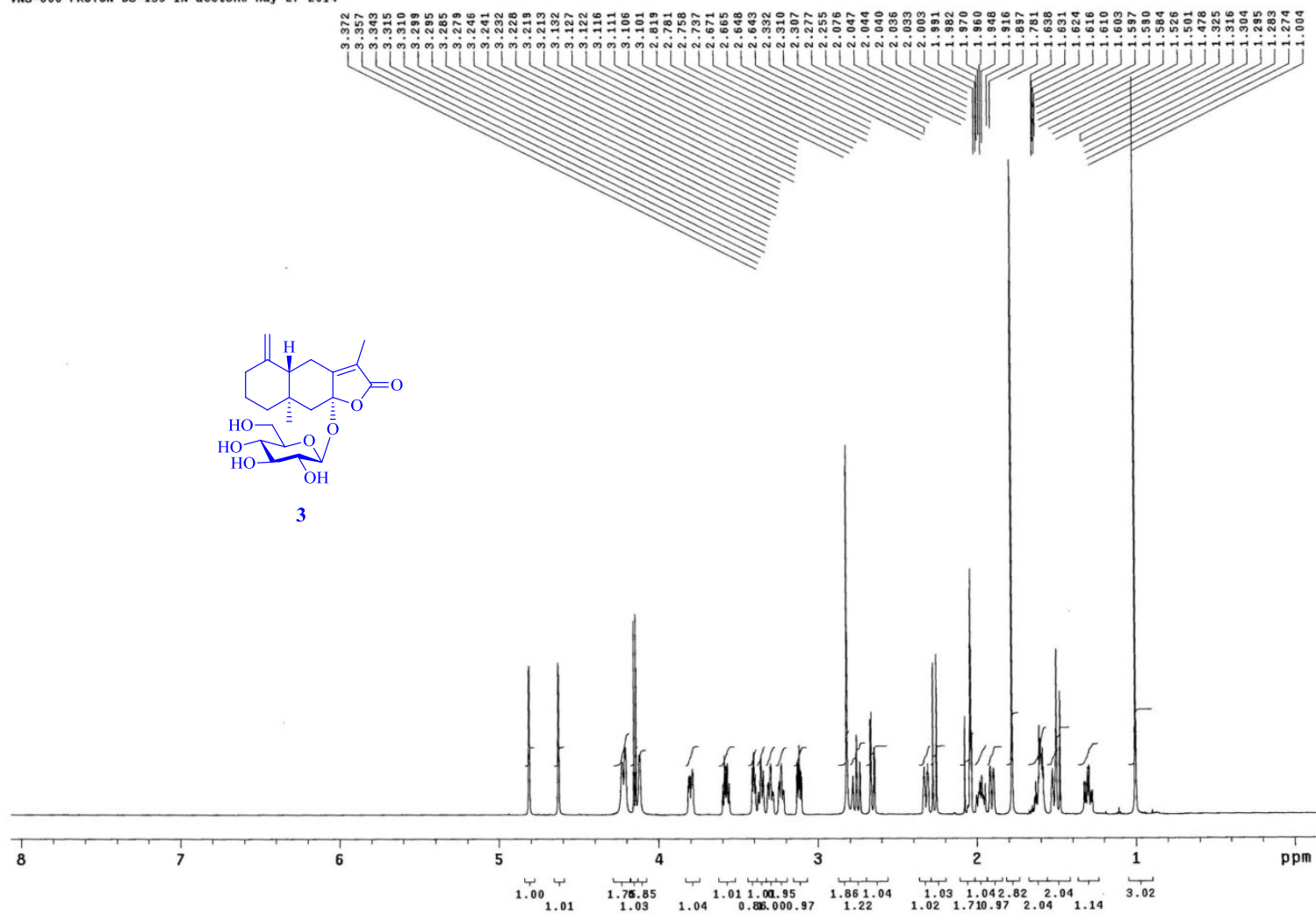
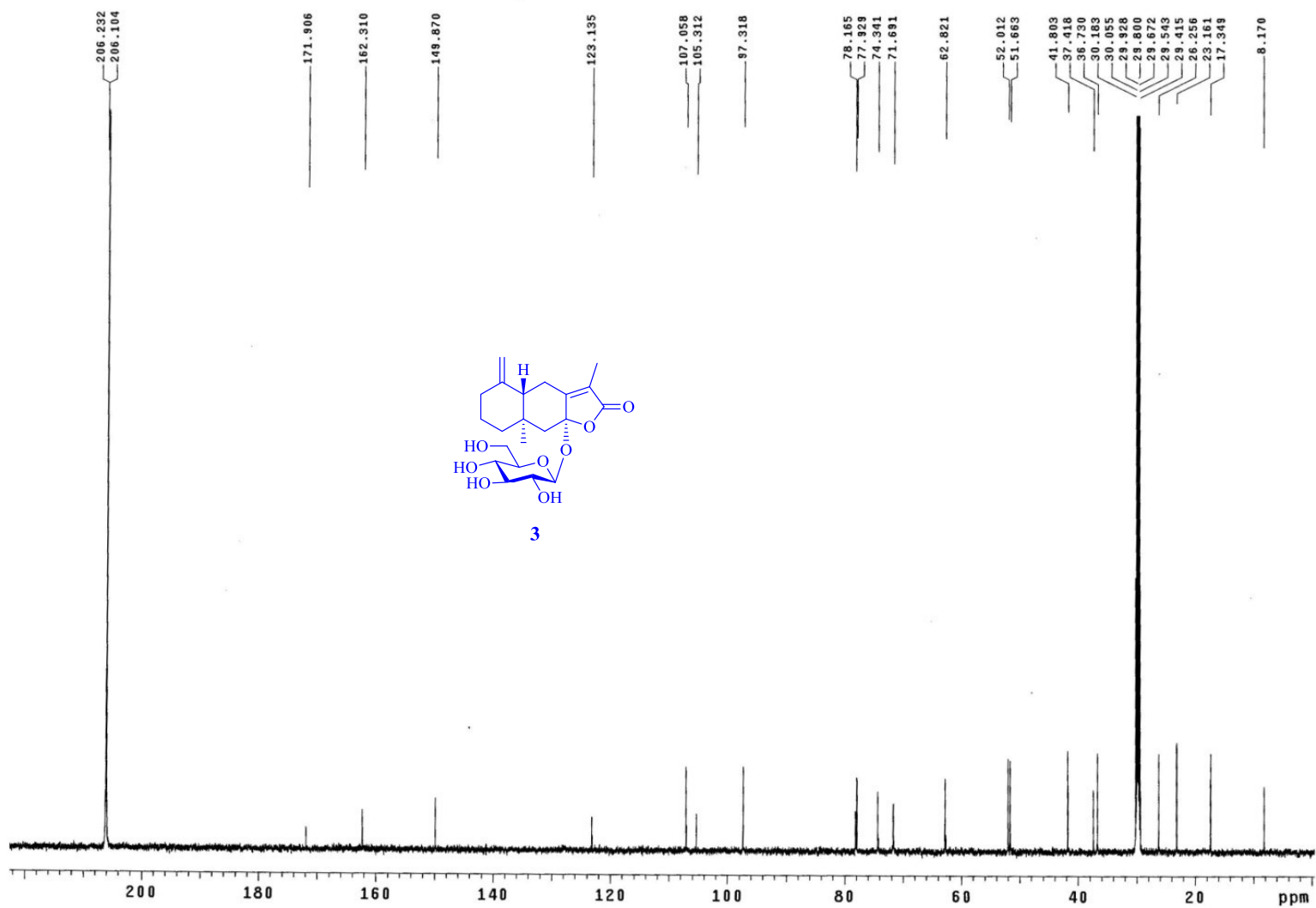
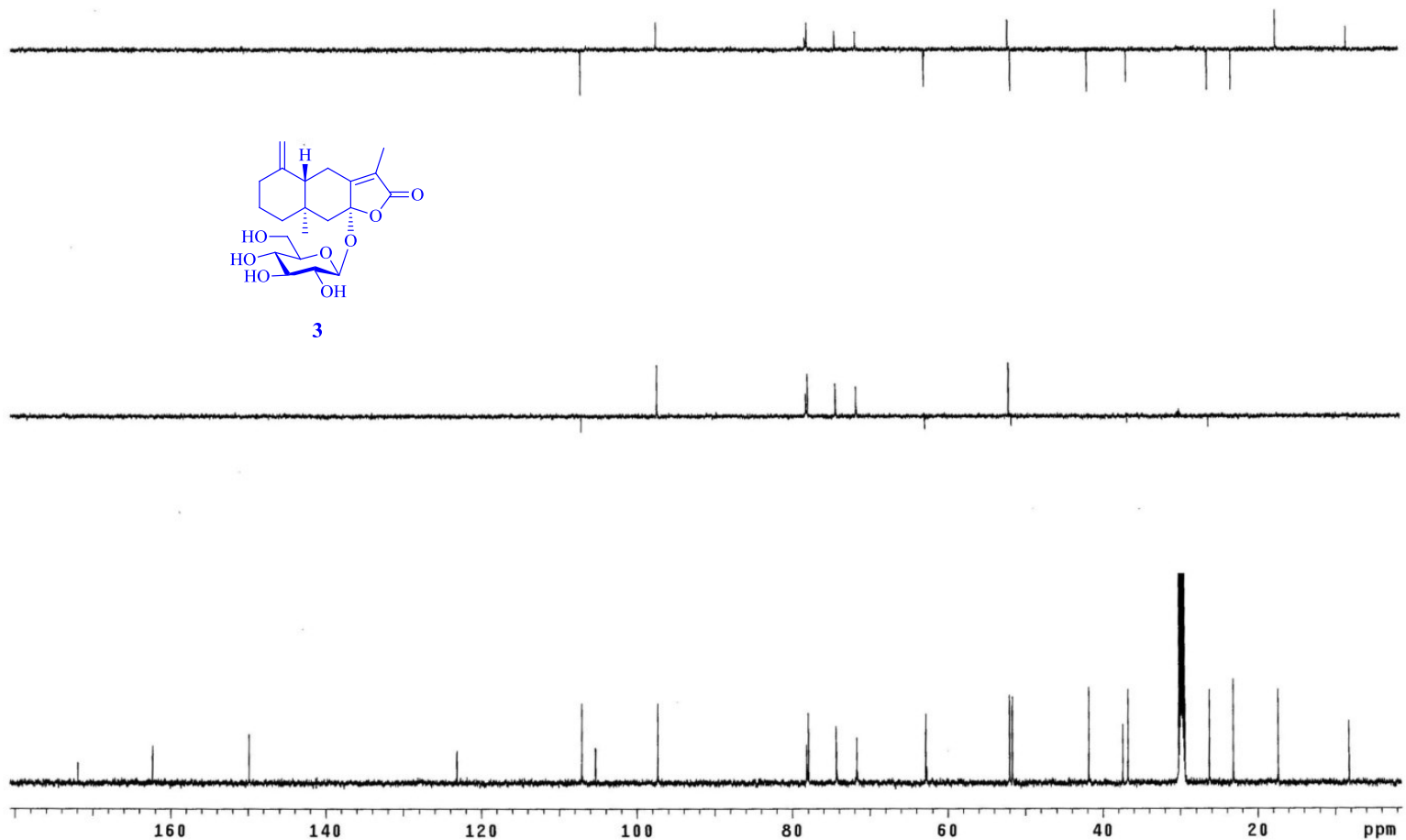


Figure S49. The <sup>1</sup>H NMR spectra of compound 3 in acetone-*d*<sub>6</sub> (600 M).



**Figure S50.** The  $^{13}\text{C}$  NMR spectrum of compound **3** in acetone- $d_6$  (150 M).





**Figure S51.** The DEPT spectra of compound **3** in acetone-*d*<sub>6</sub> (150 M).

VNS-600 gCOSY DS-135 IN acetone Jun 5 2014

Temp. 25.0 C / 298.1 K  
Sample #8, Operator: vjwalk

Relax. delay 1.000 sec  
Acq. time 0.150 sec  
Width 5364.8 Hz  
2D Width 5364.8 Hz  
2 repetitions  
200 increments  
OBSERVE H1, 599.6908008 MHz  
DATA PROCESSING  
Sq. sine bell 0.075 sec  
F1 DATA PROCESSING  
Sq. sine bell 0.021 sec  
FT size 2048 x 2048  
Total time 8 min 36 sec

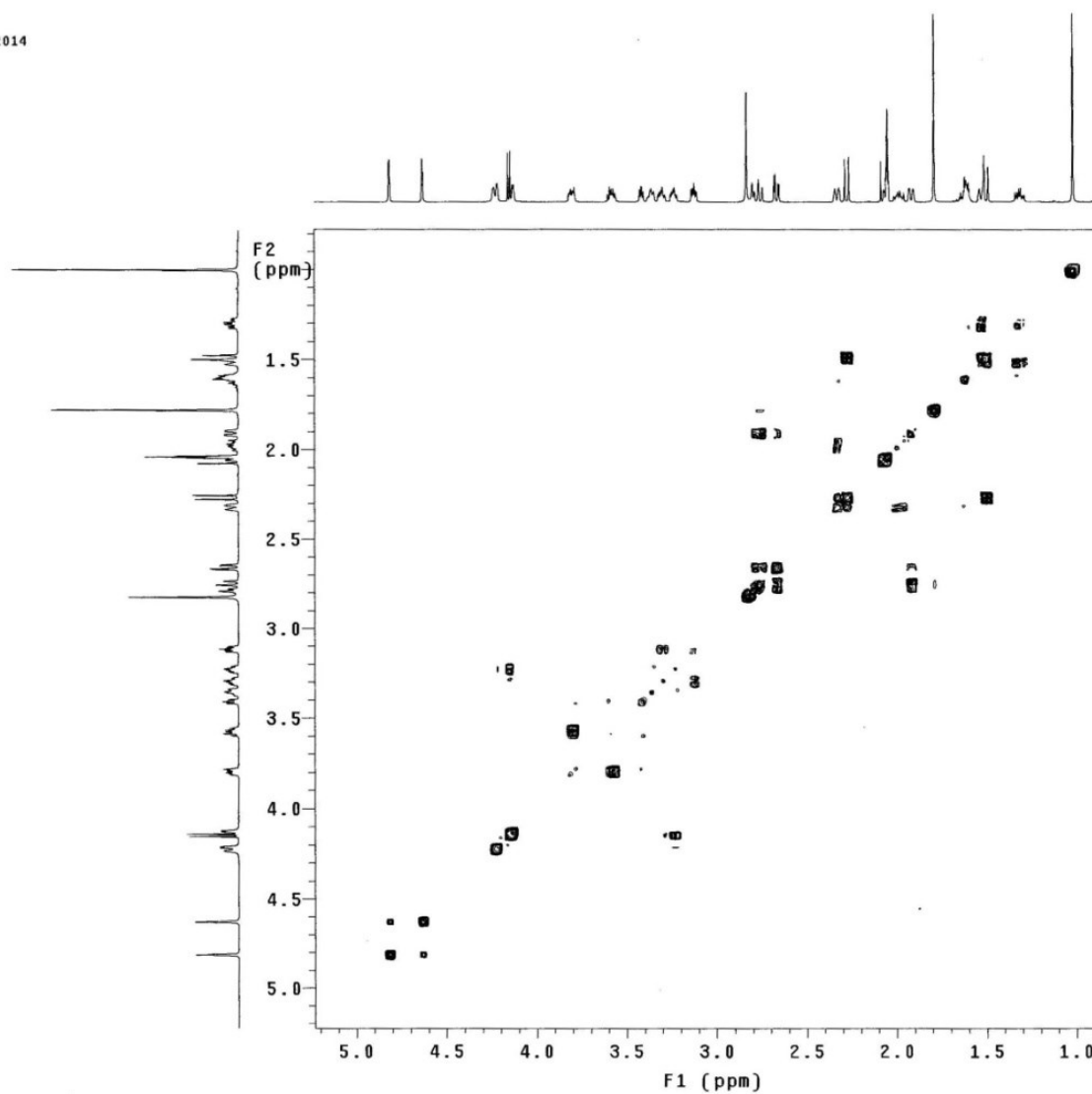
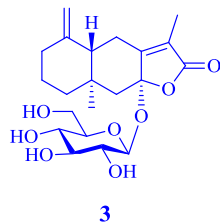


Figure S52. The  $^1\text{H}$ - $^1\text{H}$  gCOSY spectrum of compound **3** in acetone- $d_6$  (600 MHz).

VNS-600 gHSQCAD DS-135 IN acetone Jun 5 2014

Temp. 25.0 C / 298.1 K  
Sample #8, Operator: vjwalk

Relax. delay 1.000 sec  
Acq. time 0.150 sec  
Width 5364.8 Hz  
2D Width 30154.5 Hz  
12 repetitions  
140 increments  
OBSERVE H1, 599.6908056 MHz  
DECOUPLE C13, 150.8059420 MHz  
Power 36 dB  
on during acquisition  
off during delay  
W40\_NEW-SW modulated  
DATA PROCESSING  
Sine bell 0.036 sec  
F1 DATA PROCESSING  
Sine bell 0.005 sec  
FT size 4096 x 2048  
Total time 34 min

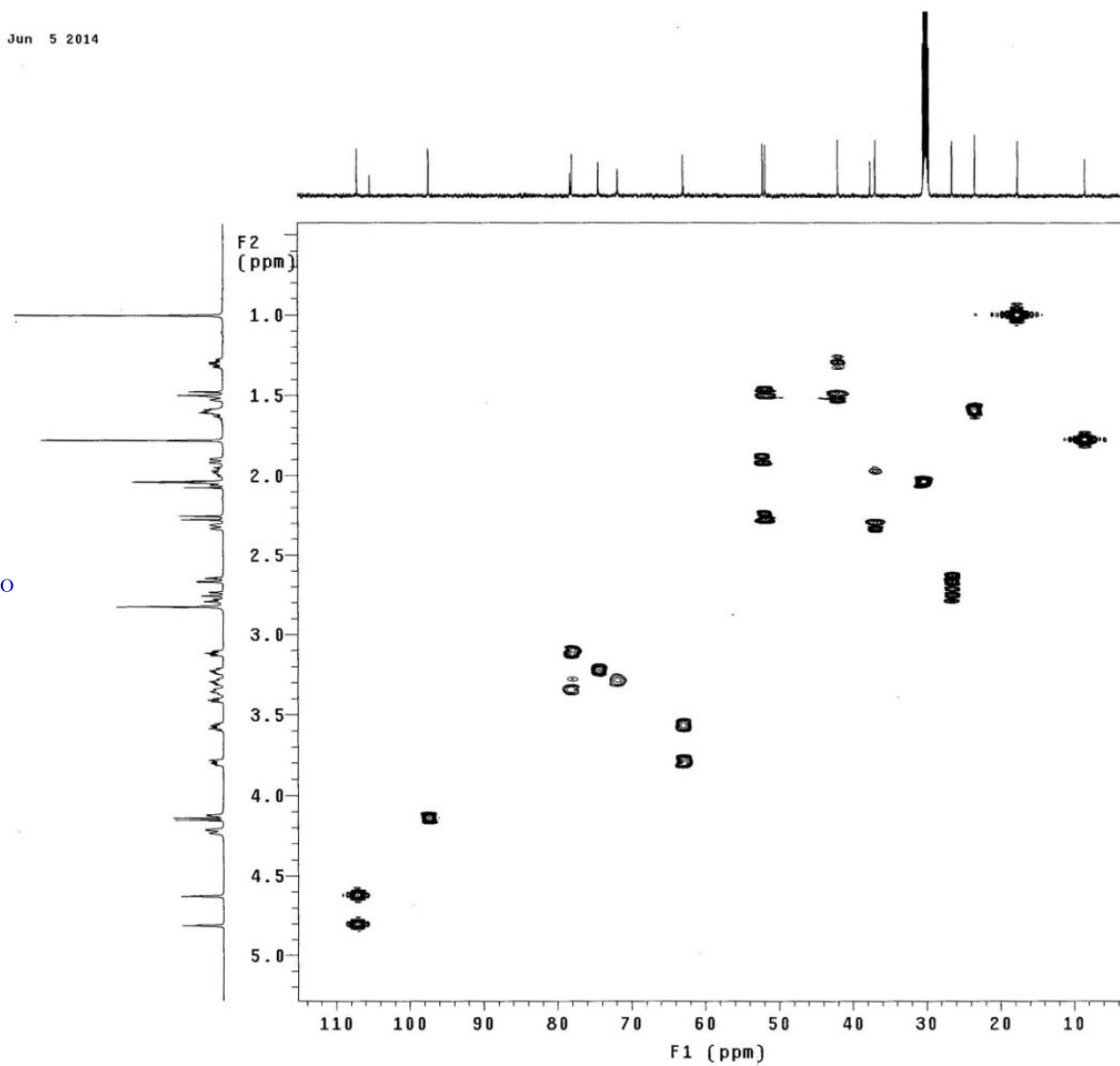
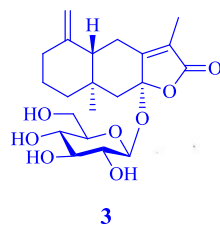


Figure S53. The gHSQC spectrum of compound **3** in acetone- $d_6$  (600 MHz for  $^1\text{H}$ ).

VNS-600 gHMBCAD DS-135 IN acetone Jun 5 2014

Temp. 25.0 C / 298.1 K  
Sample #8, Operator: vjwalk

Relax. delay 1.000 sec  
Acq. time 0.150 sec  
Width 5364.8 Hz  
2D Width 36182.7 Hz  
24 repetitions  
2 x 80 increments  
OBSERVE H1, 599.6908047 MHz  
DATA PROCESSING  
Sq. sine bell 0.075 sec  
F1 DATA PROCESSING  
Gauss apodization 0.002 sec  
FT size 4096 x 2048  
Total time 1 hr, 19 min

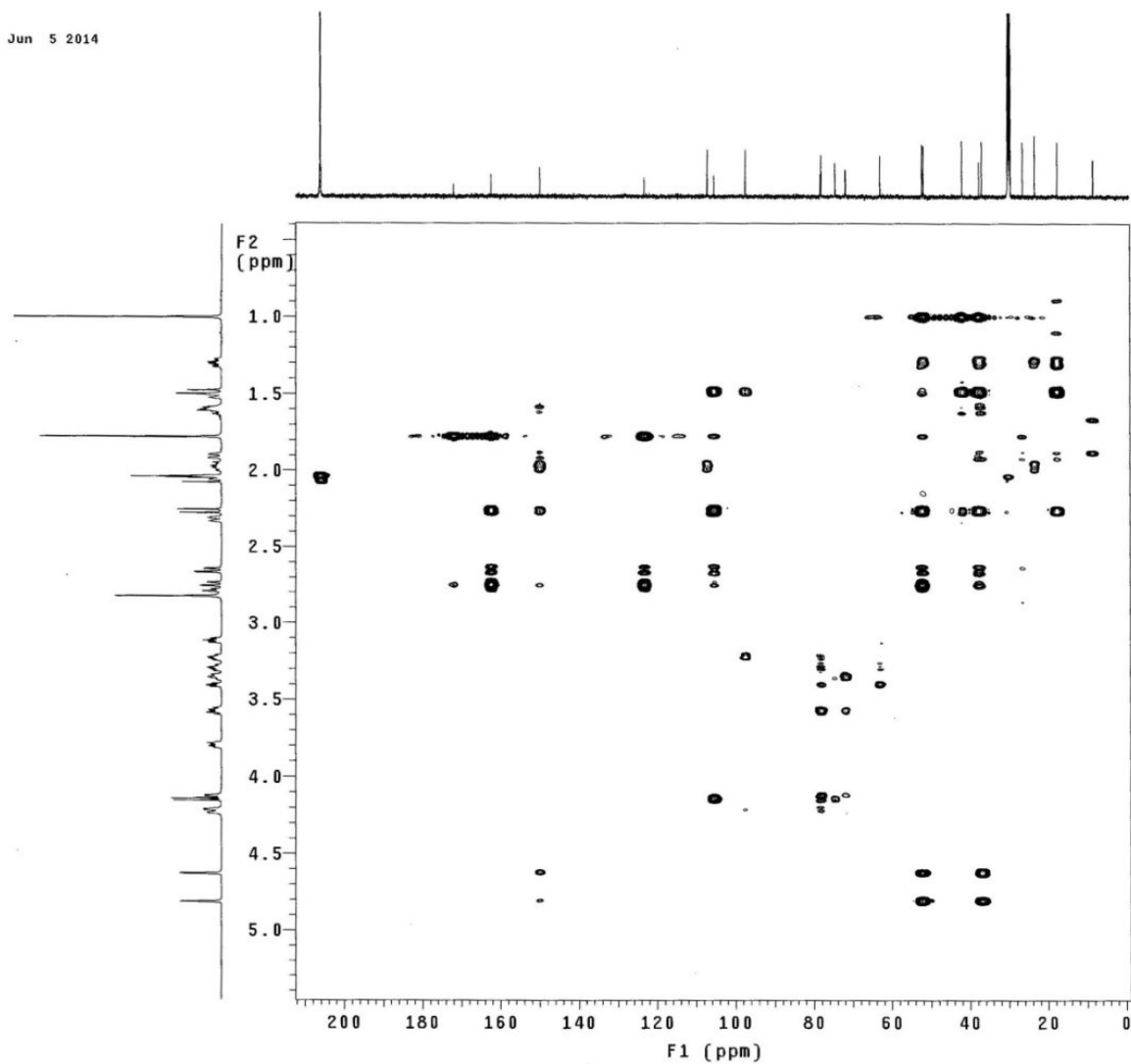
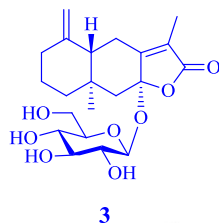
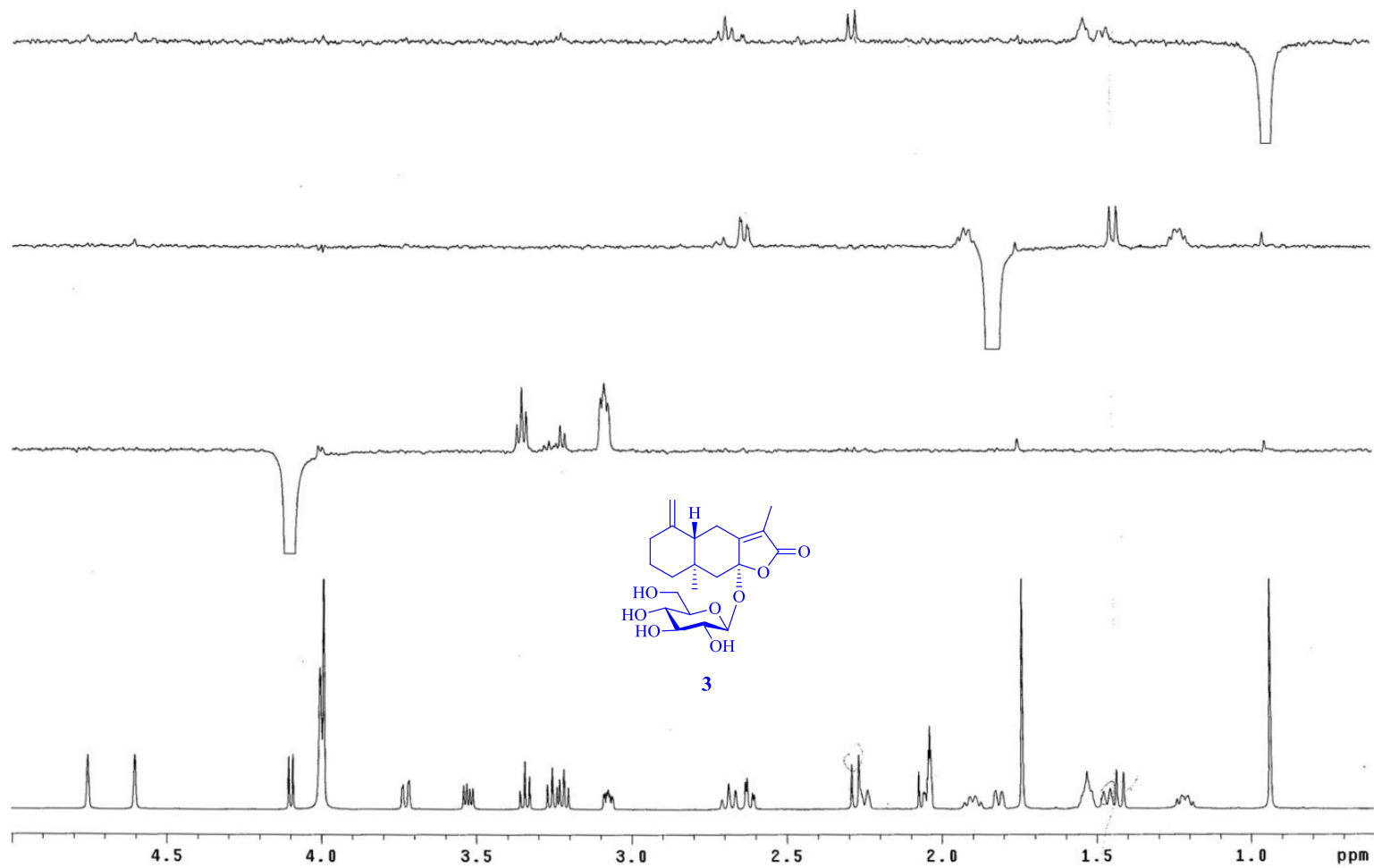


Figure S54. The gHMBC spectrum of compound **3** in acetone- $d_6$  (600 MHz for  $^1\text{H}$ ).



**Figure S55.** The NOE difference spectrum of **3** in acetone-*d*<sub>6</sub> (600 MHz).

Bruker AVIIIHD 600 20141027 DS-135a  
 PROTON Acetone D:\\ DATA2014 3

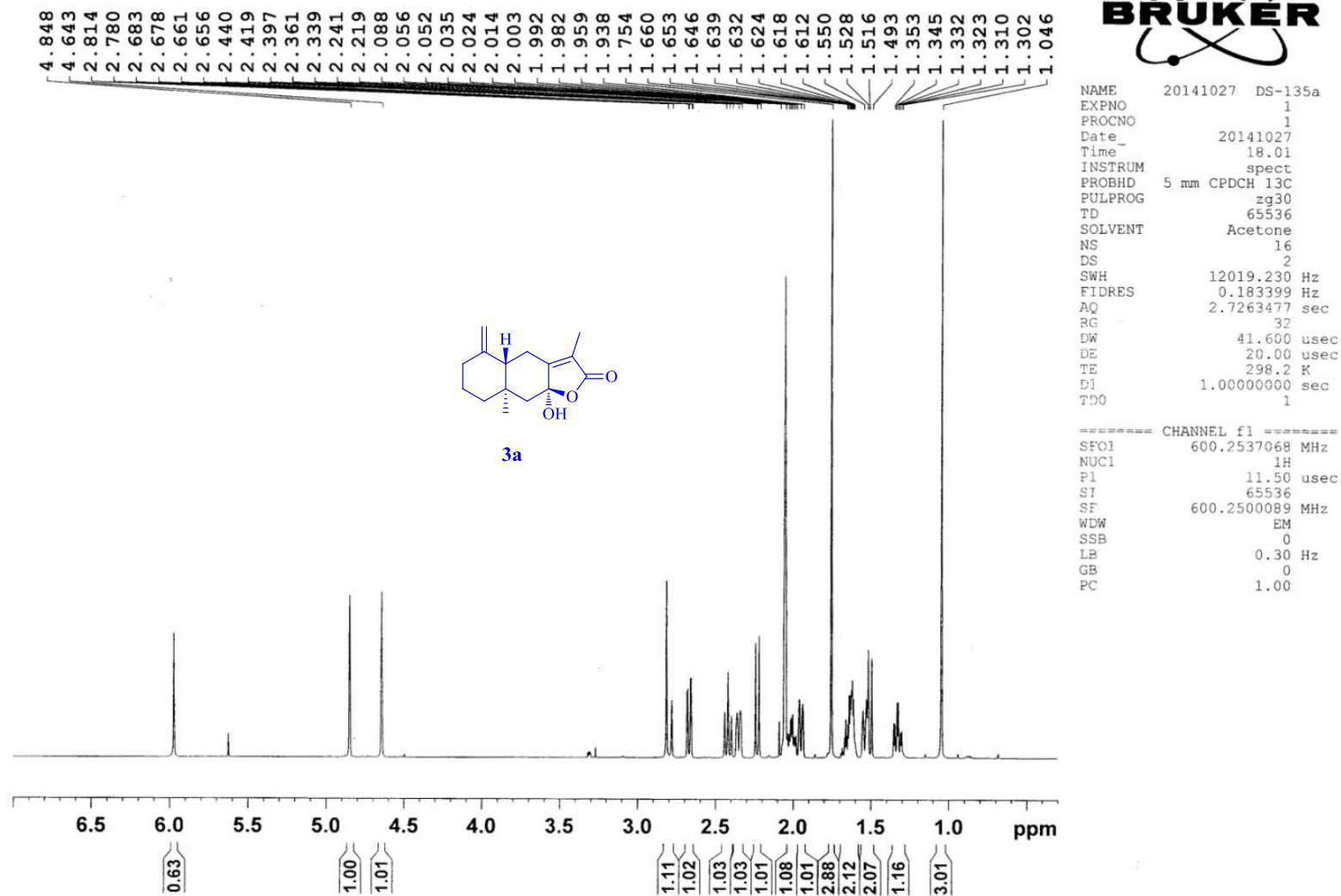
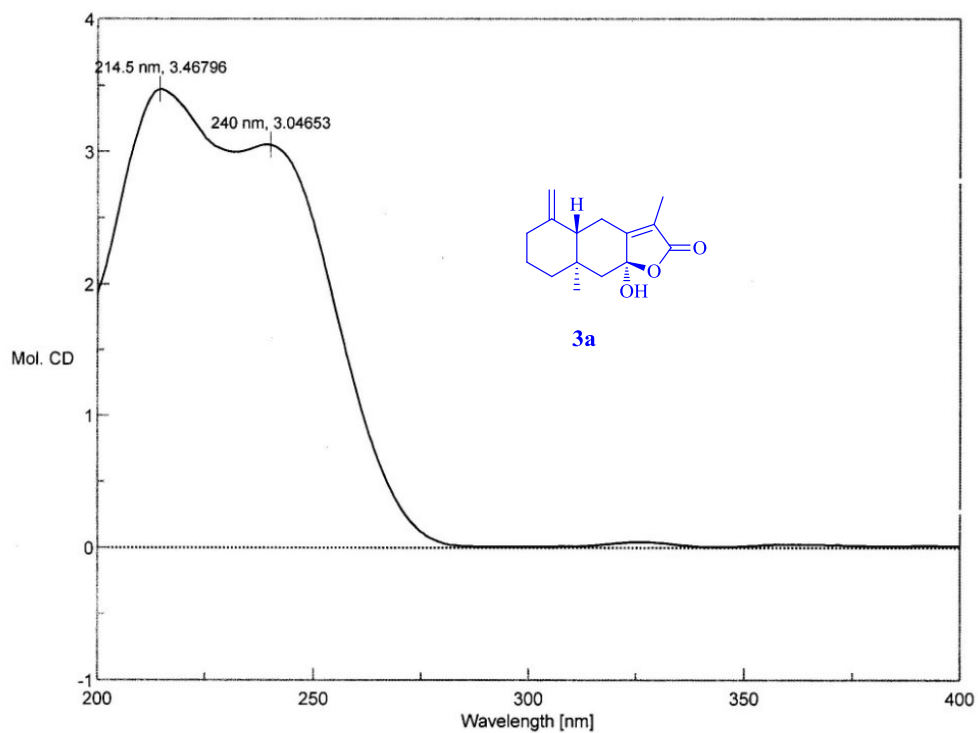


Figure S56. The  $^1\text{H}$  NMR spectrum of the aglycone of **3** in acetone- $d_6$  (600 M).



[Comments]  
 Sample name DS-F1  
 Comment  
 User  
 Division  
 Company dell

[Measurement Information]  
 Instrument Name J-815  
 Model Name J-815  
 Serial No. A024461168

Accessory Standard  
 Accessory S/N A024461168  
 Cell Length 1 mm

Photometric Mode CD, HT, Abs  
 Measure Range 400 - 200 nm  
 Data pitch 0.5 nm  
 Sensitivity Standard  
 D.I.T. 2 sec  
 Band width 2.00 nm  
 Start Mode Immediately  
 Scanning Speed 100 nm/min  
 Baseline Correction Baseline  
 Shutter Control Auto  
 PMT Voltage Auto  
 Accumulations 3  
 Solvent MEOH  
 Concentration 0.41 (w/v)%

[Detailed Information]  
 Creation date 2014-5-19 16:07

Data array type Linear data array \* 3  
 Horizontal axis Wavelength [nm]  
 Vertical axis(1) Mol. CD  
 Vertical axis(2) HT [V]  
 Vertical axis(3) Abs  
 Start 400 nm  
 End 200 nm  
 Data interval 0.5 nm  
 Data points 401

**Figure S57.** The CD spectrum of the aglycone of **3** in MeOH.

Bruker AVIIIHD 600 20141027 DS-135G  
PROTON D2O D:\ DATA2014 2

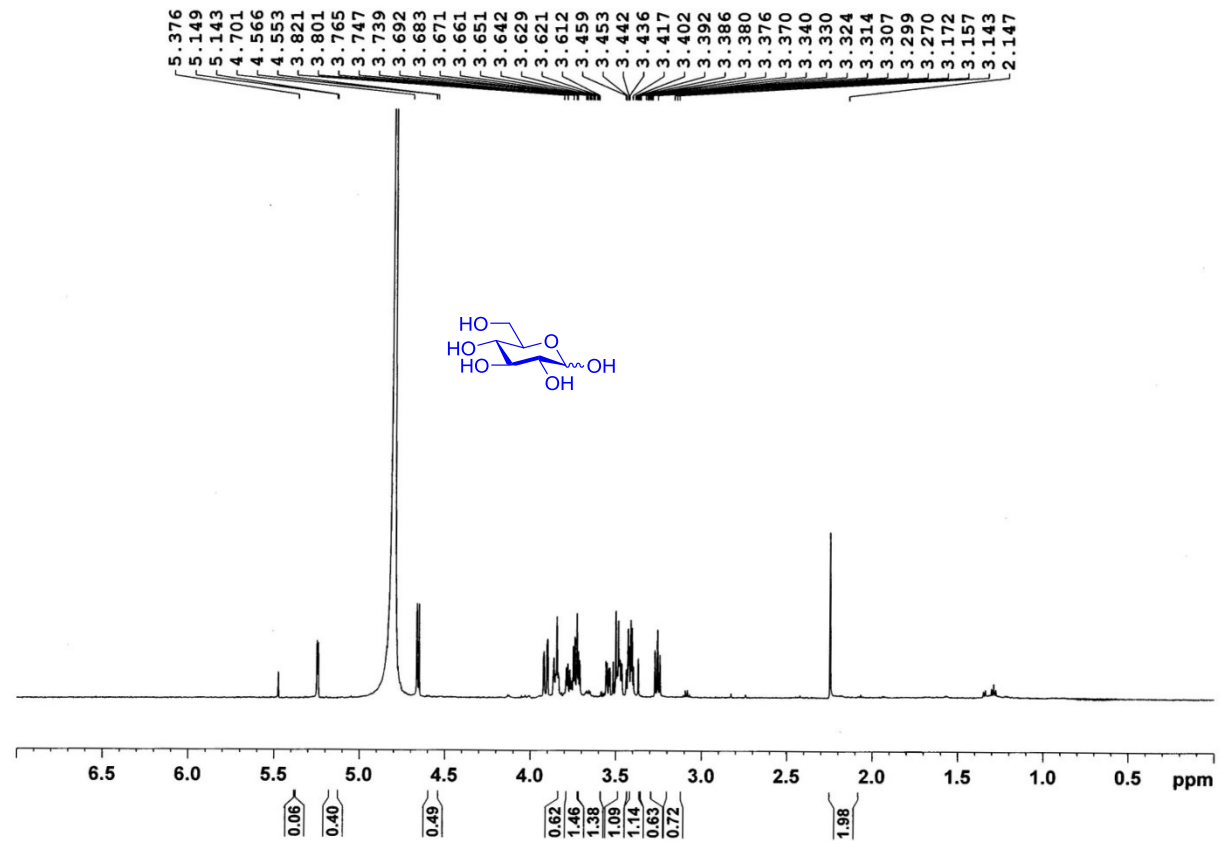


Figure S58. The <sup>1</sup>H NMR spectrum of D-glucose isolated from the hydrolysate of **3** in D<sub>2</sub>O (600 M).



Bruker AVIIIHD 600 20141017 DS-D-Glu  
PROTON D2O D:\\ DATA2014 22

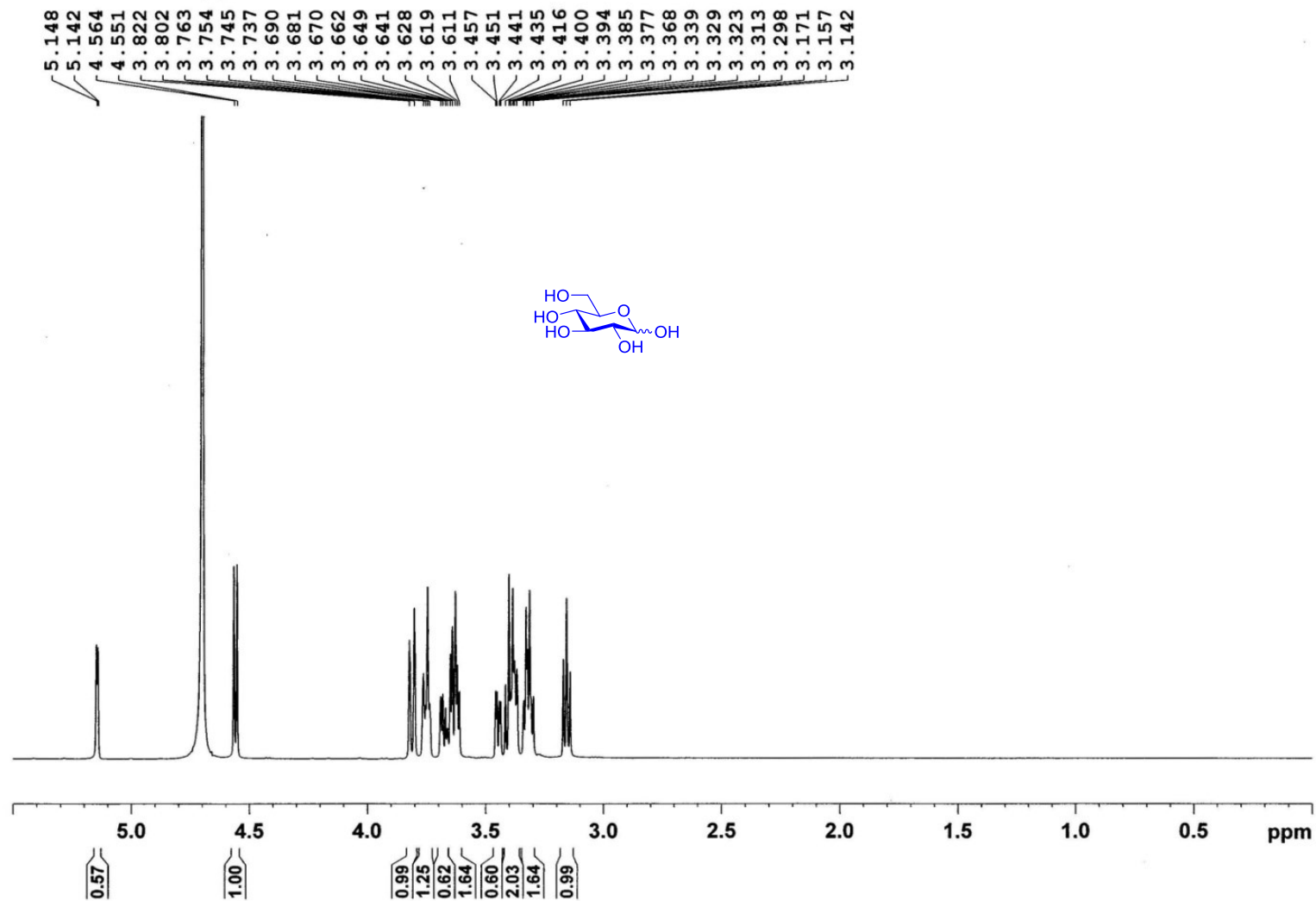


Figure S59. The  $^1\text{H}$  NMR spectrum of the authentic D-glucose in  $\text{D}_2\text{O}$  (600 M).