

**Rspo1-activated signalling molecules are sufficient to induce ovarian differentiation in XY medaka (*Oryzias latipes*)**

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## Supplementary figures

Figure S1. **Transgene construction and assessment of transgenesis.** (A) Schematics of pIRES-hrGFP1a-Rspo1 overexpression plasmid. (B) Genotyping and integration check of *Rspo1*-overexpressed adults. Upper panel, abundant amplifications were detected in both F<sub>0</sub> and F<sub>1</sub> (XY fish) injected with pIRES-hrGFP1a-Rspo1, but no amplification was found in control XX and XY fish. Lower panel, genetic sexing of control and transgenic medaka by genomic PCR, where 2 bands (Dmy and Dmrt1) indicates genetic male, while 1 band (Dmrt1) represents female. (C) Analysis of tissue specificity of hCMV promoter driven *Rspo1* transgene in F<sub>0</sub> generation. Real-time *Rspo1* mRNA expressions were highly localized in gonads and brain, while significantly low expressions were recorded in the remaining body (RB). Data are shown as mean  $\pm$  S.E. of three independent experiments and the letters (a, b, c, etc.) above the bars indicate that these groups differ significantly from each other at  $p < 0.05$ .

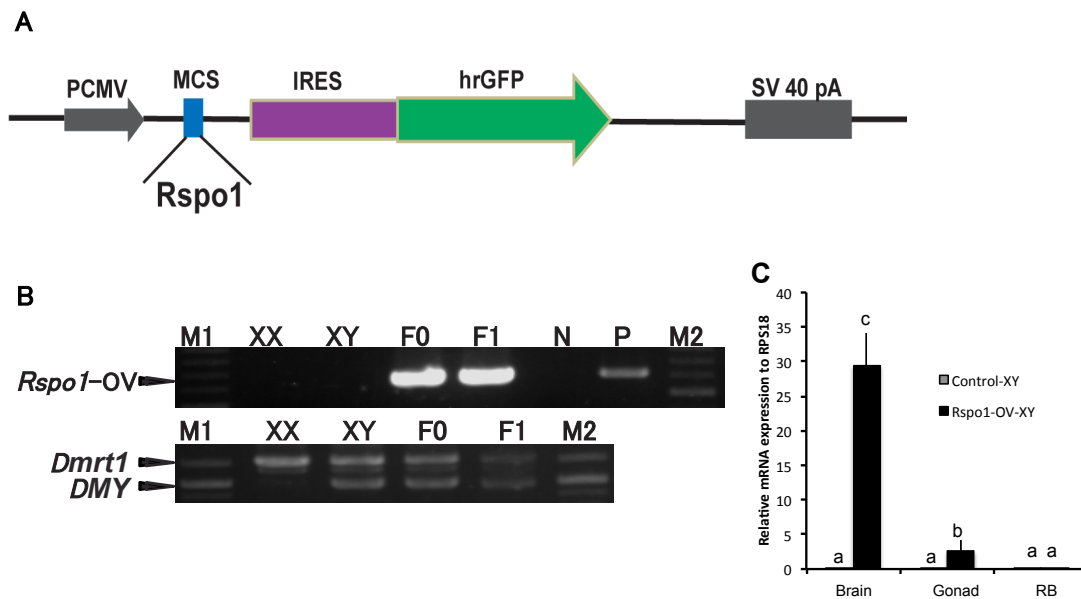
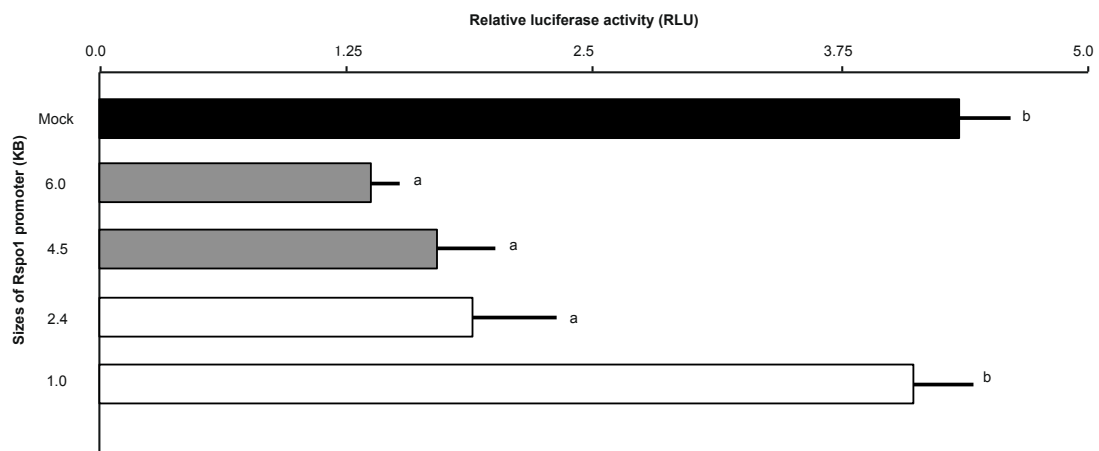


Figure S2. **Minimal promoter analysis of *Rspo1*.** *Dmy* expression plasmid were co-transfected with different sized medaka *Rspo1* (1-6 kb) promoter constructs, and relative luciferase activity (RLU) was measured after 48 hours. The RLUs of each expression plasmid (s) were plotted on Y-axis to prepare the graph. Data are shown as mean  $\pm$  S.E. of three independent experiments and the letters (a, b, c, etc.) above the bars indicate that these groups differ significantly from each other at  $p < 0.01$ . Note: the length of different promoter constructs were determined empirically using the availability of potential *DM* domain binding sites.



## Supplementary table

**SI Table 1: Screening of *Rspo1*-OV-XY F<sub>0</sub> progenies.**

Genotype	Phenotype		Fertility		
	XY	female	male	spawning	non-spawning
<b>38</b>	<b>21</b>	<b>17</b>	<b>9 (24%)</b>	<b>12 (32%)</b>	<b>17 (44%)</b>

**SI Table 2: List of major primers used in this study.**

Primer name	Primer sequence (5' to 3')	Purpose
<i>Rspo1</i> -GFP-F	GAGCTCCCGCGGATGCAGCTGGGACTGGTG	Amplification of <i>Rspo1</i>
<i>Rspo1</i> -GFP-R	GGATCCGAATTCGGTGACGGAGCTGGTTGTG	
<i>Rspo1</i> -gPCR-F	ACATGTGGGTTTAAGAAAGGC	Genomic PCR of <i>Rspo1</i> transgene
<i>Rspo1</i> -gPCR-R	GTCCTTATCATCGTCGTCTT	
<i>DMY</i> -gPCR-F	CCG GGTGCCCAAGTGCTCCCGCTG	Genomic PCR of <i>DMY</i>
<i>DMY</i> -gPCR-R	GATCGTCCCTCCACAGAGAAGAGA	
<i>β-catenin</i> -qPCR-F	ACTGGATATCGGAGCACAGG	Real-time PCR
<i>β-catenin</i> -qPCR-R	CAGGGAGAGCATCAGTGTGA	
<i>Dmrt1</i> -qPCR-F	TCCTCCTACTATGGAAACCTGTACCA	Real-time PCR
<i>Dmrt1</i> -qPCR-R	GAAGGAGTGCATGCGGTACTG	
<i>Cyp19a1a</i> -qPCR-F	AGCTTATTTTTGCCCAAGGCC	Real-time PCR
<i>Cyp19a1a</i> -qPCR-R	TTGAGCAGCAGGAGCATGAAA	
<i>Gsdf</i> -qPCR-F	GGGCTGGACACTATTCGAGA	Real-time PCR
<i>Gsdf</i> -qPCR-R	CATGACACAGAGGAGCTGGA	
<i>Dmy</i> -qPCR-F	TCCTATTATGGAAACCTGCACAACCTAC	Real-time PCR
<i>Dmy</i> -qPCR-R	GAAGGAGTGCATGCGGTACG	
<i>Sox9</i> -qPCR-F	CTCCGACGCTCCCAGTCCCA	Real-time PCR
<i>Sox9</i> -qPCR-R	GACGGCCGGGTGTTTTCGGT	
<i>Wnt4b</i> -qPCR-F	CTGGAAAGTCATGCCTCCAT	Real-time PCR
<i>Wnt4b</i> -qPCR-R	TACGCGTTAAGCGAACCTCT	
<i>EF1 α</i> -F14	CAGCTTCAACGCTCAGGTCAT	Real-time PCR
<i>EF1 α</i> -R14	TGAACTTGCAGGCGATGTGA	
<i>Rspo1</i> -pro-F	GGGTACCACGCGTGCCTGCCCTTATGGAGCAG	Amplification of promoter region
<i>Rspo1</i> -pro-R	TAATACTACC GAGATCTCTCGAGGATGATCCCGCTTTTCTCCTG	