

Autologous skeletal muscle derived cells expressing a novel functional dystrophin provide a potential therapy for Duchenne Muscular Dystrophy

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Supplementary material

Cloning of mini-dystrophin constructs

The truncation in C1 comprises 3672 bases between nucleotide 1619 and 5290, and 6801 bases in C2 between nucleotide 2199 and 8999 (GenBank NM 004006) (Figure 1A, supplementary materials). The 7.4 kb C1/ Δ R3-R13 and the 4.2 kb C2/ Δ H2-R23 mini-dystrophins were generated by splicing by overlapping extension (SOE-PCR) using human full-length dystrophin cDNA as template. There are 2 polymorphisms in C1 constructs: G/A in position 142 of dystrophin Exon 59, which is a non-pathogenic SNP (rs 1800280), and an extra –GCC inframe insertion at position 31 of Exon 47, which predicted by Mutationtaster website to be polymorphism. (<http://www.mutationtaster.org/cgi-bin/MutationTaster/MutationTaster69.cgi>). There are also 2 polymorphisms in the C2 construct: G/A in position 142 of dystrophin Exon 59; C/T in position 160 of Exon 59 which is a variant with low frequency in the ExAC database, predicted to be polymorphism.

The amplification products and the vector were digested with restriction enzymes *NgoMIV* and *ClaI*, purified and ligated back into the cDNA. The resulting constructs were fully sequenced (MWG Biotech, Ebersberg, Germany) for verification of the correct sequence and reading frame.

Generation of lentiviral vectors

An *AgeI* site was inserted into pCMV. Δ H2R23, immediately upstream of the dystrophin 5'UTR. pRRL.desmin.C2, was then assembled by cloning the C2 mini-dystrophin from the *AgeI*-modified pCMV. Δ H2R23 into pRRL.sin.cppt.desmin.GFP using sites *AgeI-SalI*.

pRRL.desmin.C1 was constructed by cloning the respective regions from pCMV. Δ R3R13 into the pRRL.desmin.C2 construct using *NsiI* and *SalI* restriction sites. A GFP tag was then introduced to all constructs by replacing the C-terminal region of each construct with that of pCihDystrophinGFP using the *BstBI* and *SalI* restriction sites for C2 and the *BstEII* and *SalI* sites for C1. For promoter comparisons, the SFFV promoter was PCR cloned from pRRL.sin.cPPT.SFFV.NIGW and inserted in place of the desmin promoter. The final clones were verified by DNA sequencing.

Titration of lentiviruses

Viral RNA titres were measured in the purified viral harvests by quantitative reverse transcription polymerase chain reaction (qRT-PCR) using a commercial kit (CloneTech cat no. 631235). Viral RNA was extracted directly from the viral preparation and DNase treated prior to qRT-PCR, which was then carried out in a single reaction. Complementary DNA was generated during an initial reverse transcription step (42°C 5 min, 95°C 10 sec), which was directly followed by real time PCR amplification (40 cycles of: 95°C 5sec; 60°C 30sec). The titre was then calculated by extrapolation of a standard curve generated from RNA standards

of known copy number.

Proviral titre was determined using genomic DNA extracted from transduced HeLa cells, adapted from a reported method⁴³. Briefly, 1×10^5 HeLa cells were plated into each well of a 6 well plate and transduced with a range of volumes of the concentrated lentivirus. After 72 hours, the transduced HeLa genomic DNA was extracted and analysed by SYBR Green amplified qPCR. The lentiviral copy number was determined by PCR directed against the LTR using the primers 5'-AGCTTGCCTTGAGTGCTTCAA-3' and 5'-AGGGTCTGAGGGATCTCTAGTTACC-3'. The proviral copy number was normalised to the total allelic copy number by PCR of human albumin using primers 5'-GCTGTCATCTCTTGTGGGCTGT-3' and 5'-ACTCATGGGAGCTGCTGGTTC-3'.

Integrated proviral copy numbers were calculated by Alu-PCR, adapted from a reported protocol⁴⁴. Briefly, 10ng of virally-transduced HeLa gDNA was amplified using conditions (95°C 15 minutes; 25 cycles of 95°C 30 sec, 55°C 30 sec, 72°C 3min30sec; 72°C 10mins) and primers Alu1 (5'-TCC CAG CTA CTG GGG AGG CTG AGG-3'), Alu2 (5'-GCC TCC CAA AGT GCT GGG ATT ACA G-3') and M667-L (5'-ATG CCA CGT AAG CGA AAC TCT GGC TAA CTA GGG AAC CCA CTG-3) to amplify integrated lentiviruses. A control reaction was processed omitting the Alu1 and Alu2 primers to account for any linear, single-stranded amplification from the M667-L oligo. The resulting amplicons were then diluted 1/100 in water and 5µl was used in a nested qPCR reaction which was amplified with a SYBR Green master mix under conditions (95°C 15 minutes; 25 cycles of 95°C 30 sec, 55°C 30 sec, 72°C 3min30sec; 72°C 10mins) and the primers AA55M (5'-GCT AGA GAT TTT CCA CAC TGA CTA A-3') and Lamda T (5'-ATG CCA CGT AAG CGA AAC T-3') to detect the incidence of LTR copies in the products. The relative level of integrated copies were calculated by $2^{-\Delta \Delta ct}$, in which the ct value for Alu-containing reactions was subtracted

from the control reaction, before this value was subtracted from the non-transduced sample value. The integrated proviral titre was then expressed relative to a sample of known titre.

The functional titre of the lentivirus was determined according to GFP output by transducing HeLa cells with serial dilutions of vector preparations and measuring the percentage of GFP positive cells by flow cytometry.

Sequence of C1 construct (from ATG to EGFP)

ATGCTTTGGTGGGAAGAAGTAGAGGACTGTTATGAAAGAGAAGATGTTCAAAG
AAAACATTCACAAAATGGGTAAATGCACAATTTTCTAAGTTTGGGAAGCAGCAT
ATTGAGAACCTCTTCAGTGACCTACAGGATGGGAGGCGCCTCCTAGACCTCCTCG
AAGGCCTGACAGGGCAAAAAGTCCAAAAGAAAAGGATCCACAAGAGTTCAT
GCCCTGAACAATGTCAACAAGGCACTGCGGGTTTTGCAGAACAATAATGTTGATT
TAGTGAATATTGGAAGTACTGACATCGTAGATGGAAATCATAAACTGACTCTTGG
TTTGATTTGGAATATAATCCTCCACTGGCAGGTCAAAAATGTAATGAAAAATATC
ATGGCTGGATTGCAACAAACCAACAGTGAAAAGATTCTCCTGAGCTGGGTCCGA
CAATCAACTCGTAATTATCCACAGGTTAATGTAATCAACTTCACCACCAGCTGGT
CTGATGGCCTGGCTTTGAATGCTCTCATCCATAGTCATAGGCCAGACCTATTTGA
CTGGAATAGTGTGGTTTGCCAGCAGTCAGCCACACAACGACTGGAACATGCATT
CAACATCGCCAGATATCAATTAGGCATAGAGAACTACTCGATCCTGAAGATGT
TGATAACCACCTATCCAGATAAGAAGTCCATCTTAATGTACATCACATCACTCTTC
CAAGTTTTGCCTCAACAAGTGAGCATTGAAGCCATCCAGGAAGTGGAAATGTTG
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CACTATTCTCAACAGATCACGGTCAGTCTAGCACAGGGATATGAGAGAACTTCTT
CCCCTAAGCCTCGATTCAAGAGCTATGCCTACACACAGGCTGCTTATGTCACCAC
CTCTGACCCTACACGGAGCCCATTTCTTCACAGCATTGGAAGCTCCTGAAGAC

AAGTCATTTGGCAGTTCATTGATGGAGAGTGAAGTAAACCTGGACCGTTATCAAA
CAGCTTTAGAAGAAGTATTATCGTGGCTTCTTTCTGCTGAGGACACATTGCAAGC
ACAAGGAGAGATTTCTAATGATGTGGAAGTGGTGAAGACCAGTTTCATACTCA
TGAGGGGTACATGATGGATTTGACAGCCATCAGGGCCGGGTTGGTAATATTCTA
CAATTGGGAAGTAAGCTGATTGGAACAGGAAAATTATCAGAAGATGAAGAACT
GAAGTACAAGAGCAGATGAATCTCCTAAATTCAAGATGGGAATGCCTCAGGGTA
GCTAGCATGGAAAAACAAAGCAATTTACATAGAGTTTTAATGGATCTCCAGAAT
CAGAAACTGAAAGAGTTGAATGACTGGCTAACAAAAACAGAAGAAAGAACAAG
GAAAATGGAGGAAGAGCCTCTTGGACCTGATCTTGAAGACCTAAAACGCCAAGT
ACAACAACATAAGGTGCTTCAAGAAGATCTAGAACAAGAACAAGTCAGGGTCAA
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GCTGCTTTGGAAGAACAACCTAAGGTATTGGGAGATCGATTTGCAGCCATTTAC
ACAGAATTAAGACTGGAAAGGCCTCCATTCCTTTGAAGGAATTGGAGCAGTTTA
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TGAATCTGAAAGAGGAAGACTTCAATAAAGATATGAATGAAGACAATGAGGGTA
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AGAGAAAGCGAGAGGAAATAAAGATAAAACAGCAGCTGTTACAGACAAAACAT
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AAGAAGACAGCAGCATTGCAAAGTGCAACGCCTGTGGAAAGGGTGAAGCTACA
GGAAGCTCTCTCCCAGCTTGATTTCCAATGGGAAAAAGTTAACAAAATGTACAA
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GGCAGATTTCAACCGGGCTTGGACAGAACTTACCGACTGGCTTTCTCTGCTTGAT
CAAGTTATAAAATCACAGAGGGTGATGGTGGGTGACCTTGAGGATATCAACGAG
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GAACTTCTGGTGTGGCTACAGCTGAAAGATGATGAATTAAGCCGGCAGGCACC
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GGCCACGGATGAGCTGGACCTCAAGCTGCGCCAAGCTGAGGTGATCAAGGGATC
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TCTGGCCAGTAGATTCTGCGCCTGCCTCGTCCCCTCAGCTTTCACACGATGATACT
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GGATCTTATCTAAATGATAGCATCTCTCCTAATGAGAGCATAGATGATGAACATT
TGTTAATCCAGCATTACTGCCAAAGTTTGAACCAGGACTCCCCCCTGAGCCAGCC
TCGTAGTCCTGCCAGATCTTGATTTCTTAGAGAGTGAGGAAAGAGGGGAGCT
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CTCCTGAAATGATGCCACCTCTCCCCAGAGTCCCCGGGATGCTGAGCTCATTGC
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CCTGGAAGACCACAATAAACAGCTGGAGTCACAGTTACACAGGCTAAGGCAGCT
GCTGGAGCAACCCCAGGCAGAGGCCAAAGTGAATGGCACAACGGTGTCTCTCC
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GGCAGTCAAACCTTCGGACTCCATGGGTGAGGAAGATCTTCTCAGTCCTCCCCAGG
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GTTCAAGAGGAAGAAATACCCCTGGAAAGCCAATGAGAGAGGACACAATGGCC
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GTCGAGCTGGACGGCGACGTAAACGGCCACAAGTTCAGCGTGTCCGGCGAGGGC
GAGGGCGATGCCACCTACGGCAAGCTGACCCTGAAGTTCATCTGCACCACCGGC
AAGCTGCCCCTGCCCTGGCCCACCTCGTGACCACCTGACCTACGGCGTGCAGT
GCTTCAGCCGCTACCCCGACCACATGAAGCAGCACGACTTCTTCAAGTCCGCCAT
GCCCCAAGGCTACGTCCAGGAGCGCACCATCTTCTTCAAGGACGACGGCAACTA
CAAGACCCGCGCCGAGGTGAAGTTCGAGGGCGACACCCTGGTGAACCGCATCGA

GCTGAAGGGCATCGACTTCAAGGAGGACGGCAACATCCTGGGGCACAAGCTGGA
GTACAACTACAACAGCCACAACGTCTATATCATGGCCGACAAGCAGAAGAACGG
CATCAAGGTGAACTTCAAGATCCGCCACAACATCGAGGACGGCAGCGTGCAGCT
CGCCGACCACTACCAGCAGAACACCCCCATCGGCGACGGCCCCGTGCTGCTGCC
CGACAACCACTACCTGAGCACCCAGTCCGCCCTGAGCAAAGACCCCAACGAGAA
GCGCGATCACATGGTCCTGCTGGAGTTCGTGACCGCCGCCGGGATCACTCTCGGC
ATGGACGAGCTGTACAAGTAAA

Sequence of C2 construct (from ATG to EGFP):

ATGCTTTGGTGGGAAGAAGTAGAGGACTGTTATGAAAGAGAAGATGTTCAAAG
AAAACATTCACAAAATGGGTAAATGCACAATTTTCTAAGTTTGGGAAGCAGCAT
ATTGAGAACCTCTTCAGTGACCTACAGGATGGGAGGCGCCTCCTAGACCTCCTCG
AAGGCCTGACAGGGCAAAAAGTCCAAAAGAAAAAGGATCCACAAGAGTTCAT
GCCCTGAACAATGTCAACAAGGCACTGCGGGTTTTGCAGAACAATAATGTTGATT
TAGTGAATATTGGAAGTACTGACATCGTAGATGGAAATCATAAACTGACTCTTGG
TTTGATTTGGAATATAATCCTCCACTGGCAGGTCAAAAATGTAATGAAAAATATC
ATGGCTGGATTGCAACAAACCAACAGTGAAAAGATTCTCCTGAGCTGGGTCCGA
CAATCAACTCGTAATTATCCACAGGTAAATGTAATCAACTTCACCACCAGCTGGT
CTGATGGCCTGGCTTTGAATGCTCTCATCCATAGTCATAGGCCAGACCTATTTGA
CTGGAATAGTGTGGTTTGCCAGCAGTCAGCCACACAACGACTGGAACATGCATT
CAACATCGCCAGATATCAATTAGGCATAGAGAACTACTCGATCCTGAAGATGT
TGATAACCACCTATCCAGATAAGAAGTCCATCTTAATGTACATCACATCACTCTTC
CAAGTTTTGCCTCAACAAGTGAGCATTGAAGCCATCCAGGAAGTGGAAATGTTG
CCAAGGCCACCTAAAGTGACTAAAGAAGAACATTTTCAGTTACATCATCAAATG
CACTATTCTCAACAGATCACGGTCAGTCTAGCACAGGGATATGAGAGAACTTCTT

CCCCTAAGCCTCGATTCAAGAGCTATGCCTACACACAGGCTGCTTATGTCACCAC
CTCTGACCCTACACGGAGCCCATTTCCTTACAGCATTGGAAGCTCCTGAAGAC
AAGTCATTTGGCAGTTCATTGATGGAGAGTGAAGTAAACCTGGACCGTTATCAAA
CAGCTTTAGAAGAAGTATTATCGTGGCTTCTTTCTGCTGAGGACACATTGCAAGC
ACAAGGAGAGATTTCTAATGATGTGGAAGTGGTGAAAGACCAGTTTCATACTCA
TGAGGGGTACATGATGGATTTGACAGCCCATCAGGGCCGGGTTGGTAATATTCTA
CAATTGGGAAGTAAGCTGATTGGAACAGGAAAATTATCAGAAGATGAAGAACT
GAAGTACAAGAGCAGATGAATCTCCTAAATTCAAGATGGGAATGCCTCAGGGTA
GCTAGCATGGAAAAACAAAGCAATTTACATAGAGTTTTAATGGATCTCCAGAAT
CAGAACTGAAAGAGTTGAATGACTGGCTAACAAAAACAGAAGAAAGAACAAG
GAAAATGGAGGAAGAGCCTCTTGGACCTGATCTTGAAGACCTAAAACGCCAAGT
ACAACAACATAAGGTGCTTCAAGAAGATCTAGAACAAGAACAAGTCAGGGTCAA
TTCTCTCACTCACATGGTGGTGGTAGTTGATGAATCTAGTGGAGATCACGCAACT
GCTGCTTTGGAAGAACAACCTTAAGGTATTGGGAGATCGATGGGCAAACATCTGT
AGATGGACAGAAGACCGCTGGGTTCTTTTACAAGACATCCTTCTCAAATGGCAAC
GTCTTACTGAAGAACAGTGCCTTTTTAGTGCATGGCTTTCAGAAAAAGAAGATGC
AGTGAACAAGATTCACACAACCTGGCTTTAAAGATCAAAATGAAATGTTATCAAG
TCTTCAAAAACCTGGCCGTTTTAAAAGCGGATCTAGAAAAGAAAAAGCAATCCAT
GGGCAAACCTGTATTCACTCAAACAAGATCTTCTTTCAACACTGAAGAATAAGTCA
GTGACCCAGAAGACGGAAGCATGGCTGGATAACTTTGCCCGGTGTTGGGATAAT
TTAGTCCAAAAACTTGAAAAGAGTACAGCACAGACCCTTGAAAGACTCCAGGAA
CTTCAAGAGGCCATGGATGAGCTGGACCTCAAGCTGCGCCAAGCTGAGGTGATC
AAGGGATCCTGGCAGCCCGTGGGCGATCTCCTCATTGACTCTCTCCAAGATCACC
TCGAGAAAGTCAAGGCACTTCGAGGAGAAATTGCGCCTCTGAAAGAGAACGTGA
GCCACGTCAATGACCTTGCTCGCCAGCTTACCACTTTGGGCATTCAGCTCTCACC

GTATAACCTCAGCACTCTGGAAGACCTGAACACCAGATGGAAGCTTCTGCAGGT
GGCCGTCGAGGACCGAGTCAGGCAGCTGCATGAAGCCCACAGGGACTTTGGTCC
AGCATCTCAGCACTTTCTTTCCACGTCTGTCCAGGGTCCCTGGGAGAGAGCCATC
TCGCCAAACAAAGTGCCCTACTATATCAACCACGAGACTCAAACAACCTTGCTGG
GACCATCCCAAATGACAGAGCTCTACCAGTCTTTAGCTGACCTGAATAATGTCA
GATTCTCAGCTTATAGGACTGCCATGAAACTCCGAAGACTGCAGAAGGCCCTTTG
CTTGGATCTCTTGAGCCTGTCAGCTGCATGTGATGCCTTGGACCAGCACAACTC
AAGCAAAATGACCAGCCCATGGATATCCTGCAGATTATTAATTGTTTGACCACTA
TTTATGACCGCCTGGAGCAAGAGCACAACAATTTGGTCAACGTCCCTCTCTGCGT
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GATCCGTGTCCTGTCTTTTAAAACCTGGCATCATTTCCCTGTGTAAAGCACATTTGG
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GGGTGAAGTTGCATCCTTTGGGGGCAGTAACATTGAGCCAAGTGTCCGGAGCTG
CTTCCAATTTGCTAATAATAAGCCAGAGATCGAAGCGGCCCTCTTCCTAGACTGG
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GCTGCAGAACTGCCAAGCATCAGGCCAAATGTAACATCTGCAAAGAGTGTCCA
ATCATTGGATTCAGGTACAGGAGTCTAAAGCACTTTAATTATGACATCTGCCAAA
GCTGCTTTTTTTCTGGTCGAGTTGCAAAAGGCCATAAAATGCACTATCCCATGGT
GGAATATTGCACTCCGACTACATCAGGAGAAGATGTTTCGAGACTTTGCCAAGGT
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CTACCTGCCAGTGCAGACTGTCTTAGAGGGGGACAACATGGAAACTCCCGTTACT
CTGATCAACTTCTGGCCAGTAGATTCTGCGCCTGCCTCGTCCCCTCAGCTTTCACA
CGATGATACTCATTACGCATTGAACATTATGCTAGCAGGCTAGCAGAAATGGA
AAACAGCAATGGATCTTATCTAAATGATAGCATCTCTCCTAATGAGAGCATAGAT

GATGAACATTTGTTAATCCAGCATTACTGCCAAAGTTTGAACCAGGACTCCCCC
TGAGCCAGCCTCGTAGTCCTGCCAGATCTTGATTTCTTAGAGAGTGAGGAAAG
AGGGGAGCTAGAGAGAATCCTAGCAGATCTTGAGGAAGAAAACAGGAATCTGC
AAGCAGAATATGACCGTCTAAAGCAGCAGCACGAACATAAAGGCCTGTCCCCAC
TGCCGTCCCCTCCTGAAATGATGCCACCTCTCCCAGAGTCCCCGGGATGCTGA
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TGTCTCTCCTTCTACCTCTCTACAGAGGTCCGACAGCAGTCAGCCTATGCTGCTC
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CCACCGGCAAGCTGCCCCTGCCCTGGCCCACCCTCGTGACCACCCTGACCTACGG
CGTGCAGTGCTTCAGCCGCTACCCCGACCACATGAAGCAGCACGACTTCTTCAAG
TCCGCCATGCCCGAAGGCTACGTCCAGGAGCGCACCATCTTCTTCAAGGACGAC
GGCAACTACAAGACCCGCGCCGAGGTGAAGTTCGAGGGCGACACCCTGGTGAAC
CGCATCGAGCTGAAGGGCATCGACTTCAAGGAGGACGGCAACATCCTGGGGCAC
AAGCTGGAGTACAACACTACAACAGCCACAACGTCTATATCATGGCCGACAAGCAG
AAGAACGGCATCAAGGTGAACTTCAAGATCCGCCACAACATCGAGGACGGCAGC
GTGCAGCTCGCCGACCACTACCAGCAGAACACCCCCATCGGCGACGGCCCCGTG
CTGCTGCCCGACAACCACTACCTGAGCACCCAGTCCGCCCTGAGCAAAGACCCC

AACGAGAAGCGCGATCACATGGTCCTGCTGGAGTTCGTGACCGCCGCCGGGATC
 ACTCTCGGCATGGACGAGCTGTACAAGTAAA

Table S1. Primers spanning the promoter to EGFP region of the constructs used for PCR on genomic DNAs extracted from SFFV-C1-GFP and SFFV-C2-GFP transduced cells.

Cell type	PCR fragment	Forward primer	Reverse primer	Size
SC1	1	ACCCATCAGATGTTTCCAGGC	ATGCATGTTCCAGTCGTTGTG	842bp
	2	GAGCTGGGTCCGACAATCAA	CATGCTAGCTACCCTGAGGC	894bp
	3	TGGGAAGTAAGCTGATTGGAACA	CTTGCCTACGCACTGCATTC	920bp
	4	GTACAAGAGGCAGGCTGATGA	AATGCCATCCTGGAGTTCCTT	813bp
	5	CGACAAGGGCGATTTGACAG	CCTCTTCACTGGCTGAGTGG	923bp
	6	AGCAGTTCAAGCTAAACAACCG	TGACTCAAGCTTGGCTCTGG	705bp
	7	AACCGGAGGCAACAGTTGAA	GACGCTTCCACTGGTCAGAA	720bp
	8	TCCTGAGATCCCTGGAAGGTT	GACGTGGCTCACGTTCTCTT	700bp
	9	TCAAGAGGCCACGGATGAG	TATCCACGCAGAGAGGGACG	712bp
	10	CCTTGGACCAGCACAACCTC	AGAGTAACGGGAGTTTCCATGT	840bp
	11	TGCACTCCGACTACATCAGG	TGGTTGTGGCCATTGTGTCC	987bp
	12	AGCACAGGGTTAGAGGAGGT	CTCAGGTAGTGGTTGTCTGGG	715bp
SC2	1	ACCCATCAGATGTTTCCAGGC	ATGCATGTTCCAGTCGTTGTG	842bp
	2	GAGCTGGGTCCGACAATCAA	CATGCTAGCTACCCTGAGGC	894bp
	3	TGGGAAGTAAGCTGATTGGAACA	TACGGTGAGAGCTGAATGCC	1030bp
	4	AGAAAAAGCAATCCATGGGCA	TATCCACGCAGAGAGGGACG	894bp
	5	CCTTGGACCAGCACAACCTC	AGATCTGGGCAGGACTACGA	1089bp
	6	CCCTCAGCTTTCACACGATGA	CTCAGGTAGTGGTTGTCTGGG	1398bp
	7	AGCACAGGGTTAGAGGAGGT	CTCAGGTAGTGGTTGTCTGGG	715bp

Supplementary Figure legends:

Figure S1. Transduction efficiency of SFFV-C1-GFP, hDesmin-C1-GFP, SFFV-C2-GFP, hDesmin-C2-GFP, SFFV-GFP and hDesmin-GFP lentivirus in DMD pericytes, analysed by FACS.

Figure S2: Full length, uncropped western blotting image of Figure 2B showing the expression of GFP (red) and tubulin 2.1 (green) by proliferating CD133+ cells (negative

control), non-transduced pericytes (NT), pericytes transduced with SFFV-GFP (SG), hDesmin-GFP (DG), SFFV-C1-GFP (SC1), hDesmin-C1-GFP (DC1), SFFV-C2-GFP (SC2) and hDesmin-C2-GFP(DC2) lentiviruses. GFP bands of 27KD were expressed by SG and DG cells. A GFP band of 312KD was expressed by SC1 cells, and a 168KD GFP band by SC2 and DC2 cells; all of these are the expected molecular weight.

Figure S3: Detection of dystrophin C1-GFP in cells transduced with SFFV-C1-GFP (SC1) or Desmin-C1-GFP (DC1) lentiviruses, by western blot using either dystrophin or GFP antibodies. Mouse anti-GAPDH antibody was used as loading control. Cells maintained *in vitro* under both proliferating and differentiated conditions were analysed. Both dystrophin and GFP antibodies recognize the same protein at the expected molecular weight.

Figure S4: Full length, uncropped western blotting image of Figure S3 showing 2 western blots that were run in parallel, to determine the staining pattern of poly-dystrophin antibody and GFP antibody on cells transduced with SC1 or DC1 lentiviruses. Blot 1 (A, B, and C) was incubated with polyclonal dystrophin antibody (A and C, red) and the GAPDH antibody (B and C, green), blot 2 was incubated with polyclonal GFP (D and F, red), myosin (MF20, green in E and F, 200KD band) and GAPDH antibody (green in E and F, 37KD). C and F are merged image of A, B and D, E, respectively. The results show similar staining pattern of dystrophin and GFP antibody by western blot, thus GFP was subsequently used as surrogate marker for dystrophin in this study. Sample ID: Lane 1: none transduced cells; 2: SC1 transduced cells (non-differentiated); 3: DC1 transduced cells (non-differentiated); 4: SC1 transduced cells 7 days after differentiation; 5: DC1 transduced cells 7 days after differentiation.

Figure S1

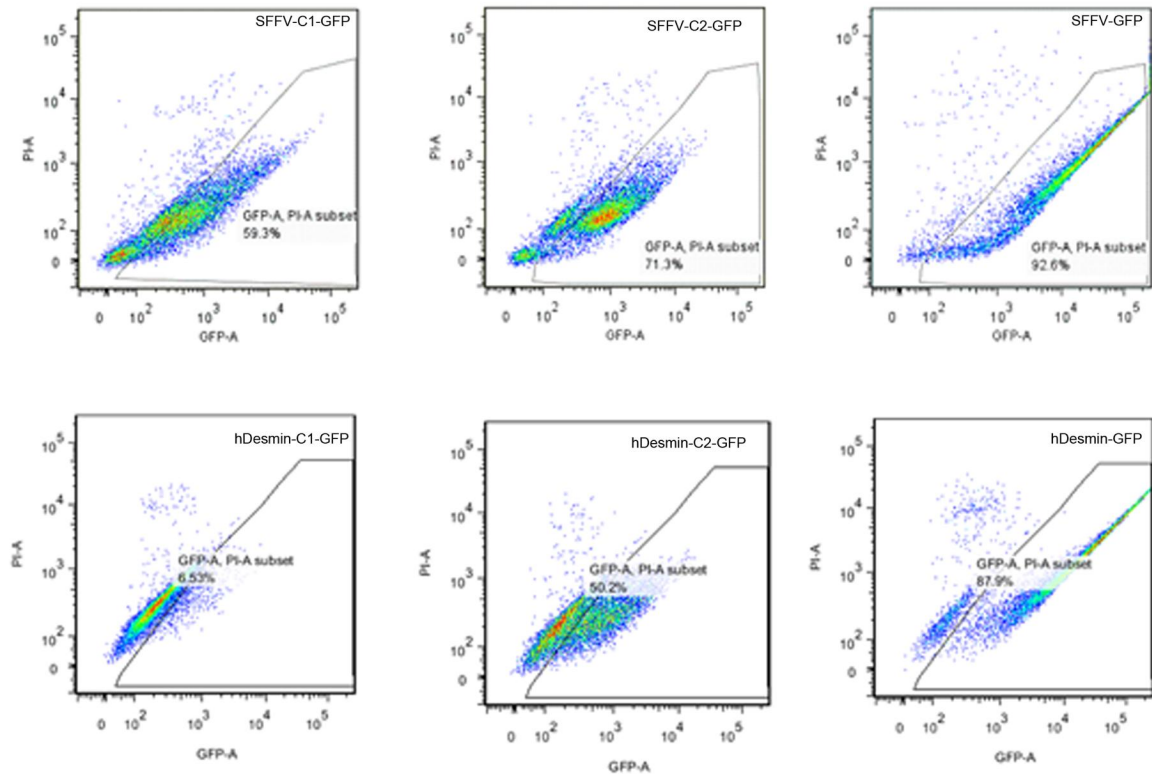


Figure S2

CD133+ NT SG DG SC1 DC1 SC2 DC2

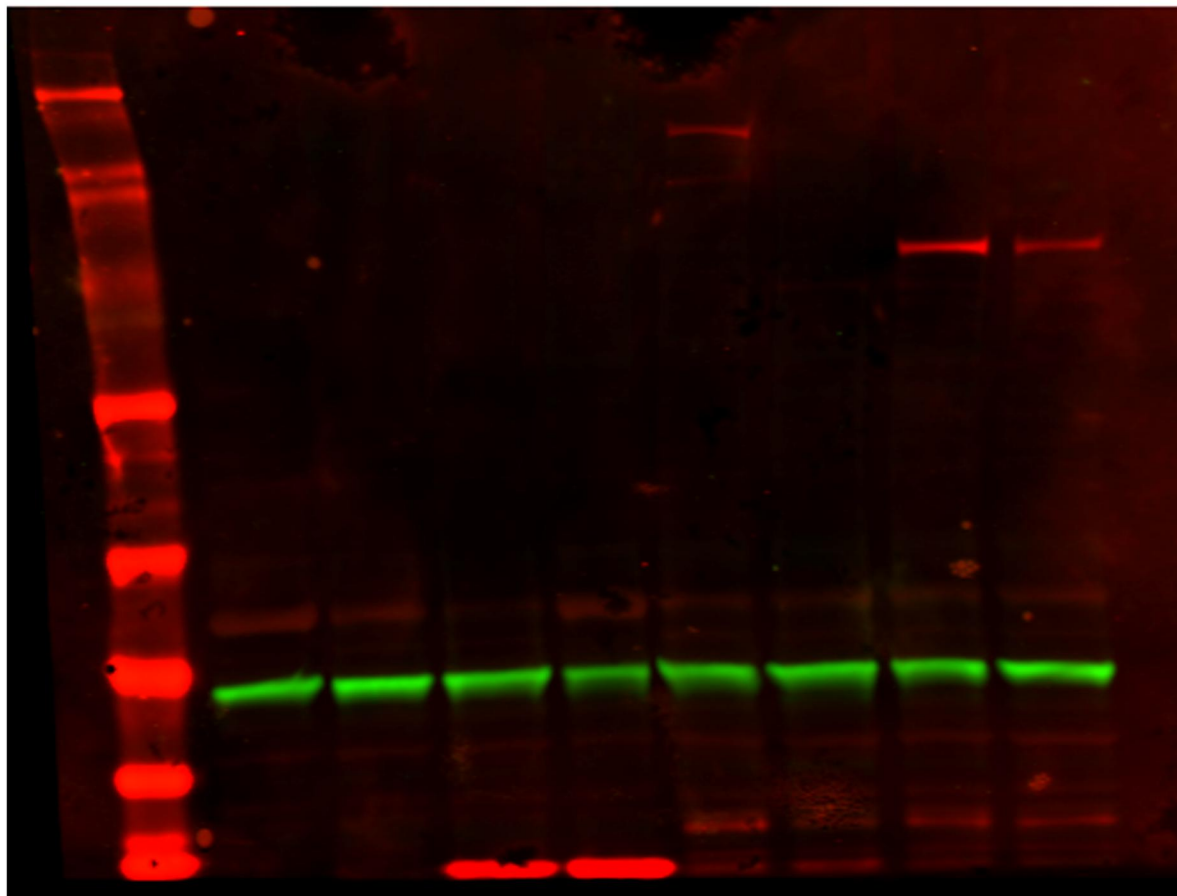


Figure S3

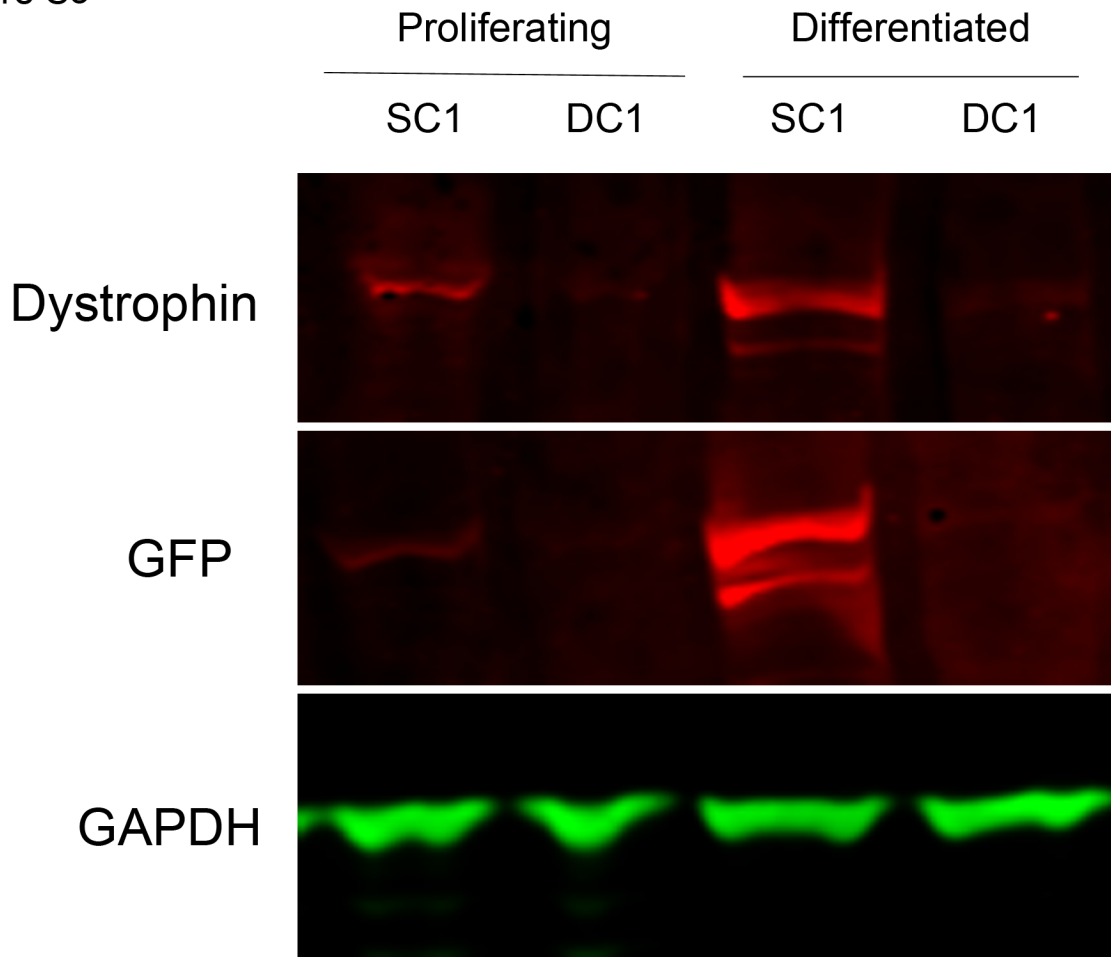


Figure S4

