

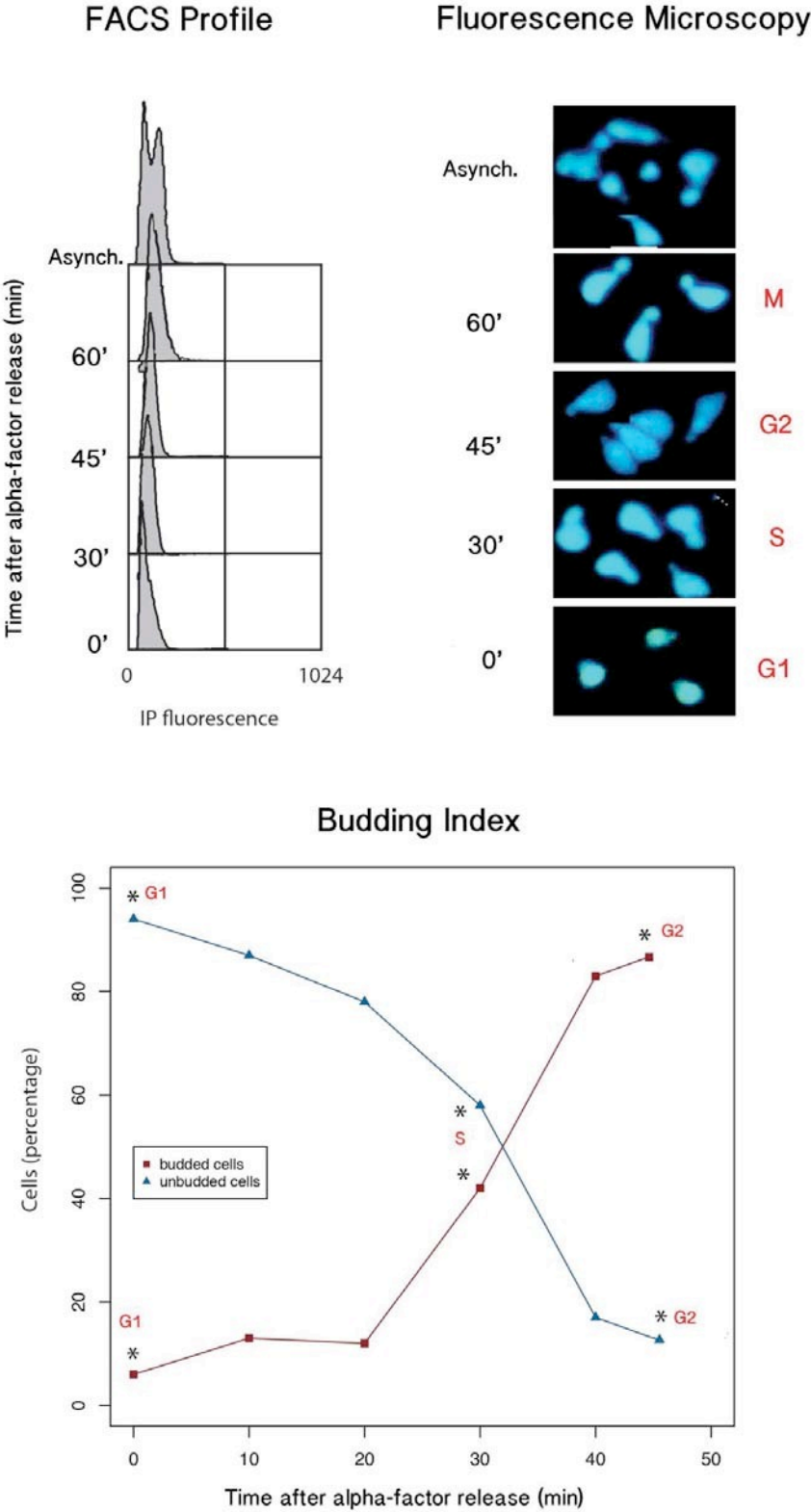
# Nucleosome architecture throughout the cell cycle

Özgen Deniz <sup>1,2,5&</sup>, Oscar Flores <sup>1,2,6&</sup>, Martí Aldea <sup>3</sup>, Montserrat Soler-López <sup>1,2,7</sup> and Modesto Orozco <sup>1,2,4\*</sup>

## Supplementary Figures & Dataset Captions

- 
- 1 Institute for Research in Biomedicine (IRB Barcelona). Baldiri Reixac 10-12. 08028 Barcelona, Spain  
2 Joint BSC-CRG-IRB Program in Computational Biology. Baldiri Reixac 10-12. 08028 Barcelona, Spain  
3 Molecular Biology Institute of Barcelona (IBMB) CSIC. Baldiri Reixac 4. 08028 Barcelona, Spain  
4 Department of Biochemistry and Molecular Biology. University of Barcelona, 08028 Barcelona, Spain  
5 Present address: Blizard Institute, 4 Newark Street E1 2AT, London, UK  
6 Present address: Genomcore SL, Jordi Girona 1, 08034 Barcelona, Spain  
7 Present address: European Synchrotron Radiation Facility (ESRF), 38000 Grenoble, France

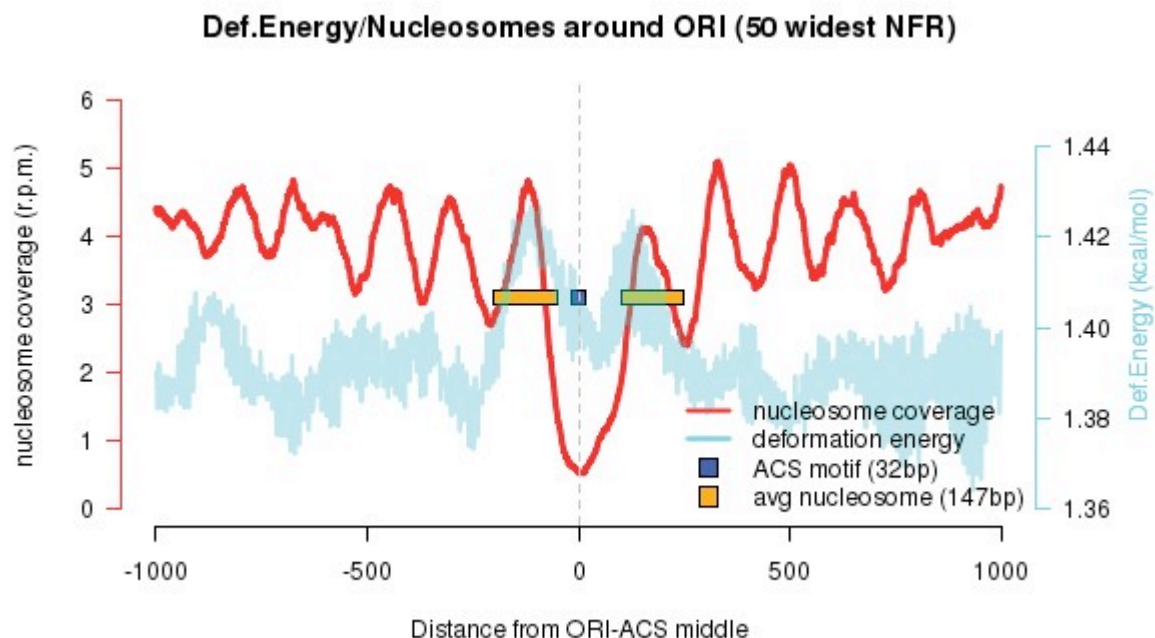
# SUP FIGURE S1



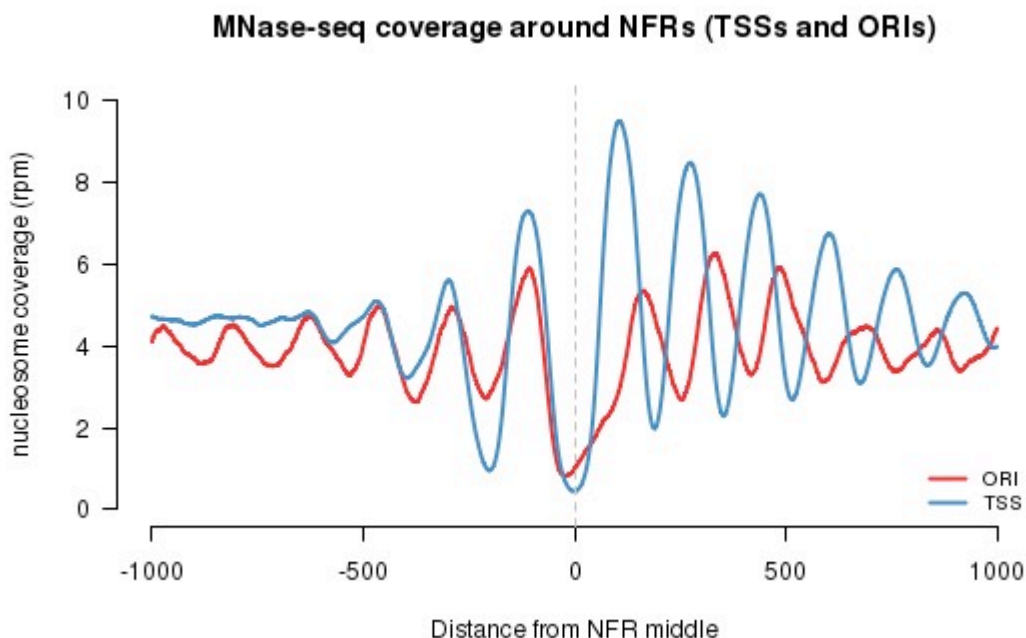
Supplementary Figure S1 – Monitoring cell cycle progress of BY4741 cells, by flow cytometry (top left), fluorescence microscopy (top right) and budding index (bottom).

## SUP FIGURE S2

a)

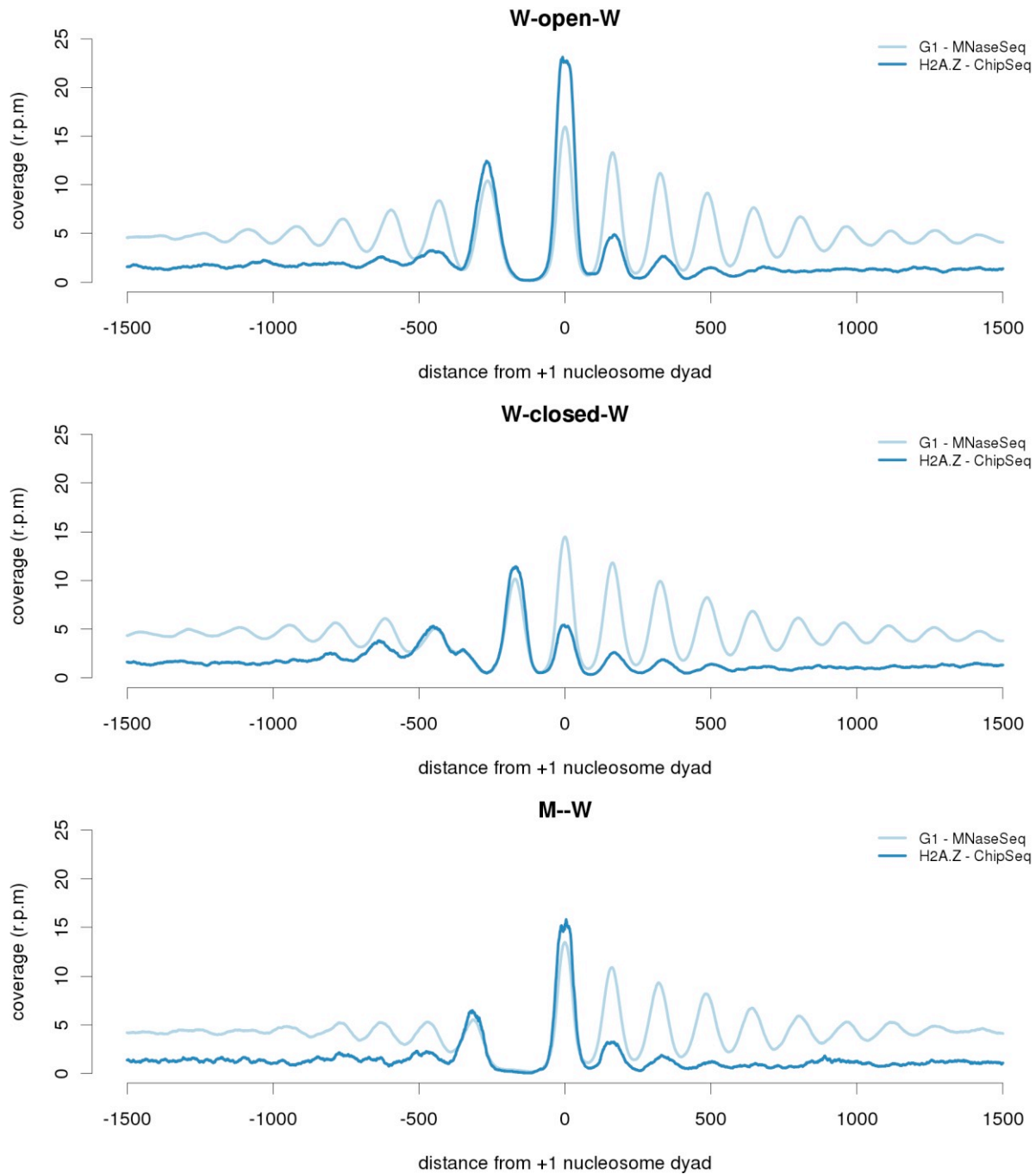


b)



**Supplementary Figure S2 – Properties of the NFRs at ORIs.** **a)** The deformation energy (blue) and coverage maps (red) were calculated and averaged across the widest NFRs at ORIs (top 50). ORI-NFRs are signaled by specific physical properties of the underlying DNA sequence (higher deformation energy shown in blue), in a similar manner to TSS-NFRs (Flores et al, 2014) **b)** Distribution of average nucleosome coverage profiles at TSSs (blue) and ORIs (red). Average nucleosome positioning at TSSs are directional and asymmetrical, meanwhile symmetry is more prevalent in the ORIs.

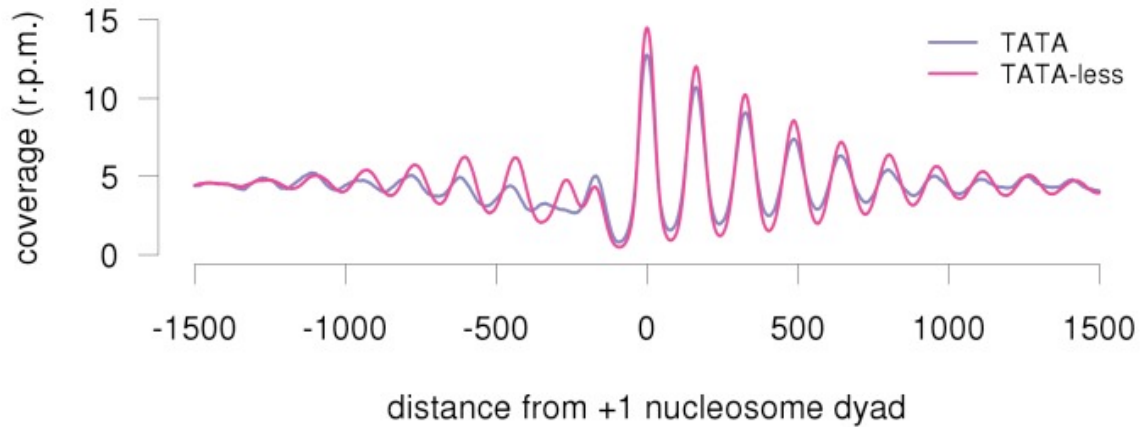
## SUP FIGURE S3



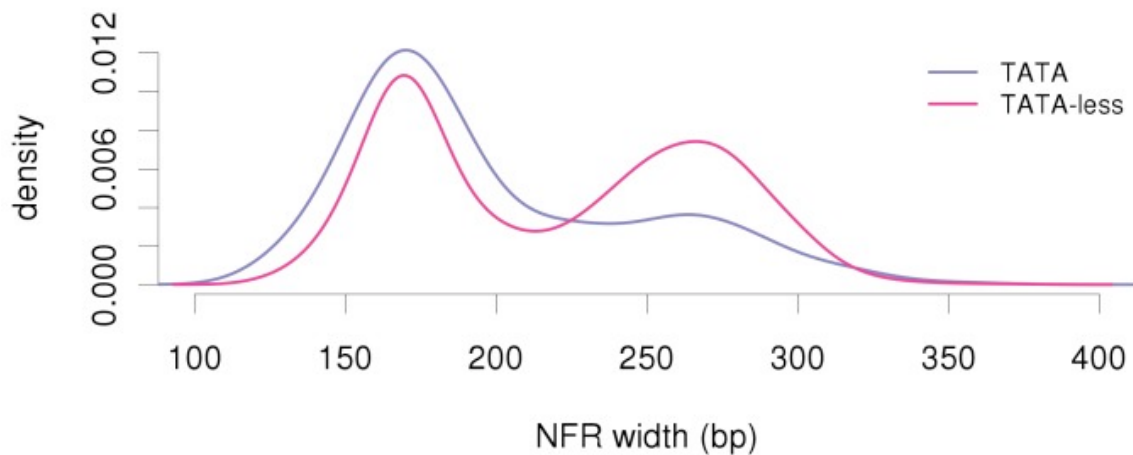
**Supplementary Figure S3 – H2A.Z histone variant enrichment in gene clusters.** H2A.Z coverage (Albert, 2007) is compared with G1 nucleosome coverage maps in a) open, b) closed and c) missing -1 nucleosome families with flanking well positioned nucleosomes.

## SUP FIGURE S4

a)

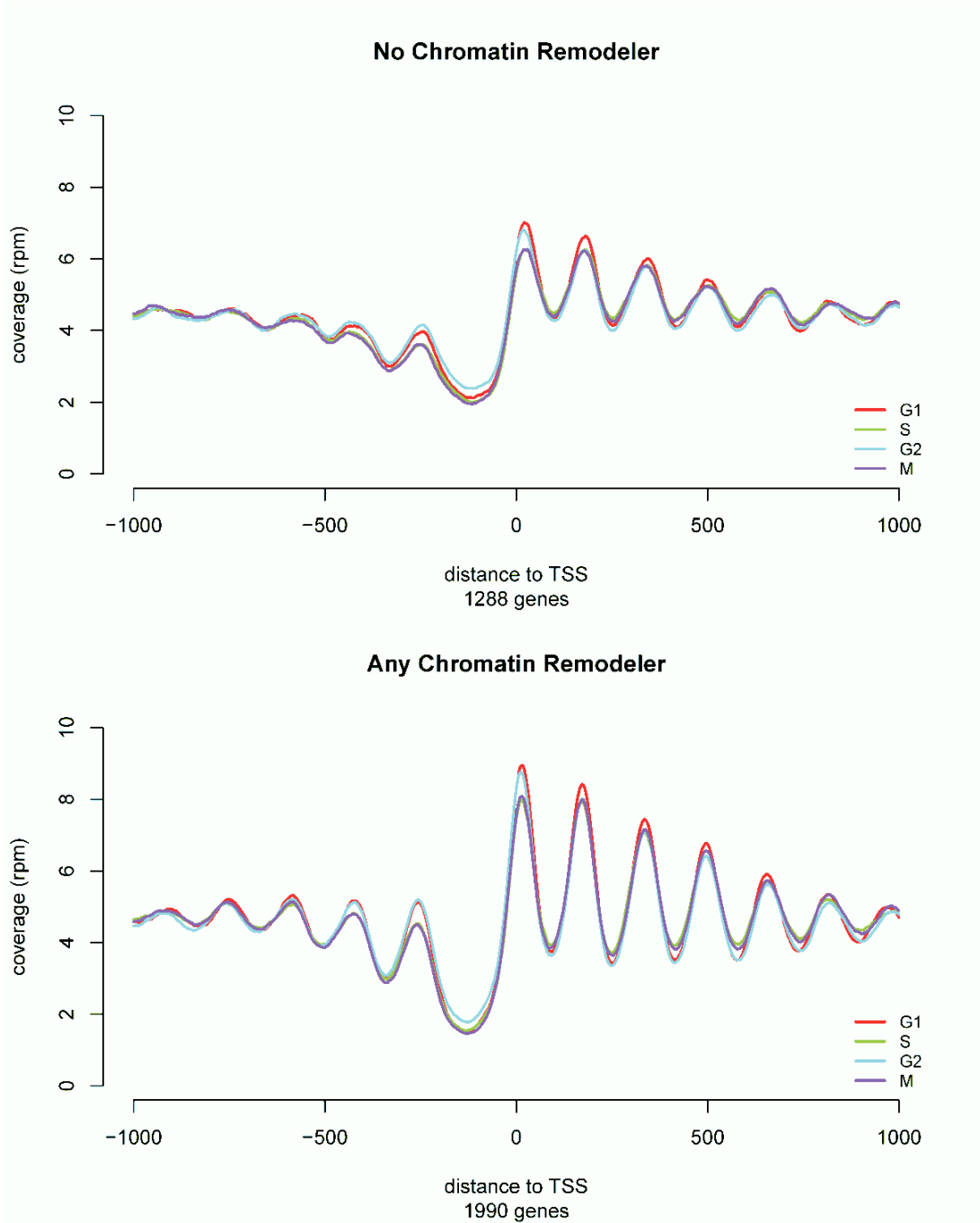


b)



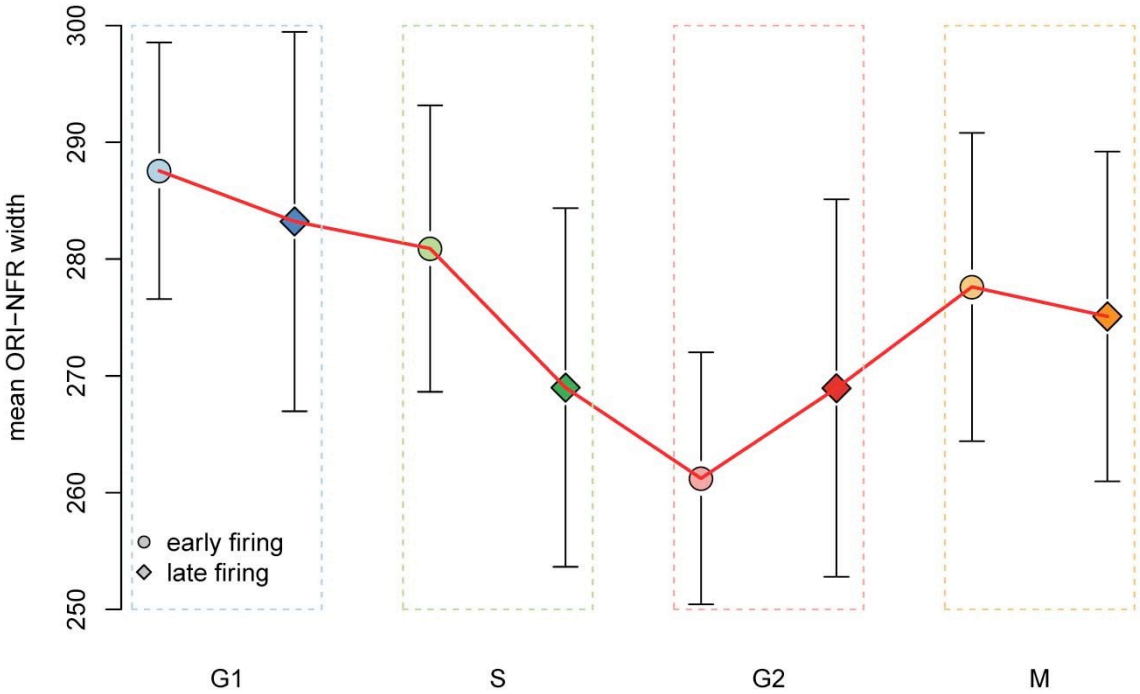
**Supplementary Figure S4** – Nucleosome coverage in TATA and TATA-less promoters. Nucleosome coverage maps of TATA-box regulated genes (pink) are compared with TATA-less genes (purple). **a)** Average coverage taking the dyad of the +1 nucleosome as reference does not display any difference in the gene body. **b)** NFR distribution in TATA and TATA-less promoters.

# SUP FIGURE S5



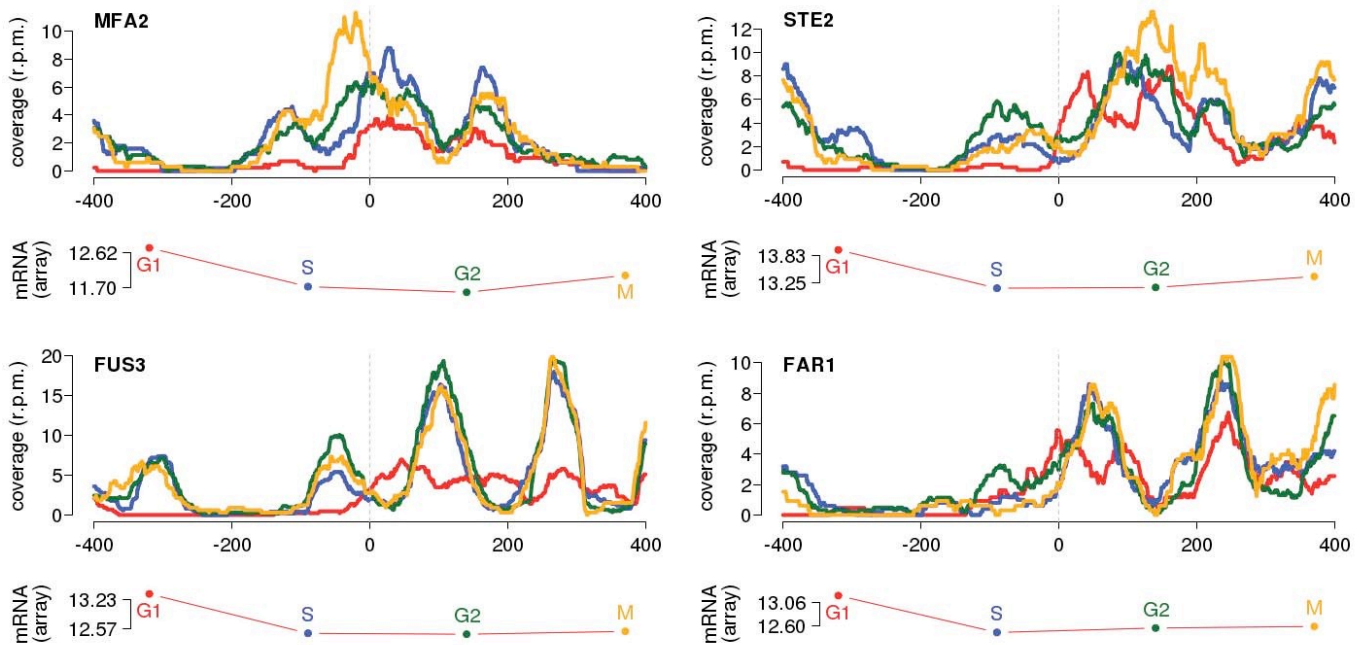
**Supplementary Figure S5. Effect of chromatin remodelers in nucleosome architecture.** Average nucleosome coverage at TSSs for genes not bound by any chromatin remodelers (top) and bound by any chromatin remodeler (bottom). The genes regulated by remodelers display a better phasing of nucleosome arrays.

# SUP FIGURE S6



**Supplementary Figure S6 – Nucleosome free region at ORIs along cell cycle.** Average width of ORI-NFRs changes during cell cycle, both in early and late firing ORIs. Error bars signal the 95% confidence interval, which is wide due to the low number of observations (N=100 in both groups)

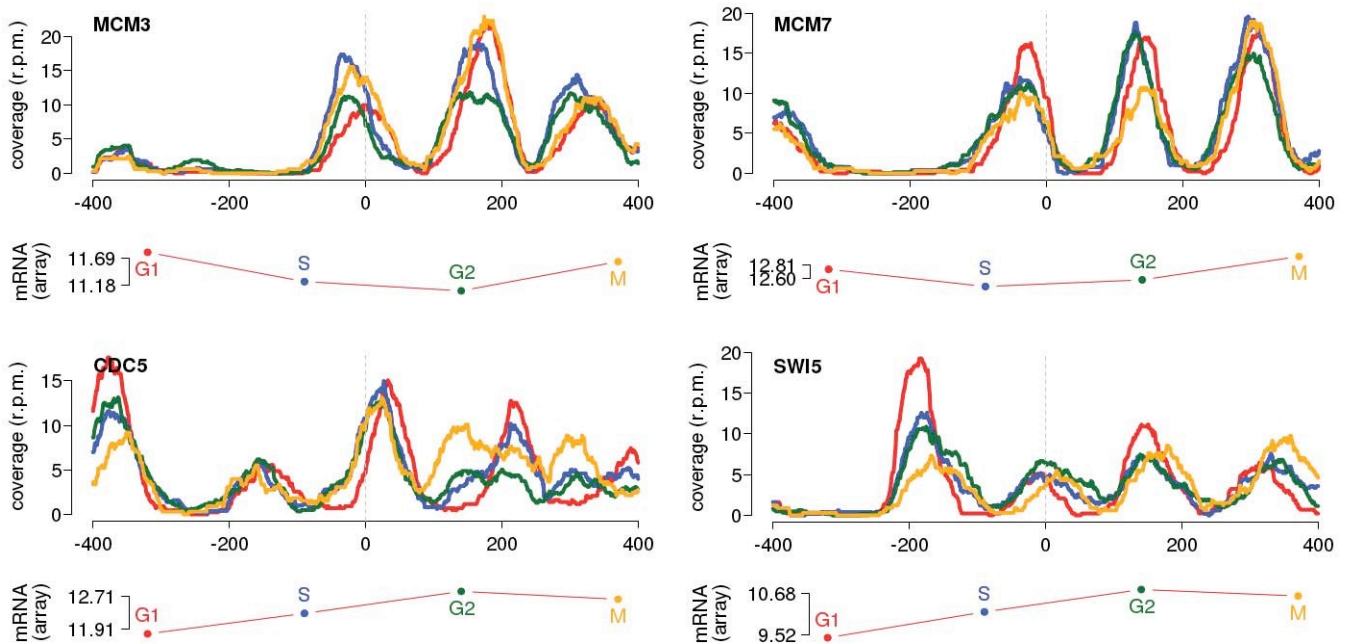
## SUP FIGURE S7



**Supplementary Figure S7 – Nucleosome architecture and expression plasticity of genes involved in alpha-factor pheromone mating and its desensitization response.** Up to bottom and left to right): MFA2, STE2, FUS3 and FAR1. Gene expression levels are indicated as log<sub>2</sub> values of the hybridization ratios from the Affymetrix GeneChip Yeast Genome 2.0 arrays.



## SUP FIGURE S8



**Supplementary Figure S8 – Nucleosome architecture and expression plasticity of genes involved in cell cycle progression.** Up to bottom and left to right): MCM3, MCM7, CDC5 and SWI5. Gene expression levels are indicated as log<sub>2</sub> values of the hybridization ratios from the Affymetrix GeneChip Yeast Genome 2.0 arrays.

### Dataset captions

**Dataset 1. Nucleosome architecture dynamics around TSS during cell cycle**

**Dataset 2. GO-term enrichment analysis for different gene clusters**

**Dataset 3. GO-term enrichment analysis for combinations of mobile genes with high expression fold-change (>1.2)**