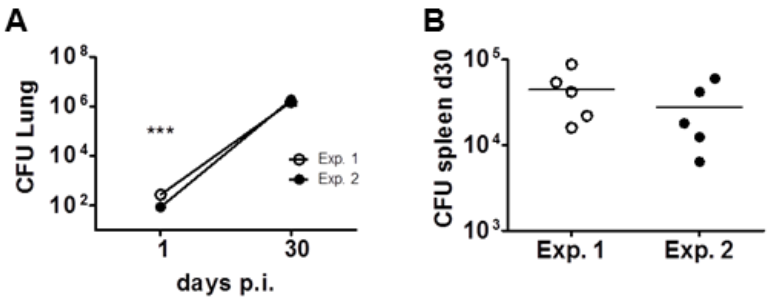
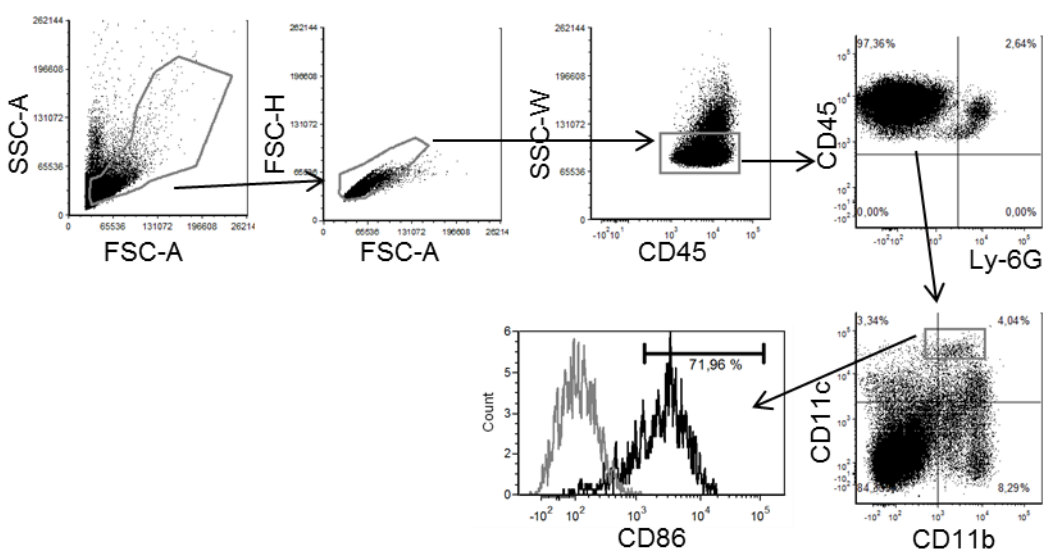
SUPPORTING INFORMATION

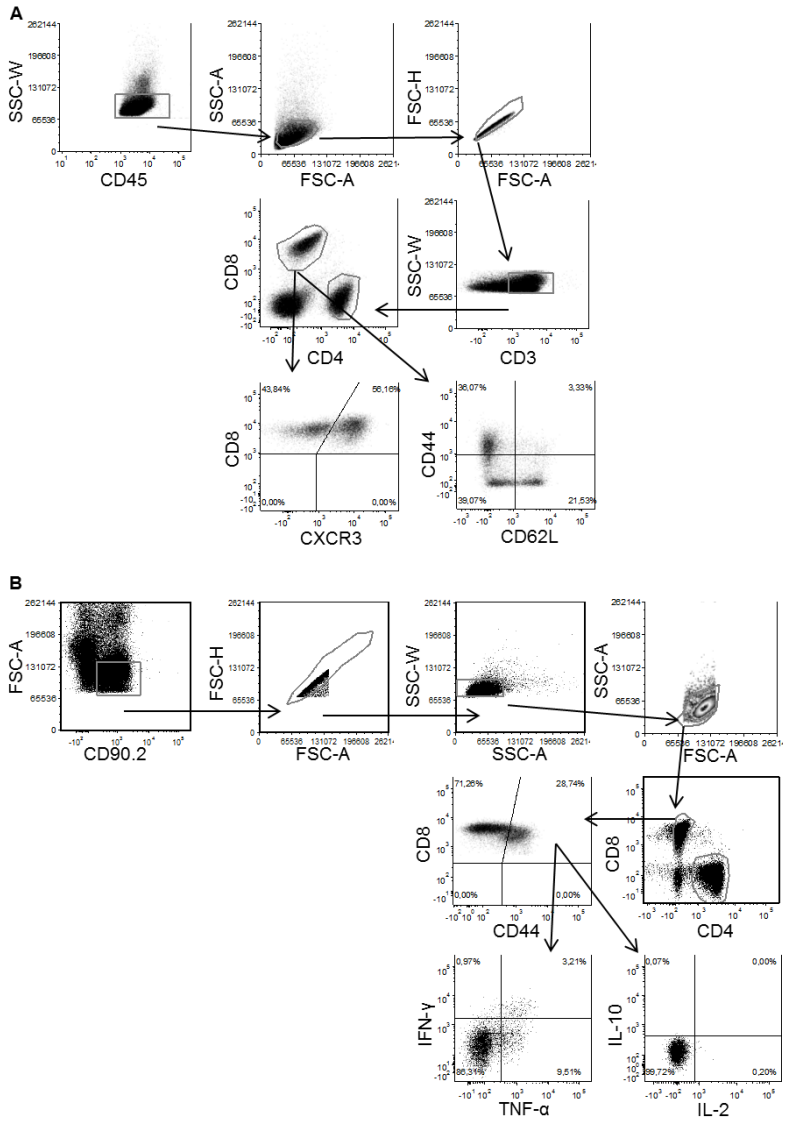


Supp. Fig. A1: Lung and spleen CFU after aerosol infection of mice with *Mtb*. C57BL/6 mice were infected via the aerosol route with *M.tb* H37Rv, and after 1 and/or 30 days the lung and spleen CFU was determined. Despite small but significant differences in initial uptake of *Mtb* into the lungs between two independent experiments, bacterial numbers in both lung and spleen are comparable on day 30 after infection, the time when mice were challenged with *Pb*ANKA. Data from two independent experiments are shown (mean ± SD, n = 5 for each group at each time point). Exp. = experiment.



Supp. Fig. 2: Gating strategy for the analysis of cDCs in the spleen.

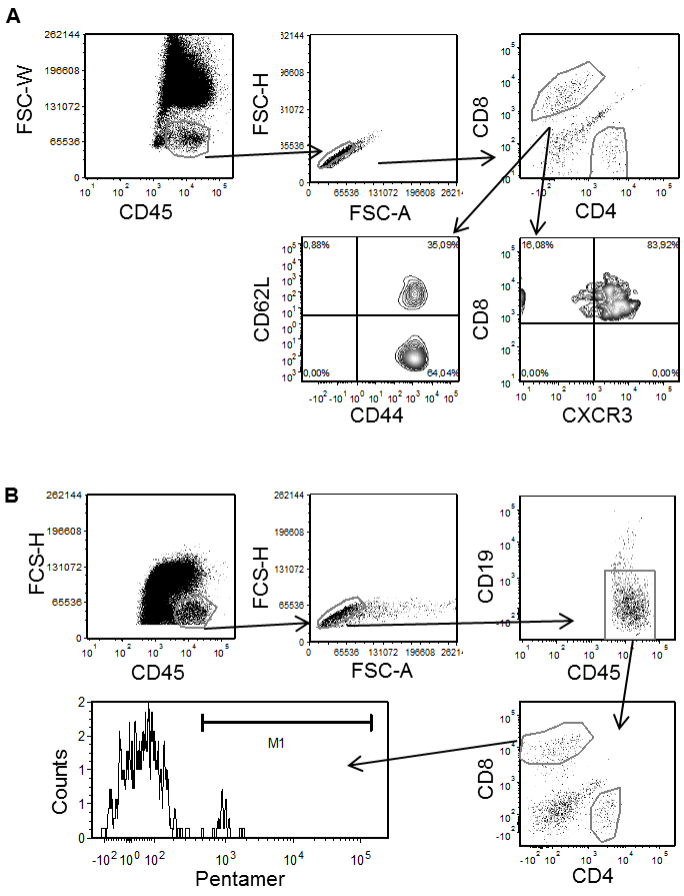
Gating strategy for CD11b^{int}CD11c^{hi} cDCs and for the expression of MHC-II and co-stimulatory molecules shown in Figure 4: Cells were stained for CD45, Ly-6G, CD11b, CD11c, MHC-II, CD80 and CD86. FCS-A versus SSC-A, FSC-A versus FSC-C and finally CD45 versus SSC-W were used to determine an accurate CD45^{+} cell population and to exclude doublets. Ly-6G versus CD45 was used to determine a Ly-6G^{-} population. Gating CD11b versus CD11c was used to determine an accurate CD11c^{hi} population which was CD11b^{int}. CD11^{int}CD11c^{hi} cells were analyzed for the expression of CD-80, CD86 or MHC-II by plotting a single-parameter histogram and using gate M1 to determine positive cells and mean fluorescence intensity (MFI), respectively. In grey: FMO control. Representative histogram for CD86 is shown.



Supp. Fig. A3: Gating strategies for the analysis of T cell responses in the spleen. A) Gating strategy for $CD8^+$ and $CD4^+$ T cells, effector memory T cells ($CD62L^-CD44^+$) and for the expression of CXCR3 shown in Figure 5 A and B: Cells were stained for CD45, CD3, CD4, CD8, CD44, CD62L and CXCR3. CD45 versus width of SSC was used to determine an accurate $CD45^+$ cell population and to exclude doublets, followed by additional doublet exclusion by gating the $CD45^+$ population in SSC-A versus FSC-A and FSC-H versus FSC-a. SSC-W versus CD3 was used to determine an accurate $CD3^+$ cell population. CD8 or CD4 cells were gated and

single and co-expression of CD62L and CD44 was determined to distinguish between specific cell phenotypes. Plotting CXCR3 versus CD8 or CD4 was used to determine an accurate CD8 or CD4 and CXCR3 co-expressing cell population. Where representative scatter plots show CD8 gating, $CD4^+$ cells were analyzed accordingly.

B) Gating strategy for analysis of T cell cytokine production shown in Figure 4 C and D: Cells were stained for CD90.2, CD4, CD8, CD44, IFN-γ, TNF-α, IL-10 and IL-2. Area of FSC versus CD90.2 was used to define a $CD90.2^+$ cell population which was subjected three different gatings for doublet exclusion. Gating CD8 versus CD4 was used to determine an accurate $CD8^+$ and $CD4^+$ cell population. $CD44^+$ cells were gated and assessed for their production of IFN-γ, TNF-α, IL-10 or IL-2. Where representative scatter plots show CD8 gating, $CD4^+$ cells were assessed accordingly.



Supp. Fig. A4: Gating strategy for the sequestration of T cells in the brain. A) Gating strategy for $CD8^+$ and $CD4^+$ T cells, effector memory T cells ($CD62L^-CD44^+$) and for the expression of CXCR3 shown in Figure 6 A and B: Cells were stained for CD45, CD4, CD8, CD44, CD62L and CXCR3. CD45 versus width of SSC was used to determine an accurate $CD45^+$ cell population and to exclude doublets, followed by additional doublet exclusion by gating the $CD45^+$ population in FSC-H versus FSC-A. Gating CD8 versus CD4 was used to determine an accurate $CD8^+$ and $CD4^+$ cell population. Single and co-expression of CD62L and CD44 was determined on $CD8^+$ or $CD4^+$ cells. Plotting CXCR3 versus CD8 or CD4 was used to determine an accurate CD8 or CD4 and CXCR3 co-expressing cell population. Where representative scatter plots show CD8 gating, $CD4^+$ cells were analyzed accordingly. B) Gating

strategy for antigen-specific $CD8^+$ T cells as shown in Figure 5 C: Cells were stained for CD45, CD19, CD4, CD8 and MHC Class I Ova. CD45 versus width of SSC was used to determine an accurate $CD45^+$ cell population and to exclude doublets, followed by additional doublet exclusion by gating the $CD45^+$ population in FSC-H versus FSC-A. CD19 versus CD45 was used to determine an accurate $CD45^+$ $CD19^-$ cell population to exclude unspecific binding of the pentamer. Gating CD8 versus CD4 was used to determine an accurate $CD8^+$ and $CD4^+$ cell population. $CD8^+$ cells were analyzed for pentamer binding by plotting a single-parameter histogram and using gate M1 to determine pentamer positive cells.