

1 **SUPPORTING INFORMATION**

2 **Table S1. *C. neoformans* strains used in this study.**

Strain	Additional information	Reference
RC2-wt (smooth phenotype)	Serotype D, Switch variant <i>MATα</i>	(1)
H99	Serotype A, <i>MATα</i>	(2)
RC2- <i>all2</i> Δ	<i>all2</i> Δ :: <i>NEO</i>	This study
RC2- <i>all2</i> Δ +P _{<i>ACT1</i>} - <i>ALL2</i>	<i>ALL2</i> promoter replaced with <i>ACT1</i> promoter to generate P _{<i>ACT1</i>} - <i>ALL2</i> - <i>NAT</i> and inserted to RC2- <i>all2</i> Δ	This study
H99- <i>all2</i> Δ	<i>all2</i> Δ :: <i>NEO</i>	This study
H99- <i>all2</i> Δ +P _{<i>ACT1</i>} - <i>ALL2</i>	<i>ALL2</i> promoter replaced with <i>ACT1</i> promoter to generate P _{<i>ACT1</i>} - <i>ALL2</i> - <i>NAT</i> and inserted to H99- <i>all2</i> Δ	This study
RC2- <i>all1</i> Δ <i>all2</i> Δ	<i>all2</i> Δ :: <i>NEO</i> <i>all1</i> Δ :: <i>NAT</i>	This study
RC2- <i>ALL1</i> ::HA	<i>ALL1</i> tagged with HA at C-terminal	This study

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4 **Table S2**
 5 **Primers used in this study.** Primers were designed using the Primer3 software
 6 package (<http://frodo.wi.mit.edu/primer3/>) for generating gene-deletion cassettes, gene-
 7 reconstitution cassettes, or real-time PCR products.
 8

Primer	Sequence (5'-3')	Function/ restriction site
All2-UpF	<u>CCATAGATTGG</u> - AGAAGAAGGGAGACGCAGCT	<i>ALL2</i> gene knock out cassette/ <i>Van911</i>
All2-UpR	<u>CCATCATTGG</u> - GTTTGGTTCTGAAGTTGATATGTGGT	
All2-DwnF	<u>CCATTTCTTGG</u> - CTGCCTAAGAAGTAGTTTGCCATAT	<i>ALL2</i> gene knock out cassette/ <i>Van911</i>
All2-DwnR	<u>CCATAAATTGG</u> - ATGGCTCACCAGGAAGAACGA	
All2-GF	<u>CATATGTCTACTGTTACTCAAGGTGTT</u> AAGGA	<i>ALL2</i> gene reconstitution/ <i>NdeI</i>
All2-GR	<u>CTCGAGCGGGGATGTGATCTGTTTTTC</u>	<i>ALL2</i> gene reconstitution/ <i>XhoI</i>
ACT1-PF	<u>TCTAGAAGGCTGCGGGAGGTGAGCT</u>	<i>ACT1</i> promoter for gene reconstitution/ <i>XbaI</i>
ACT1-PR	<u>CATATGAGACATGTTGGGCGAGTTTTA</u> C	<i>ACT1</i> promoter for gene reconstitution/ <i>NdeI</i>
ACT1-RT-f ACT1-RT-r	(3)	Real time PCR actin
All1-RT-f All1-RT-r	(3)	Real time PCR <i>ALL1</i>
All2-RTF All2-RTR	TGAAAGAAGGCCTCAAGAGCG AGTGGAAGCGGCATTGGTT	Real time PCR <i>ALL2</i>
ALL1prom- XbaI-F	<u>TCTAGAAGGTTTGGGGTTGATTTCGAA</u> GCT	<i>ALL1::HA</i>
ALL1-HA NCOI-rev	<u>CCATGGTTAAGCGTAGTCTGGGACGT</u> CGTATGGGTAAAGAGCCTGGGTCTTG CTG	
ALL1-3UTR- For-NCOI ALL1-3UTR- ECORV	<u>CCATGGAAGAATGTGTAGTAGTTTATG</u> GTATCTGAAGGTTAAAG <u>GATATCGTGC</u> ACTGAACCACAAGCTC	<i>ALL1::HA</i>
All2For-NHEI All2Rev-PSTI	<u>GCTAGCAGGCTGCGGGAGGTGAGCT</u> G <u>CTGCAGGGCAGTAGGAACTTCCTTAG</u> TGGA	<i>ALL2-mCherry</i>
P _{Act} -XbaFor mcherry rev- ECORV	<u>TCTAGAAGGCTGCGGGAGGTGAGCTG</u> <u>GATATCTTACTTGTAGAGCTCGTCCAT</u> ACCAC	<i>ALL2-mCherry</i> <i>ALL2-mCherry</i>
All2 3'UTR-for- ECORV All2 3'UTR- Rev XHOI	<u>GATATCGAAGTAGTTTGCCATATATTT</u> TTTGTCTTGGGG <u>CTCGAGTTCATCGTGC</u> GTCTTCACT TACT	<i>ALL2-mCherry</i>

11 **Table S3. Expression levels of *ALL2* as determined by real time PCR in mutants.**

Sample	- Δ C_T	2^{-ΔCT}	2^{-Δ ΔCT}
<i>ALL2</i> expression relative to reference gene – <i>ACT1</i>			
RC2 wt	3.6	0.07745	
	15.1	0.00003	0.00037
<i>all2Δ</i>			
	14.8	0.00003	0.00043
<i>all1Δall2Δ</i>			
<i>ALL2</i> expression in reconstituted strain relative to reference gene – <i>ACT1</i>			
RC2-wt	3.4	0.094732	
<i>all2Δ</i> +P _{ACT1} - <i>ALL2</i>	3.9	0.066986	1.01942
<i>ALL1</i> expression relative to reference gene – <i>ACT1</i>			
RC2-wt	2.6	0.16348	
	15.5	0.00002	0.00013
<i>all1Δall2Δ</i>			

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13 **Table S4. Characteristics of *ALL2* null mutant in H99 background.**

Characteristic	H99	H99- <i>all2</i> Δ	H99- <i>all2</i> Δ +P _{ACT1} - <i>ALL2</i>
Exo-polysaccharide viscosity (mL/g)	3238 \pm 432.6	1958 \pm 413.3	3512 \pm 426.2
H ₂ O ₂ sensitivity zone size (mm)	63	69	62
Phagocytosis index (%)	131.1 \pm 55.8	101.8 \pm 28.3	113.2 \pm 20.3

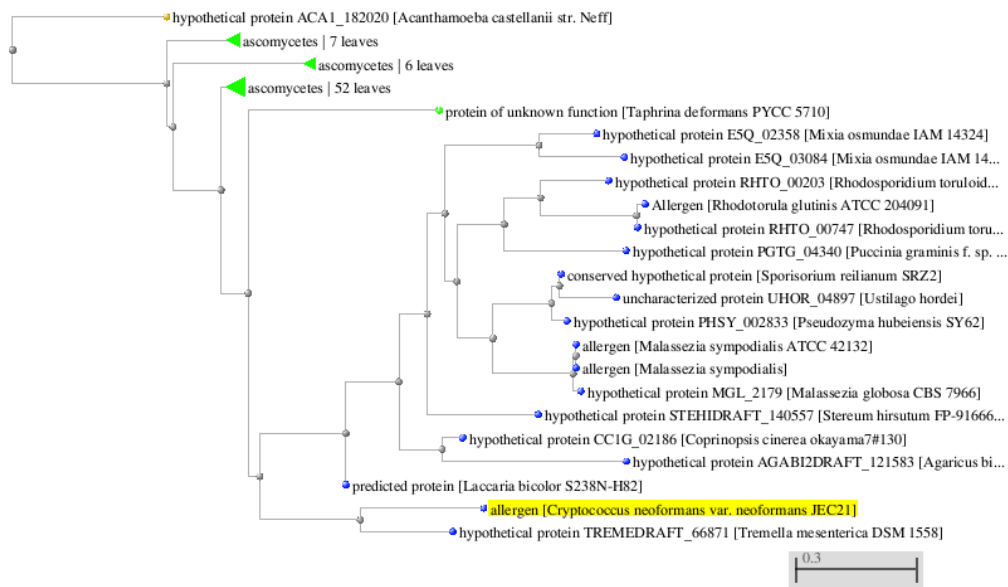
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15 To generate H99-*all2* Δ , the coding region of *ALL2* (945bp) in H99 was replaced with a
 16 neomycin resistance marker by homologous recombination as described in Materials
 17 and Methods. The exo-polysaccharide viscosity, H₂O₂ sensitivity and phagocytosis
 18 index for H99, H99-*all2* Δ , and H99-*all2* Δ +P_{ACT1}-*ALL2* were measured using similar
 19 methods described for RC2.

20 **Figure S1. All1p and All2p are homologous to fungi that affect plants.**
 21 Both All1p and All2p exhibit up to 33-38% homology with hypothetical proteins in other
 22 non-encapsulated fungi that are pathogenic to plants, but no homology to any fungi that
 23 that are pathogenic to humans.

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 25 All1p alignment tree

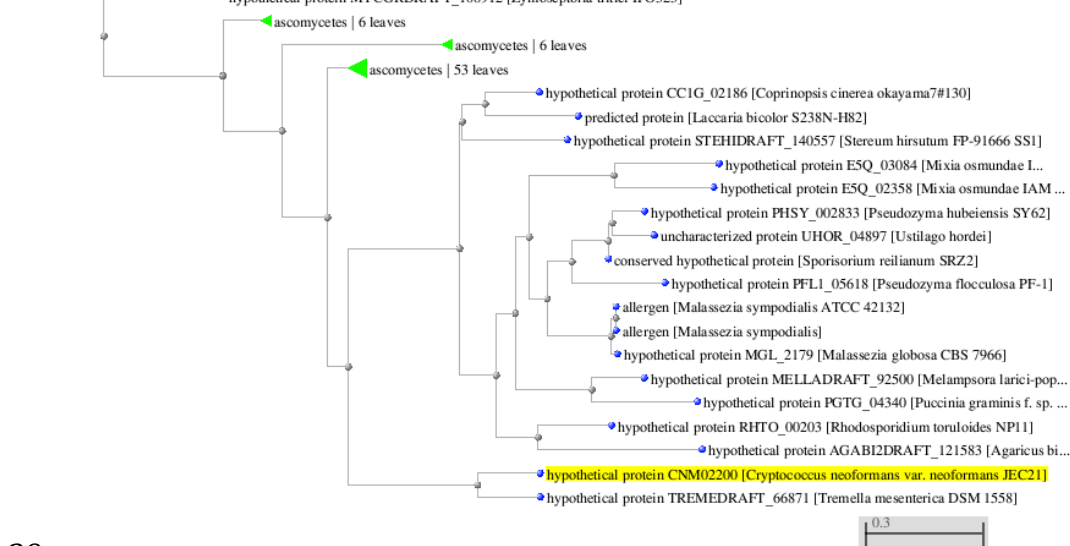
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28 All2p alignment tree

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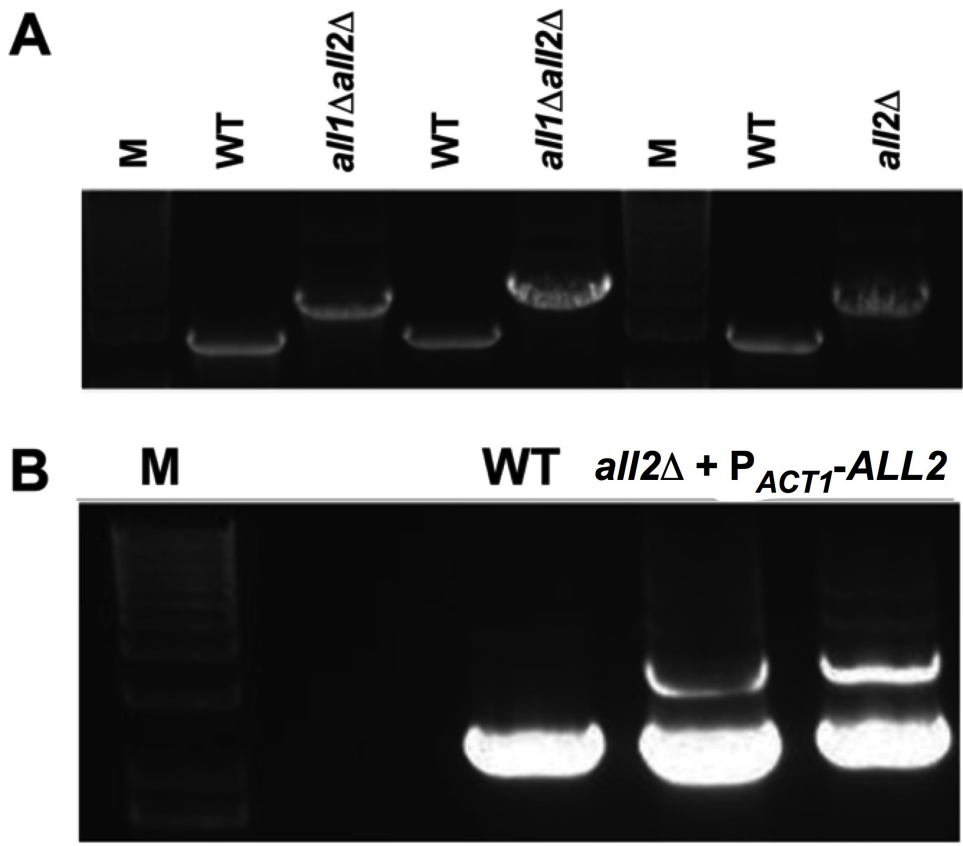


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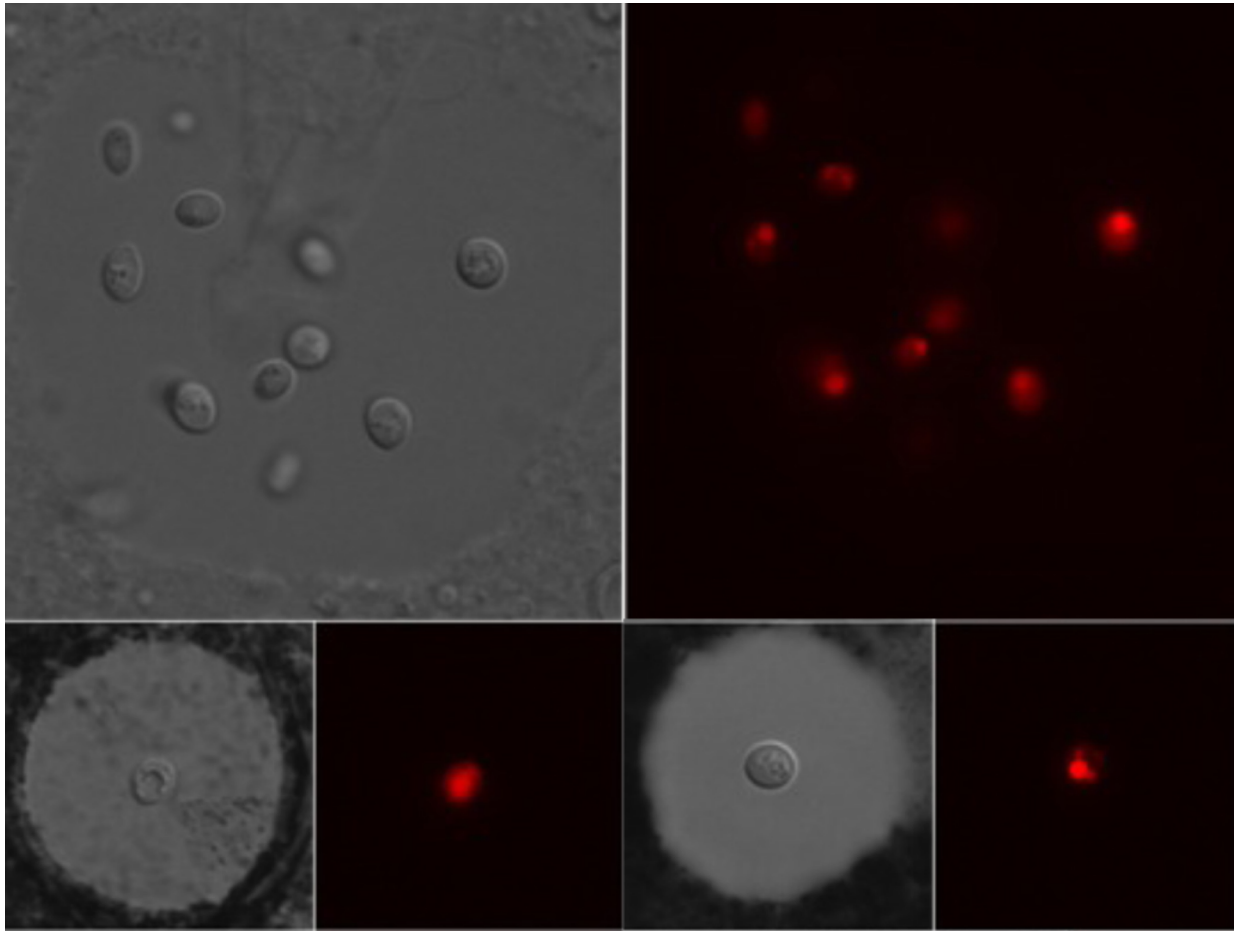
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31 **Figure S2. Validation of the *ALL2* gene disruption and complementation done by**
32 **PCR.**

33 **(A)** PCR Confirmation of the disruption of *ALL2* gene with the *NEO* disruption cassette.
34 Lane 1: 1kb DNA marker, Lane 2: PCR amplification using primers All2homo-F and
35 All2homo-R using RC2 genomic DNA yielded 3000 bp PCR product, Lane 3: PCR
36 amplification using primers All2homo-F and All2homo-R using *all1Δall2Δ* genomic DNA
37 yielded 4000 bp PCR product, Lane 4: PCR amplification using primers All1homo-F and
38 All1homo-R using RC2 genomic DNA yielded 3000 bp PCR product, Lane 5: PCR
39 amplification using primers All1homo-F and All1homo-R using *all1Δall2Δ* genomic DNA
40 yielded 4000bp PCR product, Lane 6: 1kb DNA marker, Lane 7: PCR amplification
41 using primers All2homo-F and All2homo-R using WT genomic DNA yielded 3000bp
42 PCR product, Lane 8: PCR amplification using primers All2homo-F and All2homo-R
43 using *all2Δ* genomic DNA yielded 4000bp PCR product. **(B)** PCR confirmation of
44 insertion of a copy of *ALL2* under Actin promoter to generate *all2Δ*+P_{ACT1}-*ALL2*.

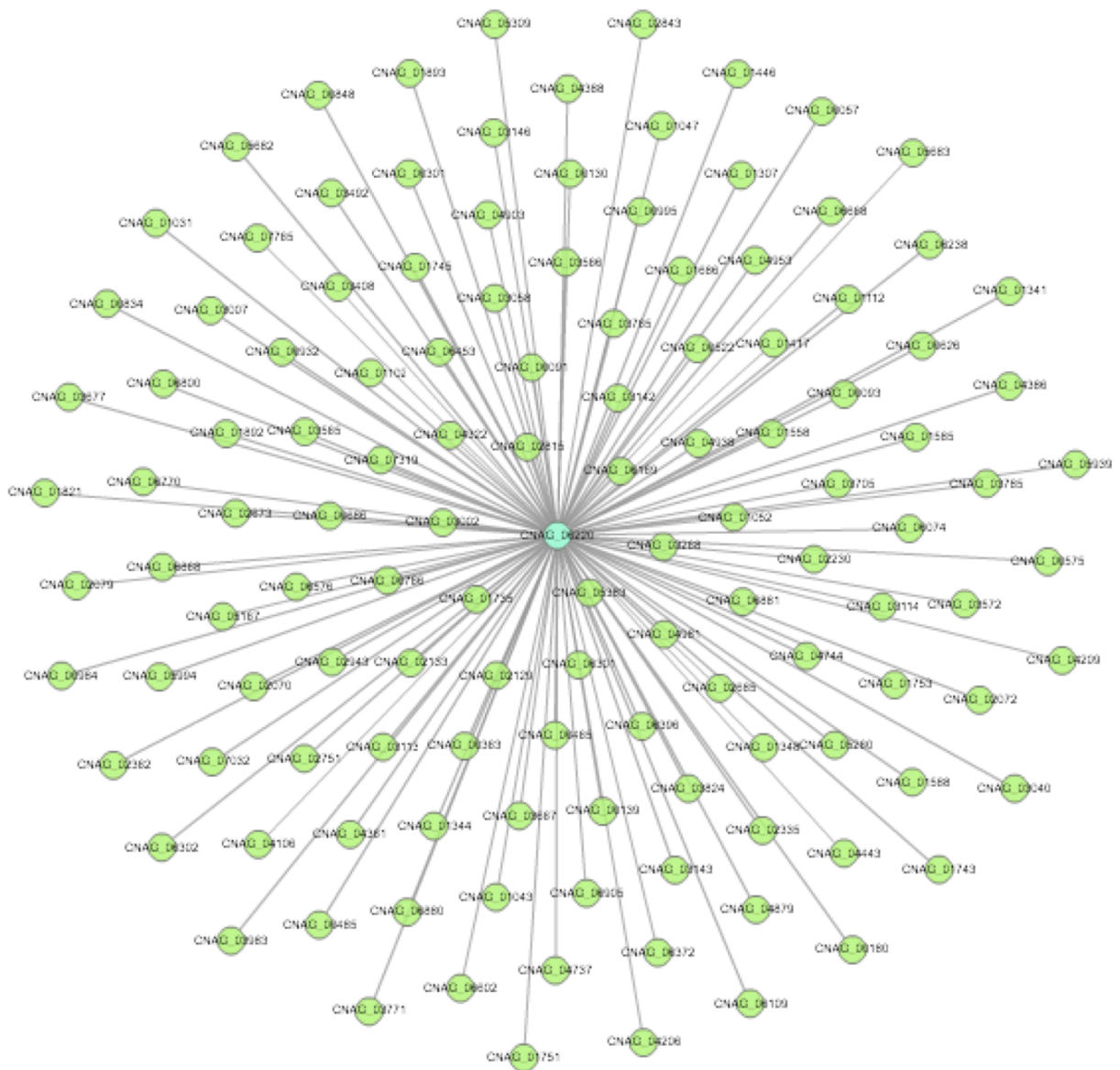


46 **Figure S3. All1-mCherry Cells recovered from murine tissue displayed All2p**
47 **localization (red) in vacuoles.**
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Figure S4. Functional network map of genes regulated by *ALL2*.



51 **References**

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59 switch variants of *Cryptococcus neoformans*. *Infect Immun* **77**:128-140.
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