

## **Supplementary Information**

### ***Detecting O<sub>2</sub> binding sites in protein cavities***

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*Supplementary materials: Discussion, Figure S1-S9, Table S1, and Movie S1-S3*

## Discussion

### Diffusion of O<sub>2</sub> into protein solution in the NMR tube.

Supplementary Figure S1 shows time-dependent changes in <sup>1</sup>H NMR spectra of <sup>15</sup>N labeled T4 lysozyme L99A when the oxygen concentration decreased from 1.8 mM to 0.27 mM over 18.7 hours at 298 K. A well-separated peak of L121 Hδ<sub>1</sub> changed its frequency (about 0.05 ppm) during 18.7 hours. We consider that this is due to the slow rate of diffusion of O<sub>2</sub> into protein solution in the NMR tube (inner diameter: 4 mm). Indeed, the time to reach equilibrium could be shortened when shaking or increasing surface area was employed (data not shown). Out of safety considerations, we did not shake the pressurized NMR tube in the present experiments.

### The potential of O<sub>2</sub>, N<sub>2</sub> and Ar gas molecules binding to protein cavities.

The mole fraction solubilities of O<sub>2</sub>, N<sub>2</sub>, and Ar in water at 298 K are  $2.3 \times 10^{-5}$ ,  $1.2 \times 10^{-5}$ , and  $2.5 \times 10^{-5}$ , respectively<sup>1</sup>. All values refer to the partial pressure of gas at one atmosphere. The van der Waals radius of Ar is 188 pm, whereas the length of the major axis of O<sub>2</sub> and N<sub>2</sub> are 218 pm and 226 pm, respectively, indicating that all three gases would fit to similar size cavities. However, in determining whether these gases possess affinity for protein cavities, their polarizability needs to be considered. It is known, for example, that the polarizability of noble gases is correlated to their affinity to protein cavities. Because electric dipole polarizabilities of N<sub>2</sub> and Ar are similar to that of oxygen (N<sub>2</sub>:1.74, Ar: 1.64, O<sub>2</sub>: 1.57 in units of  $10^{-24} \text{ cm}^3$ )<sup>2</sup>, they should be as likely to interact with the cavities as O<sub>2</sub>.

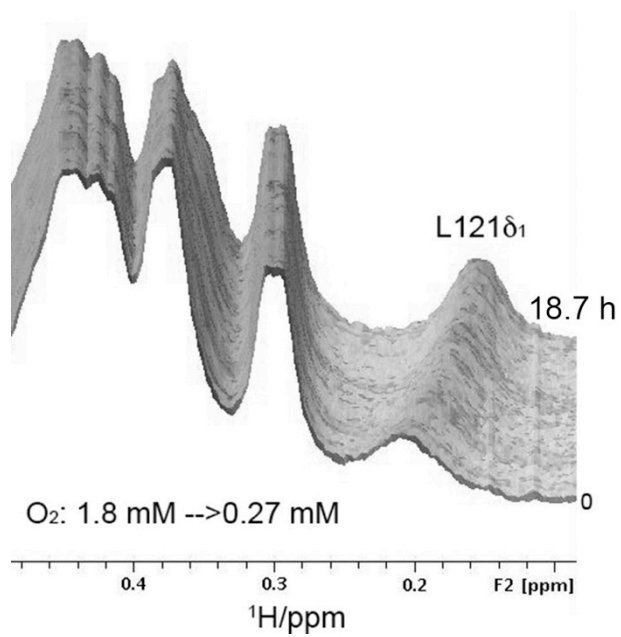


Figure S1. Time-dependent changes in  $^1\text{H}$  NMR spectra of  $^{15}\text{N}$  labeled T4 lysozyme L99A, when the oxygen concentration decreased from 1.8 mM to 0.27 mM over 18.7 hours at 298 K. The peak of L121  $\text{H}\delta_1$  is indicated.

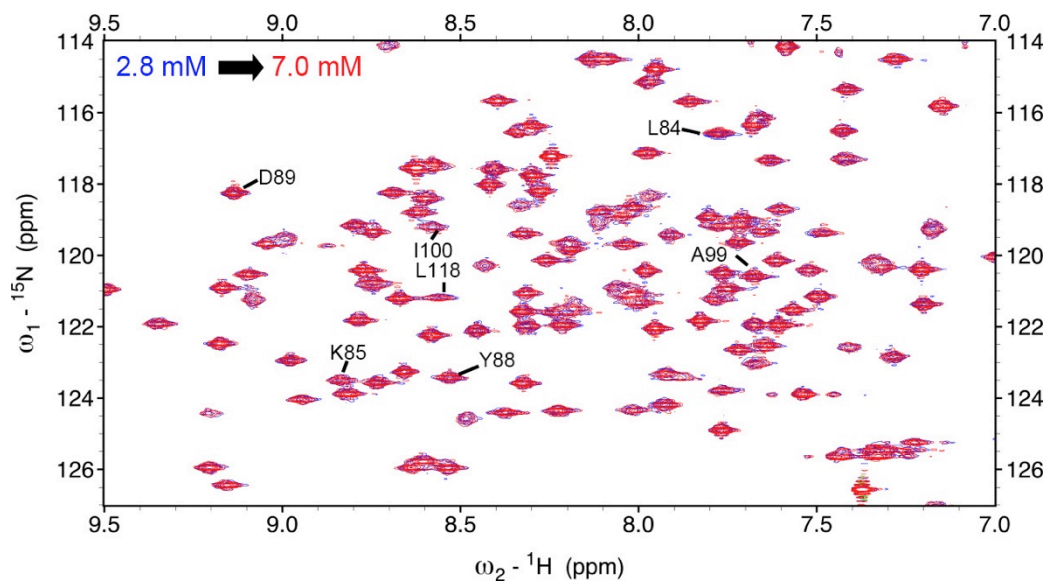


Figure S2.  $^1\text{H}/^{15}\text{N}$  HSQC spectra obtained for  $^{15}\text{N}$  labeled T4 lysozyme L99A at 2 bar (corresponding to 2.8 mM) and 5 bar (corresponding to 7.0 mM) Ar.

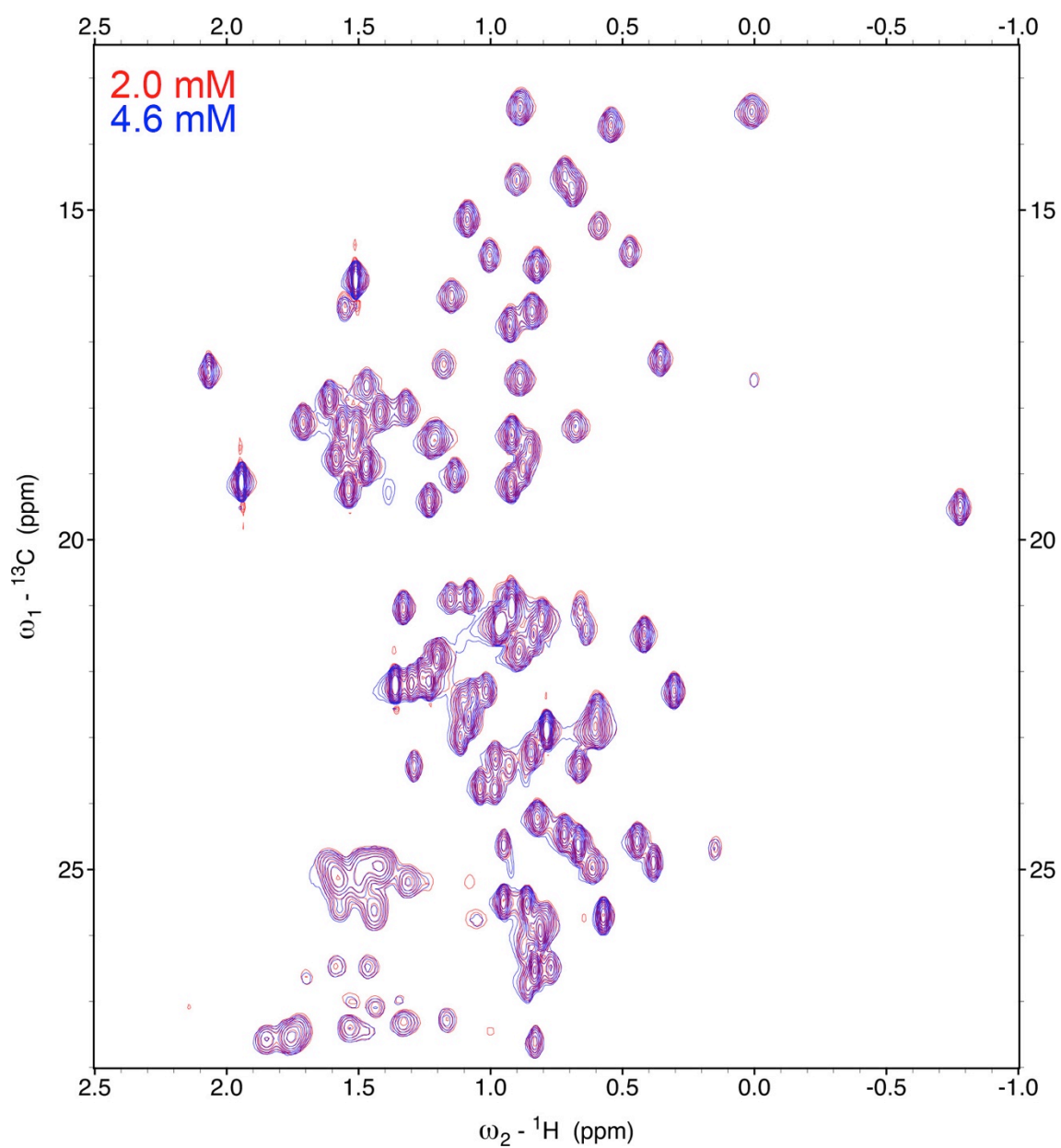


Figure S3.  $^1\text{H}/^{13}\text{C}$  CT HSQC spectra obtained for  $^{15}\text{N}/^{13}\text{C}$  labeled T4 lysozyme L99A at 3 bar (corresponding to 2.0 mM) and 7 bar (corresponding to 4.6 mM)  $\text{N}_2$ .

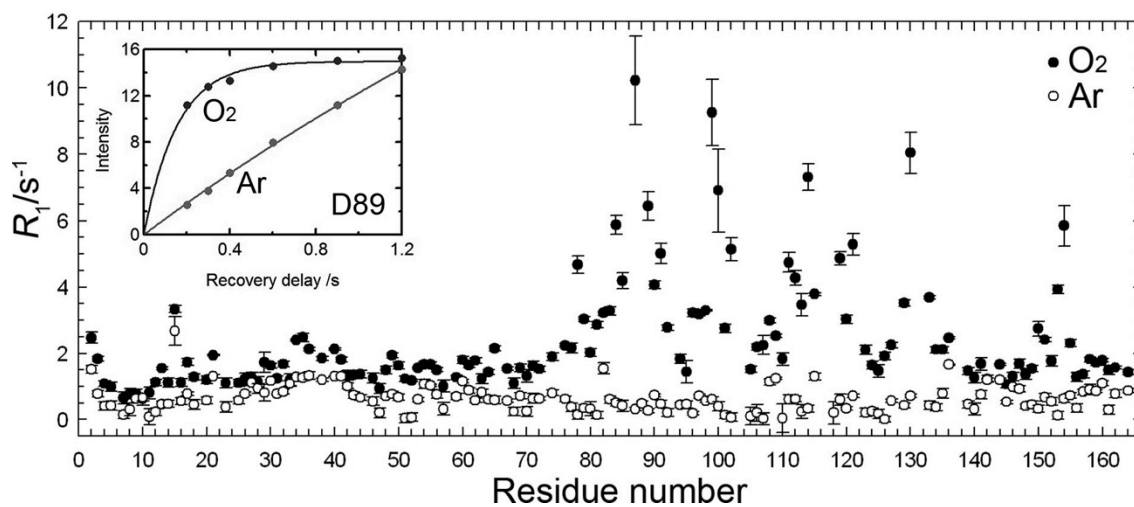


Figure S4.  $^1\text{H}$  longitudinal relaxation rate constants,  $R_1$ , for amide protons at 0 mM (Ar 2 bar, open circles) and 6.4 mM ( $\text{O}_2$  5 bar, closed circles)  $\text{O}_2$  against residue number. Error bars from curve fitting are included. Changes in crosspeak intensities of D89 in  $^1\text{H}/^{15}\text{N}$  HSQC spectra as a function of recovery delay are shown in the inset.

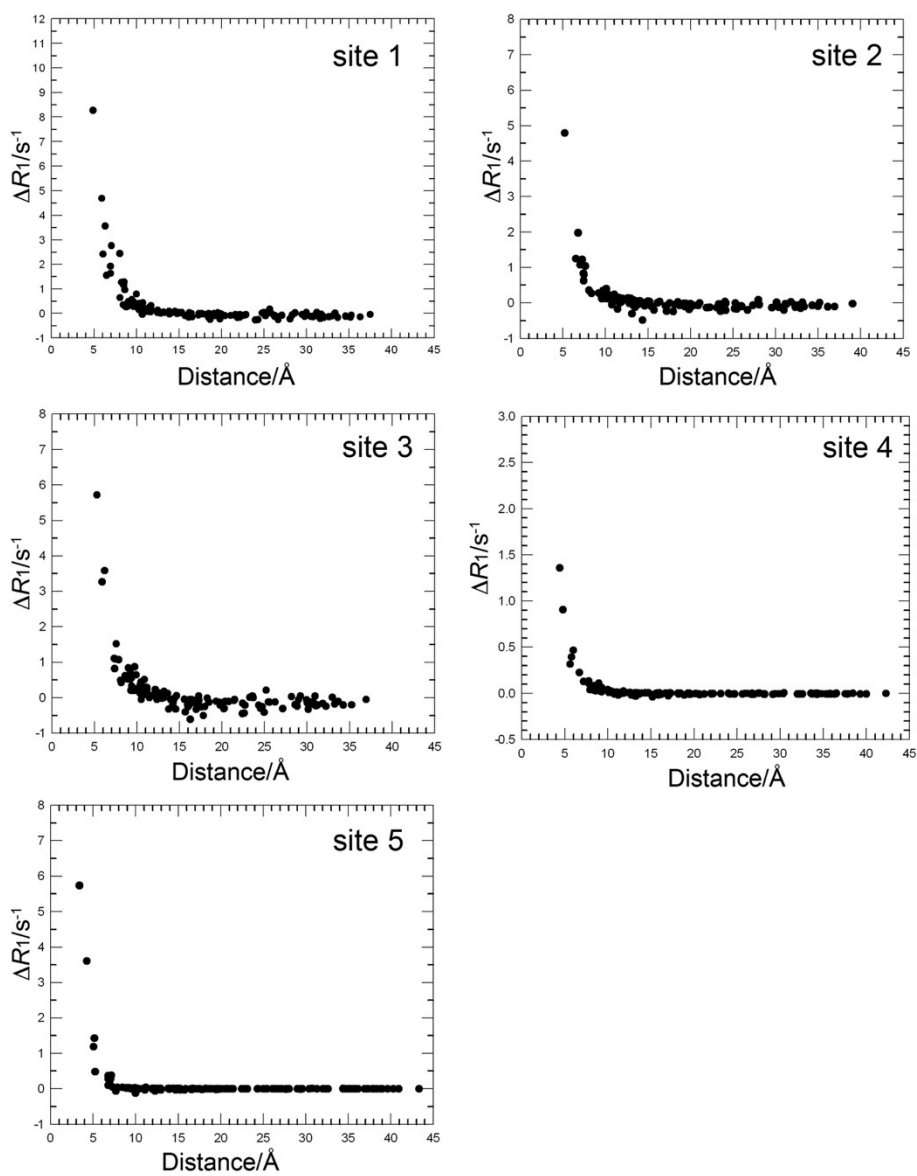


Figure S5. Predicted  $\Delta R_1$  for amide protons due to paramagnetic relaxation at each binding site (1-5) as a function of distance.  $\Delta R_1$  from each binding site 1-5 is explained in Figure 4b.

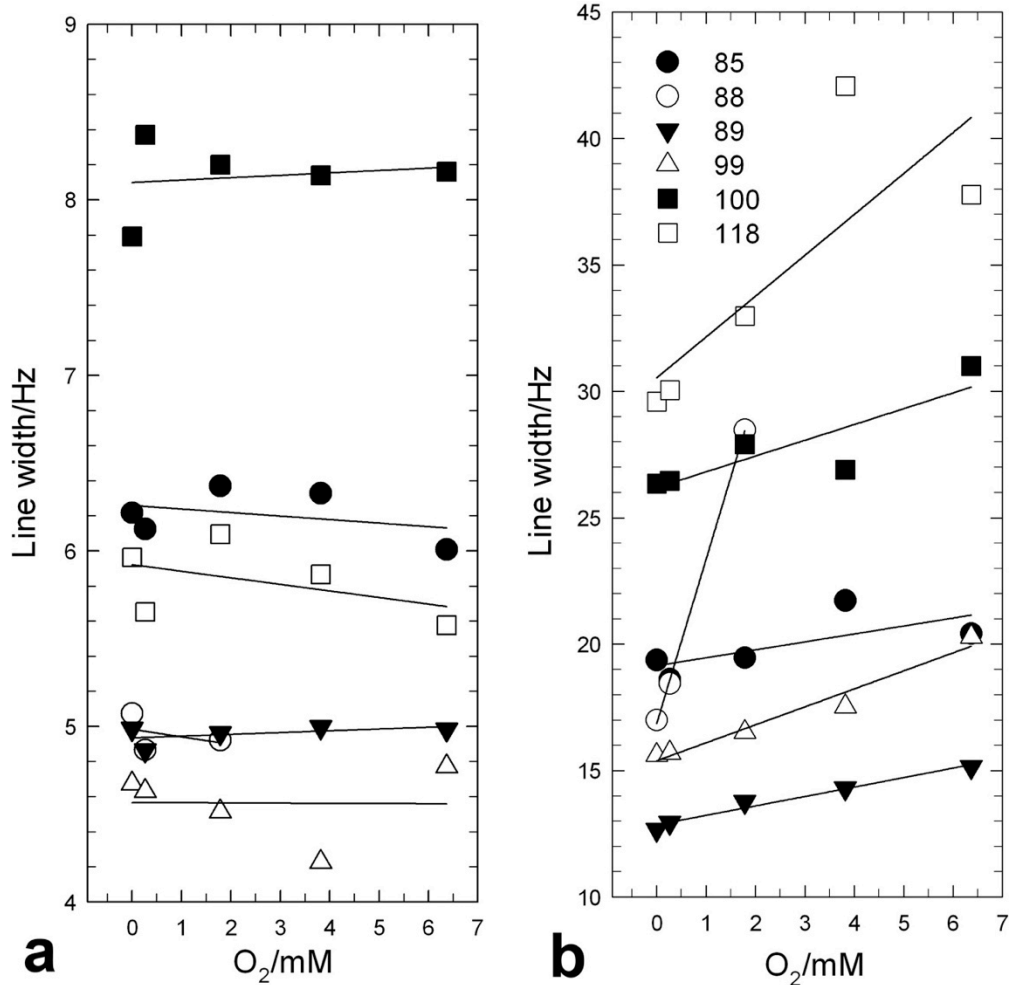


Figure S6.  $^{15}\text{N}$  (a) and  $^1\text{H}$  (b) line-widths of crosspeaks of the residues around the enlarged cavity in the refocused  $^1\text{H}/^{15}\text{N}$  HSQC spectra. The legend included in panel b is also applicable to panel a.



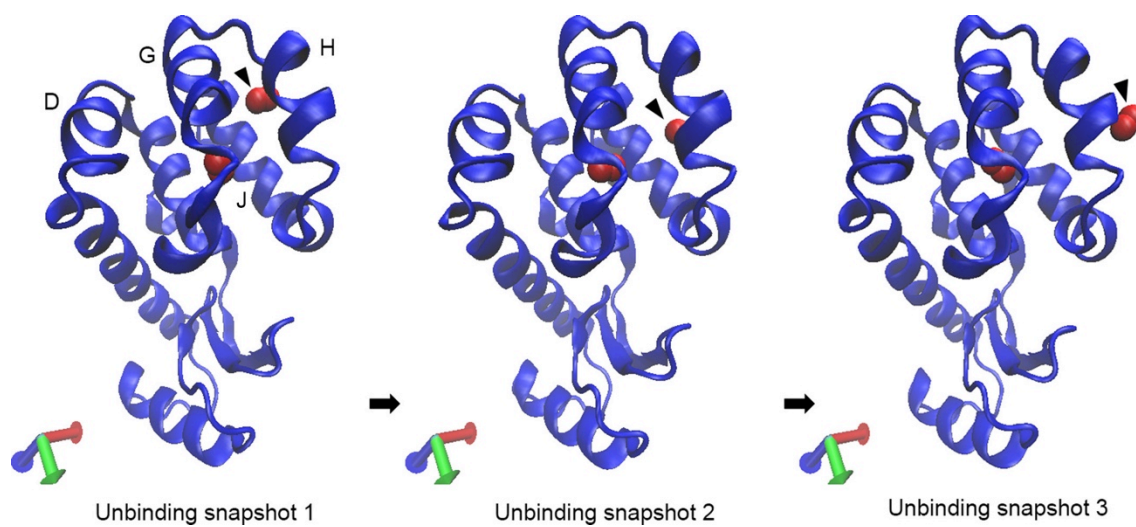


Figure S7. A series of snapshots showing egress of O<sub>2</sub> molecule from T4 lysozyme L99A. O<sub>2</sub> molecules showing egress from cavity 3 are indicated by filled triangles. The D, G, H, and J helices are labeled. The picture was prepared using VMD 1.9.2.

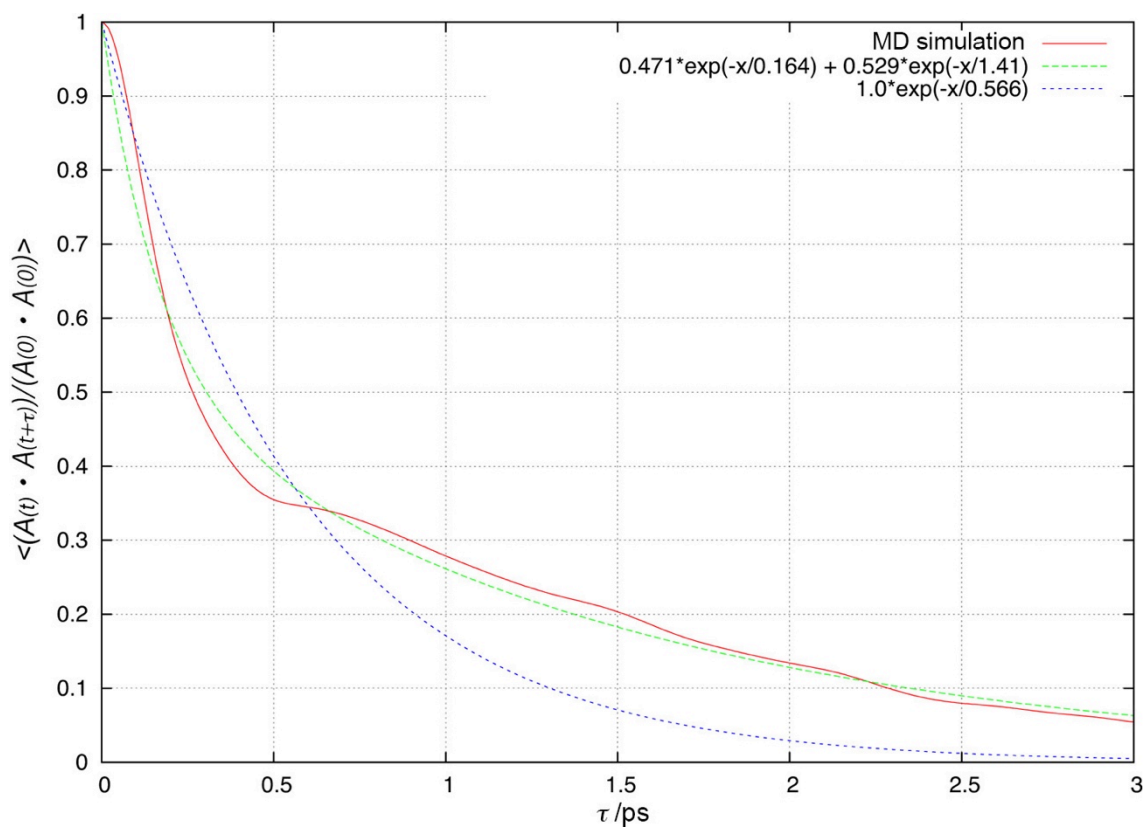


Figure S8. Estimation of the rotational correlation time for  $\text{O}_2$  in cavity 4.  $A(t)$  and  $A(t+\tau)$  are direction vectors joining the two oxygen atoms at time  $t$  and  $t+\tau$ , respectively.  $\langle (A(t) \cdot A(t+\tau)) / (A(0) \cdot A(0)) \rangle$  shows rotational correlation function of  $\text{O}_2$  molecule. Assuming single-exponential (blue) or bi-exponential (green) decays, the rotational correlation times of  $\text{O}_2$  were estimated to be  $0.6 \pm 0.1$  ps or  $0.164 \pm 0.006$  ps and  $1.41 \pm 0.02$  ps, respectively.

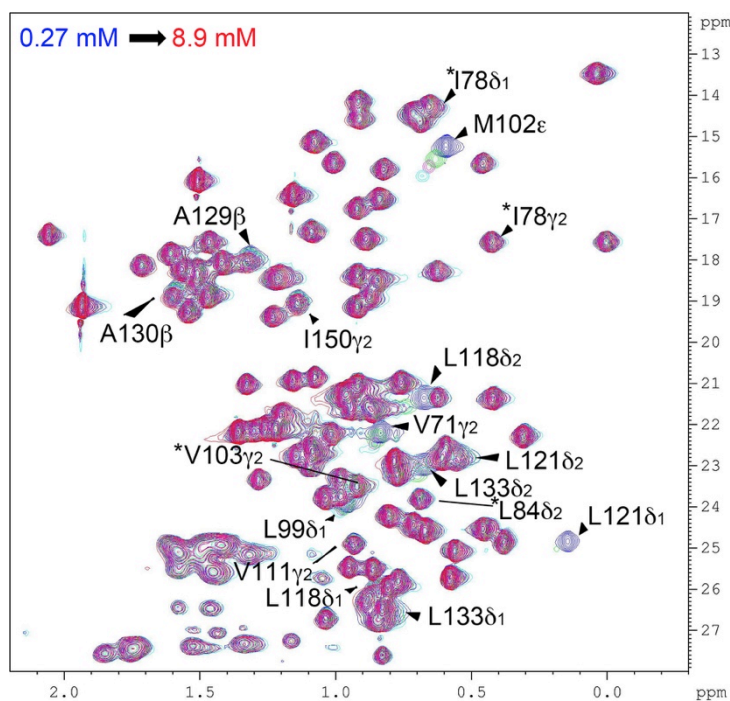


Figure S9.  $^1\text{H}/^{13}\text{C}$  constant time HSQC spectra of  $^{13}\text{C}/^{15}\text{N}$  labeled T4 lysozyme (WT\*) at different oxygen concentrations from 0.27 mM to 8.9 mM. Positive and negative crosspeaks are presented by same color. Methyl groups showing significant changes in  $^1\text{H}/^{13}\text{C}$  chemical shift and a loss of crosspeak intensities on L99A variant are indicated. Asterisks show methyl groups which exhibit significant change in chemical shifts and/or crosspeak intensities in L99A but not in WT\* protein.

Table S1. O<sub>2</sub>-induced longitudinal relaxation enhancements of methyl protons.

Group	O <sub>2</sub> (3 bar) <sup>a</sup>		N <sub>2</sub> (3 bar) <sup>a</sup>		$\Delta R_1/s^{-1}$	Error	Distance/Å <sup>c</sup>
	$R_1/s^{-1}$	Error	$R_1/s^{-1}$	Error			
M1ε	1.49	0.04	0.95	0.06	0.54	0.07	19
I3γ <sub>2</sub>	2.4	0.5	1.3	0.4	1.1	0.7	11
I3δ <sub>1</sub>	2.5	0.16	1.5	0.13	1.0	0.2	14
M6ε	1.12	0.09	0.6	0.11	0.6	0.14	11
L7δ <sub>1</sub>	2.6	0.3	2.4	0.3	0.2	0.5	13
L7δ <sub>2</sub>	2.5	0.16	2.5	0.15	0.0	0.2	13
I9γ <sub>2</sub>	2.2	0.2	1.9	0.18	0.3	0.3	19
I9δ <sub>1</sub>	1.9	0.19	1.6	0.3	0.3	0.3	20
L13δ <sub>1</sub>	2.0	0.11	1.6	0.12	0.4	0.17	26
L15δ <sub>1</sub>	3.0	0.2	2.7	0.14	0.3	0.3	30
I17γ <sub>2</sub>	2.8	0.16	2.2	0.13	0.7	0.2	30
I17δ <sub>1</sub>	2.1	0.2	1.5	0.17	0.6	0.3	29
I27δ <sub>1</sub>	2.0	0.19	1.9	0.10	0.2	0.2	29
I27γ <sub>2</sub>	2.7	0.3	2.5	0.19	0.2	0.3	25
I29γ <sub>2</sub>	1.6	0.19	1.2	0.3	0.4	0.3	19
I29δ <sub>1</sub>	1.7	0.19	1.4	0.2	0.3	0.3	22
L32δ <sub>1</sub>	2.1	0.10	1.7	0.08	0.5	0.13	21
L32δ <sub>2</sub>	2.6	0.10	2.3	0.14	0.3	0.17	19
L33δ <sub>2</sub>	3.8	0.4	2.6	0.3	1.2	0.5	26
L33δ <sub>1</sub>	3.4	0.4	3.6	0.6	-0.2	0.7	25
T34γ <sub>2</sub>	3.0	0.2	3.0	0.3	0.0	0.4	27
L39δ <sub>1</sub>	2.7	0.2	2.4	0.2	0.3	0.3	35
L39δ <sub>2</sub>	3.5	0.2	2.3	0.13	1.2	0.3	31
A41β	4.1	0.3	4.1	0.3	0.0	0.5	32
A42β	2.3	0.18	2.1	0.2	0.2	0.3	29
L46δ <sub>1</sub>	3.0	0.3	2.4	0.3	0.6	0.4	28
L46δ <sub>2</sub>	2.9	0.5	2.6	0.4	0.3	0.6	30
A49β	2.7	0.16	2.2	0.17	0.5	0.2	26

I50 $\gamma_2$	2.6	0.17	2.0	0.15	0.6	0.2	31
I50 $\delta_1$	2.6	0.16	1.4	0.17	1.2	0.2	28
T54 $\gamma_2$	2.3	0.15	1.9	0.10	0.4	0.18	33
V57 $\gamma_{1&2}$	2.93	0.06	2.33	0.05	0.60	0.08	35
I58 $\gamma_2$	1.9	0.16	1.7	0.16	0.2	0.2	30
I58 $\delta_1$	2.0	0.2	1.8	0.3	0.2	0.3	27
T59 $\gamma_2$	3.0	0.11	2.4	0.14	0.6	0.18	34
A63 $\beta$	2.6	0.3	2.3	0.3	0.3	0.4	25
L66 $\delta_1$	3.0	0.3	2.0	0.2	1.0	0.4	24
L66 $\delta_2$	3.3	0.2	1.9	0.16	1.4	0.3	24
V71 $\gamma_2$	2.9	0.2	2.4	0.16	0.6	0.3	15
A73 $\beta$	4.4	0.3	2.0	0.12	2.4	0.4	16
A74 $\beta$	4.2	0.6	2.4	0.5	1.8	0.8	11
V75 $\gamma_1$	4.6	0.3	2.6	0.18	2.0	0.3	11
V75 $\gamma_2$	3.8	0.3	1.7	0.19	2.1	0.4	11
I78 $\gamma_2$	32	10	1.2	0.17	31	10	4.6
I78 $\delta_1$	21	6	0.9	0.12	20	6	5.5
L79 $\delta_2$	3.7	0.3	1.8	0.14	1.9	0.3	8.4
A82 $\beta$	3.2	0.16	2.2	0.12	1.0	0.2	12
L84 $\delta_1$	15	1.8	2.5	0.6	12	1.9	5.7
L84 $\delta_2$	b		2.9	0.2			4.3
V87 $\gamma_1$	e		2.6	0.5			4.2
V87 $\gamma_2$	8.3	0.3	1.2	0.11	7.1	0.3	5.4
L91 $\delta_2$	9.9	0.8	1.0	0.19	8.9	0.8	7.2
A93 $\beta$	3.9	0.3	2.9	0.19	0.9	0.3	14
V94 $\gamma_1$	1.7	0.16	1.3	0.17	0.5	0.2	11
V94 $\gamma_2$	2.1	0.14	1.4	0.13	0.7	0.19	14
A97 $\beta$	4.2	0.4	2.5	0.4	1.7	0.6	12
A98 $\beta$	3.6	0.4	1.4	0.3	2.1	0.5	6.9
A99 $\beta$	b		3.5	0.8			3.6
I100 $\gamma_2$	3.4	0.3	1.7	0.3	1.7	0.4	9.9
I100 $\delta_1$	4.1	0.3	1.2	0.14	2.9	0.3	9.4

M102 $\epsilon$	23	7	0.7	0.3	22	7	4.4
V103 $\gamma_1$	4.9	0.3	2.9	0.19	2.0	0.3	8.5
V103 $\gamma_2$	e		1.8	0.5			5.4
M106 $\epsilon$	2.4	0.2	0.6	0.15	1.8	0.3	7.9
T109 $\gamma_2$	3.4	0.19	2.6	0.13	0.8	0.2	13
V111 $\gamma_1$	e		2.4	0.19			3.3
V111 $\gamma_2$	10	2	2.3	0.4	8	2	5.7
A112 $\beta$	4.5	0.3	2.0	0.2	2.5	0.4	8.6
T115 $\gamma_2$	3.8	0.3	3.0	0.16	0.9	0.3	9.9
L118 $\delta_1$	3.6	0.16	2.4	0.3	1.3	0.3	5.6
L118 $\delta_2$	b		2.1	0.5			3.8
M120 $\epsilon$	1.14	0.04	0.6	0.03	0.58	0.05	8.2
L121 $\delta_1$	b		2.4	1			3.5
L121 $\delta_2$	2.8	0.18	2.0	0.12	0.9	0.2	3.7
A129 $\beta$	8.8	0.7	1.6	0.2	7.2	0.8	3.5
A130 $\beta$	9.3	0.9	2.3	0.2	6.9	0.9	4.1
V131 $\gamma_1$	3.2	0.11	2.44	0.09	0.7	0.14	9.8
V131 $\gamma_2$	1.70	0.08	1.45	0.09	0.3	0.12	8.6
L133 $\delta_1$	12	3	2.6	0.4	9	3	3.5
L133 $\delta_2$	4.7	0.8	3.2	0.3	1.4	0.9	3.7
A134 $\beta$	2.6	0.2	1.8	0.2	0.8	0.3	8.8
T142 $\gamma_2$	3.5	0.4	2.8	0.5	0.7	0.6	16
A146 $\beta$	3	1	4	2	-1	2	10
V149 $\gamma_1$	3.6	0.3	1.5	0.17	2.2	0.3	7.6
V149 $\gamma_2$	2.3	0.3	1.5	0.3	0.8	0.4	9.8
I150 $\gamma_2$	6.0	0.6	1.5	0.2	4.5	0.6	4.5
I150 $\delta_1$	2.6	0.4	1.2	0.3	1.4	0.4	6.8
T151 $\gamma_2$	2.88	0.08	2.39	0.08	0.5	0.12	9.7
T152 $\gamma_2$	2.7	0.4	2.4	0.5	0.3	0.6	9.3
T155 $\gamma_2$	2.8	0.17	2.3	0.2	0.6	0.3	9.2
T157 $\gamma_2$	2.8	0.2	2.3	0.17	0.5	0.3	13
A160 $\beta$	2.28	0.09	2.1	0.14	0.2	0.17	11

L164 $\delta_1$	3.1	0.13	2.5	0.10	0.6	0.16	d
L164 $\delta_2$	2.70	0.07	2.20	0.07	0.5	0.10	d

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<sup>a</sup>Absolute pressure consisting of gauge pressure plus atmospheric pressure.

<sup>b</sup>Crosspeaks severely broadened at 3.8 mM O<sub>2</sub>.

<sup>c</sup>Distances from the closest xenon (one of sites 1-5) to each hydrogen in the L99A crystal structure at 8 atm xenon pressure (PDB ID, 1c6k). Hydrogen atoms were added to the crystal structure by using the WHATIF server.

<sup>d</sup>Atomic position is not available.

<sup>e</sup>Peaks are overlapped with other peaks.

Movie S1. MD simulation of 100 nanoseconds (example 1). An O<sub>2</sub> molecule was inserted in both cavities 3 and 4 of the crystal structure of L99A T4 lysozyme (PDB ID; 1c6k). The movie consists of 500 snapshots taken every 0.2 nanoseconds.

Movie S2. MD simulation of 100 nanoseconds (example 2). An O<sub>2</sub> molecule was inserted in both cavities 3 and 4 of the crystal structure of L99A T4 lysozyme (PDB ID; 1c6k). The movie consists of 500 snapshots taken every 0.2 nanoseconds.

Movie S3. MD simulation of 100 nanoseconds (example 3). An O<sub>2</sub> molecule was inserted in both cavities 3 and 4 of the crystal structure of L99A T4 lysozyme (PDB ID; 1c6k). The movie consists of 500 snapshots taken every 0.2 nanoseconds.

## References

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2. Scharlin, P., Battino, R., Silla, E., Tunon, I. & Pascual-Ahuir, J.L. Solubility of gases in water: Correlation between solubility and the number of water molecules in the first solvation shell. *Pure Appl. Chem.* **70**, 1895-1904 (1998).