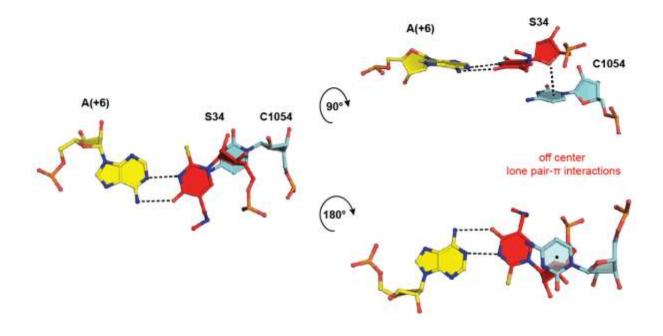


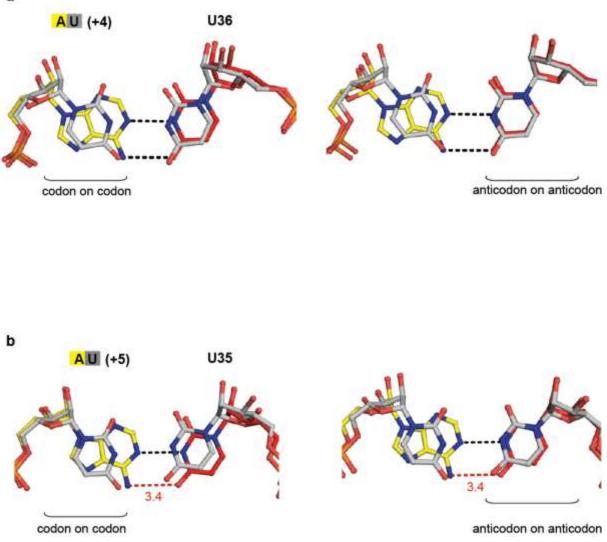
in the 30S model and, as a result, displaces towards the first nucelotide of the A-site bound codon

A1913 from the large ribosomal subunit restricts position of the anticodon loop at the t<sup>6</sup>A37 nucleotide in the 70S model

Supplementary Figure 1. Different conformations of *E.coli* tRNA<sup>Lys</sup><sub>UUU</sub> anticodon loops in the partial model of the 30S subunit and complete structure of the 70S ribosome. Comparison of the t<sup>6</sup>A37 stacking interactions in the model of the 30S subunit, which crystals were soaked with the synthetic ASL of *E. coli* tRNA<sup>Lys</sup><sub>UUU</sub>, hexanucleotide as an analogs of mRNA and miscoding antibiotic paromomycin (Paro)<sup>24</sup>, and complete system of the 70S ribosome co-crystallized with native *E. coli* tRNA<sup>Lys</sup><sub>SUU</sub> and long mRNA (current study; complex 2). The superposition shows a shift of t<sup>6</sup>A37 towards the first codon nucleoside in the 30S structure relatively to the position of t<sup>6</sup>A37 in the 70S model. The observed difference can be explained by the physical lack of a natural constraint from the large ribosomal subunit, i.e. fixation of t<sup>6</sup>A37 by A1913 from 23S rRNA, on the tRNA<sup>Lys</sup> ASL in the 30S model.



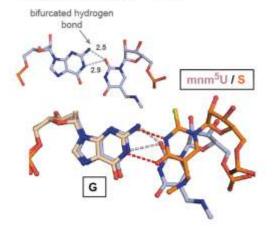
Supplementary Figure 2. Lone pair-aromatic interactions between C1054 and S34. Decoding center on the 70S ribosome controls the first anticodon nucleotide position S34 by weak lone pair- $\pi$  interactions with ribose of C1054 in 16S rRNA. Displayed are three projections, which show that the specified interaction is off-center.

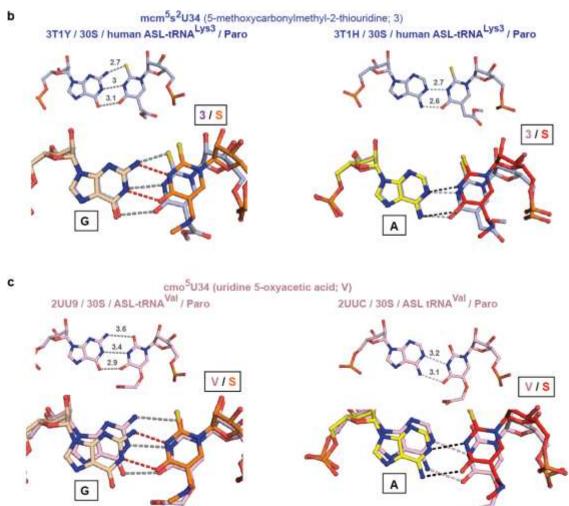


Supplementary Figure 3. Comparison of a pyrimidine-pyrimidine mismatch with the canonical Watson-Crick pair in the decoding center of the 70S ribosome. (a, b) Alignment of the Watson-Crick U•A pairs at the first (a, complex 1) and second (b, complex 2) codonanticodon positions with similar structures where the codon nucleotide was substituted to uridine (complexes 3 and 4). In both U•U mismatches the interatomic distances between Watson-Crick edges of uridines are considerably larger than in canonical pairs (see the main text).

a

mnm<sup>5</sup>U34 (5-methylaminomethyluridine) lacking s<sup>2</sup> 1XMO / 30S / ASL-tRNA<sup>Lys</sup> / Paro





Supplementary Figure 4. Comparison of S34•G and S34•A with other wobble pairs formed by hypermodified uridines. (a) Lack of the thio group in mnm<sup>5</sup>s<sup>2</sup>U leads to a drastic change in position of the uracil ring in the 30S model; refer to ref. 25, where this structural data are challenged by the same authors. (b) The mcm<sup>5</sup>s<sup>2</sup> modification specific to one of the

eukaryotic species of lysine tRNA induces a Watson-Crick-like conformation of the mcm<sup>5</sup>s<sup>2</sup>U•G wobble pair in the heterologous system with the bacterial small subunit (left). (c) The cmo<sup>5</sup>U34 of *E.coli* tRNA<sup>Val</sup><sub>UAC</sub> stimulates a C•G-like canonical interaction in the decoding center of the 30S subunit (left). In (b) and (c) the right panels show alignments of the S34•A pair with mcm<sup>5</sup>s<sup>2</sup>U34•A and cmo<sup>5</sup>U34•A. In all cases the comparison is done by superposition of codons; each panel specifies the PDB code taken for comparison as well as the modification in ASL of tRNA; Paro stands for a miscoding antibiotic paromomycin used in all models of the 30S subunit to improve ASL binding. The difference in atom positions are indicated by arrows and corresponding value in angstroms.