

Table S1. Primers used for qPCR and qRT-PCR

Name	Sequence	Target region
INTEG F*	GGCTGTCAGAATATGGGGCCGTAGTA	Intergenic region on chromosome V
INTEG R*	CACCCCGAAGCTGCTTTCACAATAC	
18S2-F**	TCCAATTGTTCTCGTTAAG	"18S" 5' Region
18S2-R**	ATTCAGGGAGGTAGTGACAA	
LYS2 BGL F	GAGTAACCGGTGACGATGATATT	"S μ " (5' of LYS2 gene; 100 nt upstream of S μ insertion site)
LYS2 BGL R	CATTAAATGACCACGTTGGTTGA	
LYS2 STP F	GGACAGGTGTTGATTGGTCTAA	"LYS3" (3' of LYS2 gene; 3.5 Kb away from S μ)
LYS2 STP R	TCATTATGAGTTGGTGGAGGTAAA	
CAN1 F	GAGTTCTGGGTCGTTCCAT	"CAN1" gene (YEL063C)
CAN1 R	GGCACCTGGGTTTCTCCAAT	
UBC6 F***	GATACTTGAATCCTGGCTGGTCTGTCTC	UBC6 gene (YER100W)
UBC6 R***	AAAGGTCTTCTGTTTCATCACCTGTATTTGC	
ALG9 F***	CACGGATAGTGGCTTTGGTGAACAATTAC	ALG9 gene (YNL219C)
ALG9 R***	TATGATTATCTGGCAGCAGGAAAGAACTTGGG	

Primers referenced from previous reports: *(1), ** (2), *** (3)

Table S2. The relative RNA levels of *lys2-GTOP* or *-GBTM* allele.

<i>lys2</i> allele	Genetic Background	Reference gene: <i>UBC6</i>		Reference gene: <i>ALG9</i>	
		% Relative RNA* - Near S μ (STD)**	% Relative RNA* - at LYS3' end (STD)**	% Relative RNA* - Near S μ (STD)**	% Relative RNA* - at LYS3' end (STD)**
<i>pTET-lys2-GTOP</i>	<i>WT</i>	100 (\pm 9.8)	100 [#] (N.D.)	100 (\pm 16)	100 [#] (N.D.)
<i>pTET-lys2-GBTM</i>	<i>WT</i>	107 (\pm 12)	75 (\pm 24)	122 (\pm 13)	94 (\pm 40)
<i>pTET-lys2-GTOP</i>	<i>top1</i> Δ	92 (\pm 31)	153 (\pm 34)	101 (\pm 14)	187 (\pm 43)
<i>pTET-lys2-GBTM</i>	<i>top1</i> Δ	88 (\pm 14)	53 (\pm 8.8)	104 (\pm 18)	68 (\pm 12)
<i>pTET-lys2-GTOP</i>	<i>rnh1</i> Δ <i>rnh201</i> Δ	91 (\pm 3.5)	165 [#] (N.D.)	105 (\pm 6.2)	196 [#] (N.D.)
<i>pTET-lys2-GBTM</i>	<i>rnh1</i> Δ <i>rnh201</i> Δ	136 (\pm 44)	93 (\pm 29)	151 (\pm 35)	110 (\pm 24)

RNA was extracted using the standard hot acidic phenol method and treated with DNase I (New England Biolabs). Relative RNA levels were determined by quantitative RT-PCR and $\Delta\Delta$ Cq analysis using *UBC6* or *ALG9* as the reference gene. qRT-PCR was carried out using SensiFAST SYBR No-ROX One-Step kit from Bioline and Biorad CFX Connect instrument. RT-PCR conditions were as follows: 45°C for 10 mins, 95°C for 2 mins followed by 40 cycles of 95°C for 5 sec, 60°C for 10 sec, and 72°C for 5 sec. Primers for amplification of “Near S μ ” (LYS2 BGL F and LYS2 BGL R) are located ~ 250 NT from the 5' end of the *LYS2* mRNA and ~ 100 NT upstream of *GTOP* or *GBTM* cassette-insertion site. Primers for the amplification of “LYS 3'” (LYS2 STP F and LYS2 STP

R) are located ~4,000 NT from the 5' end of the *LYS2* mRNA. Primer sequences are listed in Table S1. *% was calculated by normalizing to the level of RNA in WT cells with *pTET-lys2-GTOP*.

**The standard deviations (STD) were calculated from three independent experiments except when indicated (#). #Two experiments were used to calculate mean expression level (standard deviation could not be determined – N.D.). When normalized to the transcription level at S_{μ} , the relative level of transcription at *LYS3'* in WT cells with *pTET-lys2-GTOP* was $156 \pm 28\%$ (*UBC6* as control) or $145 \pm 7.7\%$ (*ALG9* as control).

SUPPLEMENTAL FIGURE LEGENDS

Figure S1. Recombination Assay to test the effect of G-run containing sequence on genome stability in yeast.

The construction of plasmids and yeast strains involved in the designing of the recombination assay was previously described in detail (4).

A. Sequence of S μ fragment. Sequence of the guanine-rich strand of the 770-bp fragment of mouse Ig S μ region (Pubmed Accession # J00442) that was inserted into the *Bgl* II site of the *LYS2* gene is shown. In this orientation, the fragment comprises 137 A, 117 C, 358 G, and 158 T. Runs of ≥ 3 guanines are indicated in red.

B. Recombination Construct. The *lys2* allele with the insertion of S μ fragment (hashed box) under the control of *pTET* promoter (*pTET-lys2-GTOP* or *-GBTM*) and the 3' truncated *lys2* allele are located on the CHR III and CHR XV, respectively. Neither allele produces functional Lys2 protein (Lys⁻). A Homologous recombination (HR) initiating at the *pTET-lys2-GTOP* or *-GBTM* construct and employing the truncated *lys2* allele on CHR XV as the donor sequence can be resolved either in crossover or gene conversion (non-crossover). Due to the orientations of the *lys2* alleles relative to the respective centromeres, crossover products of HR result in dicentric chromosome leading to inviable cells. S μ insert is lost in the gene conversion product leading to the functional *LYS2* gene and the selectable Lys⁺ phenotype.

Figure S2. Effects of Top1Y727F on the recombination rates under low transcription conditions or in WT strain background.

A. The rates of recombination under low transcription conditions. Empty vector (Vec – pRS416) or Top1 Y727F expressing plasmid was transformed into *top1* Δ cells. Growth media was synthetic media lacking uracil supplemented with 2% glucose and 2 μ g/ml doxycycline (+Dox).

B. The rates of recombination in WT background. The rates determined after growth in synthetic media lacking uracil supplemented with 2% glucose. Vec: pRS416. P values (shown above red brackets) were determined by Mann-Whitney test using the Prism software. * - significantly different; ns - no significant difference.

Figure S3. Survival of CPT-treated cells.

WT cells containing the indicated *pTET-lys2*-derived reporter construct were treated either with DMSO or CPT (see material and methods). Percent survival was calculated relative to the DMSO treated cultures. Error bars indicate standard deviation from 8 to 24 independent measurements. For low transcription conditions (black bar), 2 μ g/ml doxycycline was added to the growth media.

Figure S4. Alignment of human and yeast Top1.

The sequences of human and yeast topoisomerases I are located on the top and bottom lines of the alignment, respectively (5). Residues contacting DNA (6) are indicated in red (conserved) and in yellow (not conserved). A. The core domain (hTop1 residues 215 to 634). B. The C-terminal catalytic domain (hTop1 residues 712 to 765).

REFERENCES FOR THE SUPPLEMENTAL MATERIALS

1. Chan, Y.A., Aristizabal, M.J., Lu, P.Y., Luo, Z., Hamza, A., Kobor, M.S., Stirling, P.C. and Hieter, P. (2014) Genome-wide profiling of yeast DNA:RNA hybrid prone sites with DRIP-chip. *PLoS Genet*, **10**, e1004288.
2. El Hage, A., Webb, S., Kerr, A. and Tollervey, D. (2014) Genome-Wide Distribution of RNA-DNA Hybrids Identifies RNase H Targets in tRNA Genes, Retrotransposons and Mitochondria. *PLoS Genet*, **10**, e1004716.
3. Teste, M.A., Duquenne, M., Francois, J.M. and Parrou, J.L. (2009) Validation of reference genes for quantitative expression analysis by real-time RT-PCR in *Saccharomyces cerevisiae*. *BMC molecular biology*, **10**, 99.
4. Kim, N. and Jinks-Robertson, S. (2011) Guanine repeat-containing sequences confer transcription-dependent instability in an orientation-specific manner in yeast. *DNA Repair (Amst)*.
5. Larkin, M.A., Blackshields, G., Brown, N.P., Chenna, R., McGettigan, P.A., McWilliam, H., Valentin, F., Wallace, I.M., Wilm, A., Lopez, R. *et al.* (2007) Clustal W and Clustal X version 2.0. *Bioinformatics*, **23**, 2947-2948.
6. Redinbo, M.R., Stewart, L., Kuhn, P., Champoux, J.J. and Hol, W.G. (1998) Crystal structures of human topoisomerase I in covalent and noncovalent complexes with DNA. *Science*, **279**, 1504-1513.

Figure S1

A

			GATC	GAGCTGAGCT	GAGCTGGGTG	AGCTGAGCTG
AGCTGAGCTG	GGTGAGCTGA	GCTGAGCTGA	GCTGAGCTGG	GTGAGCTGAG	CTGAGCTGAG	CTGAGCTGAG
CTGAGCTGAG	CTGAGCTGGG	GTGAGCTGGG	CTGAGCTGAG	CTGAGCTGGG	GTGAGCTGAG	CTGAGCTGAG
CTGAGCTGAG	CTGAGCTGGG	GTGAGCTGGG	CTGAGCTGGG	GTGAGCTGGG	CTGAGCTGAG	CTGAGCTGAG
CTGGGGTGAG	CTGAGCTGAG	CTGAGCTGAG	CTGGGGTGAG	CTGAGCTGAG	CTGAGCTGGG	CTGAGCTGGG
GTGAGCTGAG	CTGGGCTGAG	CTGAGCTGGG	GTGAGCTGAG	CTGAGCTGAG	CTGAGCTGGG	CTGAGCTGGG
GTGAGCTGAG	CTGAGCTGAG	CTGGGGTGAG	CTGAGCTGGG	GTGAGCTGAG	CTGAGCTGGG	CTGAGCTGGG
GTGAGCTGAG	CTGAGCTGAG	CTGGGGTGAG	CTGAGCTGGG	GTGAGCTGAG	CTGAGCTGGG	CTGAGCTGGG
GTGAGCTGAG	CTGAGCTGGG	GTGAGCTGAG	CTGAGCTGGG	GTGAGCTGAG	CTGAGCTGGG	CTGAGCTGGG
GTGAGCTGAG	CTAGGGTGAG	CTGGGCTGGG	TGAGCTGGAG	TGAGCTGAGC	TGAGGTGAAC	TGAGGTGAAC
TGGGGTGAGC	CGGATGTTTT	GAGTTGAGCT	GGGGTAAGAT	GAGCTGAACT	GGGGTAAGAT	GGGGTAAGAT
GGGATGAGCT	GTGGTGAGGG	GAGCTGGATT	GAAGTGAAGCT	GTGTGAGCTG	AGCTGGGGTC	AGCTGGGGTC
AGCTGAGCAA	GAGTGAGTAG	AGCTGGCTGG	CCAAGAACCA	AGAATCAATT	AGGCTAAGTG	AGGCTAAGTG
AGCCAGATTG	CGCTGGGATC					

B

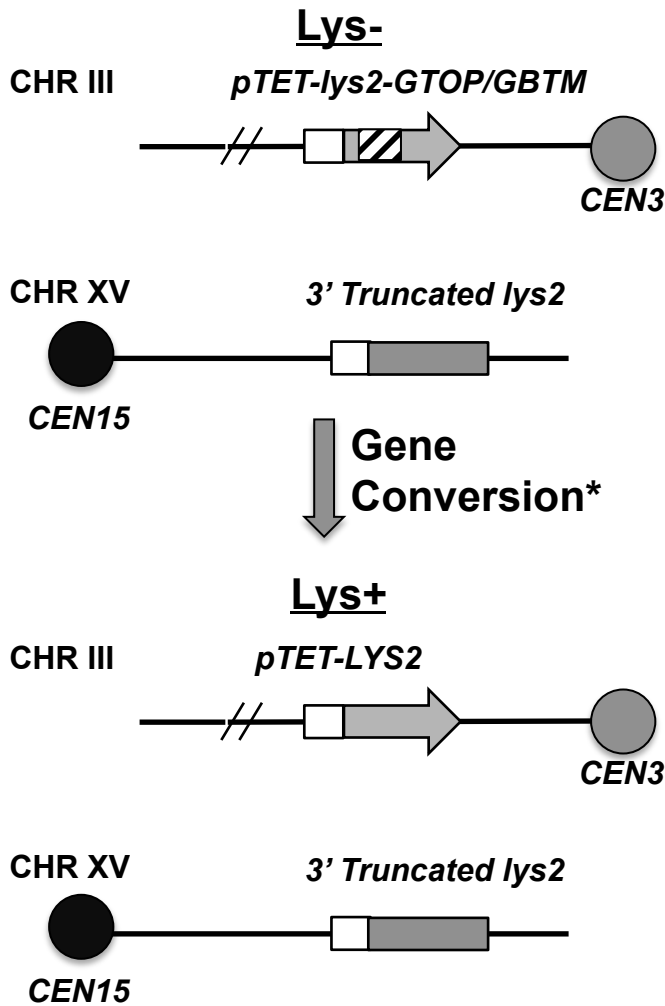


Figure S2

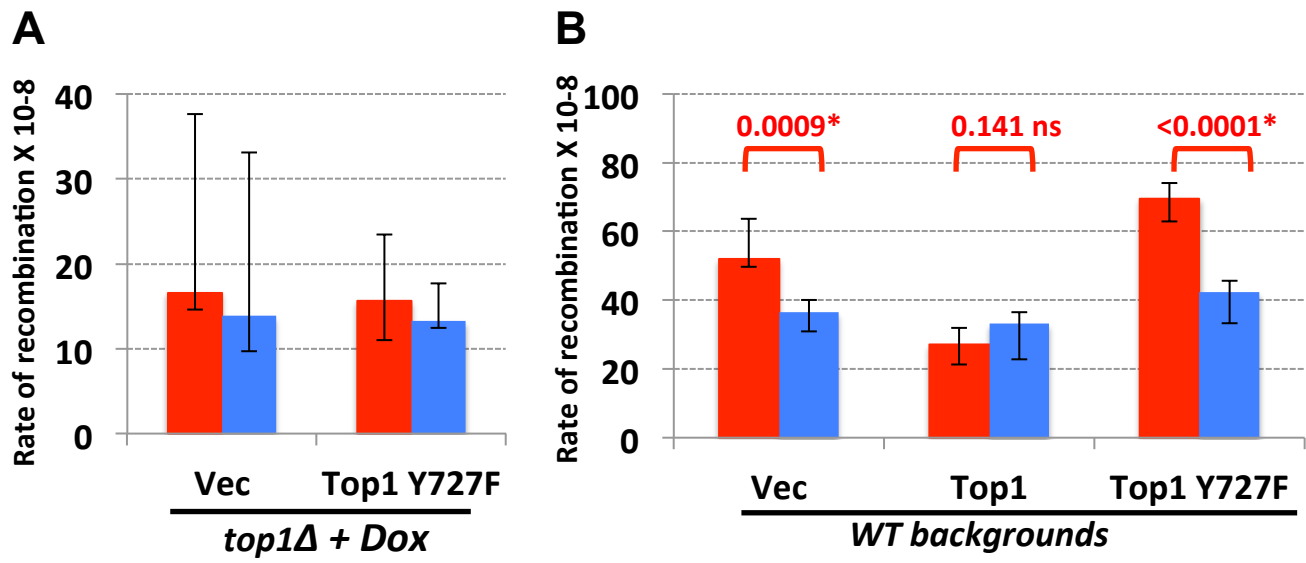


Figure S3

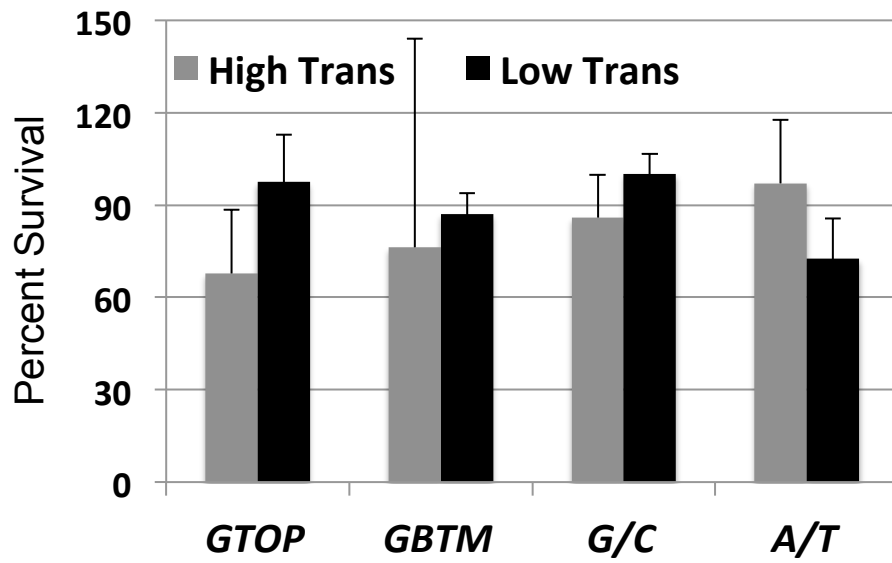


Figure S4

A. Core domain (53% identity; 73% homology)

IKWKFLEHKGPVFAPPYEPLPENVKFYDYGKVMKLSPKAEEVATFFAKMLDHEYTTKEIF
IKWVTLKHNGVIFPPPYQPLPSHIKLYDYGKPVLDLPPQAEVAGFFAALLES DHAKNPVF
*** *:*:* :*.***:***.:*:***** :.*.*:***** *** :*: :.:.: :*

RKNFFKDWRKEMTNE----EKNIIITNLSKCDFTQMSQYFKAQTEARKQMSKEEKLKIKEE
QKNFFNDFLQVLKESGGPLNGIEIKEFSRCDFTKMFDFYFQLOKEQKKQLTSQEKKQIRLE
:****:*: : :.:. : *.:*:*****:* **:*. * :*:.:.* :*: *

NEKLLKEYGFCIMDNHKERIANFKIEPPGLFRGRGNHPKMGMLKRRIMPEDI I INCSKDA
REKFEEDYKFCELDGRREQVGNFKVEPPDLFRGRGAHPKTGKLRKRRVNPEDIVLNLSKDA
. **: ::* ** :*. :.:*:.*.***** ** * *****: *****:.* *****

KVPSPPPGHKWKEVRHDNKVTVLVSWTENIQGSIKYIMLNPSSRIKGEKDWQKYETARRL
PVPPAPEGHKWGEIRHDNTVQWVLA MWRENIFNSFKYVRLAANSSLKQSDYKKFEKARQL
..* ***** :*:*:*.* **.* * **.* .*:*: * .*. * :*:.*:.*:.*.*:.*

KKCVDKIRNQYREDWKSKEKVRQRAVALYFIDKLALRAGNEKEEGETADTVGCCSLRVE
KSYIDAIRRDYTRNLKSKVMLERQKAVAIYLIDVFALRAGGEKSE-DEADTVGCCSLRYE
. :* **.*:.* .: *** * **:*:*:*:*:* :*****.*.* * : ********** *

HINLHPELDGQYVVEFDLFGKDSIRYYNKVPVEKRVFKNLQLFMEN-KQPEDDLFDRLN
HVTLKPPN-----TVIFDFLFGKDSIRFYQEVEVDKQVFKNLTIKFRPPKQPGHQLFDRLD
.::* * .* ******:*: * :*:***** :* . *** :*****:

TGILNKHLDLMEGLTAKVFR TYNASITLQOQLKELTAPDENIPAKILSYNRANRAVAIL
PSILNKYLQNYMPGLTAKVFR TYNASKTMQDQLDIPN-KGSVAEKILKYNAANRTVAIL
.*****:*: * ********** *:*:*. :. . :.***.* ***:*****

CNHQRA
CNHQRT

B. Catalytic domain (62% identity; 85% homology)

KQIALGTSKLN YLDPRI TVAWCKKWGVPIEKIYNKTQREKFAWAIDMADEDYEF
SQVSLGTSKIN YIDPRLSVVFCKKYDVPIEKIFTKTLREKFKWAIESVDENWRF
.::*****:*:*:*:*:*. :*:*:.*.*****:.* ** * ** * :.*:.*:.*