#### **Supplementary Information**

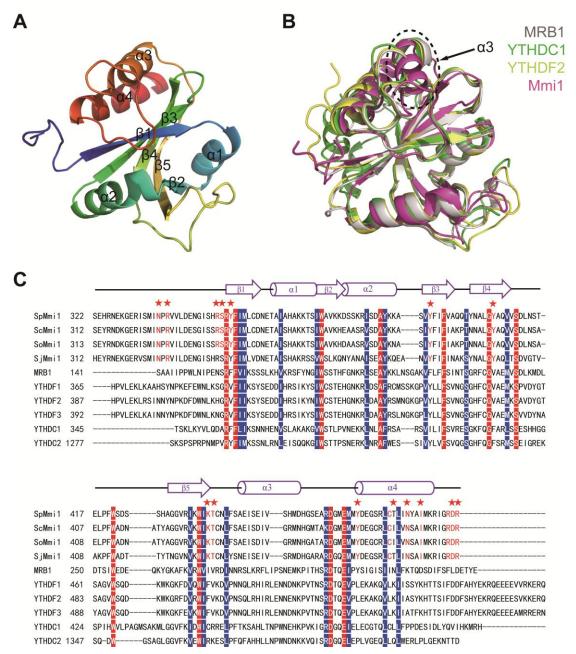
## A novel RNA-binding model of the YTH domain reveals the mechanism for recognition of determinant of selective removal by Mmi1

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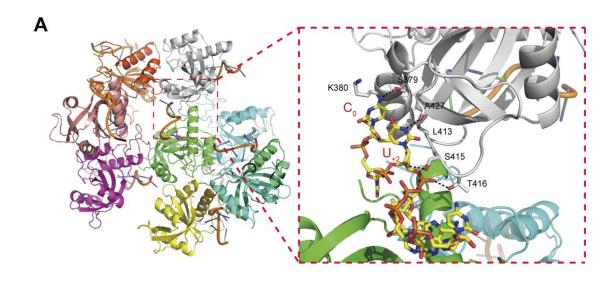
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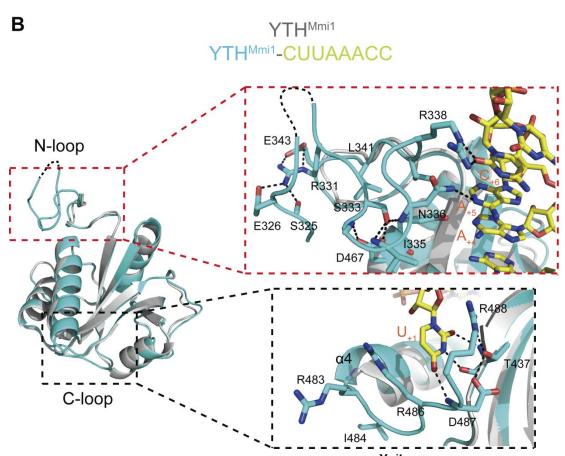
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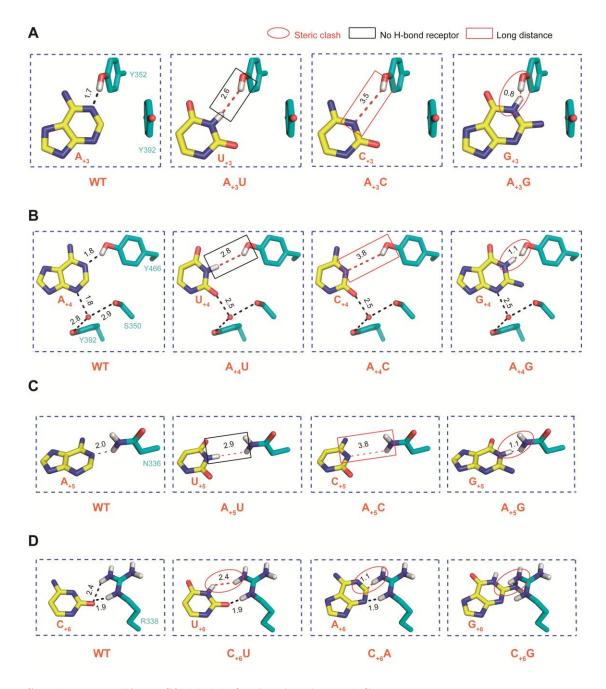


Supplementary Figure S1: Overall structure of Mmil YTH domain. (A) Structure of apo YTH<sup>Mmil</sup> is shown in cartoon. (B) Comparison of the YTH domains of YTHDC1, YTHDF2, MRB1 and Mmil. YTH domains of human YTHDC1 (PDB: 4R3H), human YTHDF2 (PDB: 4RDO) and MRB1 (PDB: 4RCM) were aligned to apo structure of the Mmil YTH domain. The significant structural difference is indicated by arrow. (C) Sequence alignment of YTH domains of human YTHDC1, human YTHDF2, *Saccharomyces cerevisiae* MRB1 and Mmil homologues in fission yeast (*Schizosaccharomyces pombe, S. japonicus, S. octosporus*, and *S. cryophilus*). Conserved and Similar residues are highlighted in blue and red, respectively. The Mmil residues binding RNA are indicated by red asterisks, which are strictly conserved in fission homologues (colored in red). The secondary structure elements of YTH<sup>Mmil</sup> is shown at the top.

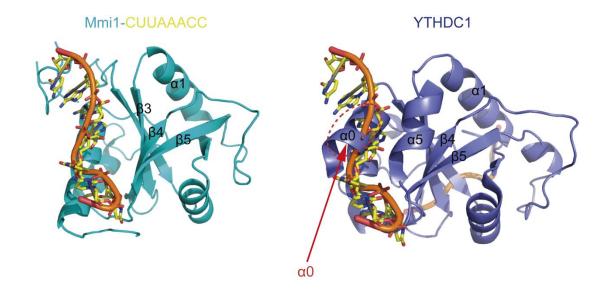


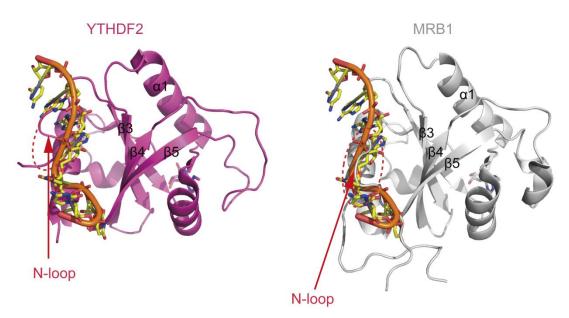


Supplementary Figure S2: The crystallization packing of YTH $^{\text{Mmi1}}$ -RNA complex and comparison of apo (grey) and RNA-bound (cyan) structures of YTH $^{\text{Mmi1}}$ . (A)  $C_0$  and  $U_{+2}$  participate in crystallization packing via their contacts with another YTH $^{\text{Mmi1}}$  molecule. Residues involved in crystallization packing are shown in stick and labled. Black dashed lines indicate the hydrogen bonds. (B) Residues with substantial conformational changes are shown and labeled. Hydrogen bonds are indicated by black dashed lines. Residues 326-329 of YTH $^{\text{Mmi1}}$  in the RNA-bound structure are also invisible in the density map and indicated with the black dash line.

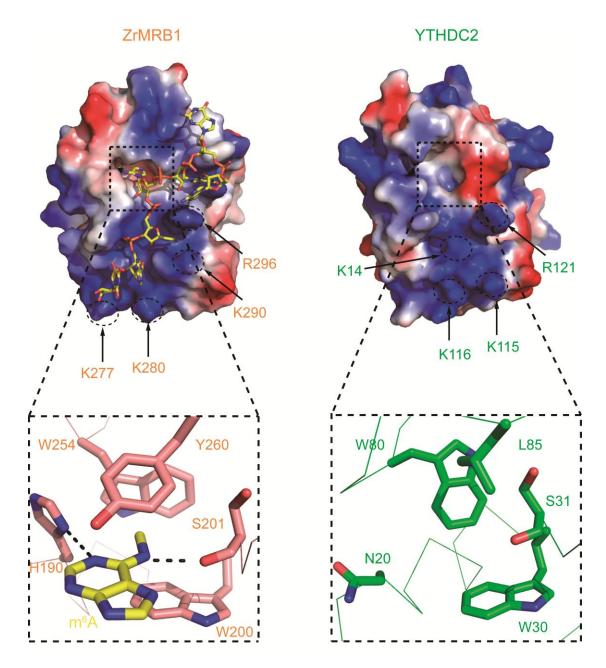


Supplementary Figure S3: Models for  $A_{+3}$ ,  $A_{+4}$ ,  $A_{+5}$ , and  $C_{+6}$  mutants. (A-D) The polar hydrogen atoms in the binding interface are shown in grey sticks. Black dashed lines indicate the hydrogen bonds. Red dashed lines indicate the distances between atoms without hydrogen bonding interactions. The steric clash is highlighted with red ovals, the loss of hydrogen bonds is highlighted within black or red rectangles.

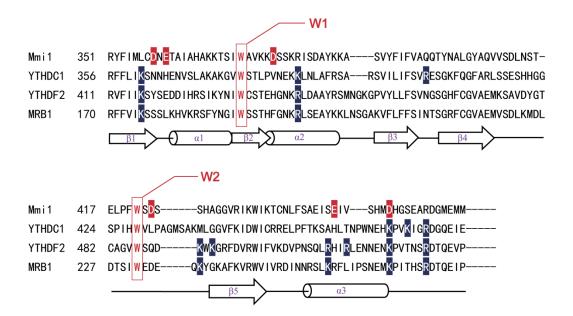




**Supplementary Figure S4: The N-terminal segments occupy the potential DSR-binding grooves in YTHDC1, YTHDF2 and MRB1.** The YTH<sup>YTHDC1</sup>-GGm<sup>6</sup>ACU complex (PDB: 4R3I), YTH<sup>YTHDF2</sup>-m<sup>6</sup>A complex (PDB: 4RDN) and YTH<sup>MRB1</sup>-m<sup>6</sup>A complex (PDB: 4RCM) are aligned to YTH<sup>Mmi1</sup>-CUUAAACC complex. The potential steric clashes are highlighted with red dashed ovals.



**Supplementary Figure S5: The aromatic cages and the surrounding grooves of YTH domains of ZrMRB1 and YTHDC2.** YTH<sup>ZrMRB1</sup> complex (PDB: 4U8T) and YTH<sup>YTHDC2</sup> (PDB: 2YU6) were aligned to YTH<sup>mmi1</sup>, as in Figure 4. The upper pictures are the electrostatic potential surface, in which positive charged residues in the m<sup>6</sup>A RNA-binding interfaces of ZrMRB1 and YTHDC2 are indicated. The lower pictures are enlarged views of the aromatic cages.



**Supplementary Figure S6: Sequence alignment of the YTH domains.** The positively and negatively charged residues in Figure 8A are highlighted in blue and red, respectively. A schematic representation of the secondary structure elements of YTH<sup>Mmil</sup> is shown below the sequences.

### Supplementary Table S1. The thermodynamic parameters of the ITC experiments.

D	RNA	ΔΗ	ΔS	$K_D$	N
Proteins	5'-3'	kcal/mol	cal/mol/K	μΜ	N
Mmi1 <sup>316-488</sup>	CC <u>UUAAAC</u> CU	-38.6	-100	0.39 ±0.01	1.1
Mmi1 <sup>338-488</sup>	CC <u>UUAAAC</u> CU			>30	
Mmi1 <sup>345-488</sup>	CC <u>UUAAAC</u> CU			ND	
	CC <u>UUAAAC</u> CU	-33.6	-83.6	0.44±0.03	1.0
	CC <u>UUAAAC</u> C	-32.2	-80.7	$1.1\pm0.02$	1.1
	C <u>UUAAAC</u> CU	-32.4	-81.5	1.2±0.03	1.1
	C <u>UUAAAC</u> C	-31.4	-79.1	$1.8 \pm 0.1$	1.0
	<u>UUAAAC</u>	-14.3	-24.5	$8.3\pm 2$	0.88
	CC <u>UUm<sup>6</sup>AAAC</u> CU	-28.3	-70.7	$5.4\pm0.3$	1.0
	CC <u>UUA<b>m</b><sup>6</sup><b>A</b>AC</u> CU	-27.7	-67.6	$2.8\pm0.09$	0.94
	CC <u>UUAA<b>m</b><sup>6</sup><b>AC</b></u> CU	-29.3	-70.1	$0.69\pm0.02$	0.98
	CC <u>UG</u> AAACCU	-32.6	-81.3	$0.70\pm0.04$	0.88
	CC <u>UA</u> AAACCU	-30.5	-74.8	$0.95 \pm 0.09$	0.97
	CC <u>UC</u> AAACCU	-32.6	-81.0	$1.0\pm0.04$	0.97
	CC <u>G</u> UAAACCU	-28.2	-70.0	$3.9\pm0.2$	0.96
$\mathrm{YTH}^{\mathrm{Mmi1}}$	CC <u>AUAAAC</u> CU	-21.1	-46.1	4.4±0.3	0.85
	CC <u>C</u> UAAACCU	-14.5	-26.2	13±2	1.2
	CC <u>UU<b>G</b>AAC</u> CU			>30	
	CC <u>UUCAAC</u> CU	-23.0	-54.8	13±1	1.0
	CC <u>UU<i>U</i>AAC</u> CU	-22.2	-50.4	$5.6 \pm 0.4$	1.1
	CC <u>UUA<b>G</b>AC</u> CU			ND	
	CC <u>UUA<b>C</b>AC</u> CU			>30	
	CC <u>UUA<i>U</i>AC</u> CU			ND	
	CC <u>UUAA<b>G</b>C</u> CU			>20	
	CC <u>UUAA<i>C</i>C</u> CU	-23.6	-54.3	3.9±0.1	0.98
	CC <u>UUAA<i>U</i>C</u> CU	-26.7	-66.3	$6.0\pm0.4$	1.1
	CC <u>UUAAA<i>G</i></u> CU	-25.9	-62.2	$4.1 \pm 0.4$	1.1
	CC <u>UUAAAA</u> CU	-26.1	-61.3	$1.9\pm0.01$	1.1
	CC <u>UUAAA<i>U</i></u> CU	-30.9	-77.8	$2.1 \pm 0.1$	0.92
	AU <u>GGAC</u> UCC			ND	
R331A(YTH <sup>Mmi1</sup> )		-38.3	-102	1.9±0.05	0.98
S333A		-36.4	-98.4	$6.2\pm0.02$	1.3
N336A		-34.3	-91.8	$8.9\pm0.2$	1.1
R338A	CC <u>UUAAAC</u> CU	-24.3	-58.6	$9.6 \pm 0.7$	0.99
Y352F		-28.8	-75.1	19±3	0.97
K436A		-18.2	-39.8	21±3	0.84
Y466F				>100	
N477A		-31.6	-82.9	$8.7 \pm 0.7$	1.0

- VTHDC1				0.010	
YTH <sup>YTHDC1</sup>		-13.1	-11.1	0.068	1.2
				±0.01	
$YTH^{YTHDF2}$	AU <u>GGm<sup>6</sup>AC</u> UCC	-9.8	-2.51	$0.25 \pm 0.03$	1.0
$YTH^{MRB1}$		-20.3	-39.3	$0.56 \pm 0.02$	1.1
YTH <sup>Mmi1</sup>				ND	
YTH <sup>YTHDC1</sup>				ND	
$YTH^{YTHDF2}$	CC <u>UUAAAC</u> CU			ND	
YTH <sup>MRB1</sup>				ND	

Bold and italic fonts are used to highlight the mutant nucleotides and the m<sup>6</sup>A nucleotides in the 10-mer DSR RNAs.

### $\label{eq:Supplementary} \textbf{ Table S2. Data collection and refinement statistics.}$

	$\mathrm{YTH}^{\mathrm{Mmil}}$	YTH <sup>Mmil</sup> -CUUAAACC		
PDB ID	5DNP	5DNO		
<b>Data Collection</b>				
Space group	C2	P6 <sub>5</sub>		
Cell dimensions				
a, b, c (Å)	100.5, 58.3, 54.3	77.5, 77.5, 65.5,		
$\alpha, \beta, \gamma$ (°)	90,108,90	90,90,120		
Wavelength(Å)	0.9795	0.9795		
Resolution* (Å)	49.79-2.30	38.76-1.80		
	(2.34-2.30)	(1.86-1.80)		
Completeness (%)	99.3(96.7)	99.9(99.6)		
Redundancy	3.5(3.5)	11.0(10.9)		
$R_{\text{sym}}$ or $R_{\text{merge}}$ (%)	12.6(64.8)	17.0(59.5)		
Ι/σΙ	13.0(2.9)	12.9(4.6)		
Refinement				
No. reflections used/free	13290/995	20917/1084		
Resolution (Å)	49.79-2.30	38.76-1.80		
$R_{ m work}/R_{ m free}$	21.2/25.6	17.6/21.6		
R.m.s.deviations				
Bondslengths (Å)	0.008	0.008		
Bond angles (°)	1.2	1.3		
<i>B</i> -factors ( $\mathring{A}^2$ )				
Protein	30.9	13.4		
RNA	N/A	20.2		
Water	24.8	22.7		
No. atoms				
Protein	2195	1304		
RNA	N/A	143		
Water	57	116		
Ramachandran plot				
Favored/ allowed/ outlier(%)	98.0/2.0/0.0	100.0/0.0/0.0		

<sup>\*</sup> Values in parentheses are for highest-resolution shell.