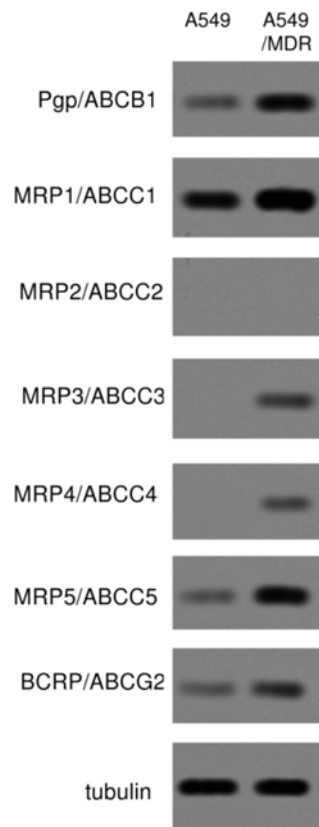
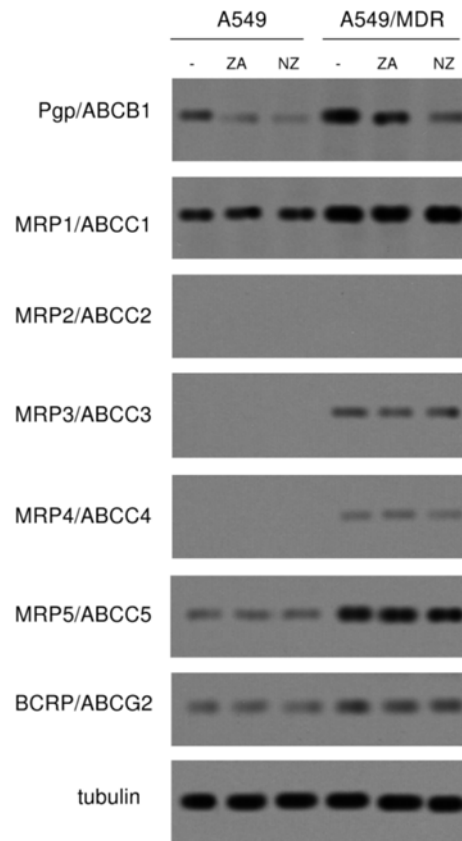


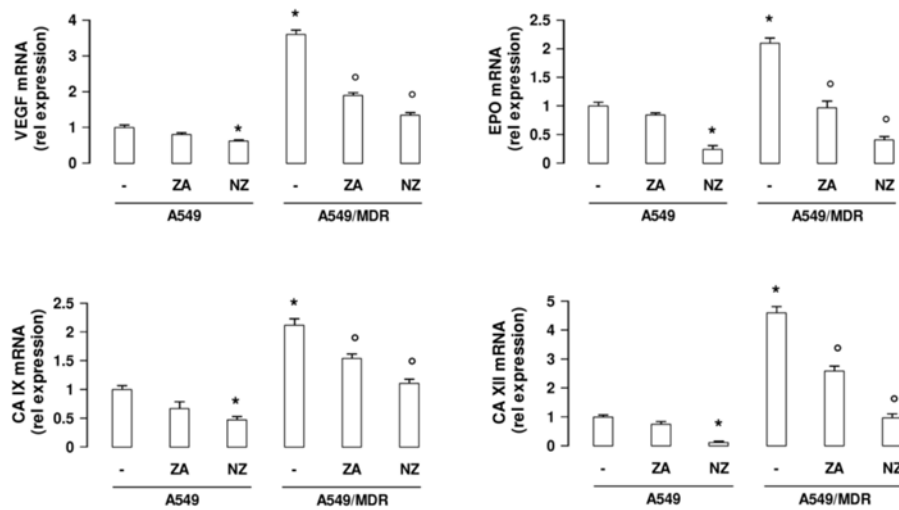
SUPPLEMENTARY FIGURES AND TABLES



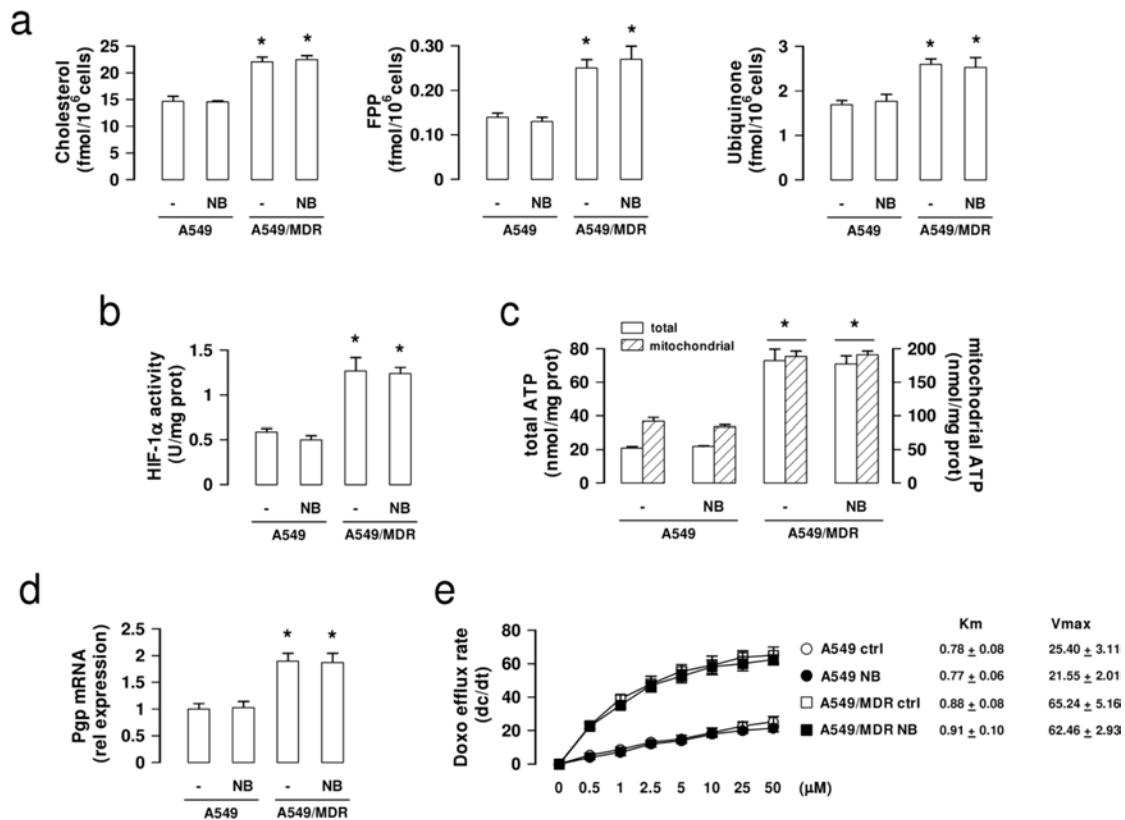
Supplementary Figure S1: Expression of ABC transporters in chemosensitive and MDR cancer cells. Chemosensitive human lung cancer A549 cells and their resistant counterpart A549/MDR cells were lysed and subjected to the Western blot analysis for Pgp/ABCB1, MRP1/ABCC1, MRP2/ABCC2, MRP3/ABCC3, MRP4/ABCC4, MRP5/ABCC5, BCRP/ABCG2. The β -tubulin expression was used as control of equal protein loading. The figure is representative of 3 experiments.



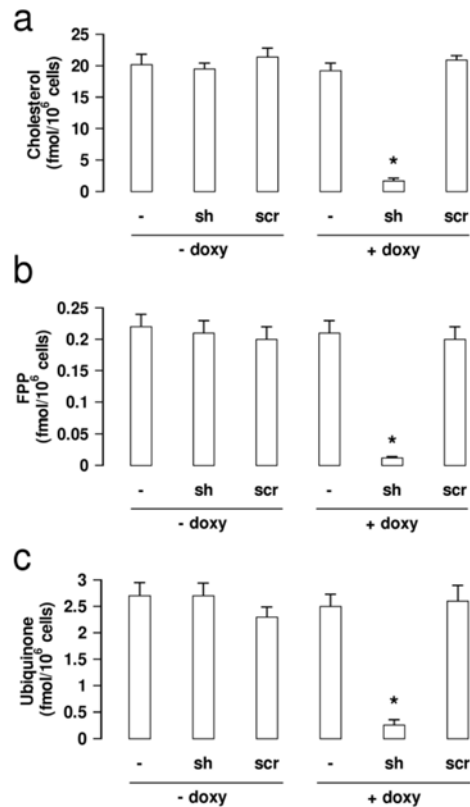
Supplementary Figure S2: Expression of ABC transporters after treatment with ZA and NZ in chemosensitive and MDR cancer cells. Chemosensitive human lung cancer A549 cells and their resistant counterpart A549/MDR cells were cultured for 48 h in fresh medium (-), in medium containing 1 μ M zoledronic acid (ZA) or 1 μ M self-assembling ZA formulation (NZ), then lysed and subjected to the Western blot analysis for Pgp/ABCB1, MRP1/ABCC1, MRP2/ABCC2, MRP3/ABCC3, MRP4/ABCC4, MRP5/ABCC5, BCRP/ABCG2. The β -tubulin expression was used as control of equal protein loading. The figure is representative of 3 experiments.



Supplementary Figure S3: Expression of HIF-1 α -target genes in chemosensitive and MDR cancer cells treated with ZA and NZ. Chemosensitive human lung cancer A549 cells and their resistant counterpart A549/MDR cells were cultured for 48 h in fresh medium (-), in medium containing 1 μ M zoledronic acid (ZA) or 1 μ M self-assembling ZA formulation (NZ). *VEGF*, *EPO*, *CA IX*, *CA XII* mRNA levels were detected in triplicate by qRT-PCR. Data are presented as means \pm SD ($n = 4$). For all panels, versus untreated A549 cells: * $p < 0.05$; versus untreated A549/MDR cells: ° $p < 0.02$.



Supplementary Figure S4: Effects of empty self assembling liposomes on chemosensitive and MDR cancer cells. Chemosensitive human lung cancer A549 cells and their resistant counterpart A549/MDR cells were cultured for 48 h in fresh medium (-) or in medium containing self-assembling formulation without zoledronic acid (blank NPs, NB) at the same final concentration used in the all the other experimental assays. **a.** Cells were radiolabelled during the last 24 h with [³H]-acetate, then the *de novo* synthesis of cholesterol, FPP and ubiquinone was measured. Data are presented as means \pm SD ($n = 3$). For all panels, A549/MDR cells versus A549 cells: $*p < 0.02$. **b.** HIF-1 α activity was measured in nuclear extracts by ELISA. Data are presented as means \pm SD ($n = 3$). A549/MDR cells versus A549 cells: $*p < 0.005$. **c.** ATP levels in whole cell (total) and isolated mitochondria (mitochondrial) were measured by a chemiluminescence-based assay. Data are presented as means \pm SD ($n = 3$). A549/MDR cells versus A549 cells: $*p < 0.001$. **d.** *Pgp* mRNA levels were detected in triplicate by qRT-PCR. Data are presented as means \pm SD ($n = 3$). A549/MDR cells versus A549 cells: $*p < 0.001$. **e.** Cells were grown for 48 h in fresh medium or in medium containing blank NPs (NB), then incubated for 20 min with increasing concentrations (0–50 μ M/L) of doxorubicin (dox). Cells were washed and tested for the intracellular drugs content. The procedure was repeated on a second series of dishes, incubated in the same experimental conditions and analyzed after 10 min. Data are presented as means \pm SD ($n = 3$). The rate of doxorubicin efflux (dc/dt) was plotted versus the initial concentration of the drug. Vmax (nmol/min/mg proteins) and Km (nmol/mg proteins) were calculated with the Enzfitter software.



Supplementary Figure S5: Effects of FPPS silencing on the synthesis of cholesterol, FPP and ubiquinone in chemoresistant cancer cells. Wild-type A549/MDR cells, cells treated with a TetON vector containing a shRNA targeting *FPPS* (sh) or with a non targeting shRNA vector (scr), were cultured 48 h in media without (- doxy) or with (+ doxy) 1 μ g/ml doxycycline. Cells were radiolabelled during the last 24 h with [³H]-acetate, then the *de novo* synthesis of cholesterol (panel a), FPP (panel b) or ubiquinone (panel c) was measured. Data are presented as means \pm SD ($n = 3$). For all panels, versus untreated (-, - doxy) cells: * $p < 0.001$.

Supplementary Table S1: IC50 (μM) of different cytotoxic drugs in HT29 and HT29/MDR cells

Drug	Transporter	HT29	HT29	HT29	HT29/MDR	HT29/MDR	HT29/MDR
		CTRL	ZA	NZ	CTRL	ZA	NZ
doxorubicin	Pgp, MRP1, MRP2, MRP3, BCRP	1.23 \pm 0.08	0.91 \pm 0.07	0.71 \pm 0.1 *	2.58 \pm 0.13 [°]	1.02 \pm 0.12 *	0.98 \pm 0.07*
vinblastine	Pgp, MRP1, MRP2	3.61 \pm 0.14	2.53 \pm 0.17	1.88 \pm 0.14 *	9.23 \pm 0.41 [°]	3.21 \pm 0.16 *	0.51 \pm 0.09*
etoposide	Pgp, MRP1, MRP2, MRP3	1.12 \pm 0.15	0.67 \pm 0.11 *	0.51 \pm 0.06 *	11.41 \pm 0.37 [°]	2.76 \pm 0.44 *	2.33 \pm 0.18*
irinotecan	Pgp, MRP1, MRP2	6.23 \pm 0.37	4.41 \pm 0.31	3.71 \pm 0.44 *	47.11 \pm 5.14 [°]	36.11 \pm 3.71	14.23 \pm 0.91*
cisplatin	MRP1, MRP2, MRP4	10.89 \pm 1.21	7.21 \pm 0.88	6.01 \pm 0.13 *	52.14 \pm 2.47 [°]	5.88 \pm 0.42 *	2.24 \pm 0.55*
oxaliplatin	MRP1, MRP4	5.87 \pm 0.61	2.43 \pm 0.52 *	0.91 \pm 0.15 *	12.21 \pm 0.18 [°]	5.21 \pm 0.23 *	0.81 \pm 0.16*
5-fluorouracile	MRP1, MRP3, MRP4, MRP5	0.83 \pm 0.11	0.61 \pm 0.14	0.65 \pm 0.13	7.53 \pm 0.71 [°]	5.44 \pm 0.23	3.12 \pm 0.14*
methotrexate	MRP4, Pgp, MRP1, MRP2, MRP3, BCRP	2.34 \pm 0.31	1.01 \pm 0.18 *	0.21 \pm 0.09 *	8.79 \pm 0.71 [°]	1.77 \pm 0.45 *	0.46 \pm 0.22*
pemetrexed	MRP5	0.74 \pm 0.01	0.65 \pm 0.17	0.12 \pm 0.07 *	8.26 \pm 0.57 [°]	5.78 \pm 0.56	1.33 \pm 0.27*
gemcitabine	MRP5	0.11 \pm 0.03	0.11 \pm 0.09	0.06 \pm 0.01 *	0.75 \pm 0.08 [°]	0.21 \pm 0.09 *	0.05 \pm 0.02*
mitoxantrone	BCRP, Pgp, MRP1	5.88 \pm 0.41	2.21 \pm 0.17 *	2.03 \pm 0.21 *	9.67 \pm 0.41 [°]	4.53 \pm 0.37 *	2.81 \pm 0.41*

Untreated (CTRL) HT29 and HT29/MDR cells, cells treated with ZA or NZ (1 μM), were incubated for 72 h with increasing concentrations of cytotoxic drugs, then stained in quadruplicate with neutral red ($n = 3$). Versus respective CTRL: * $p < 0.05$; HT29/MDR versus HT29 cells: [°] $p < 0.001$.

Supplementary Table S2: Hematochemical parameters of animals

		LDH	AST	ALT	AP	CPK	creatinine
		(U/l)	(U/l)	(U/l)	(U/l)	(U/l)	(mg/l)
A549	Ctrl	6342 ± 1567	267 ± 56	39 ± 12	75 ± 16	434 ± 68	0.05 ± 0.01
	NZ	6872 ± 1812	293 ± 24	46 ± 11	87 ± 24	511 ± 76	0.06 ± 0.02
	dox	7862 ± 2089	254 ± 34	41 ± 13	81 ± 23	897 ± 71*	0.05 ± 0.02
	NZ+dox	6723 ± 1629	309 ± 31	50 ± 22	73 ± 27	911 ± 56*	0.06 ± 0.03
	Pt	6341 ± 1098	311 ± 24	45 ± 17	76 ± 23	467 ± 71	0.09 ± 0.01*
	NZ+Pt	6009 ± 1987	298 ± 66	49 ± 17	75 ± 11	509 ± 87	0.10 ± 0.02*
A549/MDR	Ctrl	7098 ± 1803	288 ± 24	47 ± 18	78 ± 26	454 ± 91	0.04 ± 0.02
	NZ	7612 ± 2137	291 ± 34	51 ± 16	87 ± 21	459 ± 37	0.06 ± 0.02
	dox	6534 ± 1271	254 ± 71	45 ± 16	80 ± 14	434 ± 68	0.04 ± 0.01
	NZ+dox	6093 ± 1234	312 ± 45	55 ± 23	80 ± 20	987 ± 99 *	0.05 ± 0.04
	Pt	6092 ± 1261	324 ± 71	56 ± 25	77 ± 23	467 ± 93	0.08 ± 0.01*
	NZ+Pt	6873 ± 1093	321 ± 81	47 ± 16	71 ± 34	489 ± 88	0.09 ± 0.02*

Six weeks old female BALB/c mice bearing a 100 mm³-tumor of A549 or A549/MDR cells were randomly divided in the following groups (5 mice/group) and treated with saline solution (Ctrl), NZ, doxorubicin (dox), NZ + doxorubicin (NZ+dox), carboplatin (Pt), NZ + carboplatin (NZ + Pt), as detailed under Materials and Methods. The experiment was repeated 2 times. The animals were sacrificed at day 21 after randomization. Blood was collected immediately after mice euthanasia. Versus Ctrl group: * $p < 0.02$.

Supplementary Table S3: Primers sequence for qRT-PCR

Gene	Forward primer	Reverse primer
<i>GLUT1</i>	CCTGCAGTTTGGCTACAACA	TAACGAAAAGGCCACAGAG
<i>HK</i>	AGACGCACCCACAGTATTCC	CGCATCCTCTTCTCACCTC
<i>PFK1</i>	GGAGCTTCGAGAACAACCTGG	CTGTGTGTCCATGGGAGATG
<i>ALDO-A</i>	GCTATGGCCTTTTCCTTTCC	ATGCTCCCAGTGGACTCATC
<i>GAPDH</i>	GAAGGTGAAGGTCGGAGT	CATGGTGAATCATATTGGAA
<i>PGK</i>	TTCATGGATGAGGTGGTGA	CTTCCAGGAGCTCCAACTG
<i>ENO-A</i>	GCTCCGGGACAATGATAAGA	TCCATCCATCTCGATCATCA
<i>PK</i>	TGCAGTGGAGCTCAGAGAGA	GCTTCCGGTGACATAATGCT
<i>LDH</i>	TGGGAGTTCACCCATTAAGC	AGCACTCTCAACCACCTGCT
<i>VEGF</i>	ATCTTCAAGCCATCCTGTGTGC	GCTCACCGCCTCGGCTTGT
<i>EPO</i>	CAGACTTCTACGGCCTGCTG	GCTGAACACTGCAGCTTGAA
<i>CA IX</i>	GTCTCGCTTGGAAAGAAATCG	AGAGGGTGTGGAGCTGCTTA
<i>CA XII</i>	ACTGAGTCTCTGGGCATCATCC	AAAAGCCAAATGGACACCAC
<i>Pgp</i>	TGCTGGAGCGGTTCTACG	ATAGGCAATGTTCTCAGCAATG
<i>S14</i>	GGTGCAAGGAGCTGGGTAT	TCCAGGGGTCTTGGTCCTATTT