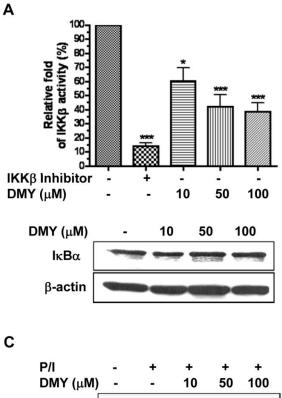
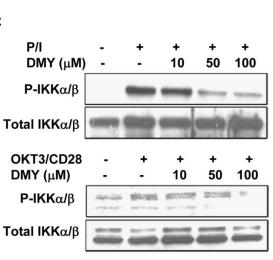
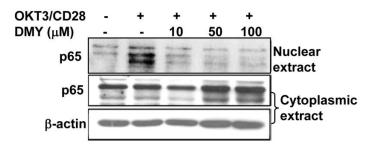
## Mutation of Cysteine 46 in IKK-beta increases inflammatory responses

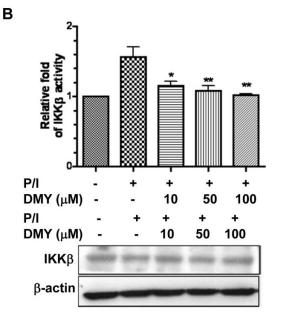
**Supplementary Material** 



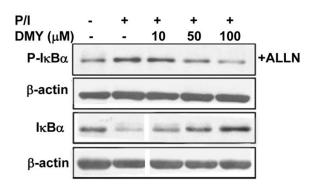




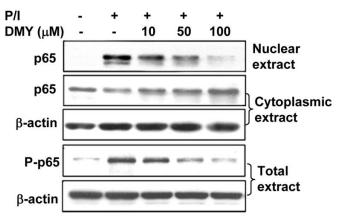




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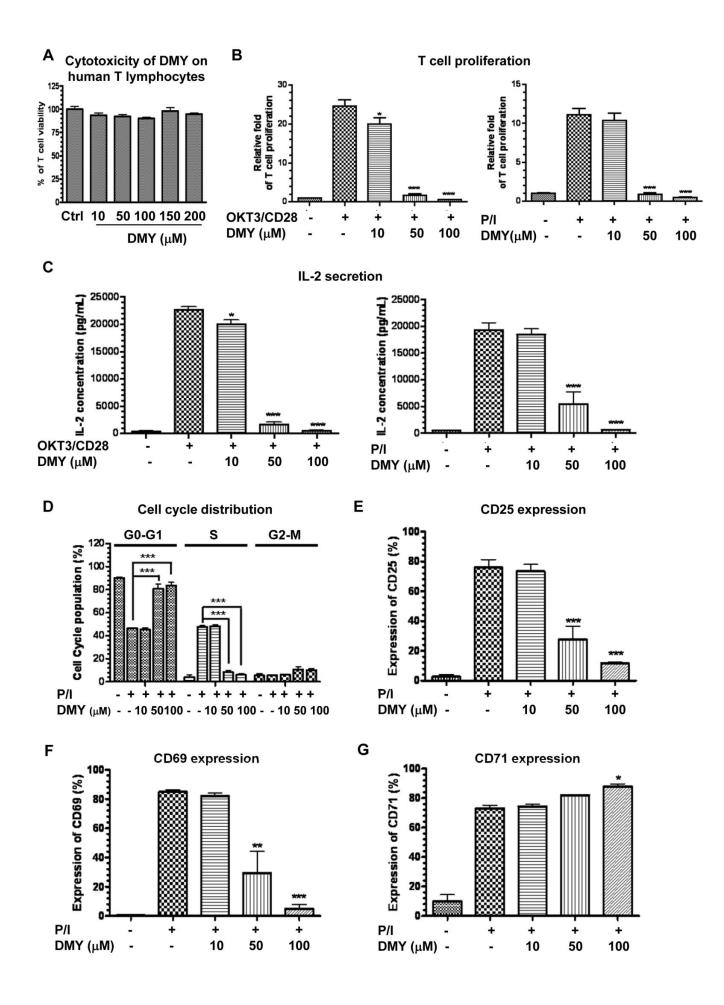


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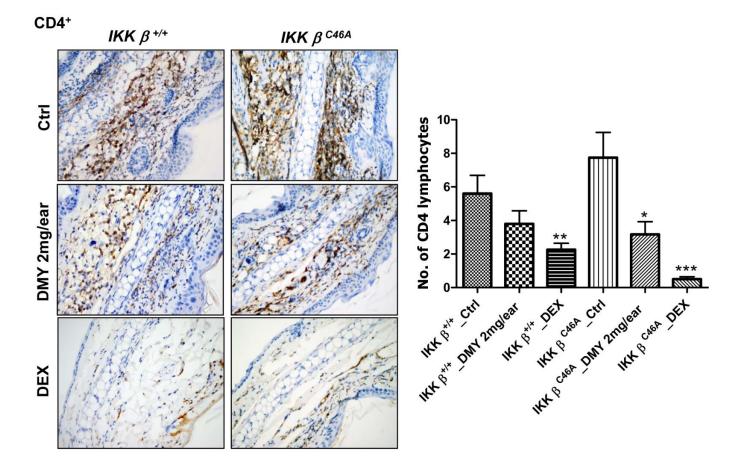
## Fig. S1. Suppression of IKK-β–NF-κB signalling by DMY.

(A) Upper panel: DMY directly inhibited IKK-β activity. His-tagged human recombinant IKK- $\beta$  (2 ng) was incubated with GST-I $\kappa$ B $\alpha$  substrate in the presence of the indicated concentrations of DMY and 10 mM ATP. IKK- $\beta$  inhibitor was the positive control provided in the screening kit. Lower panel: the expression of  $I\kappa B\alpha$ in response to DMY treatment. (B) Upper panel: DMY suppressed PMA/ionomycin (P/I)-induced IKK- $\beta$  activity in the isolated human T lymphocytes. Lower panel: the expression of IKK-B in response to DMY treatment. The isolated human T lymphocytes were treated with the indicated concentrations of DMY and then stimulated with 20 ng/ml PMA plus 1 µM ionomycin (P/I). Whole-cell extracts were harvested for IKK- $\beta$  kinase assay or Western blotting. (C) DMY suppressed P/I- or OKT-3/CD28-mediated phosphorylation of IKK- $\alpha/\beta$ . (D) DMY prevented P/I-induced phosphorylation and degradation of I $\kappa$ B $\alpha$ . (E & F) DMY abrogated the OKT-3/CD28- or P/I-mediated phosphorylation and nuclear translocation of NF-κB p65. Isolated human T lymphocytes were pretreated with the indicated concentrations of DMY, followed by co-stimulation by P/I, or the cells were treated with DMY with or without OKT-3/CD28 antibodies. For detection of phosphorylation of I $\kappa$ B $\alpha$ , the cells were first incubated with DMY plus 100  $\mu$ M ALLN prior to P/I stimulation. Data represented the mean  $\pm$  SEM of three independent experiments. \* P<0.05, \*\*P<0.01, \*\*\*P<0.001, compared with P/I treatment alone.



## Fig. S2. DMY inhibited IKK-β–NF-κB signalling and in turn suppressed human T cell activation and T cell surface marker expression.

(A) Cytotoxicity of DMY on T lymphocytes. (B) DMY inhibited OKT-3/CD28- or P/I-induced human T cell proliferation. (C) DMY inhibited OKT-3/CD28- or P/Iinduced IL-2 cytokine secretion. The isolated human T lymphocytes were costimulated with OKT-3/CD28 or P/I in the presence of the indicated concentrations of DMY. T cell proliferation was then measured by BrdU proliferation assay. For IL-2 cytokine measurement, the cell-free culture supernatants were harvested for IL-2 ELISA. (D) DMY arrested P/I-mediated cell cycle progression in isolated human T cells. Human T lymphocytes pretreated with the indicated concentrations of DMY and stimulated with or without P/I for 72 h. The cells were then fixed and stained with propidium iodide for cell cycle analysis. (E) DMY suppressed the P/I-mediated expression of the IL-2 receptor, CD25. (F) DMY suppressed P/I-mediated expression of the early-stage T cell activation marker CD69. (G) DMY slightly increased P/Imediated expression of the late-stage T cell activation marker CD71. The isolated human T cells pretreated with the indicated concentrations of DMY were costimulated with P/I. The cells were then stained with specific antibodies in the combination of CD3-PE with CD25-FITC or CD69-FITC or CD71-FITC for 30 min. The cell surface expression of CD25, CD69 and CD71 antigens were then analysed by flow cytometry. Data in each column represented the mean  $\pm$  SEM of three independent experiments. \*P<0.05, \*\*P<0.01, \*\*\*P<0.001.



**Fig. S3.** DMY treatment decreased the population of CD4<sup>+</sup> T lymphocytes in the ear tissues of DTH-*IKK*- $\beta^{+/+}$  and -*IKK*- $\beta^{C46A}$  mice. The CD4<sup>+</sup> T cells were quantitated by immunohistochemical analysis of the ear sections from DTH mice. Bar charts represented the average number of CD4<sup>+</sup> T lymphocytes found in the ear sections of both wild-type and mutant DTH animals.