

## DNA-mediated adjuvant immunotherapy extends survival in two different mouse models of myeloid malignancies

### Supplementary Material

#### Table S1. Statistical analysis of APL survival curve Figure 1B

Log-rank (Mantel-Cox) test was used to compare the percent survival of different groups. Prism software was used for the Mantel-Cox (log-rank) analysis

	log-rank (Mantel-Cox)	
	Chi-2 value	p value
ATRA+pVAX14 vs Placebo	70.96	P<0.0001
ATRA+pVAX14 vs ATRA	10.14	P<0.0014
ATRA+pVAX14 vs PVAX14	58.51	P<0.0001
ATRA+pVAX14 vs ATRA+PML-RARA	0.56	P<0.46
ATRA+PML-RARA vs Placebo	55.45	P<0.0001
ATRA+PML-RARA vs ATRA	15.52	P<0.0001
ATRA+PML-RARA vs pVAX14	45.16	P<0.0001
pVAX14 vs Placebo	4.9	P<0.03
pVAX14 vs ATRA	35.88	P<0.0001
ATRA vs Placebo	46.07	P<0.0001

**Table S2. Primer sequences for open reading frames (ORFs), *MyD88*, *PML-RARA* and *ABL***

<b>Primer</b>	<b>Sequence (5'→3')</b>	<b>Amplicon bp</b>
<b>ORF1-F</b>	<b>ATGCGATGTTTCGCTTGGTGGTCGA</b>	<b>108</b>
<b>ORF1-R</b>	<b>TCACCTTGCTCCTGCCGAGAAAGTA</b>	
<b>ORF2-F</b>	<b>ATGTTTCGCTTGGTGGTCGAATGGG</b>	<b>108</b>
<b>ORF2-R</b>	<b>TCATCTCACCTTGCTCCTGCCGAGA</b>	
<b>ORF3-F</b>	<b>ATGACAGGAGATCCTGCCCC</b>	<b>63</b>
<b>ORF3-R</b>	<b>TCACTGAAGCGGGAAGGGAC</b>	
<b>ORF5-F</b>	<b>ATGCCTGCTATTGTCTTCCCAATCC</b>	<b>180</b>
<b>ORF5-R</b>	<b>CTATTGTCTTCCCAATCCTCCCCCG</b>	
<b>ORF6-F</b>	<b>ATGCAATTCCTCATTATTATTAGGA</b>	<b>189</b>
<b>ORF6-R</b>	<b>TTACCAGAGGGCGCCCCAGCTGGCA</b>	
<b>hABL-F</b>	<b>TTCAGCGGCCAGTAGCATCTGACTT</b>	<b>263</b>
<b>hABL-R</b>	<b>GATGTAGTTGCTTGGGACCCA</b>	
<b>mAbl-F</b>	<b>GAAGACCTTGAAGGAGGACACCATG</b>	<b>183</b>
<b>mAbl-R</b>	<b>GGGTACACACCCTAGCAGCT</b>	
<b>PMLRARA-F</b>	<b>GTCTTCCTGCCCAACAGCAACC</b>	<b>190</b>
<b>PMLRARA-R</b>	<b>CTCACAGGCGCTGACCCCATAGT</b>	
<b>MyD88-F</b>	<b>CGCGCATCGAGGAGGACTGC</b>	<b>156</b>
<b>MyD88-R</b>	<b>CCGGCGTTTGTCTAGGGGGT</b>	

**Table S3. Peptide sequences predicted by open reading frames (ORFs) of pVAX14**

1, peptide 1 (ORF 1); 2, peptide 2 (ORF 2); 3, peptide 3 (ORF 3); 4, peptide 4 (ORF 4); 5, peptides 5.2, 5.3 (ORF 5); peptides 6.1, 6.2 6.3 (ORF 6). As the predicted peptides of ORFs 5 and 6 were too large to synthesize 3 overlapping peptides were designed and synthesized for each of these ORFs; overlapping sequences are shown in bold

ORF	Peptide N°	location	Amino Acid Sequence
1	1	787	MRCFAWWSNGQVAGSSVCSRRIASAMMDTFSAGAR
2	2	792	MFRLVVEWAGSRIKRMQPPHCISHDGYFLGRSKVR
3	3	896	MTGDPAPALRPIAASPFPLQ
4	4	1207	MRNDPHPVS
5	5-1	1406	<b>MPAIVFPILPLAVLPHPTPQNRMTPTQT</b>
	5-2		<b>PTPQNRMTPTQTMRCNFLILLGKDSGSG</b>
	5-3		<b>NFLILLGKDSGSGTFQGQGRHGGGLGRQ</b>
6	6-1	1495	<b>MQFPHFIRKGQWEWHLPGSRKARGRIGK</b>
	6-2		<b>GSRKARGRIGKTIAGMLGMRWALWLLL</b>
	6-3		<b>GMRWALWLLLGGFMDSKRTGIASWGALW</b>

**A**

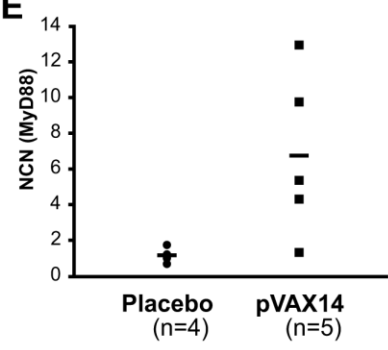
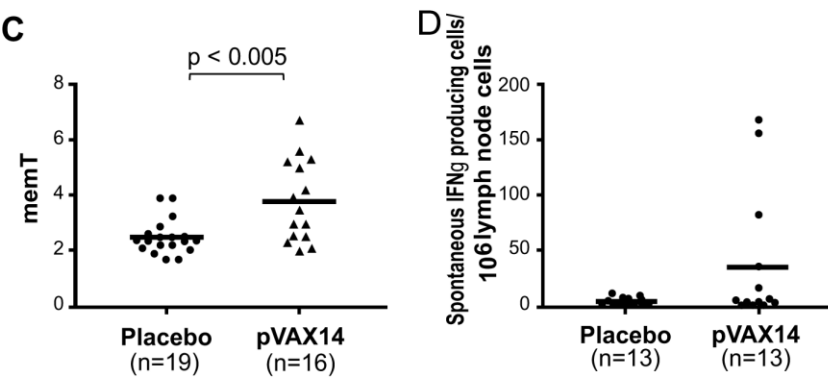
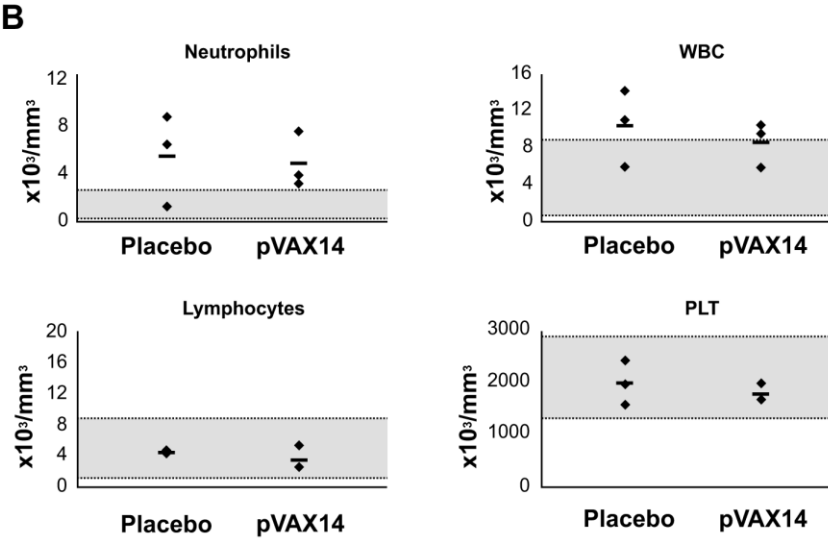
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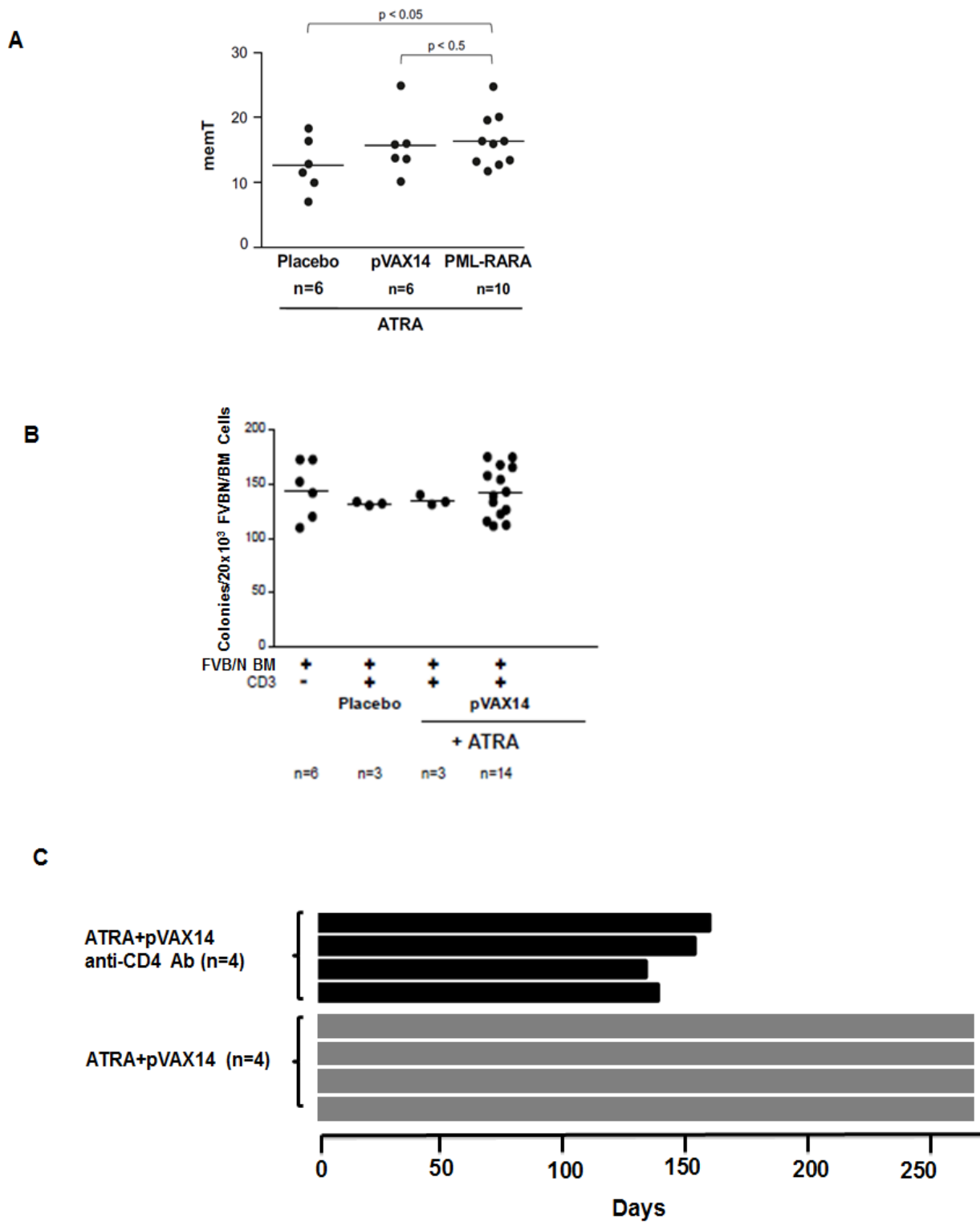
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pVax1  
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ORF 2  
ORF 3  
Insert  
ORF 4  
ORF 5  
ORF 6  
Kan<sup>r</sup>  
pVax1



## Supplementary Figure S1. Characteristics of pVAX14

A) Sequence of pVAX14 plasmid showing functional domains; grey highlighted sequences from the kanamycin (Kan) resistance gene in the antisense orientation and reversed (Kan insert originally in pVAX1 nucleotides 1226-1701), - in italics and underlined in dashed, the GC-rich sequences, - in white, the mouse and human specific consensus CpG sequences, the open reading frames (ORFs) are underlined with the start codons in bold italics, the pVAX1 sequences in the insert were originally pVAX1 nucleotide nos. 873-1225, the BamHI site (GGATC) in the multiple cloning site just precedes the insert. The entire insert of pVAX1 nucleotides 873-1701 was inverted (the antisense strand was reversed and now the coding strand) and inserted between pVAX1 nucleotide position 734 and 998; B) peripheral blood (PB) counts of normal FVB/N mice immunized with control placebo phosphate buffered saline (PBS) or pVAX14 DNA (50 µg) on day 35 of protocol illustrated on Figure 3C. The normal range is delineated in grey; C) PB memory T-cells (memT) (defined as percentage (%) of CD44<sup>hi</sup>/CD62L<sup>lo</sup> within the CD4<sup>+</sup> population) were measured on day 40. Controls were mice treated with placebo hepes buffered saline (HBSS); significant differences were observed between pVAX14 and the control ( $p < 0.005$ ); D) interferon gamma (IFN $\gamma$ ) producing cells in normal mice treated with control placebo (HBSS) and pVAX14. ELISPOT assays were undertaken on day 40 showing no significant difference between pVAX14 and placebo; E) *MyD88* expression of pVAX14 was similar to control placebo (HBSS). Nonparametric, unpaired, two tailed, student t-test was used to compare different groups. Prism software was used for the t-test analysis.



**Figure S2 DNA-based immunotherapy induced responses**

**A)** Increase in memory T-cells (memT) of pVAX14 or *PML-RARA* -treated APL mice measured on day 40; **B)** CD3<sup>+</sup> effectors from APL mice treated as shown showing no effect on FVB/N progenitors plated at an effector:target (E:T) of 10:1; **C)** The protective effect mediated by ATRA+ pVAX14 in APL mice is CD4<sup>+</sup> T-cell dependent. Long term survivors (LTS) from ATRA+pVAX14-treated mice were CD4-depleted by weekly injections of CD4-specific antibody started on day 120 after engraftment of leukemic cells. Control ATRA+pVAX14 treated mice were followed up for >500 days. Nonparametric, unpaired, two tailed, Mann-Whitney test was used to compare different groups. Prism software was used for the t-test analysis.