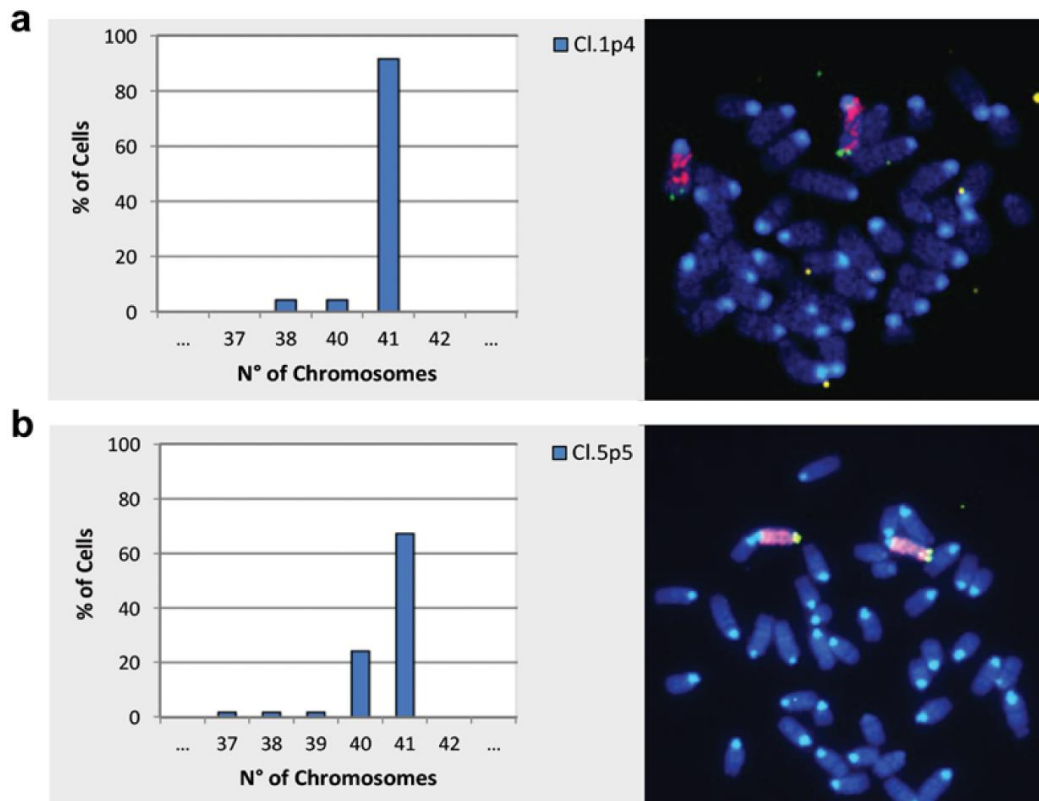
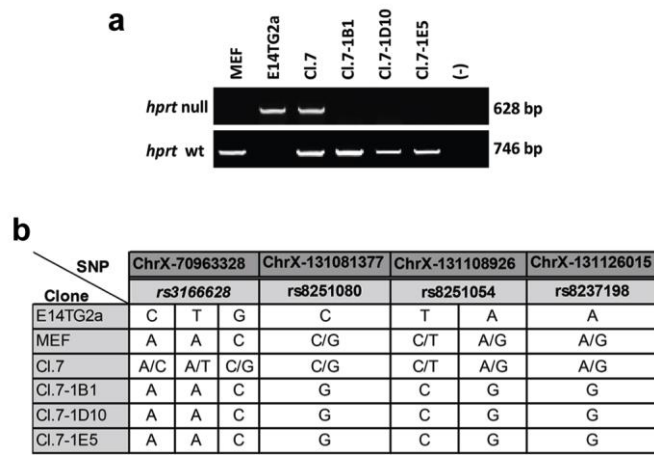


## Chromosome transplantation as a novel approach for correcting complex genomic disorders

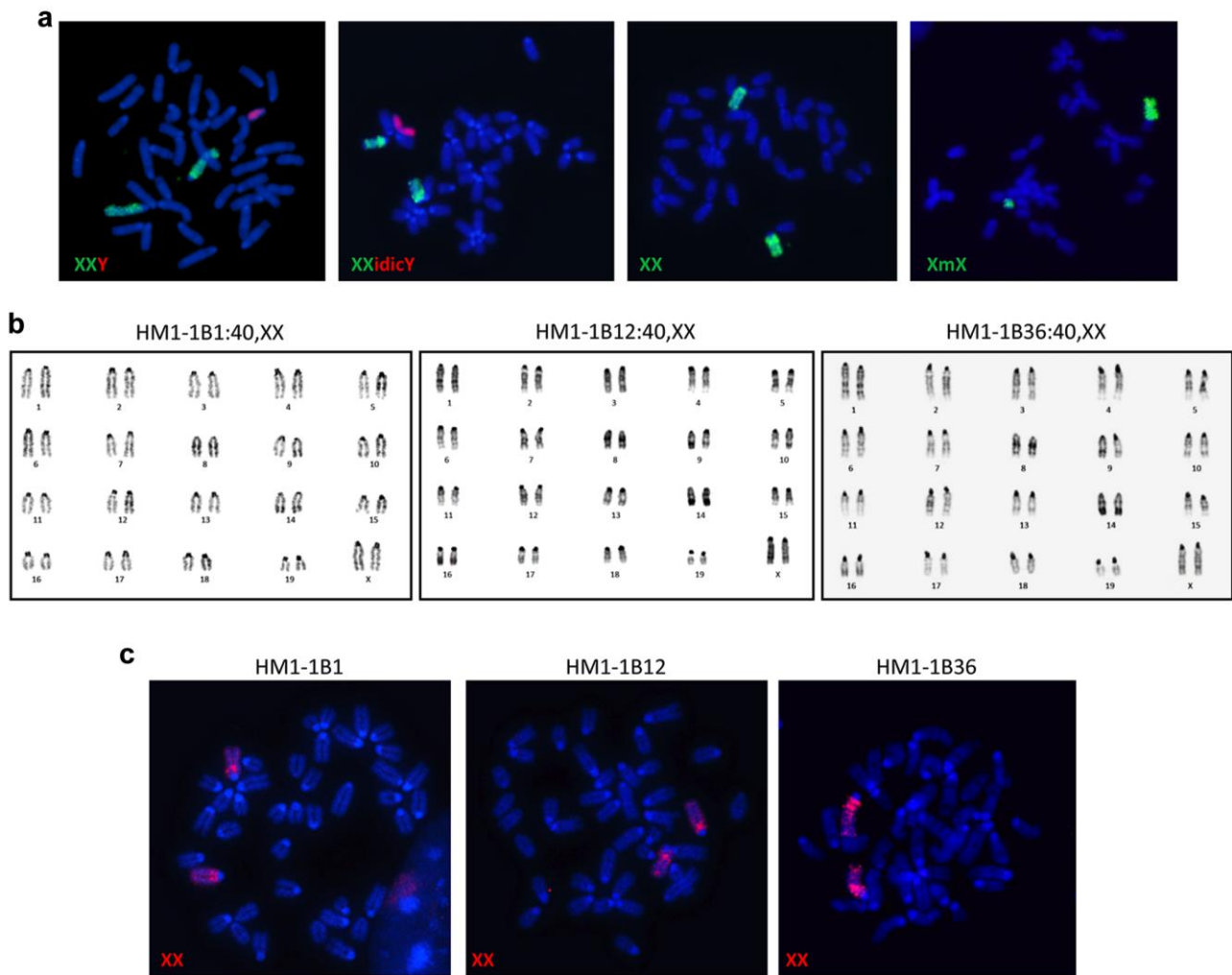
### Supplementary Material



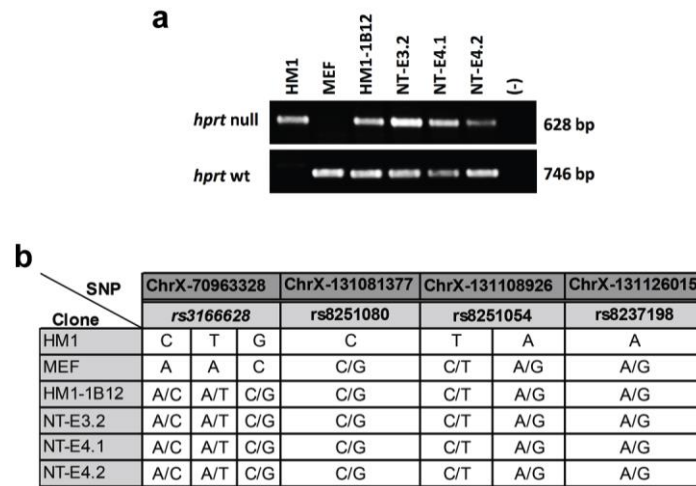
**Supplemental Figure 1: Cytogenetic analysis of HAT-resistant E14TG2a-derived clones after MMCT. (a,b)** Left: Chromosome number distributions; right: FISH metaphase spreads of the two clones. Whole mouse X chromosome painting (red) and X-linked BAC-RP23-113K2 (green) were used as probes. (n=50 each). All passage numbers are indicated with p.



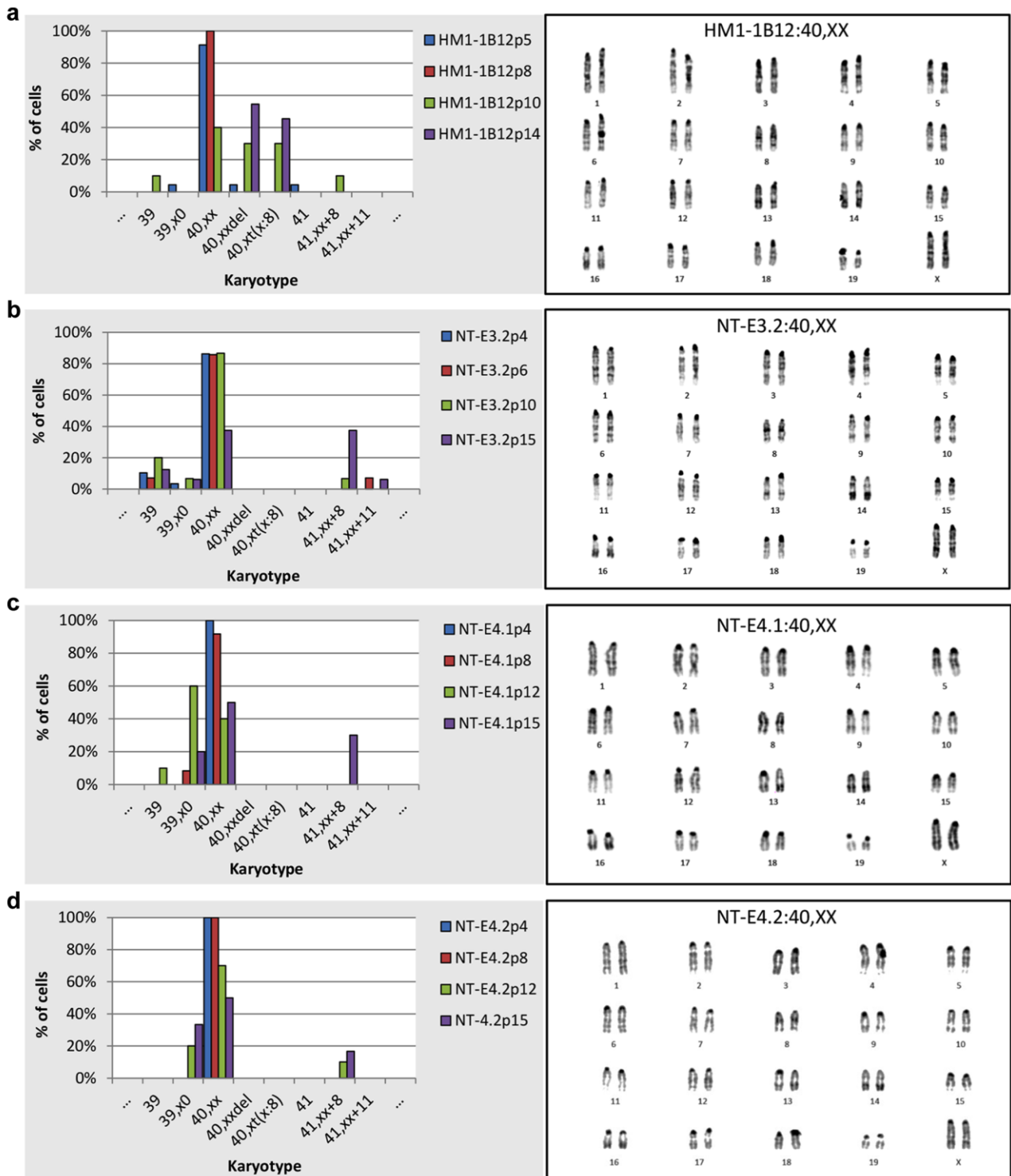
**Supplemental Figure 2: Molecular analysis of the tXY subclones.** (a) Genomic PCR of *Hprt* gene on the indicated clones. E14TG2a is the positive control of the mutated (null) locus; MEF is the positive control of the wild type (wt) locus. (b) Analysis of SNPs mapped to the X chromosome performed on the indicated samples.



**Supplemental Figure 3: Cytogenetic analysis of the sXX subclones.** (a) Representative FISH metaphase spreads of the most frequent populations of the HM1-1B clone. Whole mouse X chromosome painting (green) and Y chromosome painting (red) were used as probes. idicY: isodicentric Y chromosome; mX: X chromosome marker. (b) Representative inverted DAPI-banded karyotypes. (c) Representative FISH metaphase spreads. Whole mouse X chromosome painting (red) was used as probe.



**Supplemental Figure 4: Molecular analysis of the rejuvenated NT-clones.** (a) Genomic PCR detecting both forms of the *Hprt* locus on the indicated samples. HM1 is the positive control of the mutated (null) locus; MEF is the positive control of the wt locus; HM1-1B12 is the positive control of both loci. (b) Analysis of SNPs mapped to the X chromosome performed on the rejuvenated NT-clones.



**Supplemental Figure 5: Cytogenetic analysis of the HM1-1B12 subclone and its derivative rejuvenated NT-clones.** (a) On the left, frequency of the karyotypes of the HM1-1B12 subclone at various passages; on the right, a representative karyotype of the most frequent cell population (inverted DAPI-banded). (b,c,d) On the left, frequency of the karyotypes and on the right a representative karyotype of the most frequent cell population (inverted DAPI-banded) of the indicated NT-clones. (n= 20 each). All passage numbers are indicated with p.

**Table 1**

Primers for genomic PCR			
Name	Primer Sequence	Amplicon Size	Annealing T
rs3166628	F:5'-TACATGCACTCTCGAGTGGT-3'	261bp	55°C
	R:5'-CCAATGGTGCCATTTCTCCA-3'		
rs8251054	F:5'-TCTGCCTGAGCTCCTTCTTC-3'	200bp	54°C
	F:5'-AGTTCAATCCCAGCAACCA-3'		
rs8237198	F:5'-GGAGTCTAACGCCATTGTCC-3'	201bp	55°C
	R:5'-TGTA CTTCATGGCCTGCTGA-3'		
rs8251080	F:5'-AGGCATTGACTAGCCAGCAG-3'	295bp	55°C
	R:5'-AGCCATATTGGCATCCTCAC-3'		
mSRY	F:5'-TGCTCCTGGTATGGTGTATG-3'	863bp	58°C
	R:5'-CTGTACAACCTTCTGCAGTG-3'		

**Supplemental Table 1.** Primers for genomic PCR.

**Table 2**

Primers for RT-PCR				
Name	Accession#	Primer Sequence	Amplicon Size	Annealing T
Nestin	NM_016701	F:5'-AGGAACCAAAAGAGGCAGGT-3'	230bp	58°C
		R:5'-CTTGGGACCAGGGACTGTTA-3'		
Bmp2	NM_007553	F:5'-GCTCCACAAACGAGAAAAGC-3'	178bp	58°C
		R:5'-AGCAAGGGGAAAAGGACACT-3'		
AFP	NM_007423	F:5'-ATGAAGCAAGCCCTGTGAAC-3'	158bp	58°C
		R:5'-AGCTTGCCACAGATCCTTGT-3'		
GAPDH	NM_008084	F:5'-TGTCAGCAATGCATCCTGCA-3'	196bp	58°C
		R:5'-TGGATGCAGGGATGATGTTC-3'		

**Supplemental Table 2.** Primers for Real Time PCR.