

Cell Reports

Supplemental Information

**L-selectin Is Essential for Delivery
of Activated CD8⁺ T Cells to Virus-Infected
Organs for Protective Immunity**

Rebar N. Mohammed, H. Angharad Watson, Miriam Vigar, Julia Ohme, Amanda Thomson, Ian R. Humphreys, and Ann Ager

Supplemental Information

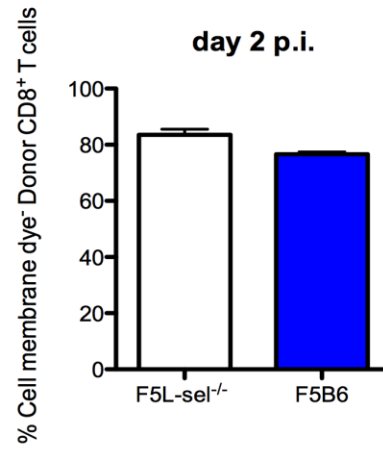
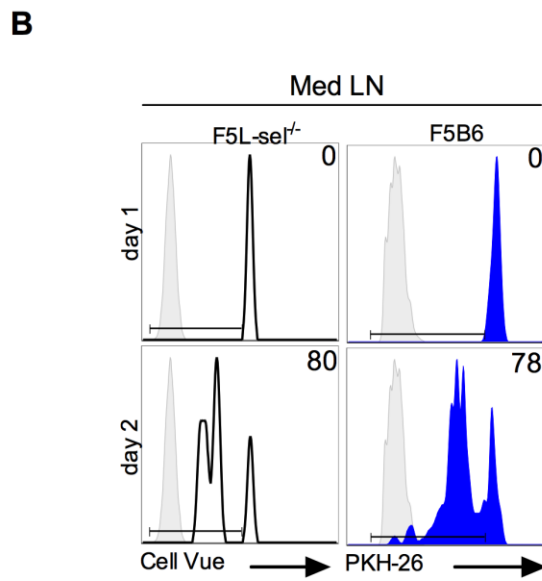
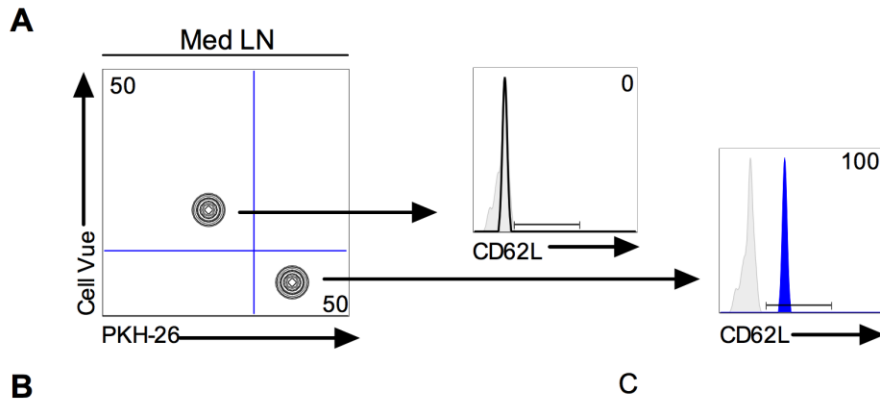


Figure S1, Related to Figure 2: L-selectin expression does not affect localization or priming of CD8⁺ T cells in the mediastinal LN. Naïve F5L-selectin^{-/-} and F5B6 CD8⁺ T cells were purified and stained with 2µM Cell Vue Claret and PKH-26 respectively. Cells were mixed 1:1 and total of 4 x 10⁶ were transferred i.v. into CD90.1 B6 mice. Mice were infected with 2 x 10⁶ pfu vaccNP or left un-infected. **(A)** Representative plot shows the percentage of F5B6 (PKH-26⁺) and F5L-selectin^{-/-} (Cell Vue⁺) CD8⁺ T cells in the Mediastinal LN in un-infected mice 24h post transfer; the donor cells were confirmed by CD62L expression. **(B)** Representative histograms show the percentage cell tracker dye dilution at days 1 and 2 p.i. compared to the unstained cell, grey histograms. **(C)** Bar chart shows the mean + SEM dye dilution (n=3).

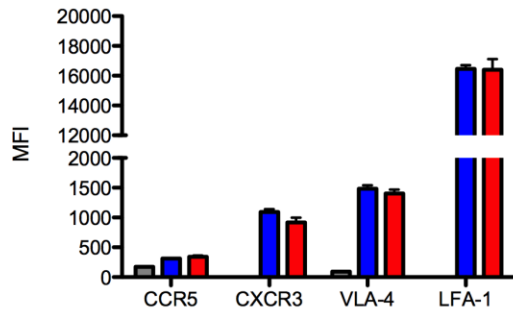
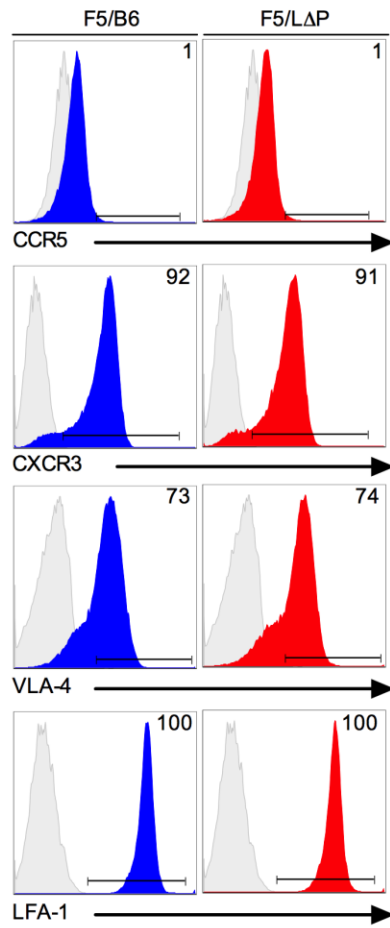
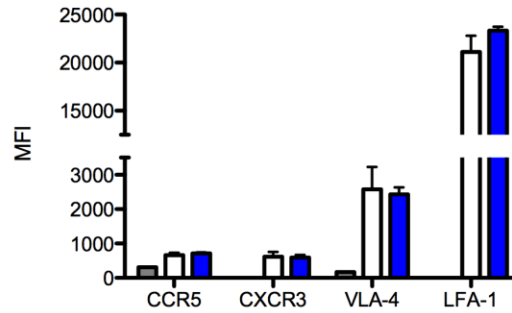
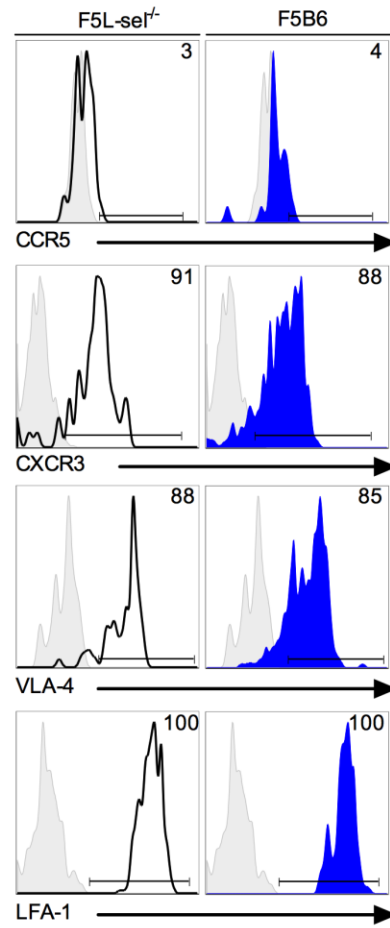
A**B**

Figure S2, Related Figure 3: L-selectin does not affect expression of inflammation associated homing molecules on virus specific CD8⁺ T cells. (A) F5/B6 (blue) and F5/LΔP (red) or (B) F5L-sel^{-/-} (open) and F5B6 (blue) CD8⁺ T cells were transferred into naïve B6 mice and 24 hours later mice given vaccNP i.p. Spleens were collected 5 day p.i. and donor cells were identified as in Fig. 2. Representative histograms show percentage positive CCR5, CXCR3, VLA-4 (CD49d) and LFA-1 (CD11a) by donor CD8⁺ T cells. Bar charts show mean \pm SEM MFI (n=4).

Table S1, Related to Experimental Procedures: Mice

Mice	F5 genotype	Endogenous L-selectin genotype	Transgenic LAP genotype	CD90 genotype	Fold increase L-selectin expression over wildtype
F5/B6	+/-	+/+	-	CD90.2	1.0
F5/LΔP	+/-	-/-	+/-	CD90.2	1.5 ^a
F5/ CD90.1B6	+/-	+/+		CD90.1/CD90.2	1.0
F5B6	+/+	+/+	-	CD90.2	1.0
F5LΔP	+/+	-/-	+/+	CD90.2	3.0 ^a
F5L-sel ^{-/-}	+/+	-/-	-	CD90.2	No expression
LΔP	-/-	-/-	+/+	CD90.2	3.0 ^a
L-sel ^{-/-}	-/-	-/-	-/-	CD90.2	No expression
C57BL/6 (B6)	-/-	+/+	-/-	CD90.2	1.0

^a Data from (Galkina et al., 2007)

Supplemental Experimental Procedures

Flow Cytometry

Single cell isolate stained with PE-conjugated H2-D^bNP68-tetramer (Influenza H17 derived nucleoprotein 366-374 ASNENMDAM) or H-2D^bNP34-tetramer (Influenza A-PR8-34 nucleoprotein 366-374 ASNENMETM) at 37°C, followed by surface stains at 4°C with FITC-anti Vβ11 (clone: KT-11, Biolegend), PerCPCy5.5-anti CD8 (53-6.7, Biolegend), PECy7-anti CD62L (MEL-14, Biolegend), APC-anti CD69 (H1.2F3, Biolegend), APCCy7-anti CD44 (IM7, Biolegend), Pacific Blue-anti CD90.2/Thy1.2 (53-2.1, Biolegend), AlexaFluor 488-anti CD90.2 (30-H12, Biolegend) and intracellular stains with PerCPCy5.5-anti IFN-γ (XMG1.2, Biolegend), PE-Cy7-anti TNF-α (TN3-19, eBioscience), eFluor-450-anti IL-2 (JES6-5H4, eBioscience) FITC anti-granzymeB (GB-11, Biolegend), Biotinylated-anti CCR5 (HM-CCR5 7A4, eBioscience), APC-anti CXCR3 (CXCR3-173, eBioscience), PE-anti CD49d (PS/2, Southern Biotechnology), PECy7-anti CD11a (M17/4, Biolegend). Fluorescence minus one (FMO) was used for non-specific, background staining. Cells were fixed and analyzed on a BD FACS Canto II flow cytometer and counted using Cytocount beads (DAKO). Figures were prepared using FlowJo software.

Short term homing assays

For *in vitro* activated CD8⁺ T cell experiment, F5LΔP and F5L-sel^{-/-} CD8⁺ T cells were purified and then re-suspended in complete medium (DMEM supplemented with 10% FCS, penicillin-streptomycin, L-glutamine, non-essential amino-acids and 2-β mercaptoethanol) and plated at 2 x 10⁶ cells/well in 24-well plates (Nunclon). Splenocytes from F5B6 mice were pulsed with 5 μg/ml NP68 peptide (Peptide Synthetics) for 1 hour at 37°C, irradiated at 3000 centigrays, 6 x 10⁶ splenocytes added per well and plates incubated at 37°C in 5% CO₂. Fresh complete medium supplemented with 360 IU/ml hrIL-2 was added after 2 days. Cells were harvested at day 7, mixed 1:1 and a total of 5.58 x 10⁶ cells (2.76 x 10⁶ F5LΔP and 2.82 x 10⁶ F5L-sel^{-/-} determined using Cytocount beads) injected i.v. into 4 vaccNP infected CD90.1⁺ B6 mice. After 3 hours, ovaries, ovdLNs, axillary lymph nodes, inguinal lymph nodes, mediastinal lymph node, spleen, peripheral blood were harvested, stained for CD8, CD90.2 and L-selectin and analyzed by flow cytometry for numbers and L-selectin expression on donor CD8⁺, CD90.2⁺ cells. For *in vivo* activated CD8⁺ T cells, F5LΔP and F5L-sel^{-/-} mice were infected with 2x10⁶ vaccNP i.p., At day 5 post infection, mice were euthanized and activated CD8⁺ T cells were isolated from spleen, mixed 1.16:0.84 and a total of 13.68 x 10⁶ cells (5.8 x 10⁶ F5LΔP and 7.87 x 10⁶ F5L-sel^{-/-}) injected into 4 vaccNP infected CD90.1⁺ B6 mice.

L-selectin inhibition and T cell proliferation in virus-infected mice

To inhibit L-selectin function, 250 μg of monoclonal antibody to mouse L-selectin (MEL-14) or rat IgG2a isotype control antibodies MAC193 (Price et al., 1997) or MAC 219 (Forsyth et al., 2000) were injected i.v. and/or i.p. either once or daily commencing 52 h post infection. The thymidine analogue 5-ethynyl-2'-deoxyuridine (EdU; Life Technology,) was re-suspended in saline and 1mg injected i.p. 24 hour prior to tissue harvest.

rIL-2

Human recombinant interleukin-2 (hrIL-2) (Proleukin, Novartis Pharma,) was diluted in saline and RAG1^{-/-} mice injected daily with 150,000 IU hrIL-2.

Supplemental References

Forsyth, I.A., Hutchings, A., and Butcher, G.W. (2000). A panel of monoclonal antibodies to ovine placental lactogen. *J Endocrinol* 165, 435-442.

Galkina, E., Florey, O., Zarbock, A., Smith, B.R., Preece, G., Lawrence, M.B., Haskard, D.O., and Ager, A. (2007). T lymphocyte rolling and recruitment into peripheral lymph nodes is regulated by a saturable density of L-selectin (CD62L). *Eur J Immunol* 37, 1243-1253.

Price, A.A., Cumberbatch, M., Kimber, I., and Ager, A. (1997). Alpha 6 integrins are required for Langerhans cell migration from the epidermis. *J. Exp. Med.* 186, 1725-1735.