Neuroimaging Genetic Risk for Alzheimer Disease in Preclinical Individuals: From Candidate Genes to Polygenic Approaches

Supplemental Information

The Genetics of Alzheimer Disease

Genome-wide association studies (GWASs) published since 2009 have added over 15 genetic loci to the list of genetic risk factors for AD (1–4). This brings the total number of genes implicated as risk genes in AD to 21 (5). These genes vary in their physiological function from synaptic proteins (*PICALM*) to co-chaperones (*CLU*) and mitochondrial transmembrane transporters (*TOMM40*). There are also autosomal dominant forms of AD caused by a mutation in one of three genes, namely *APP*, *PSEN1* and *PSEN2*, or by overexpression of *APP* caused by a duplication event or trisomy 21, in which three copies of the APP gene are present. These familial, inherited forms of AD provide unique opportunities for studying preclinical AD in mutation carriers, but questions remain as to how generalizable findings in familial AD will be to the much more common late-onset AD. Therefore, this review focuses on neuroimaging genetics of late-onset, sporadic AD which accounts for over 95% of total AD cases (6).

The International Genomics of Alzheimer's Project (IGAP) consortium published their first GWAS effort in 2013 (7). The study was the largest GWAS ever on late-onset, sporadic AD. Using the uniquely large cohort of 74,046 subjects amassed from four smaller data consortia the authors were able to confirm the association of previously implicated loci, as well as detect 11 new AD risk loci. Specifically, the first stage of analysis resulted in 15 genomic regions that showed an association to AD. These regions included 10 previously identified AD genetic risk factors, including *APOE*, and 5 newly implicated loci. The other 9 previously identified loci were *CR1*, *BIN1*, *CD2AP*, *EPHA1*, *CLU*, *MS4A6A*, *PICALM*, *ABCA7* and *CD33*. All available neuroimaging genetics findings for these replicated loci are reviewed below (*APOE*, *CR1*, *BIN1*, *EPHA1*, *PICALM*, *ABCA7*, *CD33*) or in the main text (*CLU*). The 5 new loci identified in the first

stage were *HLA-DRB5-HLA-DRB1*, *PTK2B*, *SORL1*, *SLC24A4*, *RIN3* and *DSG2*. Second stage replication analyses revealed 7 additional, novel loci that reached statistical significance for association: *INPP5D*, *MEF2C*, *NME8*, *ZCWPW1*, *CELF1*, *FERMT2* and *CASS4*. Notably, two loci from the first stage did not reach statistical significance in the second stage replication analyses: *CD33* (a previously identified risk locus) and *DSG2* (a novel locus). The authors found a total of 9 fully replicated, previously identified risk loci, including *APOE*, as well as 11 newly identified risk loci.

Variants identified in GWASs are usually commonly occurring and have low odds ratio associations with disease. Rare variants (occurring in less than 5% of the population) of the *TREM2* and the *MAPT* genes with the moderate to high effect size have also recently been associated with AD (8–10).

We now know more than ever before about the underlying genetics of AD. The growing list of AD risk genes serves to highlight the complexity of AD genetics and the need for more sophisticated experimental designs that combine multiple genetic risk loci. Next, we discuss important and valuable findings from neuroimaging genetics studies in the AD literature that focused on a single gene and then transition to polygenic approaches.

Neuroimaging Candidate Genes for AD

APOE in Older, Healthy Adult Cohorts

Hippocampal volumes have been shown to be smaller in *APOE*ɛ4 carriers compared to noncarriers in older healthy adults (11; 12). The rate of hippocampal atrophy is also higher in *APOE*ɛ4 carriers (13; 14). There is evidence that hippocampal volumes vary in an allele dosedependent manner, but it is often not possible to amass enough homozygous *APOE*ɛ4 carriers to consider them separately (15). Structural MRI (sMRI) can also be used to measure structural changes within specific areas of the hippocampus. High resolution, partial field of view sMRI allows for the segmentation of the hippocampal complex into specific subregions, such as the

entorhinal cortex, cornu ammonis (CA) subfields and the subiculum. Based on this approach, several studies have provided evidence for smaller or thinner subregions in healthy *APOE* ϵ 4 carriers. Specifically, entorhinal cortex and subiculum are thinner in healthy *APOE* ϵ 4 carriers compared to non-carriers (16). Two additional studies performed at high-field using MR images acquired at 4 Tesla found thinner CA3 and dentate gyrus subfields in *APOE* ϵ 4 carriers (17; 18). However, there are studies in which no differences in hippocampal volume between healthy *APOE* ϵ 4 carriers and non-carriers were found, although they are certainly in the minority (19).

It is not clear whether there are alterations to cortical morphology in older healthy adult carriers of the *APOE* ϵ 4 allele. One published study reported no significant differences in cortical volume between healthy *APOE* ϵ 4 carriers and non-carriers (19). However, another study found that *APOE* ϵ 4 carriers had thicker cortex in bilateral frontal and temporal regions, but a steeper longitudinal atrophic trajectory across the whole cortex (20). This supports the emerging theme that individuals with at least one copy of the *APOE* ϵ 4 allele experience an acceleration of the volume loss seen in normal aging, both in the hippocampus and across the cortex.

A caveat of structural findings in the *APOE* literature is that atrophy or volume loss is generally seen as an indication of disease processes. In contrast, increased volumes or decreased atrophy rates are usually not interpreted as a possible disease feature. Intuitive results, e.g. where *APOE*_E4 carriers have lower or smaller volumetric measurements, are likely to appear more in the published literature. In contrast, there is no such biasing intuition in fMRI, which may partially explain why results in the fMRI field comparing *APOE*_E4 carriers to non-carriers are often contradictory.

APOE is a lipoprotein that transports endogenous lipids. Because myelin is composed primarily of lipids, there is interest in better understanding the relationship between *APOE* and myelin maintenance and repair. Diffusion weighted imaging (DWI) can be used to examine the potential relationship between *APOE* and myelination, using 'white matter integrity' measured by fractional anisotropy (FA) as a proxy for myelin health. One study found that white matter

integrity in the medial temporal lobe, but not entorhinal thickness, was associated with improved performance on a verbal memory task (21). There is also evidence for a general decrease in FA in older *APOE* ε 4 carriers (22). Diffusion tensor imaging allows tractography algorithms to estimate the white matter pathways present in the brain. These tracts can then be used to apply mathematical concepts from the field of graph theory to DWI data. Brown and colleagues used graph theory to measure global integration and local interconnectivity in healthy, older subjects. *APOE* ε 4 carriers showed an age-related decrease in local interconnectivity that may indicate different aging trajectories in *APOE* ε 4 carriers and non-carriers (23). The application of graph theory to sMRI data as well as resting state fMRI data may help to elucidate the local and global network properties that are altered early in AD, but more research is needed in this area before such measures can be considered as potential biomarkers for AD.

The task-based fMRI-*APOE* literature tells a frustratingly complex and contradictory story. Some studies report increased, putatively compensatory, activity in *APOE* ϵ 4 carriers (24; 25). Others report decreased activity, putatively caused by diminished function due to disease processes (26; 27). These contradictions may be partially explained by the heterogeneity of task designs used (28). Differences between tasks can be striking. For example, it is not surprising that results from a sematic memory task and a visuospatial memory task may be difficult to summarize in a single effect of the *APOE* ϵ 4 allele on brain function (26; 25). Other possible confounding factors in task design can be more subtle. A "paired associates" memory task can actually vary widely in several ways including, but not limited to, the method of presentation of stimuli (audio, visual, or both), types of stimuli (images, words, etc.) and instructions ('pay attention' versus 'remember these pairs') (28). There are also studies in the literature in which investigators used non-episodic memory-based tasks, which complicates interpretation because there is evidence that *APOE* ϵ 4 may exert a specific effect on episodic memory systems (29). In contrast to the whole-brain approach of the studies cited above, results from studies that examined blood-oxygen-level dependent signal in the hippocampus are more cohesive, often

reporting decreases in signal in *APOEε4* carriers. In one study, with data acquired using a highresolution fMRI sequence, decreased activity in *APOEε4* carriers was reported in the CA2, CA3 and dentate gyrus subregion of the hippocampus (30). In another, decreased hippocampal activity during encoding was found in *APOEε4* carriers (31).

Results from resting state fMRI work in healthy older *APOE* ϵ 4 carriers suggest that there may be a convergence on the DMN and connectivity therein, by which *APOE* ϵ 4 carriers and non-carriers differ. In a recent study, connectivity between the posterior cingulate cortex and the hippocampus, two major nodes of the DMN, was found to be diminished in *APOE* ϵ 4 carriers (32). Another study, which focused on women, reported significantly reduced DMN connectivity in carriers of the *APOE* ϵ 4 compared to non-carriers (33). Lastly, a pattern of decreased DMN connectivity along with increased connectivity of another, opposing cognitive network, the salience network, have been described (34; 35). One theory that may explain DMN dysfunction reported in *APOE* ϵ 4 carriers is that genetic vulnerability for AD may result in a loss of appropriate hippocampal decoupling from cortical DMN regions during an active state, such as when completing a task (36). This theory is supported by a study in which a negative correlation between hippocampus-DMN synchronization and performance on a memory test was reported (36). However, more work is needed to further test this theory.

PET imaging has helped further elucidate the relationship between *APOE* and betaamyloid (A β) accumulation. While the relationship is still far from fully understood, we know the protein products of *APOE* play a role in A β clearance, with *APOE* ϵ 4 performing worse than the ϵ 3 or ϵ 2 alleles (37). This idea is supported by PET imaging studies in which the relationship between A β deposition (measured by Pittsburg compound B (PiB) or florbetapir) and *APOE* ϵ 4 carrier status is examined. Most of these studies report that healthy, older *APOE* ϵ 4 carriers have increased amyloid load compared to non-carriers (38–41). There are also differences in brain glucose metabolism, measured by fluorodeoxyglucose (FDG), between healthy *APOE* ϵ 4 carriers and non-carriers. A large study with 806 cognitively normal, PiB-PET negative subjects

recently showed that glucose metabolism in *APOE* ε 4 carriers is lower in the posterior cingulate, precuneus, lateral parietal and inferior temporal regions (42). The raw magnitude of this difference was small but it was similar to differences observed between cognitively normal *APOE* ε 4 carriers and those with mild cognitive impairment. There was also a negative correlation between average FDG uptake across the brain and age across the whole cohort, with the posterior cingulate and precuneus showing a unique vulnerability to both age and *APOE* ε 4 carrier status (42). Hypometabolism in AD vulnerable regions in healthy *APOE* ε 4 carriers has been reported before (43). However, a study of 600 cognitively healthy older subjects found no FDG-PET metabolism differences in *APOE* ε 4 carriers and non-carriers (44). This discrepancy may be due to the inclusion of PiB-PET positive subjects in the latter report, who were stratified based on tracer uptake. Perhaps when subjects are binned by amyloid burden, the power to detect *APOE* ε 4 differences in metabolism, especially in the amyloid positive groups, is reduced.

Magnetic resonance spectroscopy (MRS) is a technique that can be used to measure the relative concentrations of different hydrogen containing metabolites, each with a different peak resonance that can be plotted and quantified. Recent work using MRS in the posterior cingulate, a region particularly vulnerable to AD, has revealed that both GABA and glutamine/glutamate metabolites are reduced in individuals with MCI (45). However, the authors did not detect an association between the metabolites they measured and *APOE* ϵ 4 status or amyloid deposition, which limits their usefulness as AD-specific biomarkers. In contrast, another recent study that also focused on the posterior cingulate examined choline/creatine and myoinositol/creatine ratios and found that they were significantly higher in older adult carriers of *APOE* ϵ 4 compared to non-carriers (46). This finding supports earlier work in this field that found that myoinositol/creatine ratio is associated with neurodegenerative disease, as opposed to normal age related cognitive decline (47). Examining creatine levels alone, another study observed significantly lower creatine in *APOE* ϵ 4 carriers compared to non-carriers (48). There is

also evidence that healthy older individuals with smaller hippocampal volume have a lower Nacetylaspartate/myoinositol ratio, which has been associated with AD, compared to their peers with larger hippocampal volume (49). Taken together, these results indicate that some metabolite measures and ratios may be useful biomarkers in individuals already at increased risk for AD.

PICALM

The gene encoding phosphatidylinositol binding clathrin assembly protein (*PICALM*) was identified as an AD risk factor in 2009 (1). The original locus (rs3851179) is located upstream from *PICALM*, but subsequent studies have not only replicated this finding but also identified additional AD-risk single nucleotide polymorphisms (SNPs) within the *PICALM* gene itself (4). *PICALM* is widely expressed in the brain. It is involved in several cellular processes, but especially in the trafficking of proteins and lipids via clathrin mediated endocytosis (50). This process, which is essential to synaptic transmission, has received increased attention in the context of AD in part because of the strong association of AD with *PICALM* uncovered in GWASs (51). *PICALM* ranks third after the *APOE/TOMM40* locus and *CLU* in terms of reproducibility in GWASs (4; 7; 52; 53). Perhaps because of this relatively highly reproduced association, *PICALM*-mediated AD risk has been examined in several neuroimaging genetics studies.

In older adults ranging from cognitively healthy to diagnosed with AD, a significant association between *PICALM* (rs3851179) and hippocampal volume was reported such that carriers of the *PICALM* risk variant had lower hippocampal volume (54). The authors also found a link between *PICALM* risk and reduced entorhinal cortex thickness. The latter finding was replicated in another study that found that the *PICALM* risk allele is associated with a thinner entorhinal cortex in older adults (55). However, *PICALM* was not associated with either hippocampal or entorhinal cortex volume in a cohort of healthy young adults (56).

The relationship between *PICALM* and the functional connectivity of the hippocampus was recently reported in a resting state fMRI experiment. When compared to subjects who were homozygous for the protective allele, risk allele carriers showed weaker negative functional connectivity of the hippocampus to many cortical regions (57). However, this finding from a relatively small cohort is preliminary and needs to be replicated. Lastly, a study examining amyloid deposition as measured by florbetapir-PET found an epistatic effect involving variants of *PICALM* and *BIN1*, another AD risk gene (58). This study is further described below (see *BIN1*).

CR1

Unlike *PICALM*, expression of complement component (3b/4b) receptor 1 (*CR1*) is likely to be low in the brain (59). The CR1 protein's function is complex and varies by cell type. Generally, it is involved in the regulation of the complement cascade, a major component of the innate immune system that helps to amplify the response of the immune system to potential targets. In addition, the CR1 protein is involved in transporting opsonized immune complexes through the circulatory system for efficient removal (60). Neuroinflammation has been associated with AD for many years, but has sometimes been dismissed as a consequence, rather than a cause, of the disease (61). However, this perspective is shifting and inflammatory processes are being studied as potential pathogenic processes in AD (51). One reason, among many, why interest in neuroinflammation and AD has been renewed in recent years is the discovery of an association between a polymorphism in *CR1* and AD in a 2009 GWAS (3).

The *CR1* risk variant was associated with thinner entorhinal cortex in a study including healthy older adults (54). Interestingly, there is also evidence that the *CR1* risk variant is associated with lower entorhinal cortex volume in *young* healthy adults (62). This finding was confirmed in two independent cohorts. Additional research is needed to assess whether the relationship between *CR1* variants and entorhinal cortex is reproducible in larger samples.

BIN1

Bridge integrator 1 (*BIN1*) was reported as a risk gene for AD in 2010, after a borderline significant association was reported in one of the large 2009 GWASs (1; 52). Like *PICALM*, *BIN1* is involved with the intracellular trafficking of lipids and proteins. *BIN1* encodes a protein that has at least ten known isoforms (60). These isoforms have specific domains that influence their function. In the brain, there is one isoform of BIN1 and it contains the clathrin-associated protein-binding (CLAP) domain, which plays a role in clathrin mediated endocytosis (63). Clathrin mediated endocytosis is an essential process in synaptic vesicle recycling, which is an essential component of efficient synaptic transmission. *PICALM* and *BIN1* are implicated as molecular components of this neuronal process, which may suggest that variability in synaptic transmission efficiency contributes to AD pathology, especially during the early phase characterized by synaptic loss and neuronal death (51).

Because *PICALM* and *BIN1* are both related to synaptic transmission, one group tested for possible epistatic effects between the risk locus for each gene identified in GWASs. Hohman and colleagues used florbetapir-PET scans to test for a possible interaction effect of *BIN1* and *PICALM* on amyloid deposition (58). The authors reported that there was indeed an interaction and that this interaction was reproduced in a second, independent dataset. The *BIN1* risk variant was related to greater amyloid burden, but only in persons who were carriers of the *PICALM* protective variant. This study illustrates a limitation of candidate gene studies because both *BIN1* and *PICALM* were not related to amyloid deposition when examined on their own.

There is, however, evidence that *BIN1* genotype is directly associated with a neuroimaging-based structural biomarker of AD. Based on the preliminary evidence from the 2009 GWASs that *BIN1* may be associated with AD, Biffi and colleagues tested for an association of the *BIN1* risk variant and several neuroimaging phenotypes. The authors showed that the *BIN1* risk variant is associated with thinner temporal pole and entorhinal cortex (54).

Soon after Biffi and colleagues published their paper a *BIN1* locus reached genome-wide significance in a new AD GWAS (52).

ABCA7

ATP-binding cassette, subfamily A, member 7 (*ABCA7*) is one gene in a group of highly conserved transmembrane transporters. These transporters participate in active transport of various substrates across membranes, both at the level of the cell and the organelle (64). *ABCA* transporters are linked to cholesterol and lipid homeostasis, and appear to work directly with *APOE* by transporting lipids out of the cell to be cleared by *APOE* (65). This coordination with *APOE* may be a clue to the mechanism of the association between *ABCA7* and AD. *ABCA7* was first linked to AD in the results of a GWAS in 2011 (2).

The AD-associated locus near *ABCA7* has been studied in one recent neuroimaging study in which the authors were interested in the relationship between cholesterol levels and amyloid deposition (66). Hughes and colleagues described a greater than 2-fold increased risk of amyloid positivity, as measured by PiB-PET, in carriers of the *ABCA7* (rs3752246) risk variant.

EPHA1

EPHA1 is a member of a superfamily of proteins called the receptor tyrosine kinases and is expressed in multiple tissues including the brain (67; 68). The Eph-ephrin family of receptors and ligands are all membrane-bound proteins involved in adhesion and cell-cell contact mediated signaling, like in axonal guidance during development (68). The association between *EPHA1* and AD was first described in two GWASs published in 2011 (2; 4). The neurobiological basis of this link to AD may be related to the high expression of the *EPHA1* receptor class in the hippocampus, but the expression and function of specifically *EPHA1* in the hippocampus is not well understood (69).

One study took a neuroimaging genetics approach to describe the relationship between *EPHA1* and amyloid in the brain. Hughes and colleagues found that the *EPHA1* is associated with the likelihood of being amyloid positive, as measured by PiB-PET. In contrast to the findings for *ABCA7* described above, the authors found that risk of amyloid positivity decreased for each C allele of *EPHA1* (rs11767557) (66).

CD33

Sialic acid binding immunoglobulin-like lectin-3 (*CD33*) is a membrane-bound receptor expressed on immune cells (70). *CD33* plays a role in the differentiation of immature immune cells, as well as in the signaling of mature immune cells in the innate and adaptive immune system (70). Despite strong evidence from several GWASs that a variant of *CD33* is associated with AD, this association was not fully replicated in the IGAP consortium GWAS (2; 4; 7). This failure to replicate the association in the largest AD GWAS to date casts some doubt on the strength and reproducibility of this gene's association with AD. Perhaps the association is specific to certain regions and ancestries, even within Caucasian populations. A single study has examined the relationship between *CD33* genotype and a neuroimaging phenotype. Bradshaw and colleagues found that the risk variant of *CD33* was associated with greater, more diffuse amyloid deposition as measured with PiB-PET imaging (71).

Advanced Association Models

In addition to more traditional regression approaches, advanced association models can be used to confront the challenges of working with large datasets in neuroimaging genetics. Canonical correlation is a method for interpreting large cross-covariance matrices that maximizes correlation between linear combinations of pairs of vectors within a given matrix. Sparse canonical correlation takes this process a step further by minimizing the number of features used to find the maximum correlation structure using, for example, the well-known least

squares approach (72). Sparse canonical correlation has been used to explore genetic risk factors for AD affecting the hippocampal surface (73). Looking only at AD risk genes as listed in the AlzGene database, the authors of that study found that *APOE* and *TOMM40* were associated with hippocampal surface differences in sparse regions, including anterior and middle areas (73). Variations of the sparse canonical correlation approach have been used in two other studies focused on AD (74; 75). The first used a "knowledge-guided" algorithm that accounted linkage disequilibrium and genetic co-expression networks and examined the relationship between SNPs within *APOE* and amyloid deposition as measured by florbetapir-PET (74). This study identified only a single SNP in *APOE* that was associated with amyloid deposition, but they argue that their method can be scaled up to genome wide studies. The second study that used a similar approach and also examined *APOE* discovered an association between a specific SNP and gray matter density in right hippocampus (75).

Limitations

Clinical Utility of GWAS Loci: The Search for Causal Variants

The causal variants that give rise to the *APOE* ε 4 allele are known polymorphisms at rs429358 and rs7412. Variants at these loci alter the structure and function of the translated APOE protein (76). In fact, *APOE* ε 4 "structure correctors", which make protein products of *APOE* ε 4 behave like the more common protein products of *APOE* ε 3 are currently being developed as a possible treatment for AD (77). In contrast, many of the GWAS-identified AD risk loci are located in intronic (*CLU*, *ABCA7*) or intragenic (*BIN1*, *EPHA1*) regions with no evidence that variants affect protein structure or function. An intragenic region may play some regulatory function, but in the cases of *EPHA1* and *BIN1* there is little evidence of conservation of these intragenic regions, which makes a regulatory role in genetic expression unlikely (60). There is even some debate over whether or not genes are correctly identified when significantly associated loci reside in non-coding regions. The common approach is to report the SNP as related to the nearest gene,

but this is not necessarily the case. The search for the causal variants for these genes and for genes implicated in other common disease by GWASs is still ongoing (78). Ostensibly, the causal variant for one of these genetic risk loci will be a polymorphism in high linkage disequilibrium with the GWAS locus. In addition, the polymorphism should affect the downstream structure or function of the gene's RNA or protein product. The utility of GWAS-identified risk genes as potential drug targets is limited without first identifying the causal variants driving the association at each locus. One important step in this effort is the development of a functionally annotated genome and the tools to explore it, such as ENCODE (https://www.encodeproject.org). Using ENCODE investigators can quickly discover basic functional information about a locus of interest, perhaps one they identified in a GWAS. The functional elements annotated in the ENCODE project help investigators distinguish between, for example, regulatory elements, close-range promoters and genes that are likely to be transcribed (determined using RNA-seq and similar techniques). ENCODE is an excellent example of a large-scale collaborative project that will enhance the scientific community's ability to interpret genetic association signals.

Mechanistic Interpretations and Neuroimaging Genetics

The incorporation of human genetics into neuroimaging studies has identified brain traits that are associated with specific genetic variants or, more germane to this review, with genetic risk scores. However, as described in the text that precedes this section, the genetic loci are often identified via GWAS and thus, the causal variant is not known. This inherently limits the mechanistic insights researchers are able to gain from neuroimaging genetics studies of this kind. For *APOE*, for which there is no ambiguity about causal variants, neuroimaging has revealed that carriage of the *APOE* ε 4 allele is related to increased amyloid deposition as measured by PET imaging which, in turn, is related to neuronal death and a higher rate of cortical thinning in AD-vulnerable regions when compared to matched controls who do not carry

the risk variant. Saykin and colleagues (2015) describe a multi-step process to move from genetic signals to targeted therapeutics in which genetics and neuroimaging intersect at the first step (discovering genetic loci that are robustly associated with a relevant trait) and the final step (identifying individuals most likely to benefit from experimental therapies) (79). The middle steps include identification of causal genes, testing hypothesized mechanisms in model systems and developing therapeutics that act on these mechanisms. Thus, we believe the salient point is that neuroimaging genetics research is essential to the development and execution of therapeutic hypotheses, even if, in isolation, these studies do not always yield new mechanistic insights.

Generalizability Across Ancestries

It is important to recognize that all the largest AD GWASs used large cohorts of Caucasian European or American subjects. This creates a potential problem with generalizability to other ancestry groups, especially that of African ancestry (80). While there are published GWASs examining AD genetics in minority ancestral groups, one only of these, focused on African American participants, has topped 1,000 participants in the case and control groups ((81); see Table 1 in (82)). Thus, these groups remain understudied compared to the very large GWASs with non-Hispanic Caucasian participants. The genetic loci implicated by studies of Caucasians might fail to replicate in a cohort of subjects from a different ethnic background due to population specific variants, differing patterns of linkage disequilibrium or even a heterogeneous genetic basis of AD in different ethnic groups (83). To illustrate this issue, consider that many small GWASs have tried to replicate the association of CLU with AD in non-Caucasian cohorts. The results of these studies indicate that there is an association between CLU and AD in Chinese cohorts, but not in cohorts of non-white Americans or Europeans (53; 84; 85). The limited generalization of results from large published GWASs in AD is a problem and a greater effort must be made to amass comparably large samples of different ancestral groups for new association studies. This effort may lead to the identification of certain genes that are associated

with AD regardless of genetic background. These genes would be good candidates for increased research resources and drug targeting due to their greater generalizability. Also, importantly, further exploration of the genetic basis of AD in people of African and Hispanic descent may help elucidate any biological bases for the epidemiological differences observed in these ethnic groups, including higher incidence and earlier onset of AD (86).

Small Sample Sizes: Consequences for Neuroimaging Genetics

As eloquently described by Button and colleagues (2013), small sample sizes in neuroimaging studies decrease statistical power which leads to a decreased rate of detectable true positive results while leaving the rate of false positives unchanged (87). This has the effect of increasing the likelihood that a significant result is, in fact, spurious. Small sample sizes also bias studies toward large effect size results, as these are the only results that can be significant given the power limitations. The latter phenomenon has been dubbed the "winner's curse" and leads to studies that are very difficult to replicate (87). Given these known problems, why are neuroimaging studies with small samples still (albeit less and less so) prevalent? This is related to the relatively high cost of acquisition of neuroimaging data, the currently accepted need to "pilot" and publish new paradigms and techniques before formal funding for large-scale studies can be won and also immense pressure, especially on young investigators, to publish frequently (88). Taken together, it is clear that sample size is a very important consideration when performing a neuroimaging genetics study and robust power analyses are a crucial component of any research program.

Supplemental References

- 1. Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, *et al.* (2009): Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat Genet* 41: 1088–93.
- Hollingworth P, Harold D, Sims R, Gerrish A, Lambert J-C, Carrasquillo MM, et al. (2011): Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. Nat Genet 43: 429–35.
- Lambert J-C, Heath S, Even G, Campion D, Sleegers K, Hiltunen M, et al. (2009): Genomewide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. Nat Genet 41: 1094–9.
- 4. Naj AC, Jun G, Beecham GW, Wang L-S, Vardarajan BN, Buros J, *et al.* (2011): Common variants at MS4A4/MS4A6E, CD2AP, CD33 and EPHA1 are associated with late-onset Alzheimer's disease. *Nat Genet* 43: 436–41.
- 5. Guerreiro R, Brás J, Hardy J (2013): SnapShot: genetics of Alzheimer's disease. *Cell* 155: Elsevier968–968.e1.
- 6. Bateman RJ, Aisen PS, De Strooper B, Fox NC, Lemere CA, Ringman JM, *et al.* (2011): Autosomal-dominant Alzheimer's disease: a review and proposal for the prevention of Alzheimer's disease. *Alzheimers Res Ther* 3: 1.
- Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, *et al.* (2013): Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nat Genet* 45: Nature Publishing Group, a division of Macmillan Publishers Limited. All Rights Reserved.1452–8.
- Coppola G, Chinnathambi S, Lee JJ, Dombroski BA, Baker MC, Soto-Ortolaza AI, *et al.* (2012): Evidence for a role of the rare p.A152T variant in MAPT in increasing the risk for FTD-spectrum and Alzheimer's diseases. *Hum Mol Genet* 21: 3500–12.
- 9. Reitz C, Mayeux R (2013): TREM2 and neurodegenerative disease. *N Engl J Med* 369: 1564–5.
- 10. Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E, *et al.* (2013): TREM2 variants in Alzheimer's disease. *N Engl J Med* 368: 117–27.
- 11. Taylor JL, Scanlon BK, Farrell M, Hernandez B, Adamson MM, Ashford JW, *et al.* (2014): APOE-epsilon4 and aging of medial temporal lobe gray matter in healthy adults older than 50 years. *Neurobiol Aging* 35: 2479–85.
- 12. Lu PH, Thompson PM, Leow A, Lee GJ, Lee A, Yanovsky I, *et al.* (2011): Apolipoprotein E genotype is associated with temporal and hippocampal atrophy rates in healthy elderly adults: a tensor-based morphometry study. *J Alzheimers Dis* 23: 433–42.

- 13. Jak AJ, Houston WS, Nagel BJ, Corey-Bloom J, Bondi MW (2007): Differential crosssectional and longitudinal impact of APOE genotype on hippocampal volumes in nondemented older adults. *Dement Geriatr Cogn Disord* 23: 382–9.
- 14. Donix M, Burggren AC, Suthana NA, Siddarth P, Ekstrom AD, Krupa AK, *et al.* (2010): Longitudinal changes in medial temporal cortical thickness in normal subjects with the APOE-4 polymorphism. *Neuroimage* 53: 37–43.
- 15. Hostage CA, Roy Choudhury K, Doraiswamy PM, Petrella JR (2013): Dissecting the gene dose-effects of the APOE ε4 and ε2 alleles on hippocampal volumes in aging and Alzheimer's disease. *PLoS One* 8: e54483.
- Burggren AC, Zeineh MM, Ekstrom AD, Braskie MN, Thompson PM, Small GW, Bookheimer SY (2008): Reduced cortical thickness in hippocampal subregions among cognitively normal apolipoprotein E e4 carriers. *Neuroimage* 41: 1177–83.
- Mueller SG, Schuff N, Raptentsetsang S, Elman J, Weiner MW (2008): Selective effect of Apo e4 on CA3 and dentate in normal aging and Alzheimer's disease using high resolution MRI at 4 T. *Neuroimage* 42: 42–8.
- 18. Mueller SG, Weiner MW (2009): Selective effect of age, Apo e4, and Alzheimer's disease on hippocampal subfields. *Hippocampus* 19: 558–64.
- 19. Cherbuin N, Anstey KJ, Sachdev PS, Maller JJ, Meslin C, Mack HA, *et al.* (2008): Total and regional gray matter volume is not related to APOE*E4 status in a community sample of middle-aged individuals. *J Gerontol A Biol Sci Med Sci* 63: 501–4.
- 20. Espeseth T, Westlye LT, Fjell AM, Walhovd KB, Rootwelt H, Reinvang I (2008): Accelerated age-related cortical thinning in healthy carriers of apolipoprotein E epsilon 4. *Neurobiol Aging* 29: 329–40.
- 21. Westlye ET, Hodneland E, Haász J, Espeseth T, Lundervold A, Lundervold AJ (2012): Episodic memory of APOE ε4 carriers is correlated with fractional anisotropy, but not cortical thickness, in the medial temporal lobe. *Neuroimage* 63: 507–16.
- 22. Heise V, Filippini N, Ebmeier KP, Mackay CE (2011): The APOE ε4 allele modulates brain white matter integrity in healthy adults. *Mol Psychiatry* 16: Macmillan Publishers Limited908–16.
- 23. Brown JA, Terashima KH, Burggren AC, Ercoli LM, Miller KJ, Small GW, Bookheimer SY (2011): Brain network local interconnectivity loss in aging APOE-4 allele carriers. *Proc Natl Acad Sci U S A* 108: 20760–5.
- 24. Bondi MW, Houston WS, Eyler LT, Brown GG (2005): fMRI evidence of compensatory mechanisms in older adults at genetic risk for Alzheimer disease. *Neurology* 64: 501–8.
- 25. Bookheimer SY, Strojwas MH, Cohen MS, Saunders AM, Pericak-Vance MA, Mazziotta JC, Small GW (2000): Patterns of brain activation in people at risk for Alzheimer's disease. *N Engl J Med* 343: 450–6.

- 26. Borghesani PR, Johnson LC, Shelton AL, Peskind ER, Aylward EH, Schellenberg GD, Cherrier MM (2008): Altered medial temporal lobe responses during visuospatial encoding in healthy APOE*4 carriers. *Neurobiol Aging* 29: 981–91.
- 27. Xu G, McLaren DG, Ries ML, Fitzgerald ME, Bendlin BB, Rowley HA, *et al.* (2009): The influence of parental history of Alzheimer's disease and apolipoprotein E epsilon4 on the BOLD signal during recognition memory. *Brain* 132: 383–91.
- 28. Trachtenberg AJ, Filippini N, Mackay CE (2012): The effects of APOE-ε4 on the BOLD response. *Neurobiol Aging* 33: 323–34.
- 29. Rogalski EJ, Rademaker A, Harrison TM, Helenowski I, Johnson N, Bigio E, *et al.* (n.d.).: ApoE E4 is a susceptibility factor in amnestic but not aphasic dementias. *Alzheimer Dis Assoc Disord* 25: 159–63.
- 30. Suthana NA, Krupa A, Donix M, Burggren A, Ekstrom AD, Jones M, *et al.* (2010): Reduced hippocampal CA2, CA3, and dentate gyrus activity in asymptomatic people at genetic risk for Alzheimer's disease. *Neuroimage* 53: 1077–84.
- 31. Adamson MM, Hutchinson JB, Shelton AL, Wagner AD, Taylor JL (2011): Reduced hippocampal activity during encoding in cognitively normal adults carrying the APOE ε4 allele. *Neuropsychologia* 49: 2448–55.
- 32. Heise V, Filippini N, Trachtenberg AJ, Suri S, Ebmeier KP, Mackay CE (2014): Apolipoprotein E genotype, gender and age modulate connectivity of the hippocampus in healthy adults. *Neuroimage* 98: 23–30.
- 33. Damoiseaux JS, Seeley WW, Zhou J, Shirer WR, Coppola G, Karydas A, *et al.* (2012): Gender modulates the APOE ε4 effect in healthy older adults: convergent evidence from functional brain connectivity and spinal fluid tau levels. *J Neurosci* 32: 8254–62.
- 34. Machulda MM, Jones DT, Vemuri P, McDade E, Avula R, Przybelski S, *et al.* (2011): Effect of APOE ε4 status on intrinsic network connectivity in cognitively normal elderly subjects. *Arch Neurol* 68: 1131–6.
- 35. Sheline YI, Morris JC, Snyder AZ, Price JL, Yan Z, D'Angelo G, *et al.* (2010): APOE4 allele disrupts resting state fMRI connectivity in the absence of amyloid plaques or decreased CSF Aβ42. *J Neurosci* 30: 17035–40.
- 36. Westlye ET, Lundervold A, Rootwelt H, Lundervold AJ, Westlye LT (2011): Increased hippocampal default mode synchronization during rest in middle-aged and elderly APOE ε4 carriers: relationships with memory performance. *J Neurosci* 31: 7775–83.
- Leduc V, Domenger D, De Beaumont L, Lalonde D, Bélanger-Jasmin S, Poirier J (2011): Function and comorbidities of apolipoprotein e in Alzheimer's disease. *Int J Alzheimers Dis* 2011: 974361.

- 38. Fleisher AS, Chen K, Liu X, Roontiva A, Thiyyagura P, Ayutyanont N, *et al.* (2011): Using positron emission tomography and florbetapir F18 to image cortical amyloid in patients with mild cognitive impairment or dementia due to Alzheimer disease. *Arch Neurol* 68: 1404–11.
- 39. Morris JC, Roe CM, Xiong C, Fagan AM, Goate AM, Holtzman DM, Mintun MA (2010): APOE predicts amyloid-beta but not tau Alzheimer pathology in cognitively normal aging. *Ann Neurol* 67: 122–31.
- 40. Fleisher AS, Chen K, Liu X, Ayutyanont N, Roontiva A, Thiyyagura P, *et al.* (2013): Apolipoprotein E ε4 and age effects on florbetapir positron emission tomography in healthy aging and Alzheimer disease. *Neurobiol Aging* 34: 1–12.
- 41. Scheinin NM, Wikman K, Jula A, Perola M, Vahlberg T, Rokka J, *et al.* (2014): Cortical ¹¹C-PIB uptake is associated with age, APOE genotype, and gender in "healthy aging". *J Alzheimers Dis* 41: 193–202.
- 42. Knopman DS, Jack CR, Wiste HJ, Lundt ES, Weigand SD, Vemuri P, *et al.* (2014): 18Ffluorodeoxyglucose positron emission tomography, aging, and apolipoprotein E genotype in cognitively normal persons. *Neurobiol Aging* 35: 2096–106.
- 43. Reiman EM, Caselli RJ, Yun LS, Chen K, Bandy D, Minoshima S, *et al.* (1996): Preclinical evidence of Alzheimer's disease in persons homozygous for the epsilon 4 allele for apolipoprotein E. *N Engl J Med* 334: 752–8.
- 44. Lowe VJ, Weigand SD, Senjem ML, Vemuri P, Jordan L, Kantarci K, *et al.* (2014): Association of hypometabolism and amyloid levels in aging, normal subjects. *Neurology*. doi: 10.1212/WNL.00000000000467.
- 45. Riese F, Gietl A, Zölch N, Henning A, O'Gorman R, Kälin AM, *et al.* (2015): Posterior cingulate γ-aminobutyric acid and glutamate/glutamine are reduced in amnestic mild cognitive impairment and are unrelated to amyloid deposition and apolipoprotein E genotype. *Neurobiol Aging* 36: 53–9.
- 46. Gomar JJ, Gordon ML, Dickinson D, Kingsley PB, Uluğ AM, Keehlisen L, *et al.* (2014): APOE genotype modulates proton magnetic resonance spectroscopy metabolites in the aging brain. *Biol Psychiatry* 75: 686–92.
- 47. Kantarci K, Smith GE, Ivnik RJ, Petersen RC, Boeve BF, Knopman DS, *et al.* (2002): 1H magnetic resonance spectroscopy, cognitive function, and apolipoprotein E genotype in normal aging, mild cognitive impairment and Alzheimer's disease. *J Int Neuropsychol Soc* 8: 934–42.
- Laakso MP, Hiltunen Y, Könönen M, Kivipelto M, Koivisto A, Hallikainen M, Soininen H (2003): Decreased brain creatine levels in elderly apolipoprotein E epsilon 4 carriers. J Neural Transm 110: 267–75.
- 49. Spencer DC, Zitzelberger T, Spielman D, Kaye J (2003): MRS in relation to hippocampal volume in the oldest old. *Neurology* 60: 1194–6.

- 50. Tebar F, Bohlander SK, Sorkin A (1999): Clathrin assembly lymphoid myeloid leukemia (CALM) protein: localization in endocytic-coated pits, interactions with clathrin, and the impact of overexpression on clathrin-mediated traffic. *Mol Biol Cell* 10: 2687–702.
- 51. Morgan K (2011): The three new pathways leading to Alzheimer's disease. *Neuropathol Appl Neurobiol* 37: 353–7.
- 52. Seshadri S, Fitzpatrick AL, Ikram MA, DeStefano AL, Gudnason V, Boada M, *et al.* (2010): Genome-wide analysis of genetic loci associated with Alzheimer disease. *JAMA* 303: 1832–40.
- 53. Jun G, Naj AC, Beecham GW, Wang L-S, Buros J, Gallins PJ, *et al.* (2010): Meta-analysis confirms CR1, CLU, and PICALM as alzheimer disease risk loci and reveals interactions with APOE genotypes. *Arch Neurol* 67: 1473–84.
- Biffi A, Anderson CD, Desikan RS, Sabuncu M, Cortellini L, Schmansky N, et al. (2010): Genetic variation and neuroimaging measures in Alzheimer disease. Arch Neurol 67: 677– 85.
- 55. Furney SJ, Simmons A, Breen G, Pedroso I, Lunnon K, Proitsi P, *et al.* (2011): Genomewide association with MRI atrophy measures as a quantitative trait locus for Alzheimer's disease. *Mol Psychiatry* 16: 1130–8.
- 56. Bralten J, Franke B, Arias-Vásquez A, Heister A, Brunner HG, Fernández G, Rijpkema M (2011): CR1 genotype is associated with entorhinal cortex volume in young healthy adults. *Neurobiol Aging* 32: 2106.e7–11.
- 57. Zhang P, Qin W, Wang D, Liu B, Zhang Y, Jiang T, Yu C (2014): Impacts of PICALM and CLU variants associated with Alzheimer's disease on the functional connectivity of the hippocampus in healthy young adults. *Brain Struct Funct*. doi: 10.1007/s00429-014-0738-4.
- 58. Hohman TJ, Koran ME, Thornton-Wells T (2013): Epistatic genetic effects among Alzheimer's candidate genes. *PLoS One* 8: e80839.
- 59. Singhrao SK, Neal JW, Rushmere NK, Morgan BP, Gasque P (1999): Differential expression of individual complement regulators in the brain and choroid plexus. *Lab Invest* 79: 1247–59.
- 60. Morgan K, Carrasquillo MM (2013): *Genetic Variants in Alzheimer's Disease*. New York: Springer Science+Business Media.
- 61. Hensley K (2010): Neuroinflammation in Alzheimer's disease: mechanisms, pathologic consequences, and potential for therapeutic manipulation. *J Alzheimers Dis* 21: 1–14.
- 62. Bralten J, Franke B, Arias-Vásquez A, Heister A, Brunner HG, Fernández G, Rijpkema M (2011): CR1 genotype is associated with entorhinal cortex volume in young healthy adults. *Neurobiol Aging* 32: 2106.e7–11.

- 63. Pant S, Sharma M, Patel K, Caplan S, Carr CM, Grant BD (2009): AMPH-1/Amphiphysin/Bin1 functions with RME-1/Ehd1 in endocytic recycling. *Nat Cell Biol* 11: 1399–410.
- 64. Vasiliou V, Vasiliou K, Nebert DW (2009): Human ATP-binding cassette (ABC) transporter family. *Hum Genomics* 3: 281–90.
- 65. Hirsch-Reinshagen V, Zhou S, Burgess BL, Bernier L, McIsaac SA, Chan JY, *et al.* (2004): Deficiency of ABCA1 impairs apolipoprotein E metabolism in brain. *J Biol Chem* 279: 41197–207.
- 66. Hughes TM, Lopez OL, Evans RW, Kamboh MI, Williamson JD, Klunk WE, *et al.* (2014): Markers of cholesterol transport are associated with amyloid deposition in the brain. *Neurobiol Aging* 35: 802–7.
- 67. Hirai H, Maru Y, Hagiwara K, Nishida J, Takaku F (1987): A novel putative tyrosine kinase receptor encoded by the eph gene. *Science* 238: 1717–20.
- 68. Chen Y, Fu AKY, Ip NY (2012): Eph receptors at synapses: implications in neurodegenerative diseases. *Cell Signal* 24: 606–11.
- 69. Nakamura-Hirota T, Kadoyama K, Takano M, Otani M, Matsuyama S (2012): The expression changes of EphA3 receptor during synaptic plasticity in mouse hippocampus through activation of nicotinic acetylcholine receptor. *Neuroreport* 23: 746–51.
- 70. Crocker PR, Paulson JC, Varki A (2007): Siglecs and their roles in the immune system. *Nat Rev Immunol* 7: 255–66.
- 71. Bradshaw EM, Chibnik LB, Keenan BT, Ottoboni L, Raj T, Tang A, *et al.* (2013): CD33 Alzheimer's disease locus: altered monocyte function and amyloid biology. *Nat Neurosci* 16: 848–50.
- 72. Hardoon DR, Shawe-Taylor J (2010): Sparse canonical correlation analysis. *Mach Learn* 83: 331–353.
- 73. Wan J, Kim S, Inlow M, Nho K, Swaminathan S, Risacheri SL, *et al.* (2011): Hippocampal surface mapping of genetic risk factors in AD via sparse learning models. *Med Image Comput Comput Assist Interv* 14: 376–83.
- 74. Yan J, Du L, Kim S, Risacher SL, Huang H, Moore JH, *et al.* (2014): Transcriptome-guided amyloid imaging genetic analysis via a novel structured sparse learning algorithm. *Bioinformatics* 30: i564–71.
- 75. Du L, Jingwen Y, Kim S, Risacher SL, Huang H, Inlow M, *et al.* (2014): A novel structureaware sparse learning algorithm for brain imaging genetics. *Med Image Comput Comput Assist Interv* 17: 329–36.

- Mahley RW, Weisgraber KH, Huang Y (2009): Apolipoprotein E: structure determines function, from atherosclerosis to Alzheimer's disease to AIDS. *J Lipid Res* 50 Suppl: S183– 8.
- 77. Chen H-K, Liu Z, Meyer-Franke A, Brodbeck J, Miranda RD, McGuire JG, *et al.* (2012): Small molecule structure correctors abolish detrimental effects of apolipoprotein E4 in cultured neurons. *J Biol Chem* 287: 5253–66.
- 78. Cirulli ET, Goldstein DB (2010): Uncovering the roles of rare variants in common disease through whole-genome sequencing. *Nat Rev Genet* 11: Nature Publishing Group415–25.
- 79. Saykin AJ, Shen L, Yao X, Kim S, Nho K, Risacher SL, *et al.* (2015): Genetic studies of quantitative MCI and AD phenotypes in ADNI: Progress, opportunities, and plans. *Alzheimers Dement* 11: 792–814.
- 80. Marigorta UM, Navarro A (2013): High trans-ethnic replicability of GWAS results implies common causal variants. *PLoS Genet* 9: e1003566.
- 81. Reitz C, Jun G, Naj A, Rajbhandary R, Vardarajan BN, Wang L-S, *et al.* (2013): Variants in the ATP-binding cassette transporter (ABCA7), apolipoprotein E ε4,and the risk of lateonset Alzheimer disease in African Americans. *JAMA* 309: American Medical Association1483–92.
- 82. Reitz C, Mayeux R (2014): Genetics of Alzheimer's disease in Caribbean Hispanic and African American populations. *Biol Psychiatry* 75: 534–41.
- 83. Tian C, Gregersen PK, Seldin MF (2008): Accounting for ancestry: population substructure and genome-wide association studies. *Hum Mol Genet* 17: R143–50.
- 84. Yu J-T, Li L, Zhu Q-X, Zhang Q, Zhang W, Wu Z-C, *et al.* (2010): Implication of CLU gene polymorphisms in Chinese patients with Alzheimer's disease. *Clin Chim Acta* 411: 1516–9.
- Chen LH, Kao PYP, Fan YH, Ho DTY, Chan CSY, Yik PY, et al. (2012): Polymorphisms of CR1, CLU and PICALM confer susceptibility of Alzheimer's disease in a southern Chinese population. *Neurobiol Aging* 33: 210.e1–7.
- 86. Gurland BJ, Wilder DE, Lantigua R, Stern Y, Chen J, Killeffer EH, Mayeux R (1999): Rates of dementia in three ethnoracial groups. *Int J Geriatr Psychiatry* 14: 481–93.
- Button KS, Ioannidis JPA, Mokrysz C, Nosek BA, Flint J, Robinson ESJ, Munafò MR (2013): Power failure: why small sample size undermines the reliability of neuroscience. *Nat Rev Neurosci* 14: 365–76.
- 88. Poline J-B, Breeze JL, Ghosh S, Gorgolewski K, Halchenko YO, Hanke M, *et al.* (2012): Data sharing in neuroimaging research. *Front Neuroinform* 6: 9.