

The American Journal of Human Genetics

Supplemental Data

## **Small 6q16.1 Deletions Encompassing *POU3F2***

### **Cause Susceptibility to Obesity and Variable**

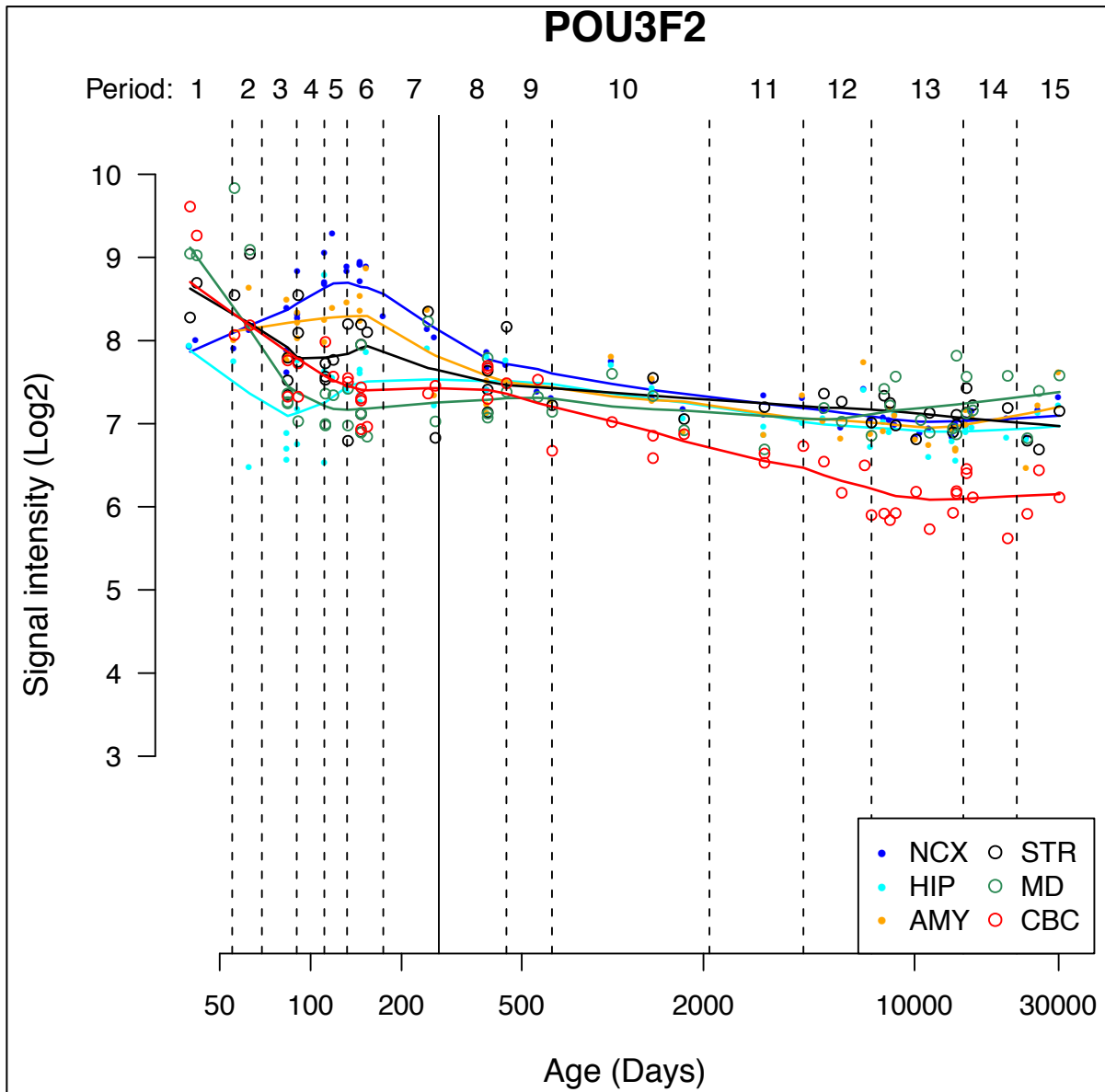
### **Developmental Delay with Intellectual Disability**

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**Figure S1. *POU3F2* is highly expressed throughout fetal and adult human brain.**

Figure generated from Human Brain Transcriptome project data (<http://hbatlas.org/>)<sup>1</sup> showing high spatio-temporal expression of *POU3F2*.

The legends and axes are labeled and are self-explanatory.



**Key:** NCX neocortex; HIP hippocampus; AMY amygdala; STR striatum; MD Mediodorsal nucleus of the thalamus; CBC cerebellar cortex.

## Figure S2: High level of expression of *POU3F2* and related genes in the human hypothalamus.

These figures were generated using Brain Explorer tool based on data from the Allen Human Brain Atlas (<http://human.brain-map.org/>)<sup>2</sup> in following steps –

- (i) Brain Explorer tool was downloaded from <http://human.brain-map.org/static/brainexplorer> and installed on a local computer.
- (ii) Allen Human Brain Atlas (37MB) – October 2013 was installed from <http://www.brain-map.org>
- (iii) Gene name was entered in the search box at URL - <http://human.brain-map.org/microarray/search> (with settings: Resolution = structures; Color Map = z-score)
- (iv) Relevant heat map was selected by clicking that activates the Brain Explorer link on the webpage.
- (v) Clicking on the activated link Brain Explorer link opens the Application and enables visualization of spatial rendition of gene expression.
- (vi) Within the application specific brain structures of interest were selected to be highlighted and to restrict the visualized expression points (in this case the hypothalamus and the hippocampal formation).
- (vii) Various brain sections were visualized and a representative image was selected and exported as an image file.

Figures are arranged to represent the Leptin>MC4R>SIM1>POU3F2>OXT pathway and show that all five genes are highly expressed in the human hypothalamus. The bottom left figure, represents expression pattern of *POU3F3*. Comparing expression patterns of human *POU3F3* and *POU3F2* shows that it is similar to what has been observed in mice<sup>3</sup>.

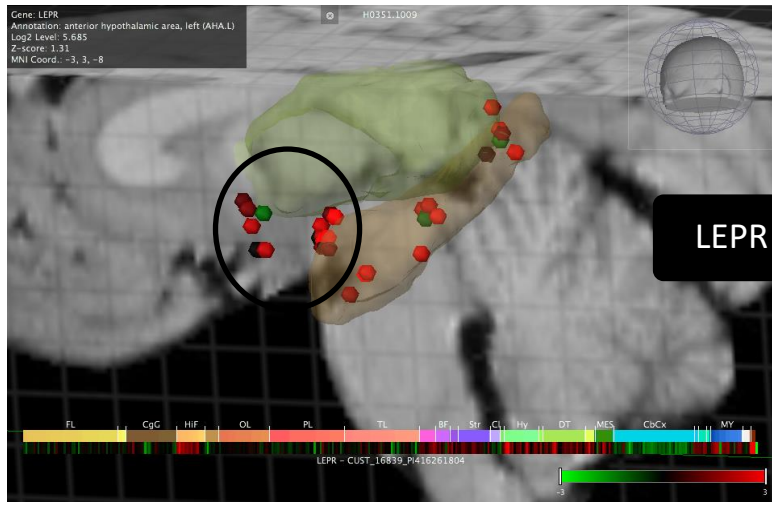
*In all the figures the inset at the top left corner provides the gene name, anatomical centre of the figure, Log2 level, Z-score and co-ordinates.*

*The top right corner inset shows a 'compass' to aid orientation of the main figure.*

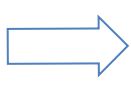
*The hypothalamic area is circled, transparent green area represents the thalamus and transparent orange area represents the hippocampal formation.*

*The expression level at each measured point is given on a green-red scale (bottom right). The two horizontal bars at the bottom provide more detailed information on the expression of the particular gene via the green-red heat map in each of the following regions of the brain – FL (frontal lobe), CgC (cingulate gyrus), HiF (hippocampal formation), OL (occipital lobe), PL (parietal lobe), TL (temporal lobe), BF (basal forebrain), Str (striatum), C (claustrum), Hy (hypothalamus), DT (dentate nucleus), MES (mesencephalon), CbCx (cerebellar cortex) and My (myelencephalon).*

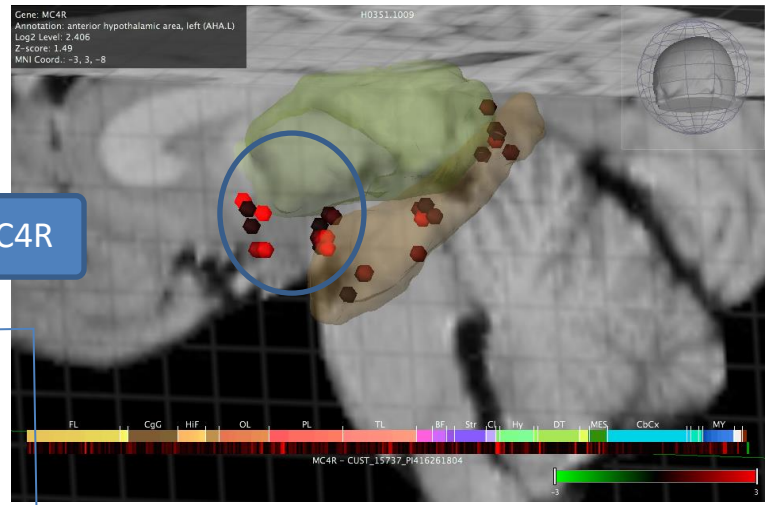
*The anatomical section and the donor sample identifiers are provided at the top centre and bottom centre respectively.*



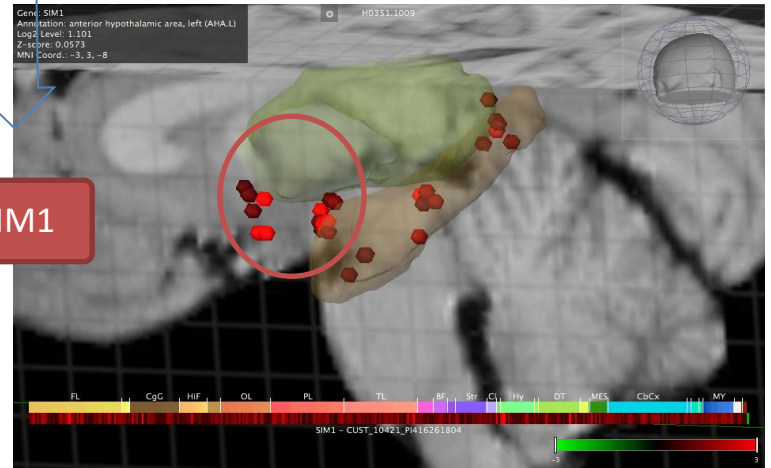
LEPR



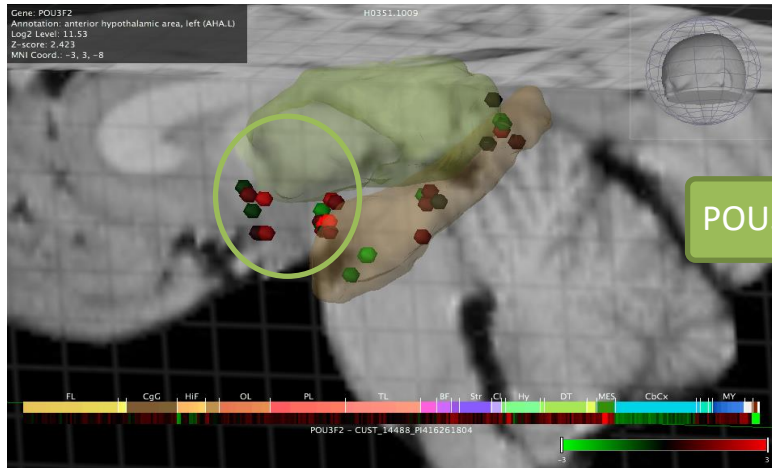
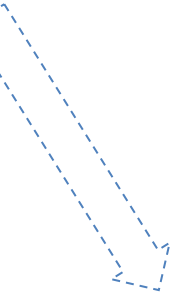
MC4R



SIM1

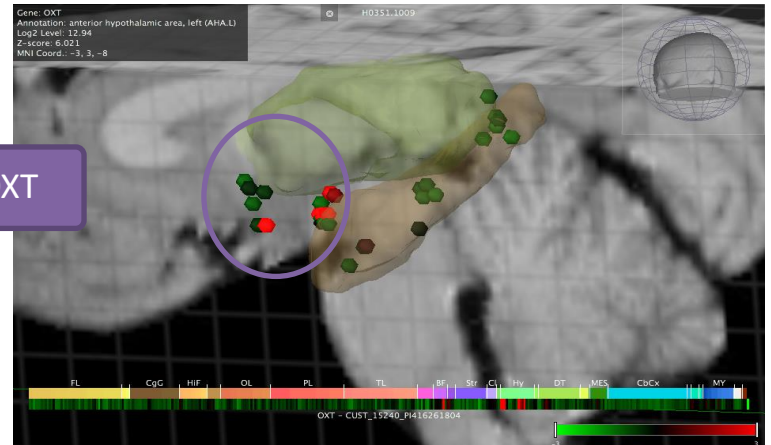
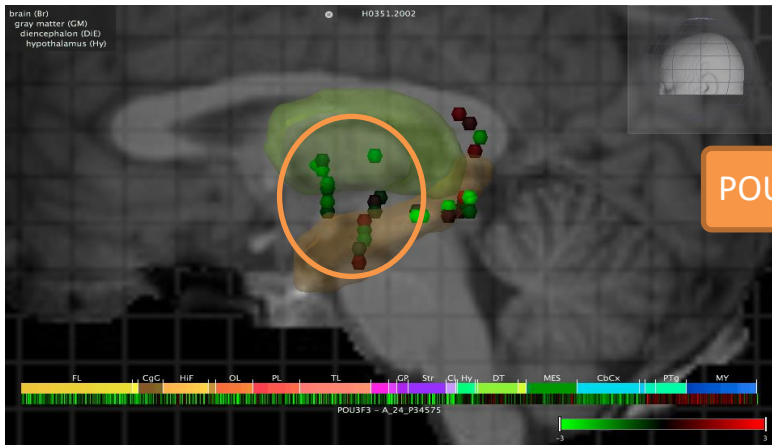


POU3F2



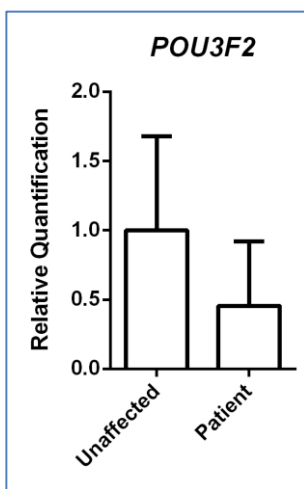
POU3F3

OXT



### Figure S3: *POU3F2* quantitative reverse transcription PCR in patient cell lines.

*POU3F2* expression was assessed in Epstein-Barr virus transformed lymphoblastoid cell lines (LCLs) derived from a patient and unaffected sibling (Family 1). LCLs were maintained as described previously<sup>4</sup>. Total RNA was extracted from LCLs ( $10 \times 10^6$  cells per sample) using TRIzol reagent (Thermo Fisher). RNA concentration was assessed using a spectrophotometer (FLUOstar Omega, Labtech). cDNA was synthesised from 1600ng RNA using the High Capacity RNA to cDNA kit (Life Technologies, UK). Quantitative reverse transcription PCR (qRT-PCR) analysis was performed using the TaqMan Universal PCR Master Mix (Applied Biosystems). Each PCR was performed three times in triplicate per cDNA sample (total of 9 technical repeats per sample). The relative abundance of *POU3F2* transcript was measured using a *POU3F2* Taqman probe (Hs00271595\_s1) following normalisation to the expression level of two 'housekeeper' genes, *HPRT1* (Hs03929096\_g1) and *18s* (Hs999999001\_s1), and assessed with the Applied Biosystems StepOne Software v2.1. Statistical significance between groups was determined by t-tests using DataAssist v2.0 (Applied Biosystems). Fold change values were plotted using Graphpad Prism v5.0 and defined as the Relative Quantification (RQ).



An approximate 50% reduction in *POU3F2* expression in LCLs derived from patient in comparison to his unaffected sibling ( $p=0.1447$ ).

Amplification of the *POU3F2* transcript occurred at very late cycle numbers, suggesting that this gene is expressed at very low levels in peripheral lymphocytes.

Indeed, we hypothesise that low expression is the most likely cause for the high variation observed, even after 9 technical repeats and using a high concentration of cDNA template.

## Figure S4: Human and zebrafish POU3F2 genes are highly conserved.

Protein sequence alignment was created using Colbalt Constraint-based Multiple Protein Alignment Tool. The sequences correspond to zebrafish Pou3f2a (NP\_571364), zebrafish Pou3f2b (NP\_571235.1), human POU3F2 (accession number: NP\_005595), and mouse POU3F2\* (accession number: NP\_032925.1), human POU3F1 (NP\_002690), human POU3F3 (NP\_006227), and human POU3F4 (NP\_000298).

The entire protein sequence is highly conserved among mice, humans, and both orthologs in zebrafish. The POU homeodomain (amino acids 95-251; Pou3f2a sequence), highlighted in red, has 92% identity and 97% similarity for all seven sequences.

FIGURE 3

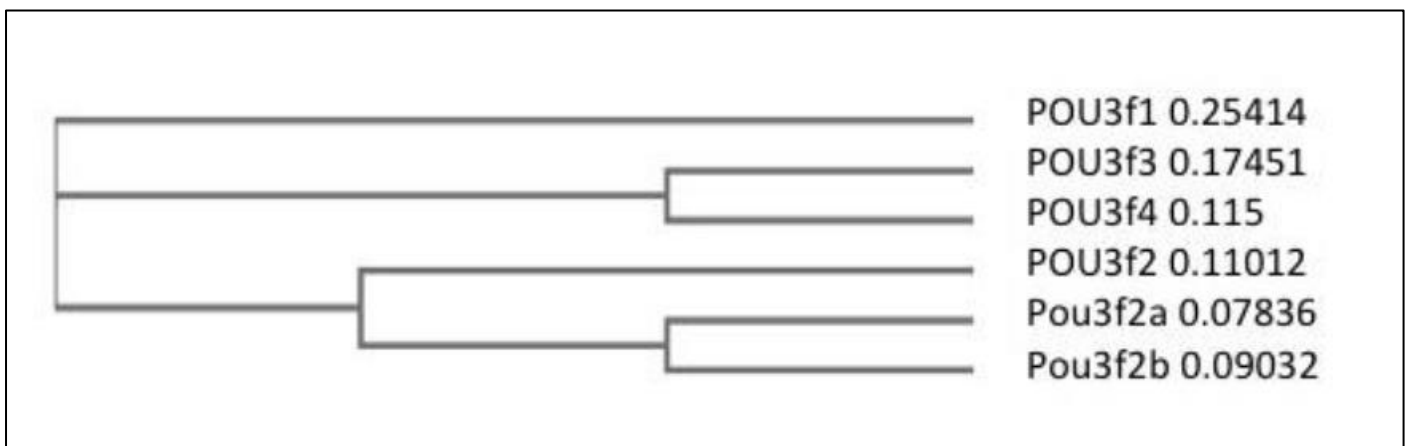
Pou3f2a1	---	LSH---			-----GgEGSPWPA	SPLGEQDIKPV-E---	E-----LQ	H	28	
Pou3f2b1	---	ALSH---			-----G-EGGPWSS	SPLGEQDIKPA-V---	QSPR-DEMHNSSLQ	H	39	
POU3F2 1	wit	ALSHGGG	GGGGGGGGGGG	GGGGGGG-DGSPWST	SPLGQPDIKPS-VvVQQGGRGDELHGPGALQ			Q	66	
POU3F2*1	wit	ALSHGGG	GGGGGGGGGGG	GGGGGGG-DGSPWST	SPLGQPDIKPS-VvVQQGGRGDELHGPGALQ			Q	66	
POU3F1 1	---	GAGHPVG [9]	GGGGGGDWAGG [5]	GKAGGGG-TGR----		-----ADGGGGGGFHA		-	54	
POU3F3 1	wit	ALSHGGG	GGGGGGGGGGG	GGGGGGG-DGSPWST	SPLGQPDIKPS-VvVQQGGRGDELHGPGALQ			Q	66	
POU3F4 1	---	-----	-----	-----LS-DGGPWSS [4]	SPLDQDQVVKPGrE----	D-----LQ [7]	R	37		
Pou3f2a29	S	P--	RQAHLVPSQ--	HHETAAWRATTTA-HMP-SMSTSNQqSL	IYSQP---		-----GY--G--EMH	77		
Pou3f2b40	Q	S--	RPPHLVHQTHGNHDSRAWRTTTAA-HIP-SMATSNQqSL	IYSQPSFS	VNGLIPGSGQ--G--IHH	101				
POU3F2 67	Q [21]	Q--	RPPHLVHHAANHHHPGPGAWRSAAAAAHLPPSMGASNG-GL	LYSQPSFT	VNGMLGAGGQPAG--LHH	152				
POU3F2*67	Q [21]	QqQ	RPPHLVHHAANHHHPGPGAWRSAAAAAHLPPSMGASNG-GL	LYSQPSFT	VNGMLGAGGQPAG--LHH	154				
POU3F1 55	-	-----	RLVHQGAH--	AGAAWAQGSTAHHLGPAMSPSPG-AS [9]	LYAQAAYP [6]	LAGMLAAGGGGAGpgLHH	129			
POU3F3 67	Q [21]	Q--	RPPHLVHHAANHHHPGPGAWRSAAAAAHLPPSMGASNG-GL	LYSQPSFT	VNGMLGAGGQPAG--LHH	152				
POU3F4 38	S	P--	HVAHHSPTHN--	HPN--AWGASPAP-N-P-SI-TSSGqPL [1]	VYSQP---	-----GF--T--VSG	83			
Pou3f2a78	H-----	EEhhs	PHLSE---HG	HPQSLH-----				<b>SDEDTPTSDDLEQFAKQF</b>	114	
Pou3f2b102	HSMRDAHEDhhs	PHLSD---HG	HPPSQHQHQSHQ-----	SHHD--				<b>HSDSDTPTSDDLEQFAKQF</b>	155	
POU3F2 153	HGLRDAHDE---	PHHAD---HH	PHPHSHPHQPPPPPPPPQPPGHPGAHHD--	PHSDSDTPTSDDLEQFAKQF					217	
POU3F2*155	HGLRDAHDE---	PHHAD---HH	PHPHSHPHQPPPPPPPPQPPGHPGAHHD--	PHSDSDTPTSDDLEQFAKQF					219	
POU3F1 130	ALHEDGHEA--Q [ 5]	PPHLG---AH	GHAHGHAHAGGLHAAAAHLHPGAGGGSSvgEHS	<b>SDEDA PSSDDLEQFAKQF</b>					202	
POU3F3 153	HGLRDAHDE---	PHHAD---HH	PHPHSHPHQPPPPPPPPQPPGHPGAHHD--	PHSDSDTPTSDDLEQFAKQF					217	
POU3F4 84	M-----	LEhgG [14]	PVLR EppdHG [4]	HHCQDH-----					<b>SDEETPTSDELEQFAKQF</b>	141
Pou3f2a115	<b>KQRRIKLGFTQADVGLALGTLYGNVFSQTTICRFEALQLSFKNMCKLKPLLNKWLEEDSTSGSPTS</b>	<b>SLDKIAAQGRKRKK</b>	194							
Pou3f2b156	<b>KQRRIKLGFTQADVGLALGTLYGNVFSQTTICRFEALQLSFKNMCKLKPLLNKWLEEDSTSGSPTS</b>	<b>SLDKIAAQGRKRKK</b>	235							
POU3F2 218	<b>KQRRIKLGFTQADVGLALGTLYGNVFSQTTICRFEALQLSFKNMCKLKPLLNKWLEEDSSSGSPTS</b>	<b>IDKIAAQGRKRKK</b>	297							
POU3F2*220	<b>KQRRIKLGFTQADVGLALGTLYGNVFSQTTICRFEALQLSFKNMCKLKPLLNKWLEEDSSSGSPTS</b>	<b>IDKIAAQGRKRKK</b>	299							
POU3F1 203	<b>KQRRIKLGFTQADVGLALGTLYGNVFSQTTICRFEALQLSFKNMCKLKPLLNKWLEETDSSSGSPT</b>	<b>NLDKIAAQGRKRKK</b>	282							
POU3F3 218	<b>KQRRIKLGFTQADVGLALGTLYGNVFSQTTICRFEALQLSFKNMCKLKPLLNKWLEEDSSSGSPTS</b>	<b>IDKIAAQGRKRKK</b>	297							
POU3F4 142	<b>KQRRIKLGFTQADVGLALGTLYGNVFSQTTICRFEALQLSFKNMCKLKPLLNKWLEEDSSTGSPTS</b>	<b>IDKIAAQGRKRKK</b>	221							
Pou3f2a195	<b>RTSIEVSVKGALESHFLKCPKPGASEINSLADSLQLEKEVVRVWFCNRRQKEKRMT</b>	<b>PPNGP-MAGSE</b>	<b>DVY</b>	G---DAS	267					
Pou3f2b236	<b>RTSIEVSVKGALESHFLKCPKPAASEITSLADSLQLEKEVVRVWFCNRRQKEKRMT</b>	<b>PPGGP-LPGTE</b>	<b>DVY</b>	G---DTP	308					
POU3F2 298	<b>RTSIEVSVKGALESHFLKCPKPSAQEITSLADSLQLEKEVVRVWFCNRRQKEKRMT</b>	<b>PPGGT-LPGA</b>	<b>EDVY</b>	GGSRDTP	373					
POU3F2*300	<b>RTSIEVSVKGALESHFLKCPKPSAQEITSLADSLQLEKEVVRVWFCNRRQKEKRMT</b>	<b>PPGGT-LPGA</b>	<b>EDVY</b>	GGSRDTP	375					
POU3F1 283	<b>RTSIEVSVKGALESHFLKCPKPSAQEITGLADSLQLEKEVVRVWFCNRRQKEKRMT</b>	<b>PAAGAgHPPM</b>	<b>DVY [7]</b>	GGGGASP	366					
POU3F3 298	<b>RTSIEVSVKGALESHFLKCPKPSAQEITSLADSLQLEKEVVRVWFCNRRQKEKRMT</b>	<b>PPGGT-LPGA</b>	<b>EDVY</b>	GGSRDTP	373					
POU3F4 222	<b>RTSIEVSVKGVLETHFLKCPKPAQEISSLADSLQLEKEVVRVWFCNRRQKEKRMT</b>	<b>PP-----</b>		G---DQQ	283					
Pou3f2a268	P	H	HGAQTPVP	277						
Pou3f2b309	P	H	HGVQTPVQ	318						
POU3F2 374	P	H	HGVQTPVQ	383						
POU3F2*376	P	H	HGVQTPVQ	385						
POU3F1 367	P [11]	H [4]	HTLPGSVQ	391						
POU3F3 374	P	H	HGVQTPVQ	383						
POU3F4 284	P	H [4]	HTVKTDTS [4]	301						

\*POU3F2 mouse

**Figure S5: Human POU3F2 is orthologous with zebrafish Pou3f2a and Pou3f2b.**

A phylogeny tree was created using MUSCLE Multiple Sequence Alignment. Protein sequences for all four human POU3 family proteins, POU3F1 (NP\_002690), POU3F2 (NP\_005595), POU3F3 (NP\_006227), POU3F4 (NP\_000298), as well as sequences for zebrafish Pou3f2a (accession number: NP\_571364) and Pou3f2b (accession number: NP\_571235.1), were used as input data. The output data cluster human POU3F2 more closely with zebrafish pou3f2a and pou3f2b than the other POU3 family proteins confirming that these genes are orthologous.

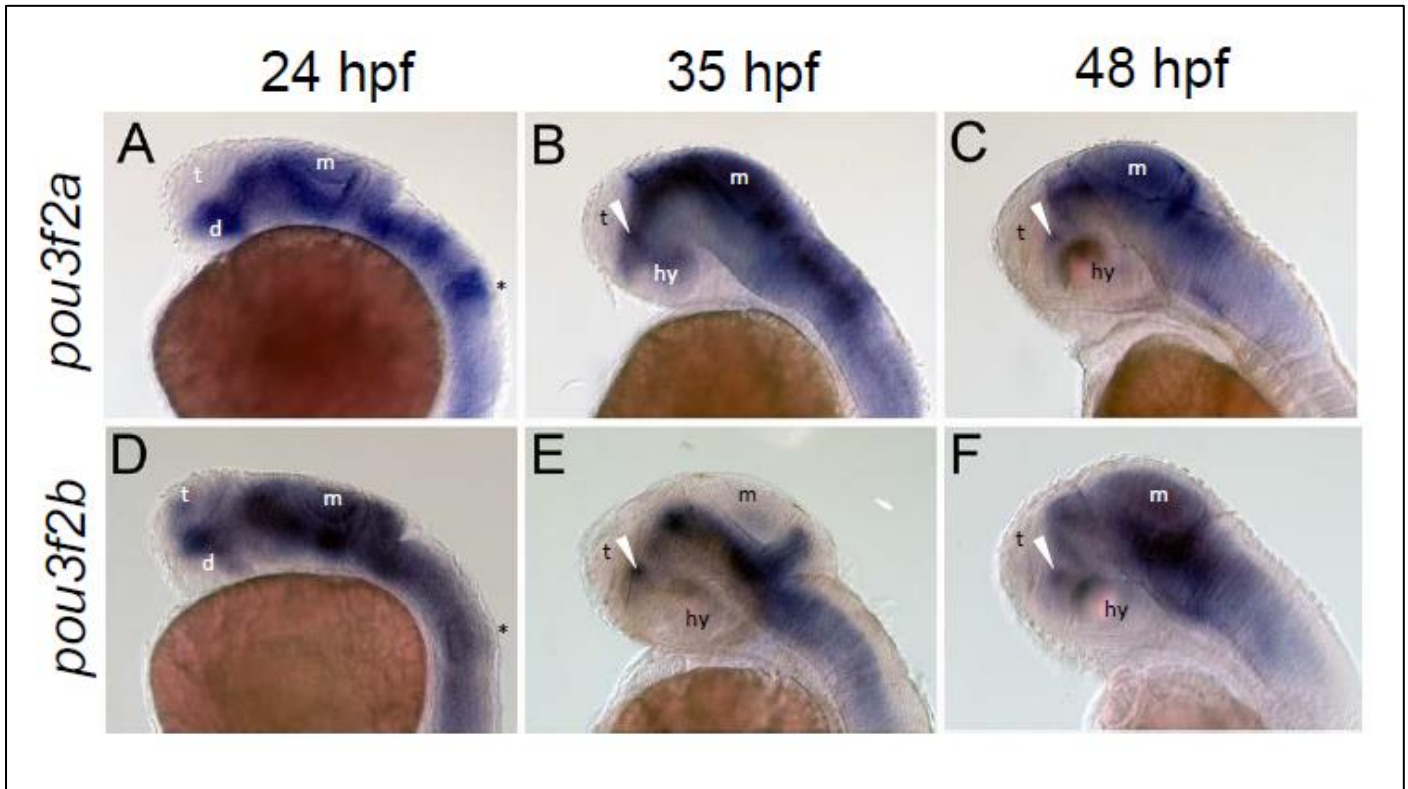
Human *POU3F2*, and zebrafish *pou3f2a*, and *pou3f2b* contain single protein coding exons, as well as conserved synteny (Synteny Database, <http://syntenydb.uoregon.edu>) (data not shown). The conserved synteny of human *POU3F2* with both *pou3f2a* and *pou3f2b* gene regions indicates that chromosomal duplication likely generated the zebrafish *pou3f2* paralogs.



**Figure S6: Developmental expression patterning of zebrafish *pou3f2a* and *pou3f2b*.**

The expression patterns of *pou3f2a* and *pou3f2b* mRNAs were determined by Whole mount in situ hybridization (WISH) at 24, 35, and 48 hours post fertilization (hpf).

Lateral views, with anterior left, and dorsal up.



Key: t, telencephalon; d, diencephalon; hy, hypothalamus; m, midbrain; \*, marks the position of rhombomere r5.

(A, D) At 24 hpf *pou3f2a* and *pou3f2b* are extensively expressed in the diencephalon, midbrain, and hindbrain. In the hindbrain, *pou3f2a* expression is concentrated in rhombomeres r1, r3 and r5.

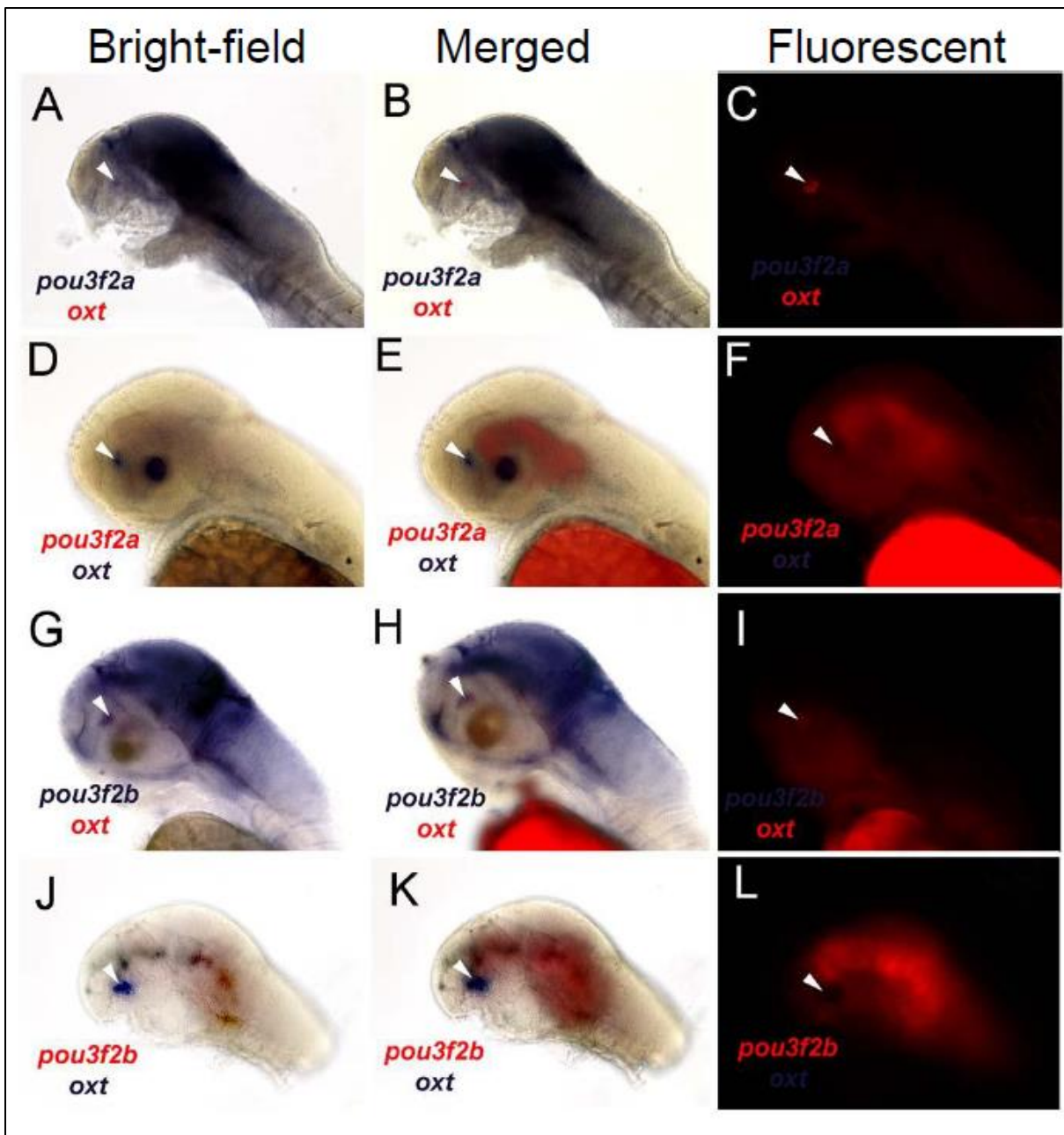
(B, E) By 35 hpf, *pou3f2a* and *pou3f2b* expression becomes weaker in the anterior diencephalon along the telencephalic-diencephalic border, and in the posterior hypothalamus. Within the anterior diencephalic domain, strong *pou3f2a* expression is maintained in the neuroendocrine preoptic area (NPO), indicated with white arrowheads.

(C, F) By 48 hpf, *pou3f2a* and *pou3f2b* expression is seen in the diencephalon, the midbrain tegmentum and throughout the hindbrain. In the diencephalon, expression further narrows and strong expression of *pou3f2a* and *pou3f2b* remains restricted to a small area of the NPO.



**Figure S7: Co-localization of *pou3f2a* and *pou3f2b* mRNAs with subsets of oxytocin cells.**

Co-expression of *pou3f2a*, *pou3f2b* and *oxt* mRNA was determined by double label WISH in 48 hpf embryos. *Pou3f2a* and *pou3f2b* probes were labelled with digoxigenin (DIG) while oxytocin probes were labeled with fluorescein (FL). DIG and FL were detected using either BM Purple or Red Fast Stain. The white arrowhead indicates the location of the *oxt* expression domain. (A, D, G, J) Bright field images (C, F, I, L) are the corresponding epifluorescence images, and (B, E, H, K) are the merged bright field and epifluorescence images.



(A-C) *pou3f2a* expression is visualized with purple staining and *oxt* expression is visualized with red staining. The eyes have been removed to better visualize the staining patterns. The purple *pou3f2a* staining quenches the red *oxt* stain, except in a small posterior region, of the *oxt* domain.

(D-F) The chromogenic staining is reversed such that *pou3f2a* expression is visualized with red staining and

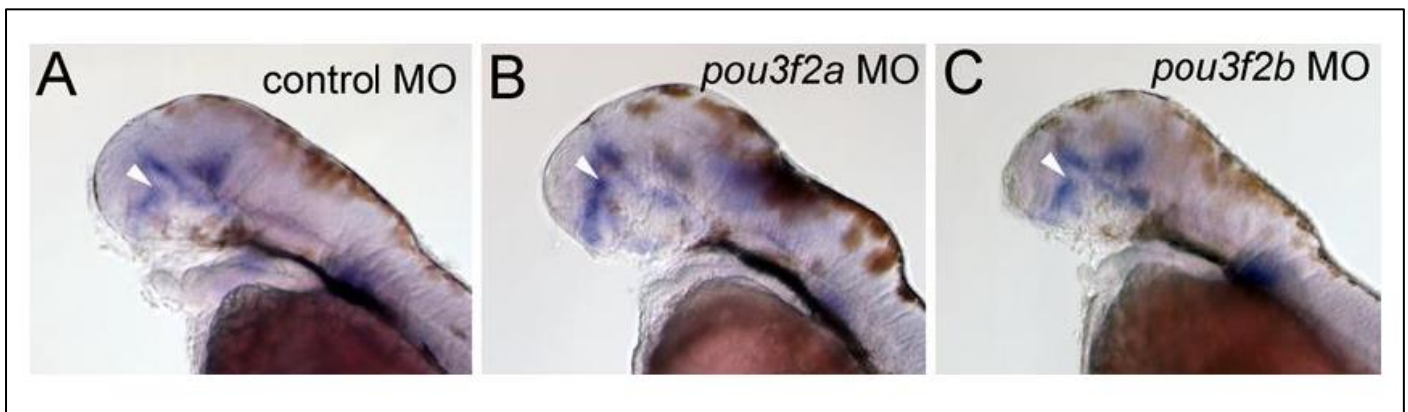
*oxl* expression is visualized with purple staining. The purple *oxl* stain quenches the entire red *pou3f2a* stain in this region. Thus, *pou3f2a* appears to be co-expressed in a subset of oxytocin cells.

(G-I) *pou3f2b* expression is visualized with purple staining and *oxl* expression is visualized with red staining. The purple *pou3f2b* staining quenches the red *oxl* stain of the *oxl* domain, except for in a small posterior region as seen with *pou3f2a*.

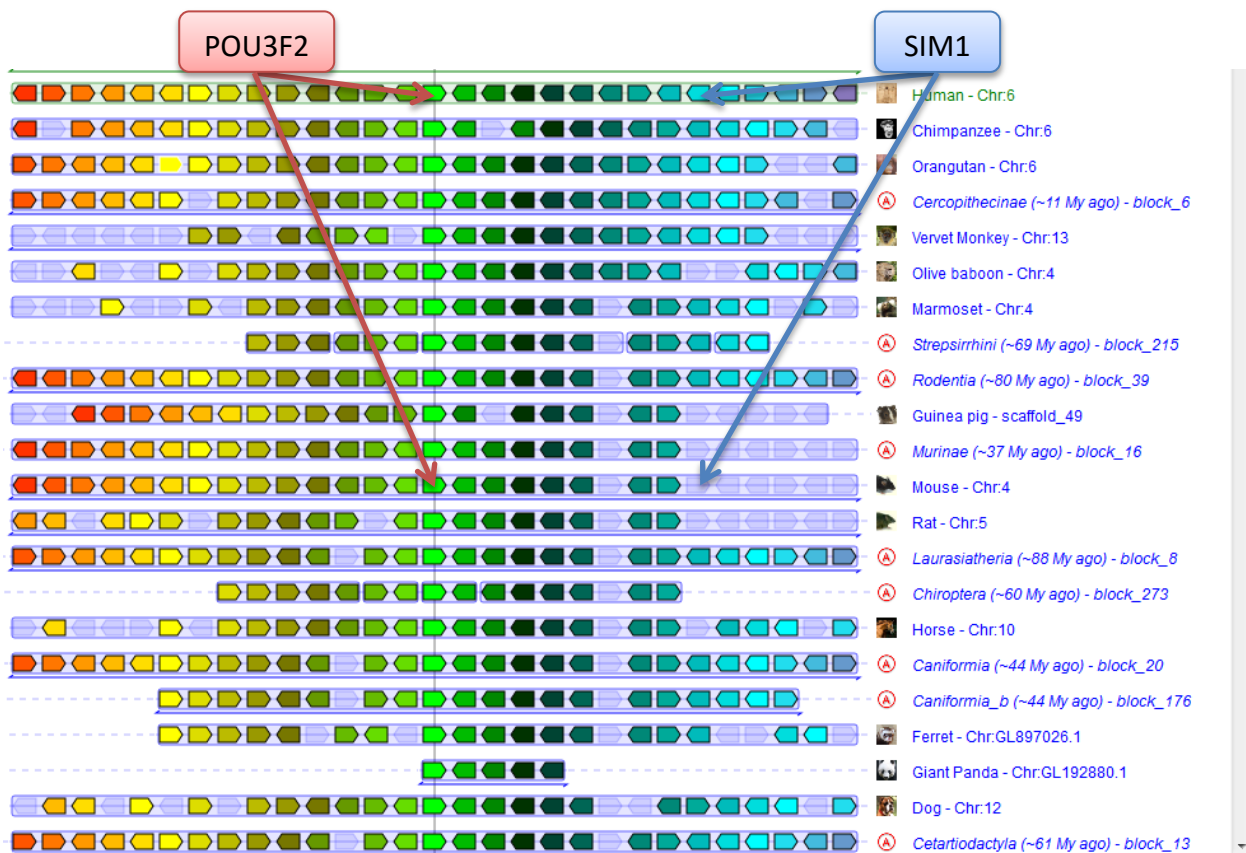
(J-L) The chromogenic staining is reversed such that *pou3f2b* expression is visualized with red staining and *oxl* expression is visualized with purple staining. The eyes have been removed to better visualize the staining patterns. The purple *oxl* stain quenches the entire red *pou3f2b* stain in this region. Thus, similar to *pou3f2a*, *pou3f2b* appears to be co-expressed in a subset of oxytocin cells.

**Figure S8: Knockdown of *pou3f2a* or *pou3f2b* has no effect on expression of *sim1a***

MO knockdown of *pou3f2a* or *pou3f2b* causes no obvious change in *sim1a* expression patterns. (A-C) Lateral views, 48 hpf embryos stained for *sim1a* expression by WISH. The location of the NPO is indicated with white arrowheads. (A) control MO injected embryos (B) *pou3f2a* MO injected embryo, (C) *pou3f2b* MO injected embryo.



**Figure S9: The evolutionary syntenic architecture of the 6q16.2 region shows that POU3F2 and SIM1 are not located on the same chromosome in a number of mammals.**



Snapshot from Genomicus genome browser. The figure is centred on POU3F2 gene which is indicated with a red box and arrow. SIM1 location and orientation is indicated by blue box and arrow. The right column gives the name of the species. Faded boxes indicate that the genes are not syntenically linked in the respective specie. Mouse Pou3f2 and Sim1 are highlighted demonstrating that unlike in humans, these two genes are on different chromosomes.



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1081 TCGGACGACCTGGAGCAGTTCGCCAAGCAGTTCAAGCAGCGGCGGATCAAACCTGGGATTT
799 TCGGACGACCTGGAGCAGTTCGCCAAGCAGTTCAAGCAGCGGCGGATCAAACCTGGGATTT
267 -S--D--D--L--E--Q--F--A--K--Q--F--K--Q--R--R--I--K--L--G--F-

1141 ACCCAAGCGGACGTGGGGCTGGCTCTGGGCACCCTGTATGGCAACGTGTTCTCGCAGACC
859 ACCCAAGCGGACGTGGGGCTGGCTCTGGGCACCCTGTATGGCAACGTGTTCTCGCAGACC
287 -T--Q--A--D--V--G--L--A--L--G--T--L--Y--G--N--V--F--S--Q--T-

1201 ACCATCTGCAGGTTTGGAGCCCTGCAGCTGAGCTTCAAGAACATGTGCAAGCTGAAGCCT
919 ACCATCTGCAGGTTTGGAGCCCTGCAGCTGAGCTTCAAGAACATGTGCAAGCTGAAGCCT
307 -T--I--C--R--F--E--A--L--Q--L--S--F--K--N--M--C--K--L--K--P-

1261 TTGTTGAACAAGTGGTTGGAGGAGGCGGACTCGTCCTCGGGCAGCCCCACGAGCATAGAC
979 TTGTTGAACAAGTGGTTGGAGGAGGCGGACTCGTCCTCGGGCAGCCCCACGAGCATAGAC
327 -L--L--N--K--W--L--E--E--A--D--S--S--S--G--S--P--T--S--I--D-

1321 AAGATCGCAGCGCAAGGGCGCAAGCGGAAAAAGCGGACCTCCATCGAGGTGAGCGTCAAG
1039 AAGATCGCAGCGCAAGGGCGCAAGCGGAAAAAGCGGACCTCCATCGAGGTGAGCGTCAAG
347 -K--I--A--A--Q--G--R--K--R--K--K--R--T--S--I--E--V--S--V--K-

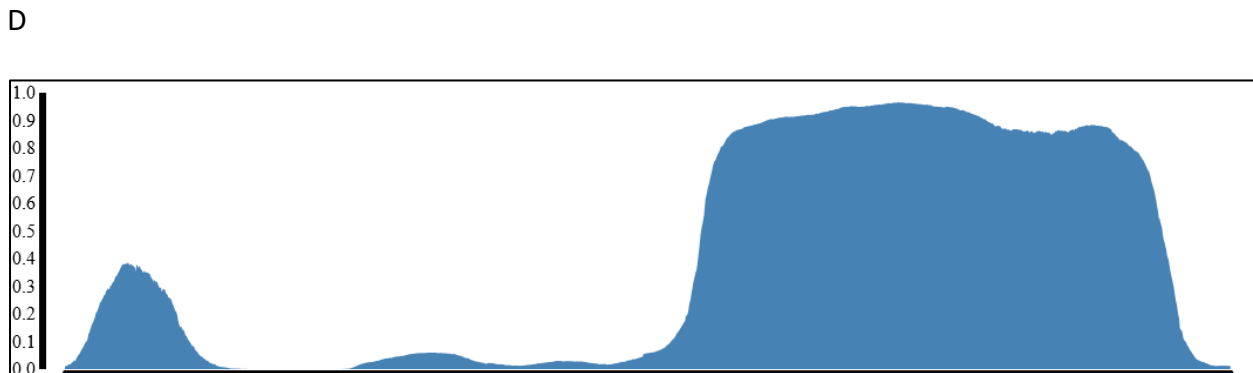
1381 GGGGCTCTGGAGAGCCATTTCTCAAAATGCCCAAGCCCTCGGCCAGGAGATCACCTCC
1099 GGGGCTCTGGAGAGCCATTTCTCAAAATGCCCAAGCCCTCGGCCAGGAGATCACCTCC
367 -G--A--L--E--S--H--F--L--K--C--P--K--P--S--A--Q--E--I--T--S-

1441 CTCGCGGACAGCTTACAGCTGGAGAAGGAGGTGGTGAGAGTTTGGTTTGTAAACAGGAGA
1159 CTCGCGGACAGCTTACAGCTGGAGAAGGAGGTGGTGAGAGTTTGGTTTGTAAACAGGAGA
387 -L--A--D--S--L--Q--L--E--K--E--V--V--R--V--W--F--C--N--R--R-

1501 CAGAAAGAGAAAAGGATGACCCTCCCGGAGGGACTCTGCCGGGCGCCGAGGATGTGTAC
1219 CAGAAAGAGAAAAGGATGACCCTCCCGGAGGGACTCTGCCGGGCGCCGAGGATGTGTAC
407 -Q--K--E--K--R--M--T--P--P--G--G--T--L--P--G--A--E--D--V--Y-

1561 GGGGGGAGTAGGGACACTCCACCACACCACGGGGTGCAGACGCCCGTCCAGTGAACTCGA
1279 GGGGGGAGTAGGGACACTCCACCACACCACGGGGTGCAGACGCCCGTCCAGTGA.....
427 -G--G--S--R--D--T--P--P--H--H--G--V--Q--T--P--V--Q--*--.....

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- Snapshot from Ensembl genome browser from Gene Summary page for *POU3F2* (*ENSG00000184486*) demonstrating that this gene has only one exon.
- POU3F2* cDNA sequence. The long repeats in the DNA sequence are highlighted.
- Domains of *POU3F2* protein
- ExAC coverage plot for *POU3F2*. X-axis denotes nucleotide positions (unnumbered) in *POU3F2* exon and y-axis number of individuals with more than 30X coverage for the corresponding nucleotide position on the X-axis.

**Table S1: Monogenic obesity syndromes in the Leptin>melanocortin>SIM1 neuro-endocrine pathway.**

Gene	Phenotype	OMIM	Reference
<i>LEP</i>	Morbid obesity due to leptin deficiency	614962	<sup>5</sup>
<i>LEPR</i>	Morbid obesity due to leptin receptor deficiency	614963	<sup>6,7</sup>
<i>POMC</i>	Obesity, adrenal insufficiency and red hair syndrome	609734	<sup>8</sup>
<i>PCSK1</i>	Obesity with impaired prohormone processing	600955	<sup>9</sup>
<i>MC4R</i>	Autosomal dominant obesity	601665	<sup>10,11</sup>
<i>MRAP2</i>	Susceptibility to obesity BMIQ18	615457	<sup>12</sup>
<i>SIM1</i>	Severe obesity	601665	<sup>13-15</sup>

**Table S2: List of genes within the deletion in Family 1 and properties of their variation.**

Table based on data accessed from <http://exac.broadinstitute.org/> on 15<sup>th</sup> September 2015.

Gene	Number of potentially truncating variants	Combined allele frequency of truncating variants	HI index	Associated disease (OMIM)	Inheritance pattern of the disease
<i>POU3F2</i>	0	0	15.18	None	Not applicable
<i>FBXL4</i>	11	$3.1 \times 10^{-4}$	10.28	Encephalomyopathic type mitochondrial DNA depletion syndrome (605654)	Autosomal recessive
<i>FAXC</i>	2	$1.7 \times 10^{-5}$	21.43	None	Not applicable
<i>COQ3</i>	19	$8.2 \times 10^{-4}$	40.90	None	Not applicable
<i>PNISR</i>	11	$2.2 \times 10^{-4}$	9.31	None	Not applicable
<i>USP45</i>	57	$1.9 \times 10^{-2}$	45.26	None	Not applicable
<i>TSTD3</i>	No data	No data	No data	None	Not applicable
<i>CCNC</i>	10	$3.8 \times 10^{-1}$	2.44	None	Not applicable
<i>PRDM13</i>	8	$8.1 \times 10^5$	56.20	None	Not applicable

**Table S3: Extent of 6q16 deletion in all the families studied in this project.**

Family #	Start co-ordinate (Hg19)	End co-ordinate (Hg19)	Size (Mb)
1	chr6:99218535	chr6:100260996	1.04
2	chr6:98583798	chr6:99582590	0.99
3	chr6:98981807	chr6:100377741	1.39
4	chr6:98981812	chr6:100261128	1.28
5	chr6:99179231	chr6:100641841	1.46
6	chr6:99218553	chr6:100319568	1.10

**Table S4: Amino acid identities in the amino terminal region of POU3 family proteins.**

Outside of the homeodomain there is 59% identity between Pou3f2a and Pou3f2b, and 50% identity between POU3F2 and Pou3f2a or Pou3f2b. The amino acid identity among Pou3f2a/b and other members of the POU family is reduced in comparison.

POU family protein	Pou3f2a	Pou3f2b
zf_Pou3f2b	59%	-
Hu_POU3F1	35%	30%
Hu_POU3F2	50%	50%
Hu_POU3F3	42%	28%
Hu_POU3F4	47%	45%

**Table S5: pou3f2a/b MO sequences.**

pou3f2a 5'	AGTGGTTGGACGCCGCGGTCGCC <u>CA</u> T
control-pou3f2a 5'	AGTcGTTGcACcCCGCcGTCc <u>CA</u> T
pou3f2b 5'	GATTGGATGCTGTAGTCGCC <u>CA</u> TGAC
control-pou3f2b 5'	GAaTGcATGCTGTAGTCGCg <u>AT</u> cAC

The underlined nucleotides mark the AUG translation initiation site, while the lower case letters in the control MO indicates the bases that are mis-matched.

**Table S6: Primers and reaction conditions for *arnt2*<sup>hi2639cTg</sup> genotyping**

Hi2639_3E01 (F1):	5' ATA CTG AGG GTG AAC GCA GAC G 3'
Hi2639_3E02 (R1):	5' TCG CTT CTC GCT TCT GTT CG 3'
<i>arnt2</i> Endogenous:	5' AAA TCG CAT CCA AGC ATC GC 3'

Embryos were genotyped for the *arnt2*<sup>hi2639cTg</sup> allele by tail clip. DNA was prepared by alkaline lysis (25 mM NaOH, 0.2 mM disodium EDTA) for 1 hour at 95° C, followed by neutralization with an equal amount of 40 mM Tris-HCl; pH5. Amplification conditions were as per ZIRC PCR protocol (<http://zebrafish.org/zirc/fish/pdf/pcr/hi2639cTg.pdf>), with the addition of a third primer to identify the endogenous allele. The PCR reaction was performed using 1X PCR Master Mix (Thermo Scientific), 0.4 uM F1 primer, 0.2 uM R1 primer, 0.2 uM Endogenous primer, and 4 µl of tail clip DNA, in a 25 µl reaction solution. The *arnt2* PCR program was run according to the following protocol: 94°C, 3 minutes; 94°C, 30 seconds; 62°C, 40 seconds; 72°C, 30 seconds; repeat steps 2-4, 34 cycles, 72°C, 5 minute extension. The entire reaction was run on a 2% agarose gel, producing a mutant band (*arnt2*<sup>hi2639cTg</sup> = 342 bp), a wild-type band (wild-type = 237 bp) or both for the heterozygous genotype.



## Supplementary References

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