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Supplemental Information

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of Gene Expression in Human Peripheral Blood**

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Supplemental Data

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Zhao *et al*, 2015. **Figure S1**

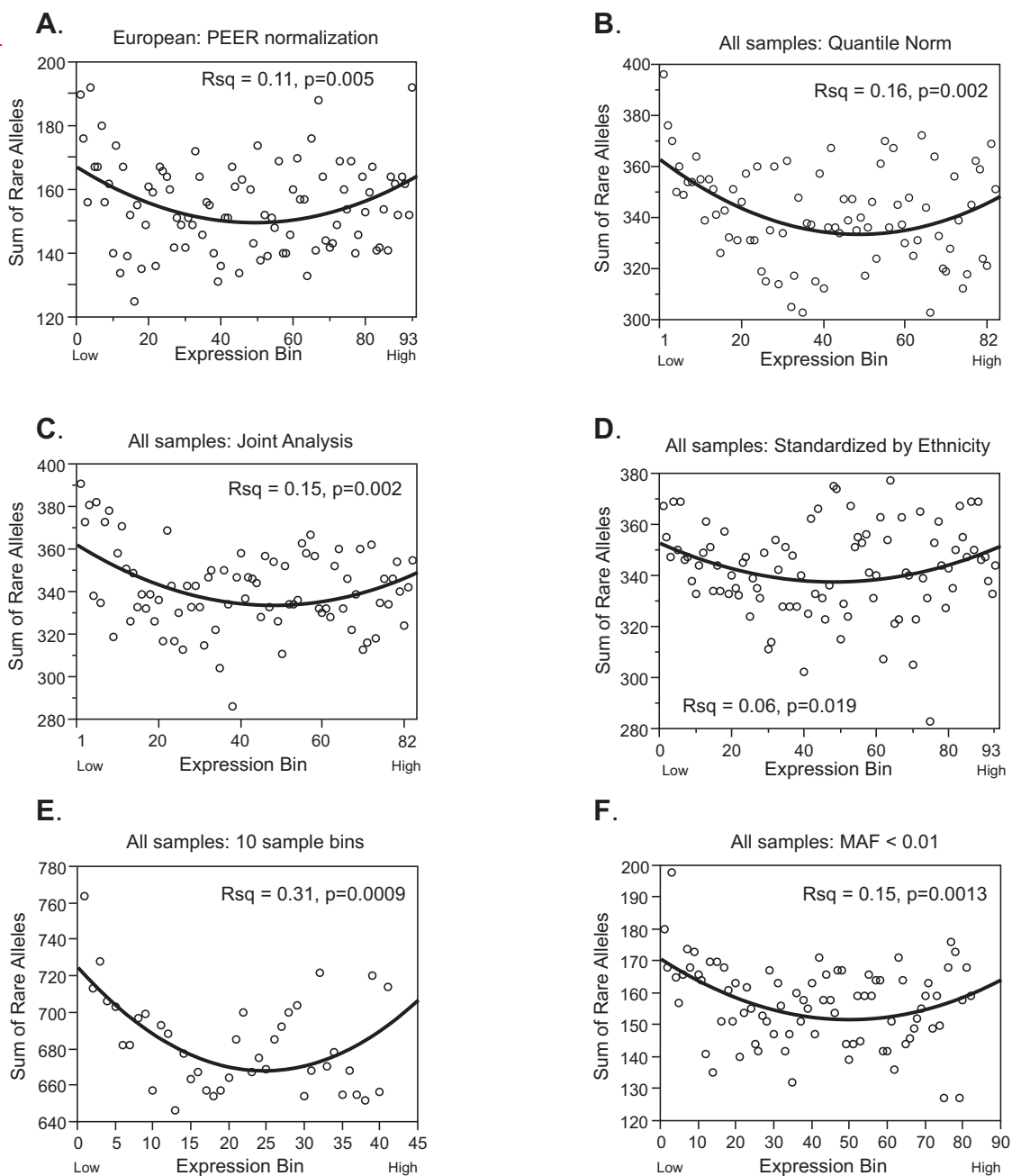
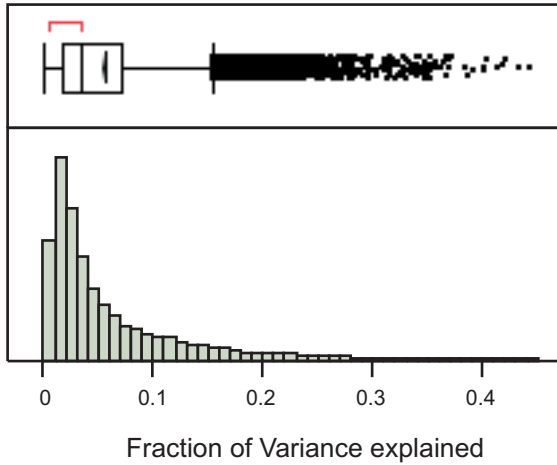


Figure S1. Quadratic regression plots with alternate normalization, bin size or MAF. As in Figure 1, each plot shows the quadratic regression fit of the sum of rare variants (A-E: MAF < 0.05; F, MAF < 0.01) in equal-sized expression bins from low to high (A-D,F: 5 individuals; E: 10 individuals). Rsq and p-values refer to the full model R-squared and p-value from a quadratic regression. (A) European only analysis with PEER normalization; (B) Quantile of all samples, no adjustment; (C) Joint analysis of z-scores of all genes; (D) SNM normalization of all samples after standardization by ancestry; (E) as in Figure 1A but with bin size of 10; (F) All samples SNM but with MAF<0.01.

A. Blood cell counts



B. Axes of Variation

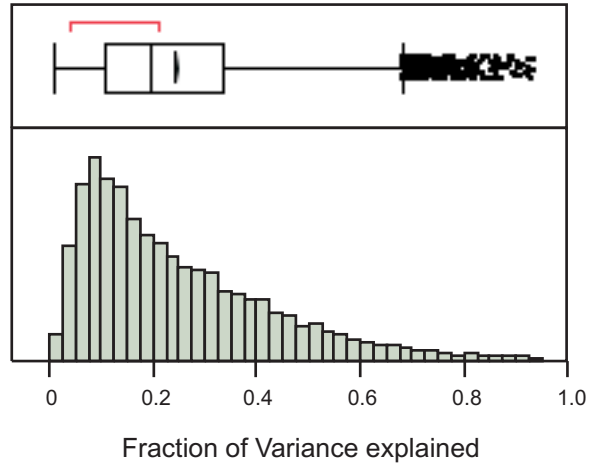


Figure S2. Variance explained by Blood Counts and Axes of Variation in CHDWB cohort. Histograms show the proportion of genes with the indicated fraction of variance explained by blood counts (lymphocytes, neutrophils, monocytes, red blood cells, platelets) and seven common Axes of variation respectively in (A) and (B), as the R-squared for the full model in a multiple regression. Boxes show the mean and interquartile range (25th to 75th percentile), and whiskers show 1.5X the respective IQR with outlier points shown as small squares.

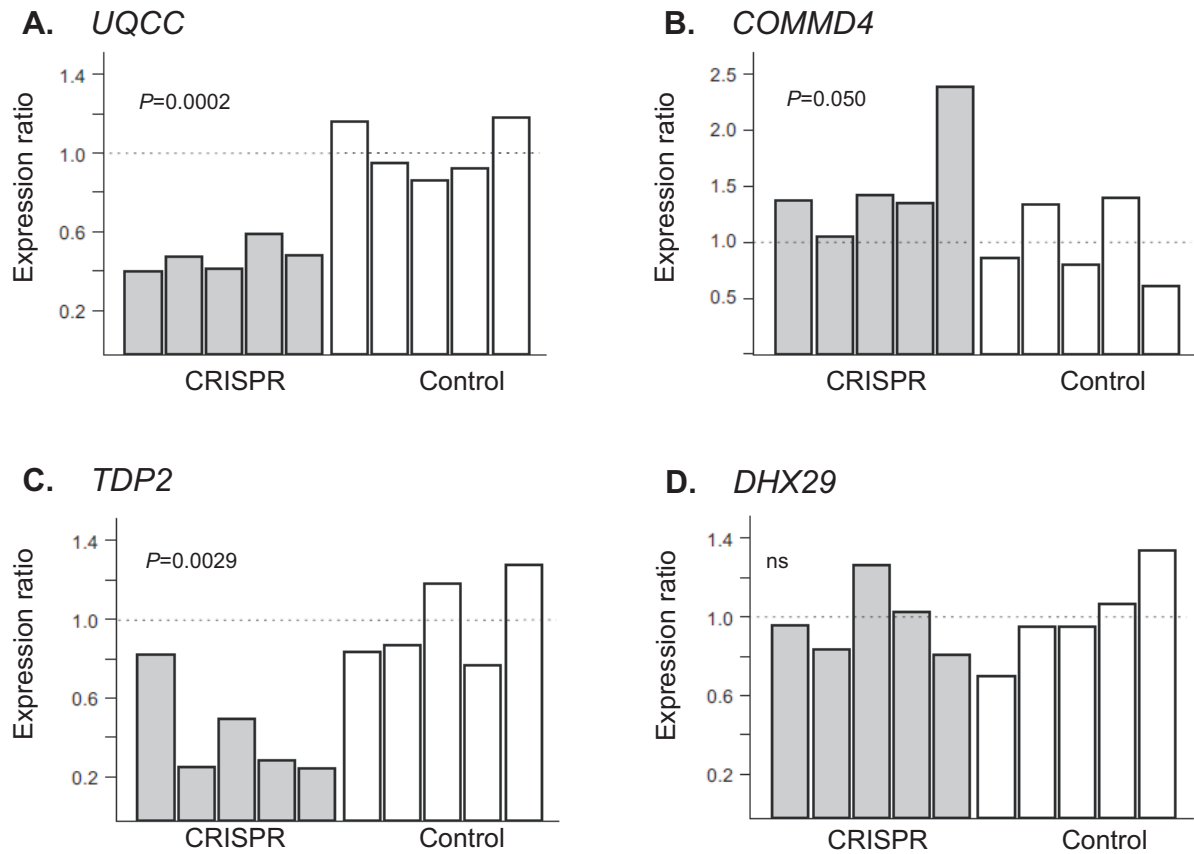


Figure S3. CRISPR / Cas9 mutagenesis validation of rare SNP regulatory effects. Average relative expression measures for the indicated transcript for five CRISPR mutagenized K562 10-cell clones in the same gene (gray) and in a different negative control gene (white). Dotted line represents mean expression of the control clones, to which the mutant clones are compared. See Supplementary methods for details of analysis.

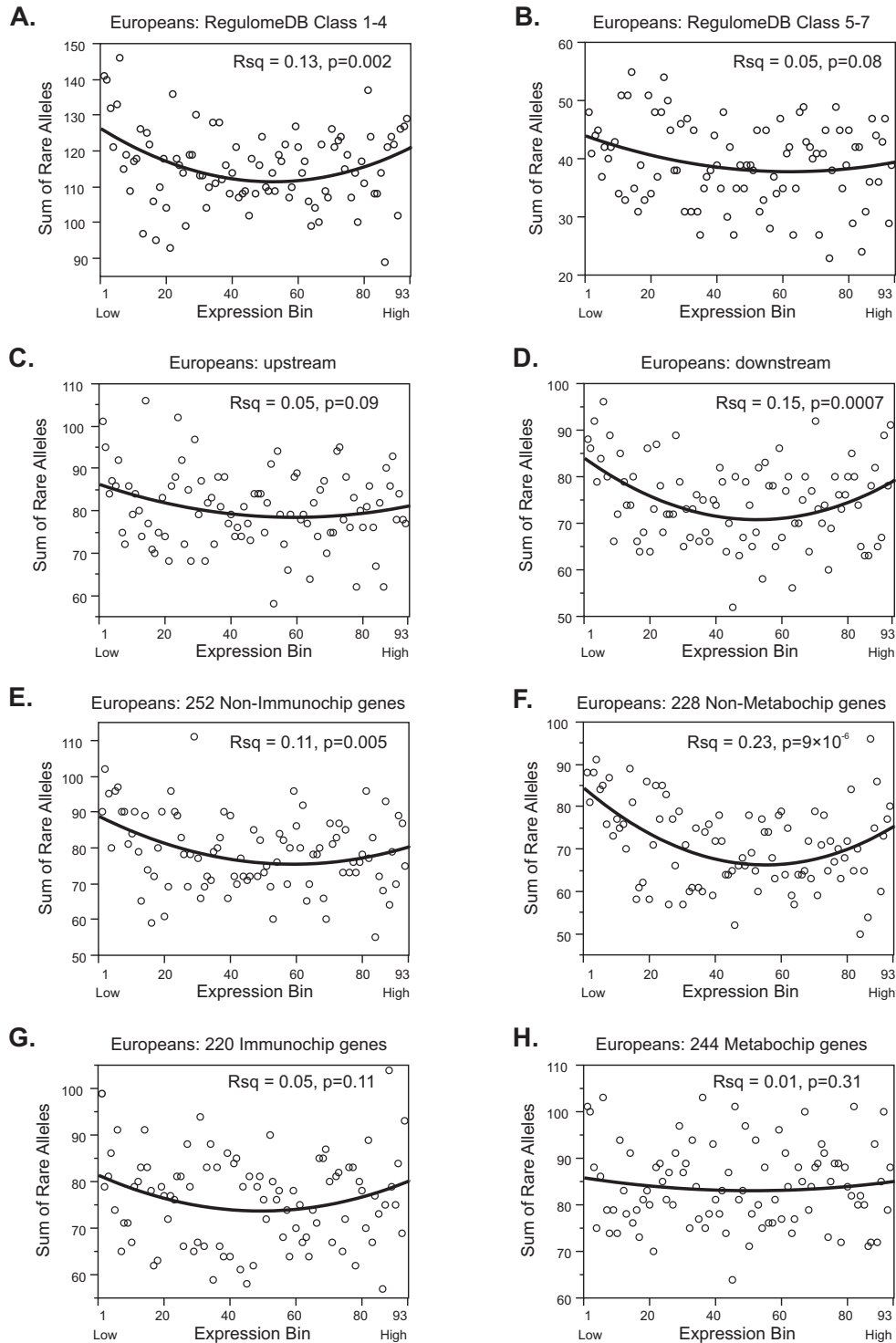


Figure S4. Quadratic regression plots for select sub-sets of SNPs or genes. Rsq and p -values refer to the full model R -squared and p -value from a quadratic regression. See Suppl. Table 1 for significance of linear and quadratic terms separately.

Zhao *et al*, 2015. **Figure S5**

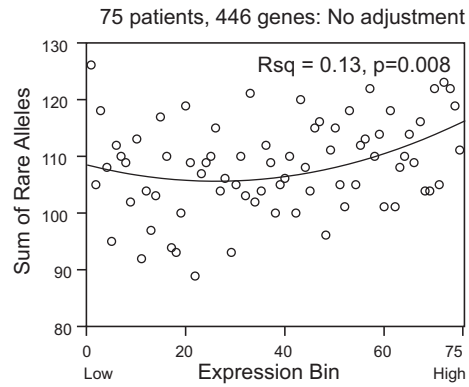


Figure S5. Quadratic regression plots for Replication dataset with 446 genes only, namely the same genes as in the CHDWB that are also present in the RNASeq dataset. Figure 1C shows an expanded replication dataset

Table S1 Comparisons of rare variant burden in subsets of SNPs and Genes

Comparison	First set				Second set			
	AvgCt ¹	Model ²	Linear ³	Quad ⁴	AvgCt ¹	Model ²	Linear ³	Quad ⁴
High polymorphism (242) vs Low polymorphism (230)	104.9	14.4**	0.032	0.002	50.1	0.03 ^{ns}	0.50	0.12
High expression (236) vs Low expression (236)	74.9	10.7*	0.35	0.002	80.2	14.5**	0.0008	0.08
Metabochip (244) vs non-Metabochip (228)	83.9	0.7 ^{ns}	0.81	0.45	71.1	22.7***	0.006	<0.0001
Immunochip (220) vs non-Immunochip (252)	76.1	4.8 ^{ns}	0.72	0.04	78.9	11.2*	0.021	0.017
With eQTL (207) vs No eQTL (265)	68.7	15.3*	0.027	0.001	86.4	4.9 ^{ns}	0.33	0.06
Upstream (472) vs Downstream (472)	80.5	5.1 ^{ns}	0.12	0.12	74.5	15.0*	0.12	0.0004
RegulomeDB 1-4 (472) vs RegulomeDB 5-7 (472)	115.7	12.7*	0.16	0.001	39.4	5.5 ^{ns}	0.08	0.15

¹ Average count of rare variants in each bin in the dataset

² Model R-squared, with significance: * 0.01<p<0.001; ** 0.001<p<0.0001; *** p<0.0001

³ p-value for linear term in the model

⁴ p-value for quadratic term in the model

Table S3. Validation of SNP effects by CRISPR/Cas9

Gene	SNP	Estimated effect ¹	Change in ddPCR ²	p-value	N clones Δ ³
UQCC	chr20:33999719	0.17X	0.48X	0.0002	5 of 5
TDP2	chr6:24667167	0.11X	0.46X	0.0029	4 of 5
COMMD4	rs182080358	12.5X	1.53X	0.050	4 of 5
DHX29	chr5:54603837	12.9X	0.98X	0.43	1 of 5

¹ Inferred from difference in log₂ fluorescence intensity between the individual with the mutation and all other 409 samples

² Average change in ratio of drop digital PCR counts relative to two reference genes normalized to unity

³ Number of the 5 clones deviant in the expected direction (see Suppl. Fig. 6).

APPENDIX: R code

//Code for PEER Normalization

Input is 279 Caucasian samples with 11,056 genes, Quantile normalized with SAS/JMP

//std for genes within batches(279, 11056G), get lists of std by batch

```
res <- apply(qnm_edata, 2, tapply, qnm_edata$Batch, scale)
```

//combine each gene, then combine all genes

```
std=matrix(rep(0),279,11056)
for(i in 1:11056){
tmp=do.call("rbind",res[[i]])
std[,i]=tmp[order(row.names(tmp)),]}
colnames(std)=colnames(qnm_edata)[2,11057]
rownames(std)=rownames(qnm_edata)
```

//ComBat fit batch

```
library(SVA)
edata=t(std)
batch=pheno$Batch
modcombat = model.matrix(~1, data=pheno)
combat_edata = ComBat(dat=edata, batch=batch, mod=modcombat, par.prior=TRUE, prior.plots=FALSE)
```

//transpose data

```
edata_ComBat=as.matrix(combat_edata)
transpose_ComBat_edata=t(edata_ComBat)
ComBat_edata=as.data.frame(transpose_ComBat_edata)
```

//Fit Age and Sex

```
rr=matrix(rep(0),279,11056)
for(i in 1:11056){
ff=lm(ComBat_edata[,i]~pheno$Gender+pheno$Age)
rr[,i]=ff$res
}
colnames(rr)=colnames(transpose_ComBat_edata)
rownames(rr)=rownames(transpose_ComBat_edata)
```

//Fit Axes (analysis not reported in paper, for comparison with SVA)

```
exp=rr
res=matrix(rep(0),279,11056)
```

```

Pmin=rep(1,11056)
Cmin=rep(1,11056)

for(i in 1:11056){
  Pmin[i]=1
  Cmin[i]=1
  for(j in 1:7){
    fit=lm(exp[,i]~Axes[,j])
    if(summary(fit)$coef[2,4]<Pmin[i]){
      Cmin[i]=j
      Pmin[i]=summary(fit)$coef[2,4]
      res[,i]=fit$res}
    }
  }
}
Bind=cbind(Cmin,Pmin)
colnames(res)=colnames(transpose_ComBat_edata)
rownames(res)=rownames(transpose_ComBat_edata)
write.csv(Bind,file="279CAU_qnm_std_ComBat_fitAgeSex_fitAxis_coef.csv")
write.csv(res,file="279CAU_qnm_std_ComBat_fitAgeSex_fitAxis.csv")

```

```
//PEER -- NK20
```

```

library(peer)
expr=res
model = PEER()
PEER_setPhenoMean(model,as.matrix(expr))
PEER_setNk(model,20)
PEER_update(model)
factors = PEER_getX(model)
weights = PEER_getW(model)
precision = PEER_getAlpha(model)
residuals = PEER_getResiduals(model)
pdf('PEER_Nk20_Model.pdf')
PEER_plotModel(model)
dev.off()
pdf('PEER_Nk20_Precision.pdf')
plot(precision)
dev.off()

```

```

//SNM normalization (on 546 samples, 14111 probes)

//Take mean of probes for each gene (on 546 samples, 14111 probes) using apply function in R

//Remove duplicate sample GG2_0043 and American Indian sample(GG1-000149) -->
544sample_SNM.csv

//Read tab-delimited files for expression, biological, adjustment and samples

chdwb.snm = read.csv("C:/Documents/544sample_SNM.csv", header=T, row.names=1)
chdwb.bio = read.csv("C:/Documents/chdwb_ageBethnR_bio.csv", header=T, row.names=1)
chdwb.adj = read.csv("C:/Documents/chdwb_ageBethnR_adj.csv", header=T, row.names=1)
chdwb.int = read.csv("C:/Documents/chdwb_ageBethnR_int.csv", header=T)

//Create model matrices and run SNM

int.var = chdwb.int
int.var$Array = as.factor(int.var$Array)
adj.var = model.matrix(~.,chdwb.adj)
bio.var = model.matrix(~.,chdwb.bio)
raw.data = as.matrix(chdwb.snm)
snmR.chdwb = snm(raw.data,bio.var,adj.var,int.var,rm.adj=TRUE,num.iter=10)
write.table(snmR.chdwb$norm.dat, file = "C:/Documents/544sample_SNM_stdBatch.csv", sep="," ,
col.names=NA)

//For SVA analysis, this step was included to use a linear model to fit age and gender to 472 genes and
411 samples among 544 which have genotyping information

//input "544sample_SNM" to R as "exp"
//input phenotype file to R as "pheno"
rr=matrix(rep(0),411,472)
for(i in 1:472){
ff=lm(exp[,i]~pheno$Gender+pheno$Age)
rr[,i]=ff$res
}
write.table(rr, file = "C:/Documents/544sample_SVA_stdBatch_agegender.csv", sep="," , col.names=NA)

//Fit Independent Common eQTL (within 1kb upstream and gene region) for 411 samples with
genotyping information

//Prepare eQTL files with row as samples, column as each common eSNP (order by coordinates) -->
input to R as "gen"

//Prepare expression file with 207 genes with common cis-eQTLs (order by coordinates)
--> input to R as "exp"

```



```

//Prepare another expression file with the left 265 genes without cis-eQTLs

//Prepare a file with two columns, the first column is gene name, the second column is the number of
common eQTLs. --> input to R as "count"

j=1
res=exp
Pmin=rep(1,207)
Cmin=rep(1,207)
for(i in 1:207){
  Cend=j+count[i,2]-1
  for (m in j:Cend){
    fit=lm(exp[,i]~gen[,m])
    if(summary(fit)$coef[2,4]<Pmin[i]&&summary(fit)$coef[2,4]<0.05){
      Cmin[i]=m
      Pmin[i]=summary(fit)$coef[2,4]
      res[,i]=fit$res}
    }
  exp[,i]=res[,i]
  j=Cend+1}
  //Check Pmin, if there is at least one Pmin<0.05, then re-do the whole steps, until all
Pmins>=0.05 for all genes
  //Output res, combine with the 265 gene expression.

//385 samples who have expression levels.were included in 411 samples -->
385sample_SNM_stdBatch_fitEqtl

```

//Fit the most significant Axis to 385 samples

```

  //input "385sample_SNM_stdBatch_fitEqtl" to R as "exp"

  //Prepare axis file with 7 axes --> input to R as "Axes"

res=matrix(rep(0),385,472)
Pmin=rep(1,472)
Cmin=rep(1,472)
for(i in 1:472){
  Pmin[i]=1
  Cmin[i]=1
  for(j in 1:7){
    fit=lm(exp[,i]~Axes[,j])
    if(summary(fit)$coef[2,4]<Pmin[i]){
      Cmin[i]=j
      Pmin[i]=summary(fit)$coef[2,4]
      res[,i]=fit$res}
    }
  }
}

```

```
//res --> expression data for 385 samples after normalization and adjustment
//279 are Caucasians among 385 samples. --> 279CAU_SNM_stdBatch_fitEqtlAxis
```

//Rare variant association test

```
    //input "279CAU_SNM_stdBatch_fitEqtlAxis" as "exp"
    //input rare variant counts per gene per sample as "snp"
    //the below code split genes to 93 bins with 3 genes in each bin for each sample

count=c(rep(0,93))
for(i in 1:472){
Rk=rank(exp[i,],ties.method="first")
  for(j in 1:279){
    zz=as.integer((Rk[j]+2)/3)
    if(snp[i,j]!=0){
      count[zz]=count[zz]+snp[i,j]}
    j=j+1}
i=i+1}
perc=1:93

    //"count" --> rare variant counts in each bin
    //"perc" --> bin number (expression bin from lowest to highest)
```

//Perform quadratic test on count and perc

```
SAS/JMP -- Analyze -- Fit Y by X -- Fit polynomial – quadratic
```