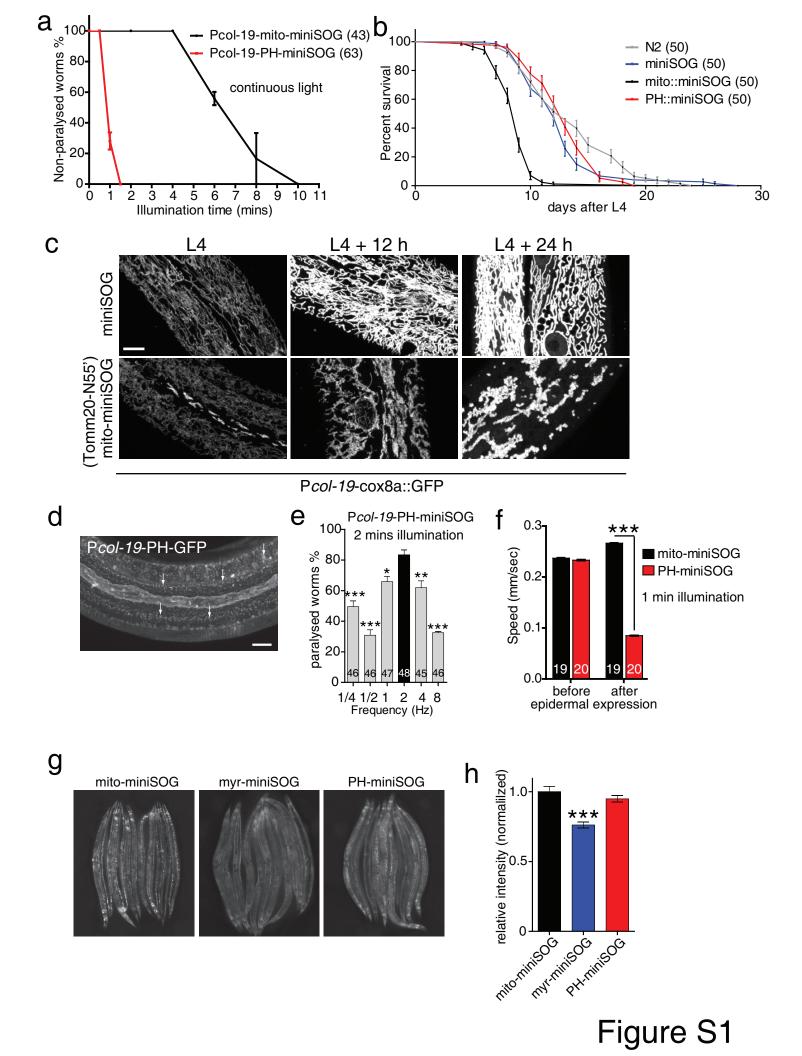
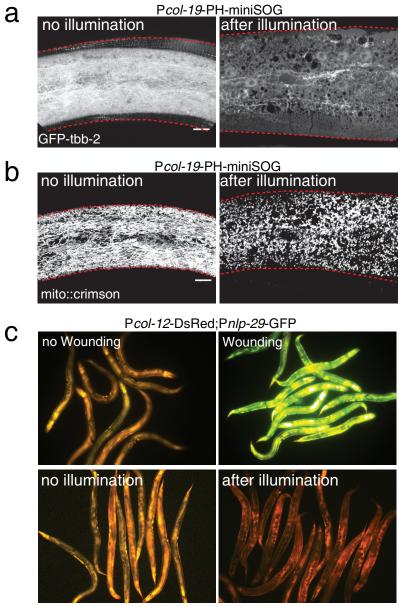
Highly efficient optogenetic cell ablation in *C. elegans* using membrane-targeted miniSOG

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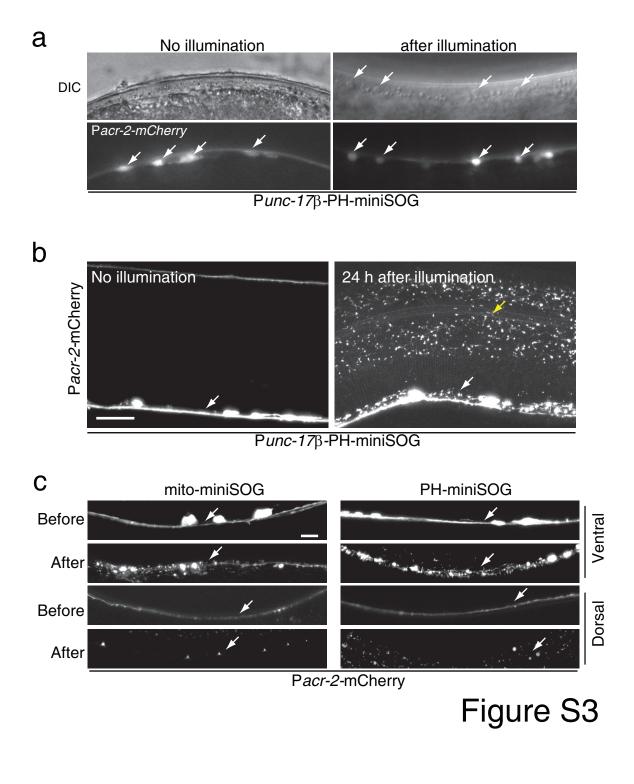
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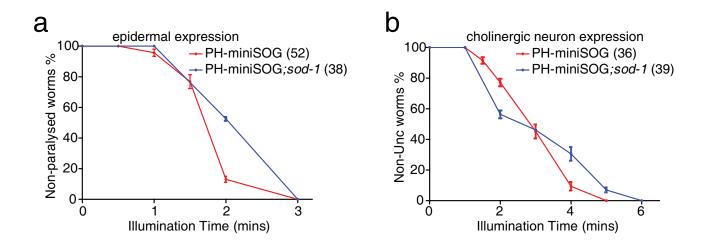
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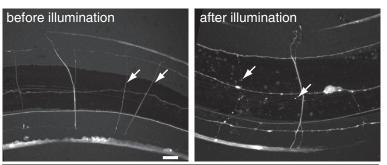


Pcol-19-PH-miniSOG





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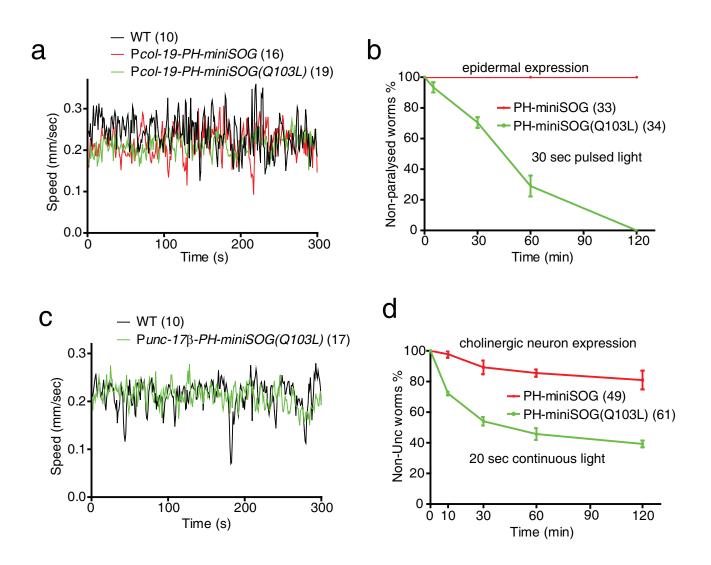


Punc-25-PH-miniSOG;Prgef-1-mCherry

b

before illumination AVM	before illumination PVM	
ALM _	PLM	
after illumination	after illumination	

Pmec-4-PH-miniSOG;Pmec-4-mCherry



Supplemental Information

Table S1. New miniSOG transgenic strains and plasmids

Cell types	Strains	Genotype/Transgenes ¹	Plasmid ²	Concentra tion of miniSOG constructs
Epidermis	CZ16052	Pcol-19-miniSOG (juEx4532)	pCZGY2137	10 ng/μl
	CZ16855	Pcol-19-cox8-miniSOG(juEx4973)	pCZGY2136	10 ng/μl
	CZ16054	Pcol-19-mito-miniSOG (juEx4534)	pCZGY2138	10 ng/μl
	CZ23271	Pcol-19-myr-miniSOG (juEx7095)	pCZGY2847	10 ng/μl
	CZ16493	Pcol-19-PH-miniSOG (juEx4771)	pCZGY2141	10 ng/μl
	CZ16236	Pcol-19-PH-miniSOG/Pcol-19- tdTomato (juls400)	PCZGY2141/ pCZGY1575	10 ng/μl of each
	CZ23277	Pcol-19-PH-miniSOG(Q103L) (juEx7101)	pCZGY2850	10 ng/μl
Body wall muscle	CZ15071	Pmyo-3-mito-miniSOG/Pmyo-3- mCherry (juEx4076)	pCZGY1555/ pCZGY872	1 ng/μl of each
	CZ22703	Pmyo-3-PH-miniSOG/Pmyo-3- mCherry (juEx6916)	pCZGY2838/ pCZGY872	5 ng/μl of each
Cholinergic motor neuron	CZ15033/ CZ15844	Punc-17β-mito-miniSOG/Pacr-2- mCherry (juls385)	pCZGY1558/ pCZGY847	50 ng/µl of each
	CZ23273	Punc-17β-myr-miniSOG/Pacr-2- mCherry (juEx7097)	pCZGY2848/ pCZGY847	20 ng/μl of each
	CZ22692	Punc-17β-PH-miniSOG/ Pacr-2- mCherry (juEx6905)	pCZGY2844/ pCZGY847	20 ng/μl of each
	CZ23279	Punc-17β-PH-miniSOG(Q103L)/ Pacr-2-mCherry (juEx7103)	pCZGY2851/ pCZGY847	20 ng/μl of each
GABAergic motor neuron	CZ19758	Punc-25-mito-miniSOG/Punc-25- crimson (juEx5983)	pSK9/pSK10	50 ng/μl of each
	CZ22698	Punc-25-PH-miniSOG/Prgef-1- mCherry (juEx6911)	pCZGY2842/ pCZGY922	20 ng/μl of each

Interneuron	CZ14478	Pnmr-1-mito-miniSOG/Pnmr-1- mCherry (juEx3771)	pCZGY1552/ pCZGY903	50 ng/μl of each
	CZ22695	Pnmr-1-PH-miniSOG/Pnmr-1- mCherry (juEx6908)	pCZGY2845/ pCZGY903	20 ng/μl of each
Touch neuron	CZ23283	Pmec-4-mito-miniSOG/Pmec-4- mCherry (juEx7107)	pCZGY2852/ pCZGY546	20 ng/μl of each
	CZ23281	Pmec-4-PH-miniSOG/ Pmec-4- mCherry (juEx7105)	pCZGY2840/ pCZGY546	20 ng/μl of each
Microtubule and mitochondria marker	CZ21790	Pcol-19-GFP-tbb-2;Pcol-19-mito- crimson (juEx6578)	pCZ899/pCZ GY2160	10 ng/μl of each

Notes:

- 1. Co-injection markers were Pttx-3-GFP or Pttx-3-RFP (50 ng/µl)
- 2. All miniSOG clones used DNA codons optimized for mammals ¹.

Supplemental Figure Legends

Figure S1. Comparison of the effects of mito-miniSOG and PH-miniSOG in epidermis

a. Quantitation of paralysis in epidermal mito-miniSOG and PH-miniSOG expressing animals immediately after continuous blue light illumination for the indicated period. All transgenes use *col-19* promoter. Numbers are the animals that were analyzed in four independent experiments.

b. Lifespan of wild type and miniSOG transgenic animals. Only mito-miniSOG transgenics displayed significantly reduced lifespan compared to the WT. p < 0.001, survival test, error: 95% CI. n = 50 animals for each strain.

c. Mito-miniSOG disrupts the tubular structure of mitochondria in the epidermis during development. Representative confocal images of epidermal mitochondria (P*col-19*-mito-GFP) in transgenic animals expressing cytosolic miniSOG and mito-miniSOG at different stages. Scale, 10 μ m.

d. The PH domain targets fusion proteins to the epidermal cell membrane. Representative confocal image of PH-GFP expressed in the epidermis. Arrows indicate apical puncta. Scale, 10 μ m.

e. Quantitation of paralysis in P*col-19*-PH-miniSOG animals immediately after 2 min blue light illumination at different frequencies. Numbers are the animals that were analyzed in three independent experiments. *, P < 0.05, **, P < 0.01, ***, P < 0.001, One-way ANOVA. f. Quantitation of locomotion velocity immediately after blue light illumination. P*col-19*-PH-miniSOG transgenic animals displayed significantly reduced speed compared to P*col-19*-mito-miniSOG transgenics. Numbers are the animals that were analyzed in 2 independent experiments. ***, P<0.001, t-test.

g. Green fluorescence images of epidermal miniSOG transgenic animals. Scale, 250 μ m.

h. Quantitation of green fluorescence intensity in panel f. ***, P < 0.001, One way ANOVA. mito-miniSOG fluorescence intensity was normalized to 1.

Figure S2. Membrane targeted miniSOG expressed in the epidermis causes cell disruption after blue light illumination.

a. Animals expressing P*col-19*-PH-miniSOG the epidermis display disrupted microtubule structure (P*col-19*-GFP-TBB-2 marker) after 2 min blue light illumination. Red dashed lines indicate the outlines of the animal. Scale, 10 μm.

b. Animals expressing PH-miniSOG in the epidermis display disrupted mitochondrial structure (P*col-19*-mito::crimson) after 2 min blue light illumination. Scale, 10 μm.

c. PH-miniSOG activation in the epidermis does not induce antimicrobial peptide expression (P*nlp-29-*GFP reporter *frls7*, containing P*col-12-*dsRed internal control). Top: needle wounding triggers P*nlp-29-*GFP induction in the epidermis 4 h later. Bottom: PH-miniSOG expression in the epidermis paralyzed animals but did not induce P*nlp-29-*GFP, 4 h after 2 min blue light illumination.

Figure S3 Effects of PH-miniSOG on cholinergic neuron morphology

a. DIC and fluorescence images of Punc-17 β -PH-miniSOG transgenic animals before and 2 h after illumination (2 min and 2 Hz). Arrows indicate rounded-up soma in the VNC in the DIC images, and Pacr-2-mCherry in the fluorescence image.

b. Representative confocal images of *Pacr-2-mCherry*; Punc-17 β -PH-miniSOG before and 24 h after blue light illumination. White arrows indicate VNC and yellow arrowheads indicate mCherry aggregates in the epidermis. Scale, 10 μ m.

c. Representative confocal images of *Pacr-2-*mCherry before and 24 h after blue light illumination. Both mito-miniSOG and PH-miniSOG (under the control of *unc-17* β promoter) induce cholinergic neuron degeneration after blue light illumination. White arrows indicate VNC and DNC. Scale, 10 µm.

Figure S4. Membrane targeted miniSOG-mediated cell killing is not enhanced in superoxide dismutase *sod-1* mutant

a. Quantitation of paralysis in WT and *sod-1(tm776)* mutant animals expressing epidermal PH-miniSOG immediately after 2 Hz blue light illumination, for the indicated times. Note *sod-1* mutant does not enhance the paralysis of PH-miniSOG transgenic animals after blue light illumination. b. Quantitation of Unc animals in WT and *sod-1(tm776)* mutant expressing PH-miniSOG immediately after 2 Hz blue light illumination for the indicated times. PH-miniSOG is under the control of *unc-17* β promoter. The *sod-1* mutant does not enhance the Unc phenotype of PH-miniSOG transgenic animals after blue light illumination.

Figure S5 PH-miniSOG efficiently kills GABAergic motor neurons and touch neurons after blue light illumination.

c. Pan-neuronal morphology (labeled by Prgef-1-mCherry) in Punc-25-PH-miniSOG transgenic animals before and 24h after 4 min blue light illumination. Note that PH-miniSOG activation kills motor neurons but not other nearby neurons. White arrows indicate motor neurons. Scale, 10 μ m.

d. PH-miniSOG expression in touch neurons caused ALM, AVM, PLM, and PVM cell degeneration 24 h after 4 min blue light illumination. White arrows indicate degenerated cells. Scale, 10 μ m.

Figure S6. The Q103L miniSOG variant enhances cell ablation efficacy

a. Animals expressing PH-miniSOG or PH-miniSOG (Q103L) in the epidermis were indistinguishable from wild type before blue light illumination. Quantitation of locomotion velocity of PH-miniSOG, PH-miniSOG (Q103L) transgenic and WT animals, using the multi worm tracker. Numbers are the animals that were analyzed. b. Onset of paralysis in animals expressing epidermal PH-miniSOG or PH-miniSOG (Q103L), after 30 sec 2 Hz blue light illumination. Numbers are the animals that were analyzed in three independent experiments.

c. Animals expressing PH-miniSOG (Q103L) in the cholinergic motor neurons are superficially wild type prior to blue light illumination. Quantitation of locomotion velocity of Punc-17 β -PH-miniSOG (Q103L) transgenic and N2 (WT) animals using the multi worm tracker. Numbers are the animals that were analyzed.

d. Onset of paralysis in animals expressing cholinergic PH-miniSOG and PH-miniSOG (Q103L) animals after 20 sec continuous blue light illumination. Numbers are the animals that were analyzed in three independent experiments.

Supplemental Movie Legends

Movie S1. P*col-19*-miniSOG. With and without blue light illumination. These transgenic animals display normal locomotion before and after illumination. Time: min.

Movie S2. P*col-19*-mito-miniSOG. With and without blue light illumination. These transgenic animals display normal locomotion before illumination and are paralyzed immediately after 10 min blue light (2 Hz) illumination. Time: min.

Movie S3. P*col-19*-PH-miniSOG. With and without blue light illumination. These transgenic animals display normal locomotion before illumination and are paralysed immediately after 2 min blue light (2 Hz) illumination. Time: min.

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Movie S4. P*col-19*-myr-miniSOG. With and without blue light illumination. These transgenic animals display normal locomotion before illumination and are paralyzed immediately after 4 min blue light (2 Hz) illumination. Time: min.

Movie S5. Punc-17 β -PH-miniSOG. With and without blue light illumination. These transgenic animals display normal locomotion before illumination and are Unc immediately after 3 min blue light (2 Hz) illumination. Time: min.

Movie S6. Punc-25-PH-miniSOG. With and without blue light illumination. These transgenic animals display normal locomotion and are Unc immediately after 4 mins blue light illumination. Time: min.

Movie S7. Pmyo-3-PH-miniSOG. With and without blue light illumination. These transgenic animals display normal locomotion before illumination and are paralyzed immediately after 4 min blue light (2 Hz) illumination. Time: min.

References:

1 Qi, Y. B., Garren, E. J., Shu, X., Tsien, R. Y. & Jin, Y. Photo-inducible cell ablation in *Caenorhabditis elegans* using the genetically encoded singlet oxygen generating protein miniSOG. *Proc Natl Acad Sci U S A* **109**, 7499-7504 (2012).