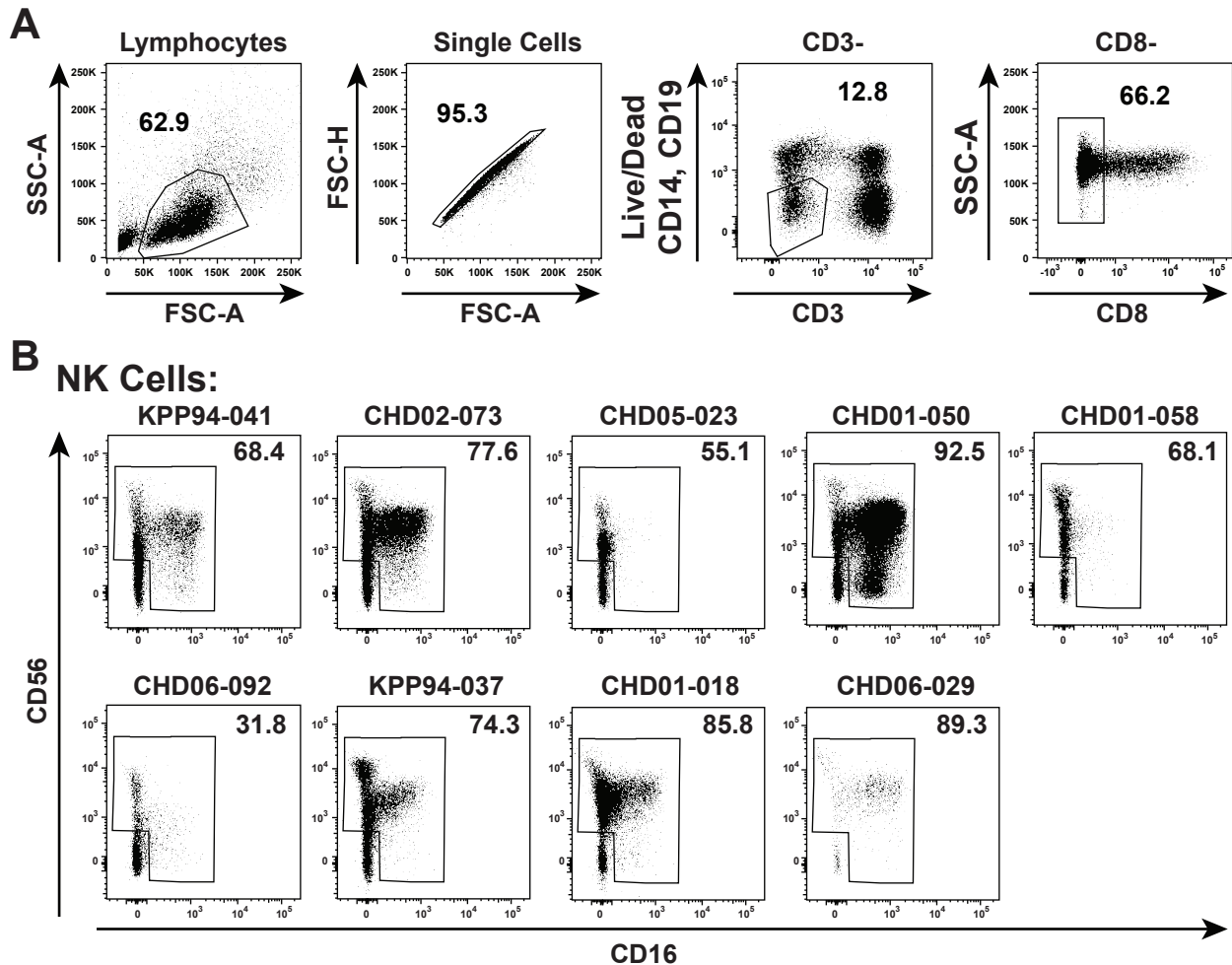
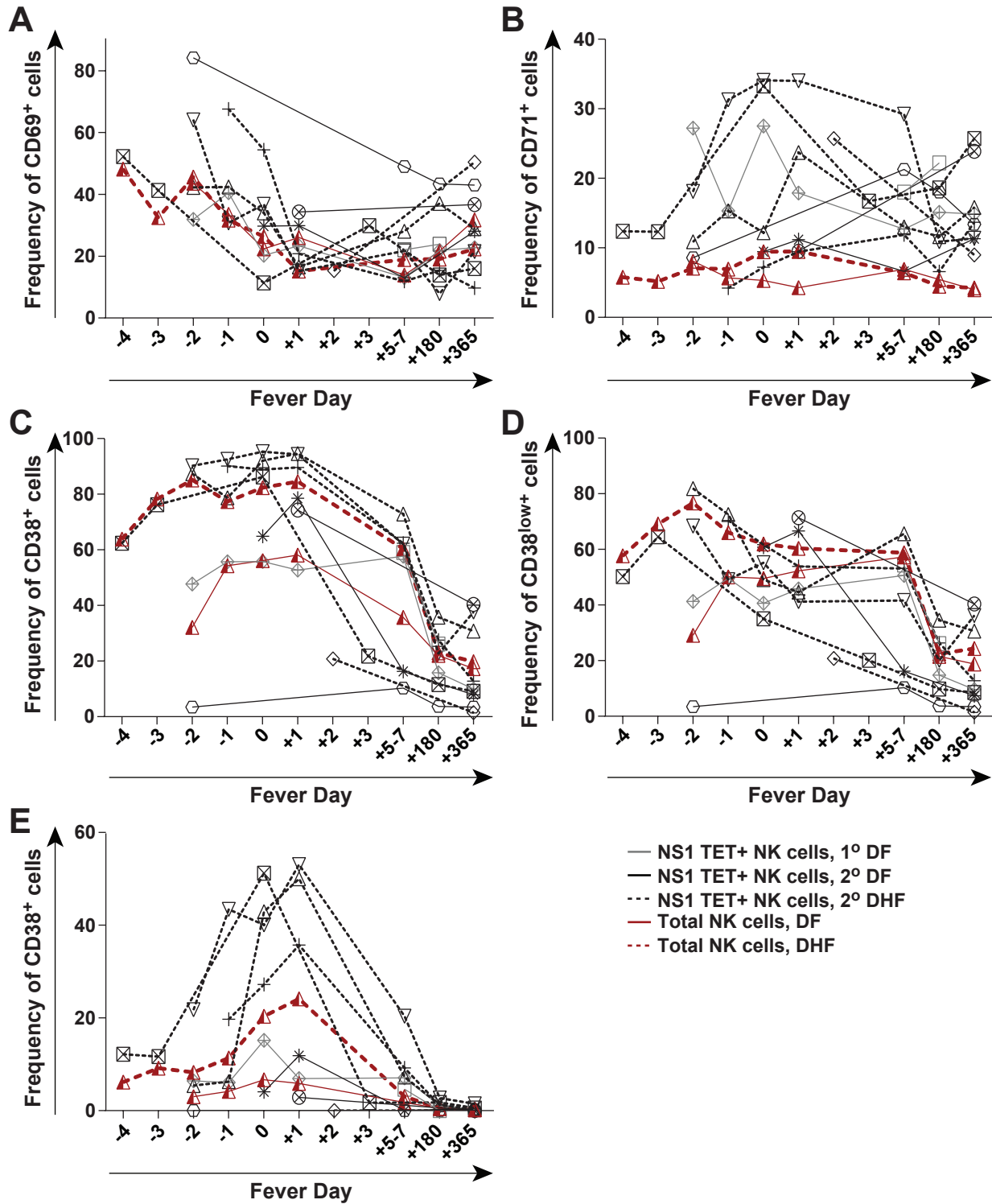


## Supplemental Figure 1



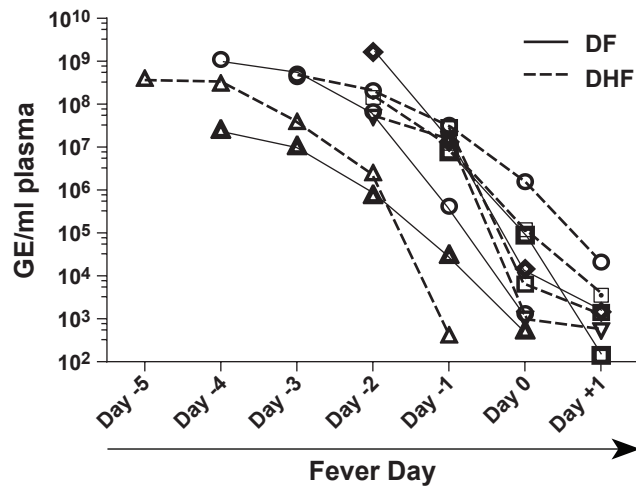
**Supplemental Figure S1.** Frequencies of NK cells in the CD3-CD8-CD14-CD19<sup>-</sup> gate. (A) Gating strategy to identify CD3-CD8-CD14-CD19<sup>-</sup> cells. Cells were first selected within the lymphocyte gate as defined by forward and side scatter profiles. Singlets were then identified and live CD3-CD14-CD19<sup>-</sup> cells were selected in a dump (LIVE/DEAD<sup>®</sup> Green with  $\alpha$ CD14 and  $\alpha$ CD19) versus CD3 bivariate plot. CD8<sup>-</sup> cells were gated within this population. (B) Frequencies of CD56<sup>+</sup> and/or CD16<sup>+</sup> NK cells in PBMCs collected from Thai cohort subjects 2 years after acute DENV infection. Plots are gated on live CD3-CD8-CD14-CD19<sup>-</sup> cells.

Supplemental Figure 2



**Supplemental Figure S2.** Activation of NS1 TET<sup>+</sup> and total NK cells over the course of acute dengue illness. Kinetics of CD69 (A), CD71 (B), total CD38 (C), CD38<sup>low</sup> (D), and CD38<sup>hi</sup> (E) expression on NS1 TET<sup>+</sup> and total NK cells during acute dengue illness and convalescence. The average frequencies of CD69<sup>+</sup>, CD71<sup>+</sup>, total CD38<sup>+</sup>, CD38<sup>low</sup>, and CD38<sup>hi</sup> total NK-enriched cells are shown using a solid red line for subjects with DF and a dashed red line for subjects with DHF. Symbols distinguish subjects with primary ( $n=2$ , grey symbols) versus secondary ( $n=8$ , black symbols) DENV infections and lines distinguish those with DF ( $n=5$ , black line) versus DHF ( $n=5$ , dashed line).

### Supplemental Figure 3



**Supplemental Figure S3.** Magnitude of DENV viremia by day of illness. Levels of DENV genome equivalent (GE) cDNA (copies/mL) were determined in serial plasma samples from HLA-B57<sup>+</sup> patients. Symbols denote individual subjects and lines distinguish those with DF ( $n=4$ , black line) versus DHF ( $n=5$ , dashed line).

SUPPLEMENTAL TABLE 1: Antibodies used for flow cytometry studies

<b>Marker</b>	<b>Clone</b>	<b>Manufacturer</b>	<b>Fluorochrome</b>
<b>CD3</b>	UCHT1	BD Biosciences	V500
		BioLegend	BV510
	OKT3	BioLegend	BV510
<b>CD8</b>	SK1	Invitrogen	PE-AlexaFluor610
<b>CD45RA</b>	HI100	BD Pharmingen	APC-H7
<b>CCR7</b>	150503	BD Horizon	V450
<b>CD69</b>	CH/4	Invitrogen	PE-Cy5.5
	FN50	BioLegend	BV650
<b>CD38</b>	HB7	eBioscience	eFluor@650NC
<b>CD57</b>	HCD57	BioLegend	PerCP/Cy5.5 (Lightening Link)
<b>CD71</b>	OKT9	eBioscience	PE-Cy7 (Lightening Link)
	CY1G4	BioLegend	APC
<b>CD28</b>	CD28.2	BioLegend	AlexaFluor700
<b>CD56</b>	B159	BD Biosciences	AlexaFluor700
<b>CD19</b>	HIB19	BD Biosciences	FITC
<b>CD14</b>	HCD14	BioLegend	FITC
<b>CD56</b>	HCD56	BioLegend	BV711
<b>CD16</b>	3G8	BD Horizon	APC-H7
<b>NKp30</b>	P30-15	BioLegend	APC
<b>NKp46</b>	9E2	BioLegend	BV421
<b>CD161</b>	HP-3G10	BioLegend	BV605
<b>NKG2D</b>	1D11	BD Biosciences	PE-CF594
<b>KIR3DL1</b>	DX9	Beckman Coulter	PE
		BioLegend	PE
<b>KIR3DL1/S1</b>	Z27	Beckman Coulter	APC
<b>HLA-A,B,C</b>	W6/32	BD Biosciences	APC, PE, FITC
		BioLegend	PE
<b>HLA-B57</b>	BIH0243	One Lambda	PE-NeutrAvidin