

Supplementary Information

Inhibition of mechanical allodynia in neuropathic pain by TLR5-mediated A-fiber blockade

Zhen-Zhong Xu, Yong-Ho Kim, Sangsu Bang, Yi Zhang, Temugin Berta,
Fan Wang, Seog-Bae Oh, & Ru-Rong Ji

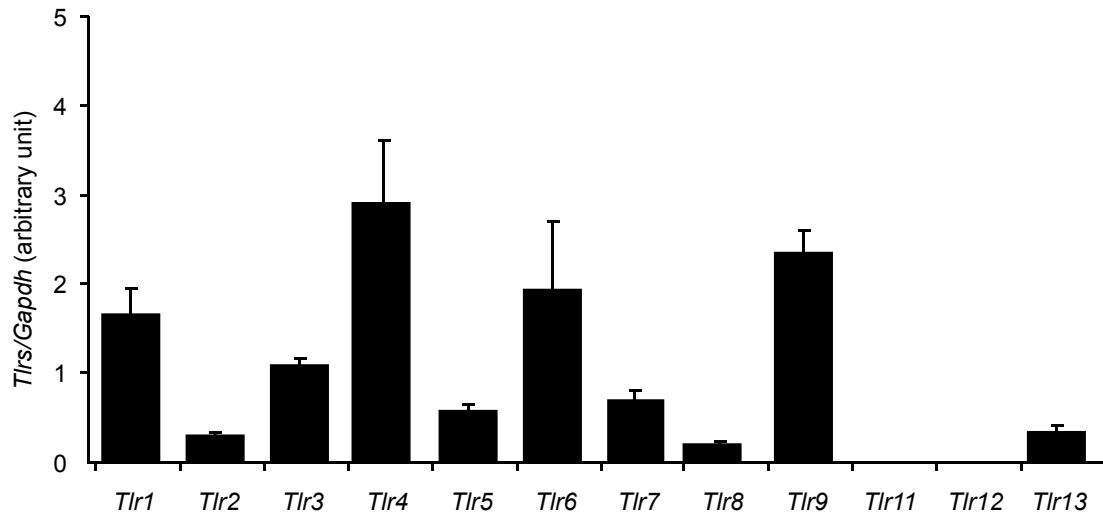
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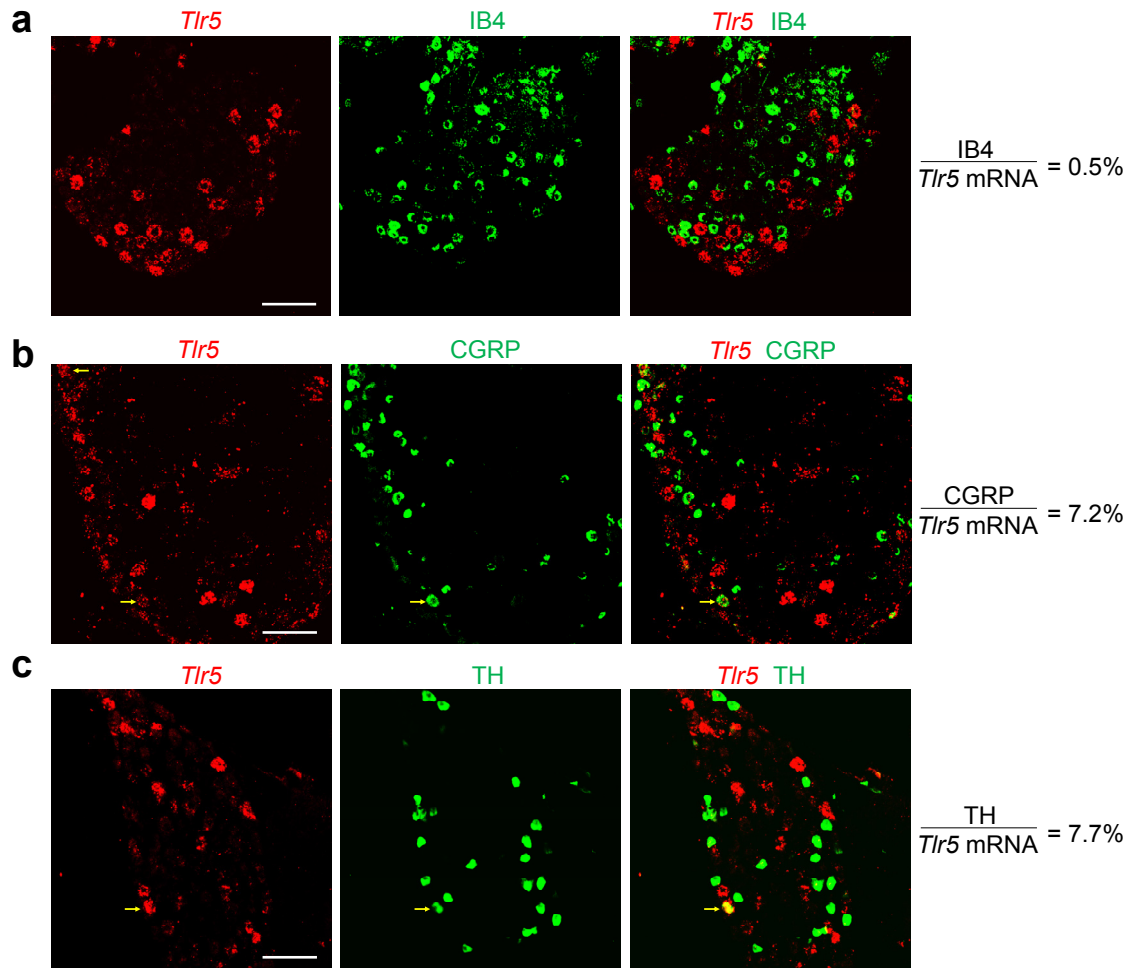
Supplementary Table 1. Mouse sequences of *Tlr* primers used for quantitative RT-PCR.

List of primer sequences designed for quantitative real-time RT-PCR.

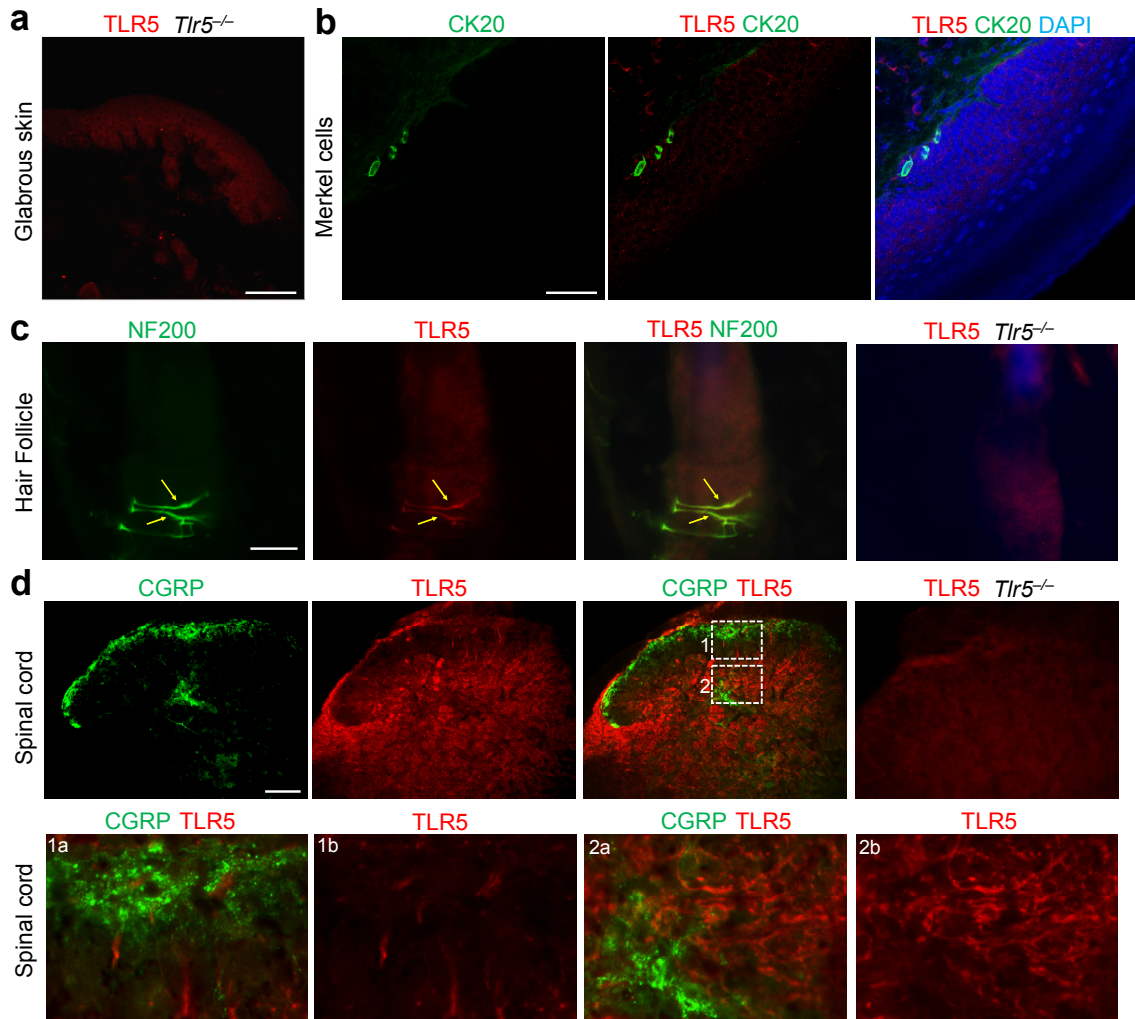
	Forward primers	Reverse primers	Accession No.
<i>Gapdh</i>	TTGATGGCAACAATCTCCAC	CGTCCCGTAGACAAAATGGT	NM_001001303
<i>Tlr1</i>	ATGTGAGCTGAGGGACTTTG	GGATAGTGGAGACATGTGGAAG	NM_030682
<i>Tlr2</i>	ACCAAGATCCAGAAGAGCCA	CATCACCGGTCAGAAAACAA	NM_011905
<i>Tlr3</i>	GCGTTGCGAAGTGAAGAACT	TTCAAGAGGAGGGCGAATAA	NM_126166
<i>Tlr4</i>	TTCAGAACTTCAGTGGCTGG	TGTTAGTCCAGAGAACTTCCTG	NM_021297
<i>Tlr5</i>	GCAGGATCATGGCATGTCAAC	ATCTGGGTGAGGTTACAGCCT	NM_016928
<i>Tlr6</i>	CCAAGAACAAAAGCCCTGAG	TGTTTTGCAACCGATTGTGT	NM_011604
<i>Tlr7</i>	GATGTCCTTGGCTCCCTTC	TTTGTCTCTCCGTGTCCAC	NM_133211
<i>Tlr8</i>	CGTTTTACCTTCCTTTGTCTATAGAAC	CGTCACAAGGATAGCTTCTGG	NM_133212
<i>Tlr9</i>	AACCGCCACTTCTATAACCAG	GTAAGACAGAGCAAGGCAGG	NM_031178
<i>Tlr11</i>	CAGGCTGGGATTGCTCATC	CCAGTCAAGGTAAGGCTCAC	NM_205819
<i>Tlr12</i>	GCTCTGATTCTCTGGTGTAG	AGAATGTGAAATAGCGGGAGAC	NM_205823
<i>Tlr13</i>	GGAGCGCCTTGATCTAACTAACA	TCAGGTGGGTCAGAGAAACCA	NM_205820



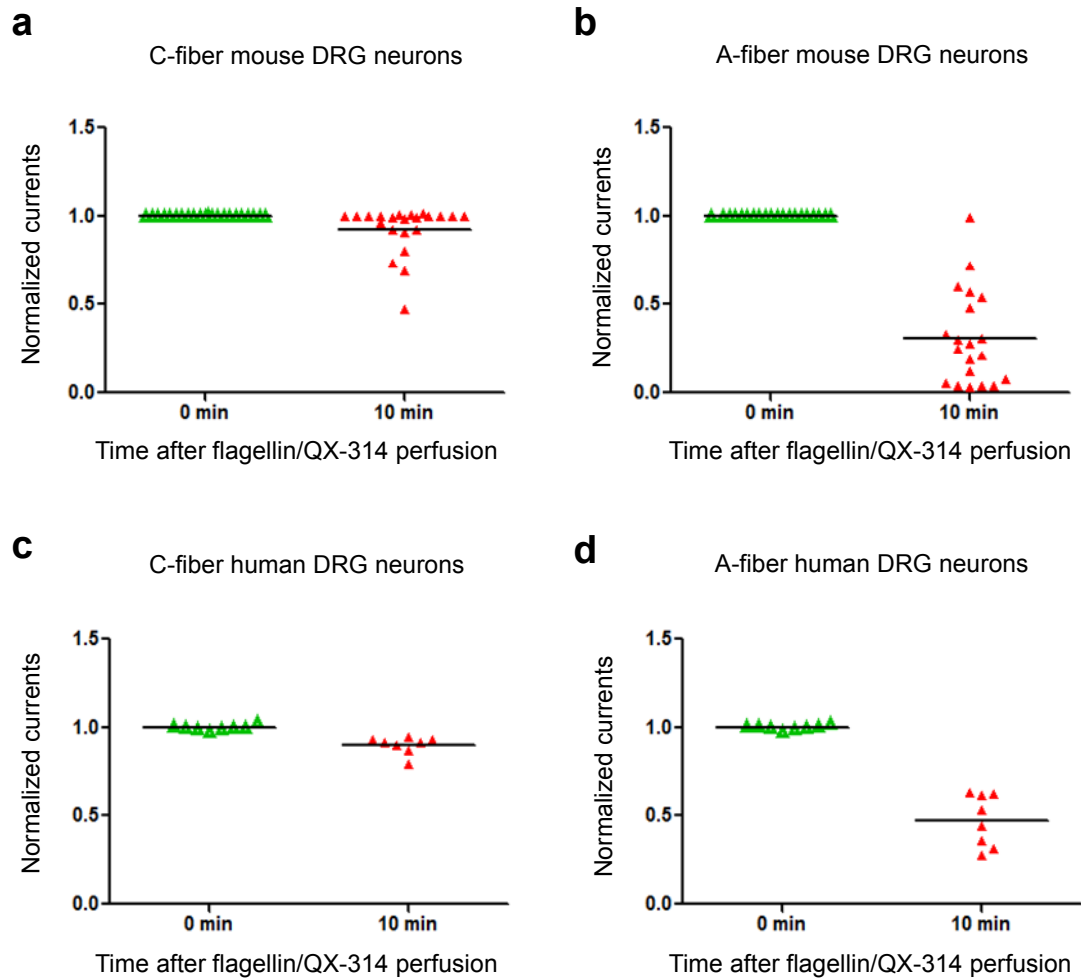
Supplementary Fig. 1. Quantitative RT-PCR analysis shows relative expression of *Tlr* mRNAs in mouse DRG tissues. The values are normalized to GAPDH. $n = 3-4$ mice/group. All data were expressed as mean \pm s.e.m



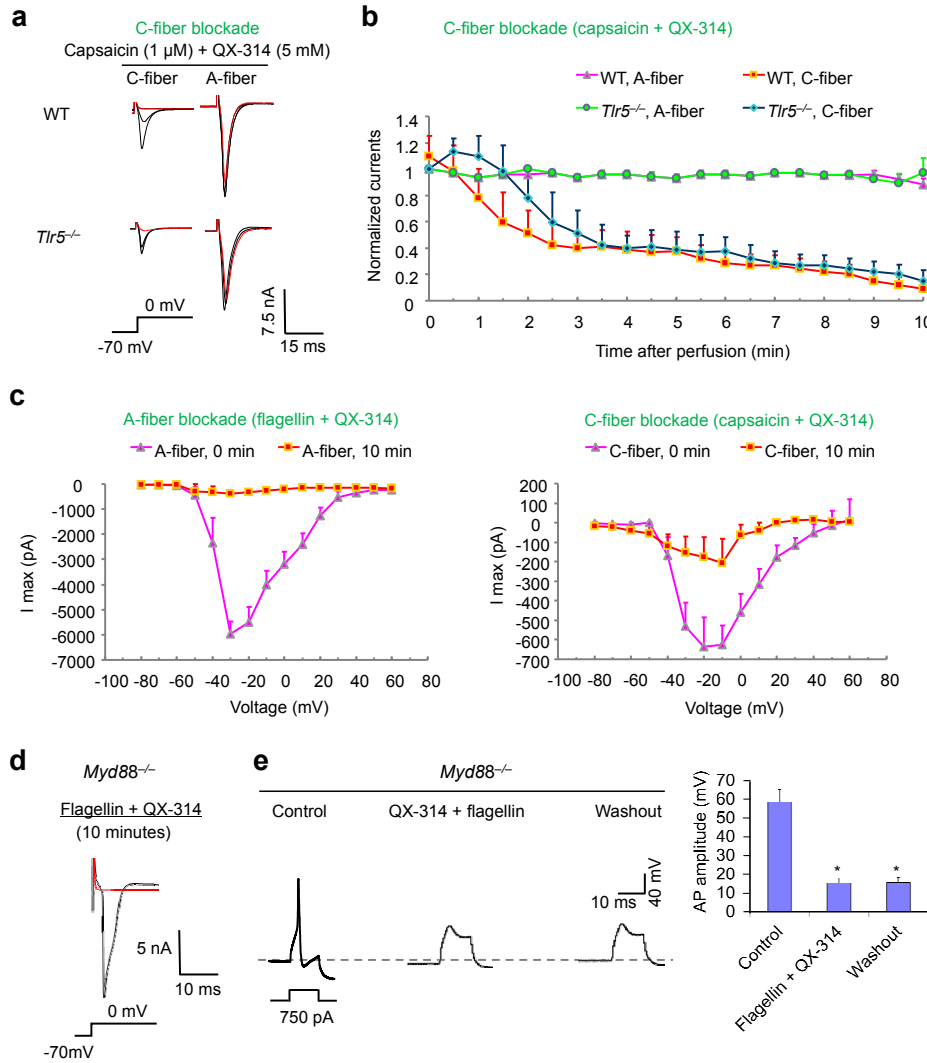
Supplementary Fig. 2. Co-expression of *Tlr5* mRNA with IB4 (a), CGRP (b), and TH (c) in mouse DRG neurons. *Tlr5* mRNA expression was revealed by in situ hybridization. Within the *Tlr5* population, 7.2% of DRG neurons express CGRP and 7.7% of DRG neurons express tyrosine hydroxylase (TH), but only 0.5% of DRG neurons bind IB4. Scale, 100 μm . 4-5 DRG sections per mouse and 3 mice per group were used for percentage quantification.



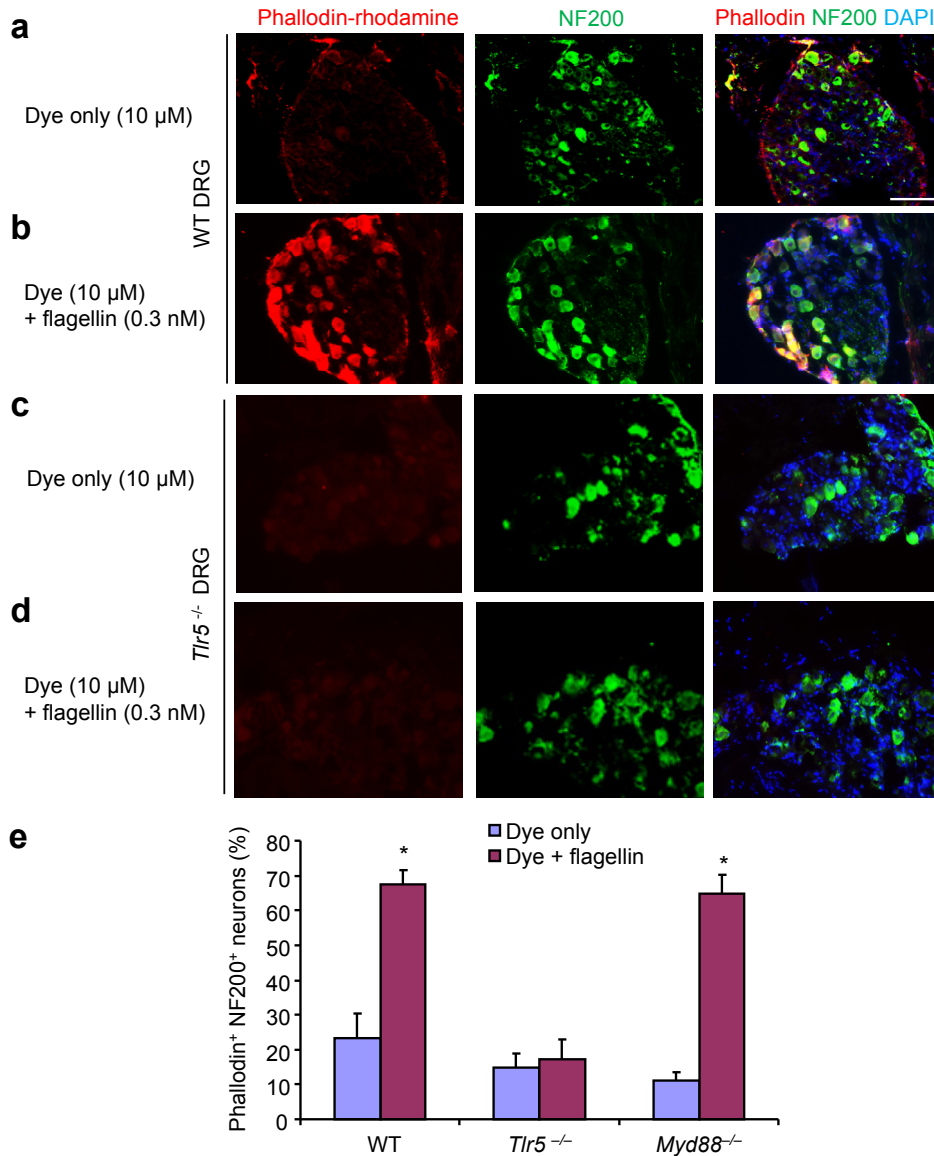
Supplementary Fig. 3. TLR5 expression in skin and spinal cord of WT and *Tlr5*^{-/-} mice. (a) TLR5 immunostaining is absent in glabrous skin of *Tlr5*^{-/-} mice. (b) Double staining of TLR5 and CK20, a marker for Merkel cells, in hindpaw skin. Note there is no TLR5 staining around Merkel cells. Scale, 100 μ m. The skin morphology was revealed by DAPI nuclear staining. (c) Double staining of TLR5 and NF200 in hairy skin of WT and *Tlr5*^{-/-} mice. TLR5-IR is present in nerve fibers surrounding the base of hair follicle. Yellow arrows indicate double-labeled nerve fibers. TLR5 staining is absent in *Tlr5*^{-/-} mice. Scale, 20 μ m. (d) Double immunostaining of TLR5 and CGRP in spinal cord dorsal horn. Box-1 and Box-2 in the dorsal horn are enlarged in low panels. 1a and 2a are merged images, and 1b and 2b are TLR5 single staining images. Note there is no obvious co-localization of TLR5 and CGRP in spinal cord axons and axonal terminals. Scale, 100 μ m.



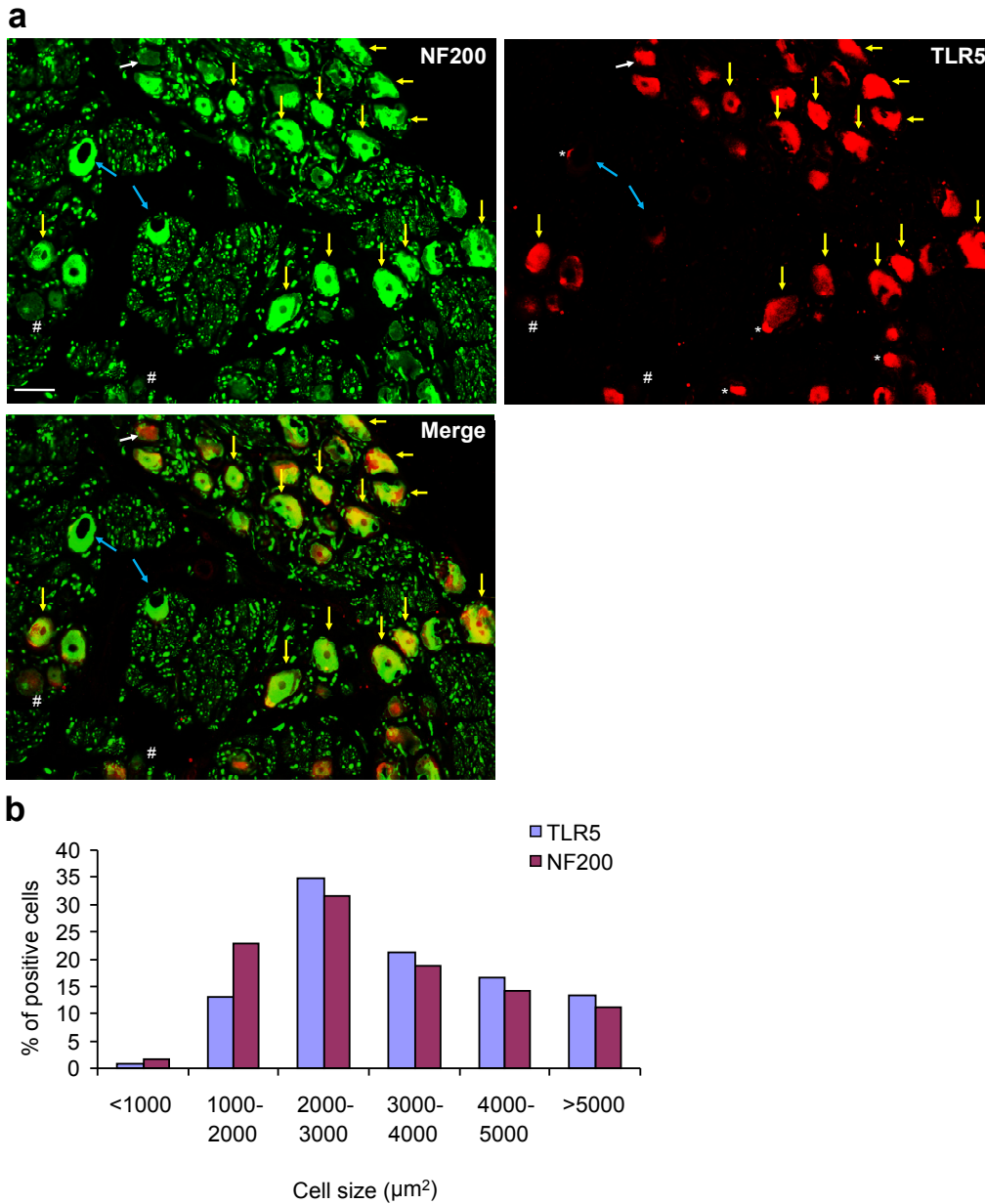
Supplementary Fig. 4. Summary of mouse and human DRG neurons' responses to A-fiber blockade. **(a)** C-fiber and **(b)** A-fiber DRG neurons' responses to A-fiber blockade (0.3 nM flagellin and 5 mM QX-314) in mice. $n = 22$ neurons for C-fibers and $n = 20$ neurons for A-fibers. Y-axis shows normalized sodium currents (amplitude, normalized to the baseline) at 0 min and 10 min after QX-314/flagellin treatment. Note a marked reduction in currents in A-fiber neurons (30-55 μm). A few C-fiber (10-25 μm) neurons also display moderate inhibition. **(c)** C-fiber and **(d)** A-fiber human DRG neurons' responses to A-fiber blockade (0.9 nM flagellin and 12 mM QX-314). $n = 8$ neurons per group.



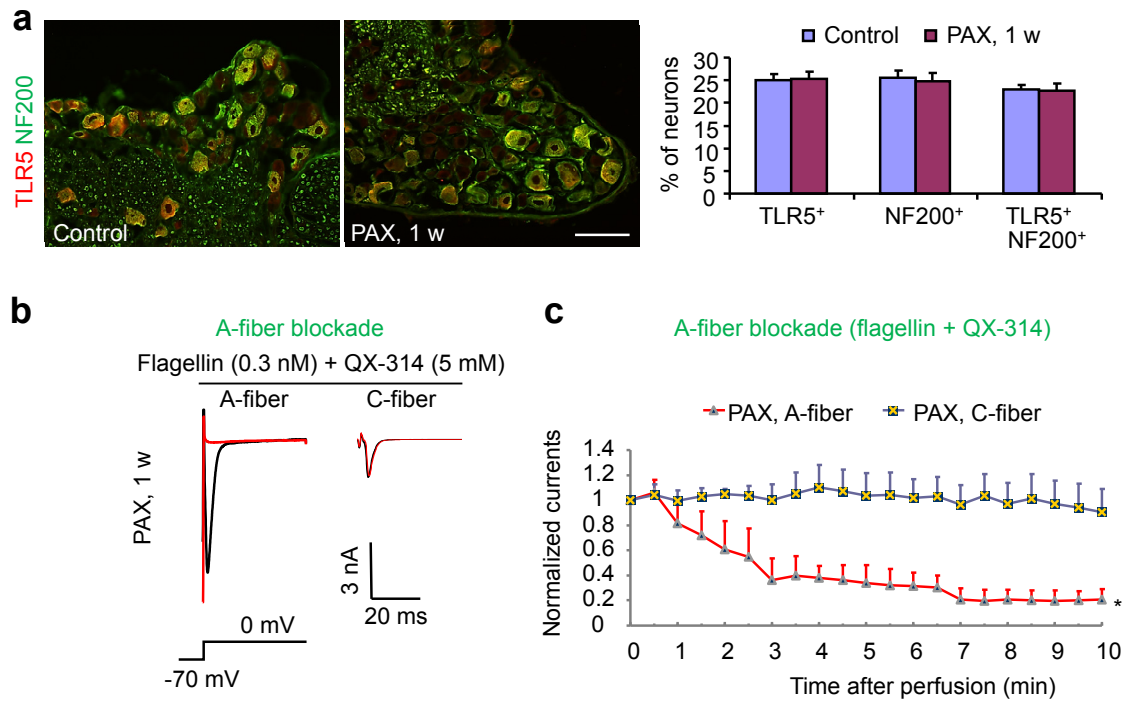
Supplementary Fig. 5. Effects of C-fiber blockade on sodium currents in small C-fiber and large A-fiber DRG neurons in WT and *Tlr5*^{-/-} mice and effects of A-fiber blockade in *Myd88*^{-/-} mice. **(a)** Traces of transient sodium currents in C- and A-fiber neurons of WT and *Tlr5*^{-/-} mice, before and after C-fiber blockade (1 μ M capsaicin / 5 mM QX-314). **(b)** Time course of sodium currents in C- and A-fiber neurons after C-fiber blockade. C-fiber blockade via capsaicin/QX-314 suppresses sodium currents and action potentials in small C-fiber but not large A-fiber DRG neurons. Note that the effect of C-fiber blockade is not affected in *Tlr5*-deficient neurons. * $P < 0.05$, two-way ANOVA. $n = 10$ neurons/group. **(c)** I/V curves of A-fiber blockade and C-fiber blockade. **(d)** Traces of transient sodium currents in A-fiber neurons of *Myd88*^{-/-} mice before and after flagellin/QX-314 (A-fiber blockade). **(e)** Traces of action potentials in A-fiber neurons of *Myd88*^{-/-} mice before and during flagellin/QX-314 treatment and after washout. Right, amplitude of action potentials. Note that A-fiber blockade retains its efficacy in *Myd88*^{-/-} mice. * $P < 0.05$, vs. control, $n = 10$ neurons/group. All data were expressed as mean \pm s.e.m.



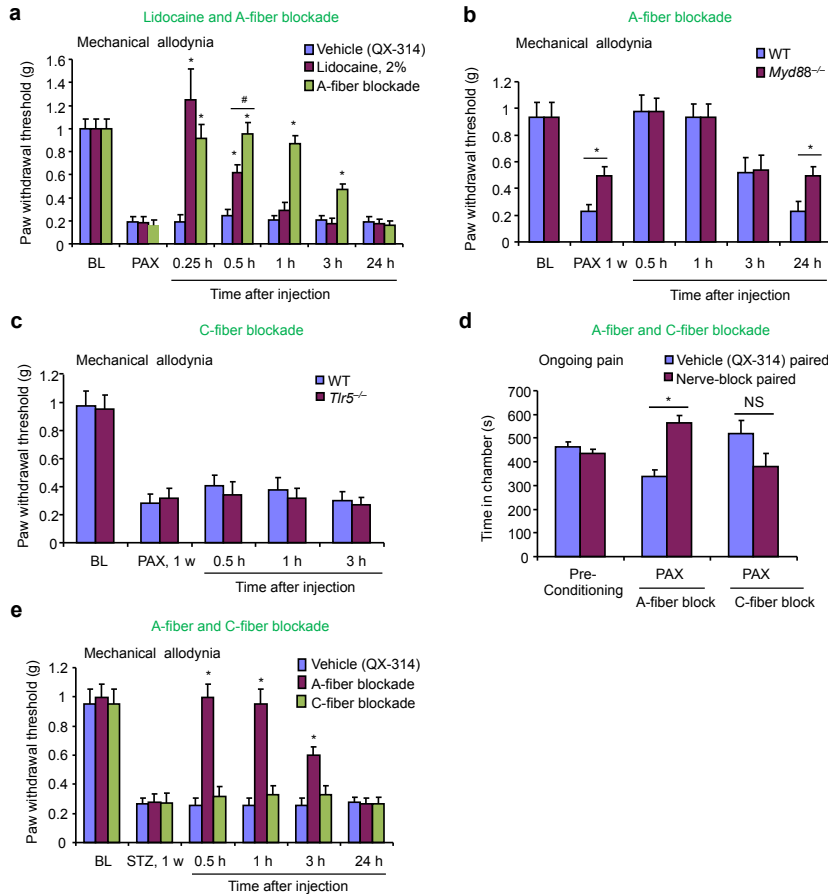
Supplementary Fig. 6. Flagellin/QX-314 treatment causes entry of phalloidin-rhodamine into large A-fiber DRG neurons via TLR5 activation. **(a-d)** Double labeling of phalloidin-rhodamine (red) and NF200 (green) in DRG sections of WT mice **(a,b)** and *Tlr5*^{-/-} mice **(c,d)** after treatment of flagellin **(a,c)** and flagellin/QX-314 **(b,d)**. Whole mount DRGs were treated with 0.3 nM flagellin or 0.3 nM flagellin / 5 mM QX-314 for 10 min. After the treatment, the DRGs were fixed and processed for NF200 immunostaining and DAPI DNA staining (blue). Note that only flagellin/QX-314 co-application induces phalloidin entry into NF200-expressing neurons in WT but not *Tlr5*^{-/-} mice. Scale, 100 μ m. **(e)** Percentage of phalloidin-rhodamine/NF200 double-labeled neurons following dye incubation with and without flagellin/QX-314 in WT, *Tlr5*^{-/-} mice, and *Myd88*^{-/-} mice. Note that phalloidin-rhodamine entry is blocked after *Tlr5* but not *Myd88* deficiency. **P* < 0.05, *n* = 3 mice/group. All data were expressed as mean \pm s.e.m.



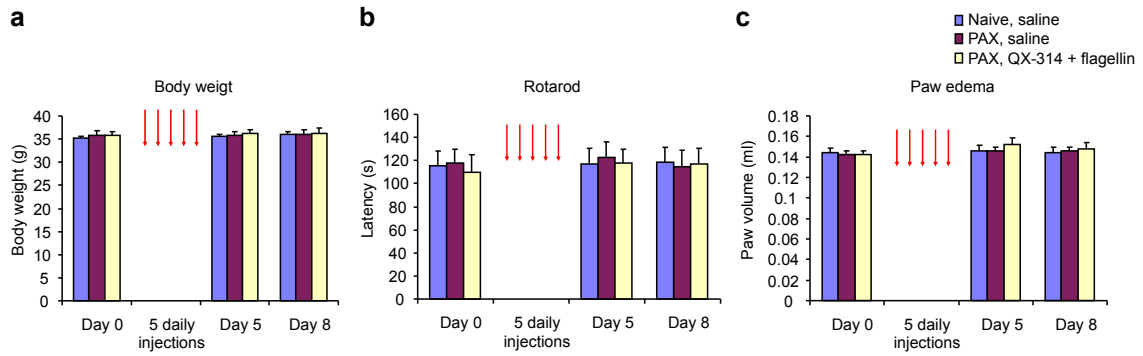
Supplementary Fig. 7. Co-localization of TLR5 with NF-200 in human DRG. **(a)** Double staining of TLR5 and NF200 in human L4 DRG section. Yellow, white, and blue arrows show double-labeled neurons, TLR5 single-labeled neurons, and NF200 single-labeled neurons, respectively. # denotes neurons that are TLR5- and NF200-negative. * indicates possible artifacts related to autofluorescence. Scale, 100 μm . **(b)** Size distribution of TLR5-IR and NF-200-IR neurons in human L4-L5 DRGs.



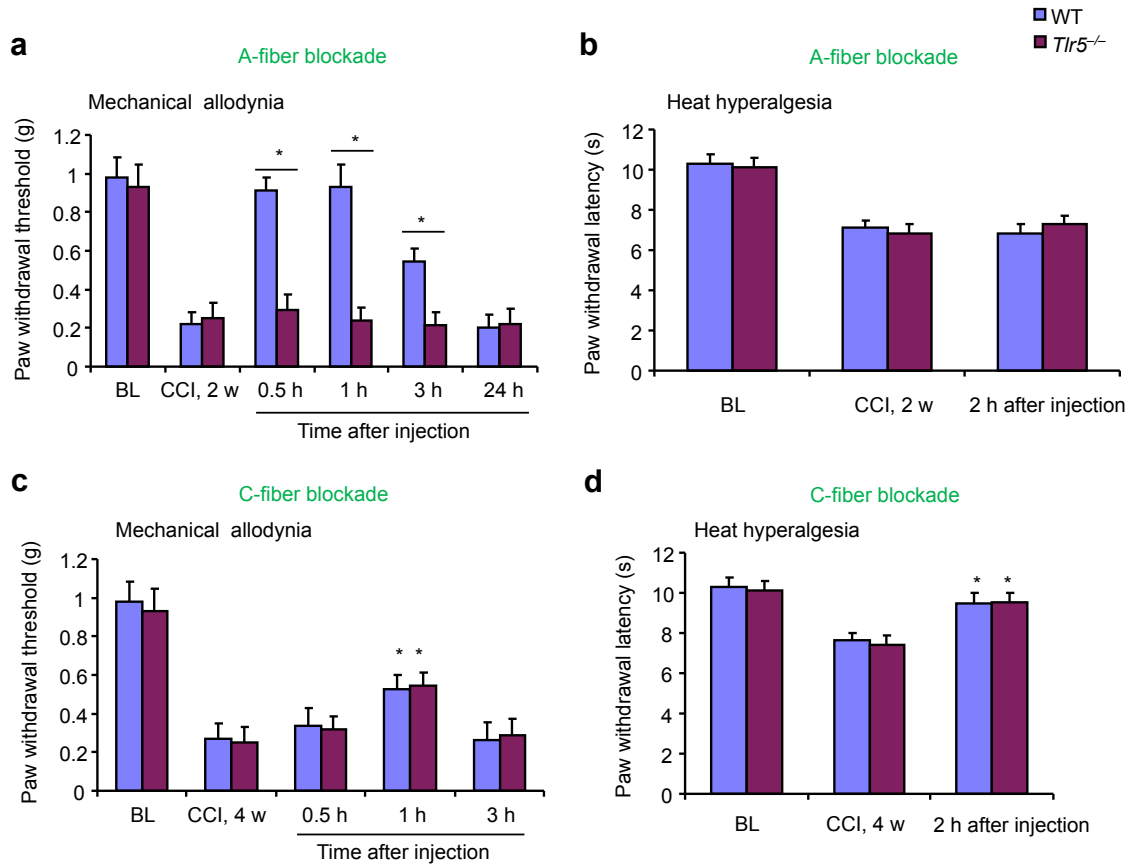
Supplementary Fig. 8. QX-314/flagellin treatment blocks A-fiber but not C-fiber activities in DRG neurons one week after paclitaxel (PAX) treatment. **(a)** Co-localization of TLR5 (red) and NF200 (green) in DRG sections of control mice and PAX-treated mice (1 w). Note that the percentage of DRG neurons expressing TLR5, NF200, and TLR5/NF200 is similar in control and PAX-treated mice. $n = 4$ mice / group. Scale, 100 μm . **(b)** Traces of transient sodium currents in C- and A-fiber neurons of WT mice, before and after the flagellin/QX-314 treatment. **(c)** Time course of sodium currents in C- ($n = 5$) and A-fiber ($n = 8$) neurons after the A-fiber blockade. $*P < 0.05$, two-way ANOVA. All data were expressed as mean \pm s.e.m.



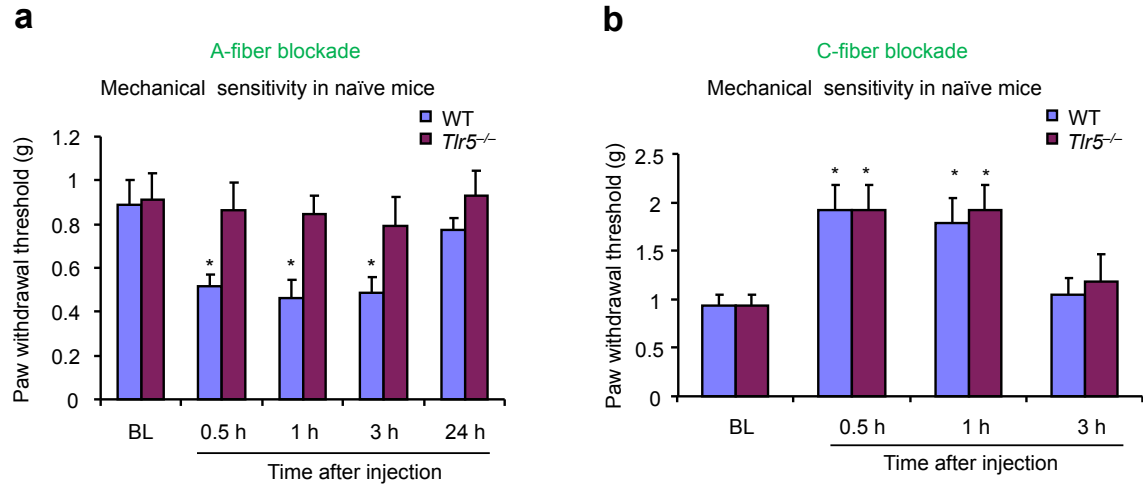
Supplementary Fig. 9. Effects of A-fiber blockade, C-fiber blockade, or lidocaine on paclitaxel (PAX)-induced mechanical allodynia and ongoing pain in WT, *Tlr5*^{-/-}, *Myd88*^{-/-} mice. **(a)** Comparison of the effects of flagellin/QX-314 (A-fiber block) and 2% lidocaine on PAX-induced mechanical allodynia in WT mice. Note that lidocaine only produces a very transient reversal of allodynia (< 1 h). Behavior was tested 15 min after lidocaine treatment to avoid lidocaine-evoked motor impairment (<10 min). **P* < 0.05, vs. vehicle, #*P* < 0.05, two-way ANOVA followed by post-hoc Bonferroni test. *n* = 5 mice/group. **(b)** Reversal of PAX-induced mechanical allodynia by A-fiber blockade in *Myd88*^{-/-} mice. PAX-induced mechanical allodynia is partially reduced after *Myd88* deletion. **P* < 0.05, vs. vehicle, two-way ANOVA followed by post-hoc Bonferroni test. *n* = 5 mice/group. **(c)** No effects of C-fiber blockade on PAX-induced mechanical allodynia in WT and *Tlr5*^{-/-} mice. *n* = 5 mice/group. **(d)** CPP shows that mice spend more time in A-fiber blockade (*n* = 7 mice) paired chamber. Note C-fiber blockade (*n* = 5 mice) fails to show significant effects on CPP. **P* < 0.05, N.S, no significance. A-fiber and C-fiber blockade was induced by flagellin (0.3 μg) / QX-314 (0.2%) or capsaicin (10 μg) / QX-314 (0.2%), respectively, via intraplantar route. Lidocaine was also given via intraplantar injection. **(e)** Distinct effects of intraplantar A-fiber blockade vs. C-fiber blockade on STZ-induced mechanical allodynia. **P* < 0.05, two-way ANOVA followed by post-hoc Bonferroni test. *n* = 5 mice/group. All data were expressed as mean ± s.e.m.



Supplementary Fig. 10. Effects of repeated intraplantar injections of flagellin (0.3 μ g) and QX-314 (0.2%) on body weight, motor function (evaluated by Rotarod test), and paw inflammation (assessed by paw volume) in naïve and paclitaxel (PAX)-treated mice. n=5 mice/group. All data were expressed as mean \pm s.e.m.



Supplementary Fig. 11. Distinct effects of A-fiber and C-fiber blockade on CCI-induced mechanical allodynia and heat hyperalgesia in WT and *Tlr5*^{-/-} mice. **(a,b)** Reversal of CCI-induced mechanical allodynia **(a)** but not heat hyperalgesia **(b)** by A-fiber blockade in WT but not *Tlr5*^{-/-} mice. **P* < 0.05, two-way ANOVA followed by post-hoc Bonferroni test. *n* = 5 mice/group. **(c,d)** Effects of C-fiber block on CCI-induced mechanical allodynia **(c)** and heat hyperalgesia **(d)** in WT and *Tlr5*^{-/-} mice. Note that C-fiber block produces complete reversal of CCI-induced heat hyperalgesia but only mild reversal of CCI-induced mechanical allodynia and the blocking effect is *Tlr5*-independent. **P* < 0.05, two-way ANOVA followed by post-hoc Bonferroni test. *n* = 5 mice/group. A-fiber and C-fiber blockade was induced by flagellin (0.3 μg) / QX-314 (0.2%) or capsaicin (10 μg) / QX-314 (0.2%), respectively, via intraplantar route. All data were expressed as mean ± s.e.m.



Supplementary Fig. 12. Different effects of intraplantar A-fiber blockade (**a**) and C-fiber blockade (**b**) on baseline mechanical sensitivity in naïve WT and *Tlr5*^{-/-} mice. A-fiber blockade causes *Tlr5*-dependent mechanical hypersensitivity (allodynia). In contrast, C-fiber blockade causes *Tlr5*-independent mechanical hyposensitivity (analgesia) in naïve mice. A-fiber and C-fiber blockade was induced by flagellin (0.3 μ g) / QX-314 (0.2%) or capsaicin (10 μ g) / QX-314 (0.2%), respectively, via intraplantar route. * $P < 0.05$, compared to baseline (BL), two-way ANOVA followed by post-hoc Bonferroni test. $n = 5$ mice/group. All data were expressed as mean \pm s.e.m.