

Supplementary information

Critical POU domain residues confer Oct4 uniqueness in somatic cell reprogramming

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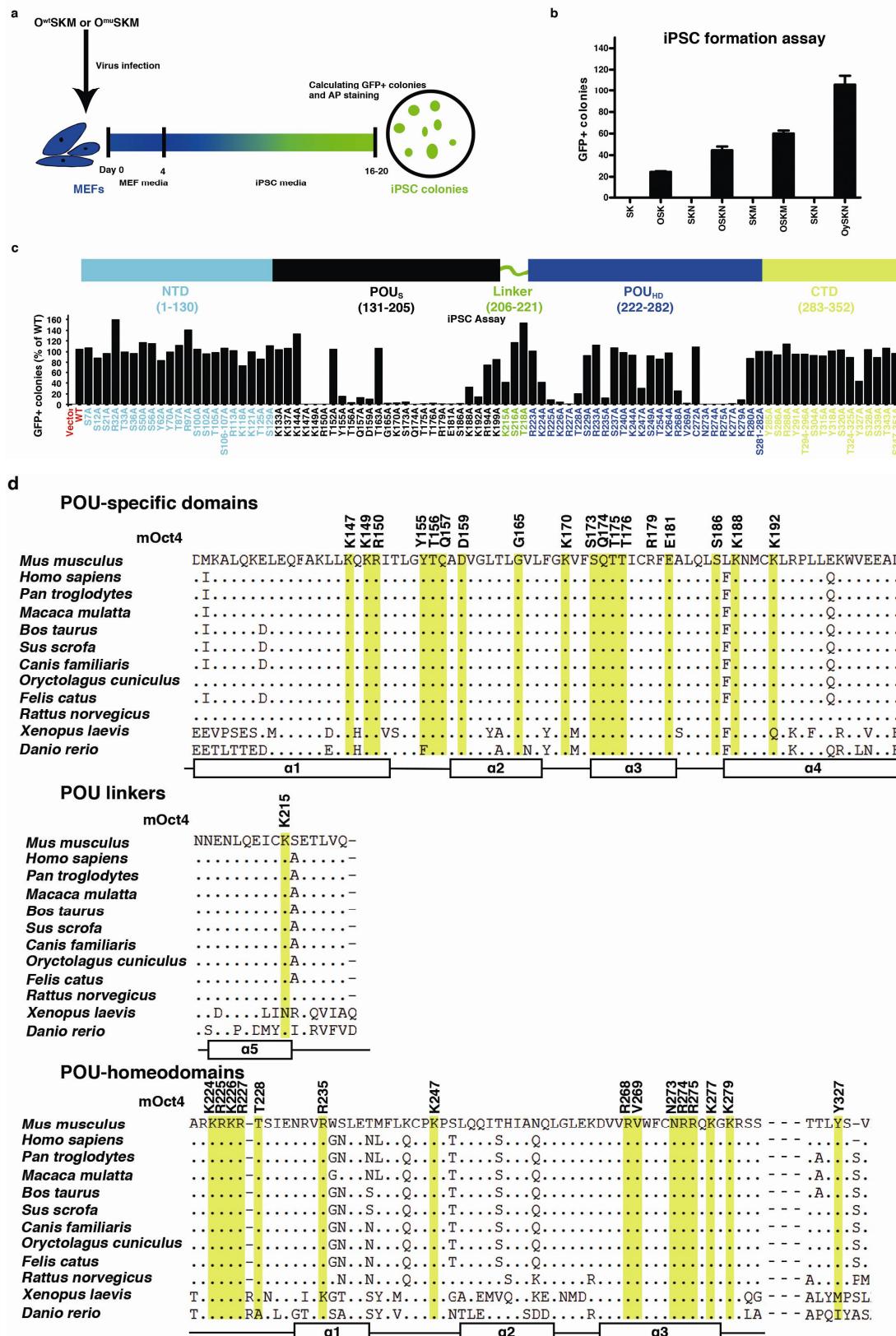
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Supplementary figures



e

POU-specific domains

	mOct4	K147 K149 R150	Y155 Y156 Q157 D159	G165	K170	S173 Q175 T176	R179	E181	S186 K188	K192
pou5f1	DMKALQKELEQ-FAKLLKQKRITLGYTCADVGLTLG--VLFGK-VFSQTTICR-----FEALQLSLKNMCKLRPL-LEKWVEEAD---									
pou5f1(n)	I.....-.....	K149 R150						F.....	Q.....	-Q.....
pou1f1	MDSPEIR.....NEF.VR.K.....TN.EA.A--AVH.S-E.....							N.....F.....A.....KAI-S.L..E-----		
pou2f1	EEPSDLE.....TF.R.K.F.G...AM--K.Y.N-D.....							N.....F.....K.....LND.E-----		
pou2f2	EEPSDLE.....RTF.R.K.F.G...AM--K.Y.N-D.....							N.....F.....K.....LND.E-----		
pou3f1	.EPTDLE...K-TF.R.K.F.G...AM--K.Y.N-D.....							N.....F.....K.....LND.E-----		
pou3f2	EDTPSDD...-QF.R.K.F.....A--T.Y.N.....							F.....K-N.L.....		
pou3f3	EDTPSDD...-QF.R.K.F.....A--T.Y.N.....							F.....K-N.L.....		
pou3f4	EETPTSD...-QF.R.K.F.....A--T.Y.N.....							F.....K-N.L.....		
pou4f1	.SDTDPR...A-ERF.R.K.V...SA.ANLKIP.VGSL..S.....							S.T.HN.IA.K.I-QA.L..E-----		
pou4f2	.VD.DPRD..A-ERF.R.K.V...SA.ANLKIP.VGSL..S.....							S.T.HN.IA.K.I-QA.L..EKSHR		
pou4f3	.VESDPR...A-ERF.R.K.V...AA.ANLKIP.VGSL..S.....							S.T.HN.IA.K.V-QA.L..E-----		
pou5f2	.VS.I...M-L.E.R.M.S...FAV--AM--L.....							Q.....A.W.....KM.L.V.....		
pou6f1	EDGINLE.IRE...NF.IR.LS.L.TQ.QA.T--ATE.P-AY..SA.....							K.DITP.SAQ.K.V....IM.E-----		
pou6f2	VDGVNLE.IRE...AF.IR.LS.L.TQ.QA.S--ATE.P-AY..SA.....							H--R.K.DITP.SAQ.IK.V...R.MA..E		
pou2af1	-EYVSHEAVSCPYSTEMVQVCPS..VVGPSSV.T--YASPPLITNV.FRST-----ATPAVGQELEGFEHQAPLTYYFPWPQ-----									

POU linkers

	mOct4	K215
pou5f1	NNENLQEI K SETLVQ-----	
pou5f1(n)A.....	
pou1f1	QVGA.YNEKVGAN-----	
pou2f1	.LSSDSTASSPSA.NSPGLGAEGLN-	
pou2f2	TMSVDSLPSPNQ.SSPSGLFDGLPG	
pou2f3	SSPSDPSASTPSSYPTLSEVFG---	
pou3f1	SSSGSPTNL K IAAQG-----	
pou3f2	SSSGSPTSID K IAAQG-----	
pou3f3	SSTGSPTSID K IAAQG-----	
pou3f4	SSTGSPTSID K IAAQG-----	
pou4f1	GAQREKMNKP E LFNGG-----	
pou4f2	EKLTKP.LF N GA-----	
pou4f3	AAYREKNSK P ELFNGS-----	
pou5f2	EKNL.GICRMEMI.E-----	
pou6f1	LRNQEQQNIM.FVGGEPS-----	
pou6f2	ARHRAGMQN I T.FIGSEPS-----	
pou2af1	PLST.PTSS L QYQPP-----	

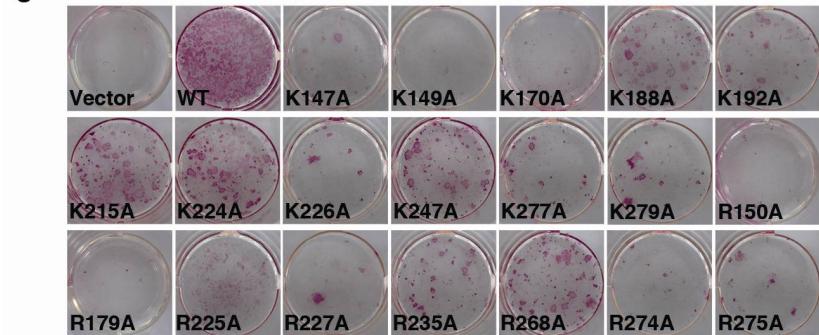
POU-homeodomains

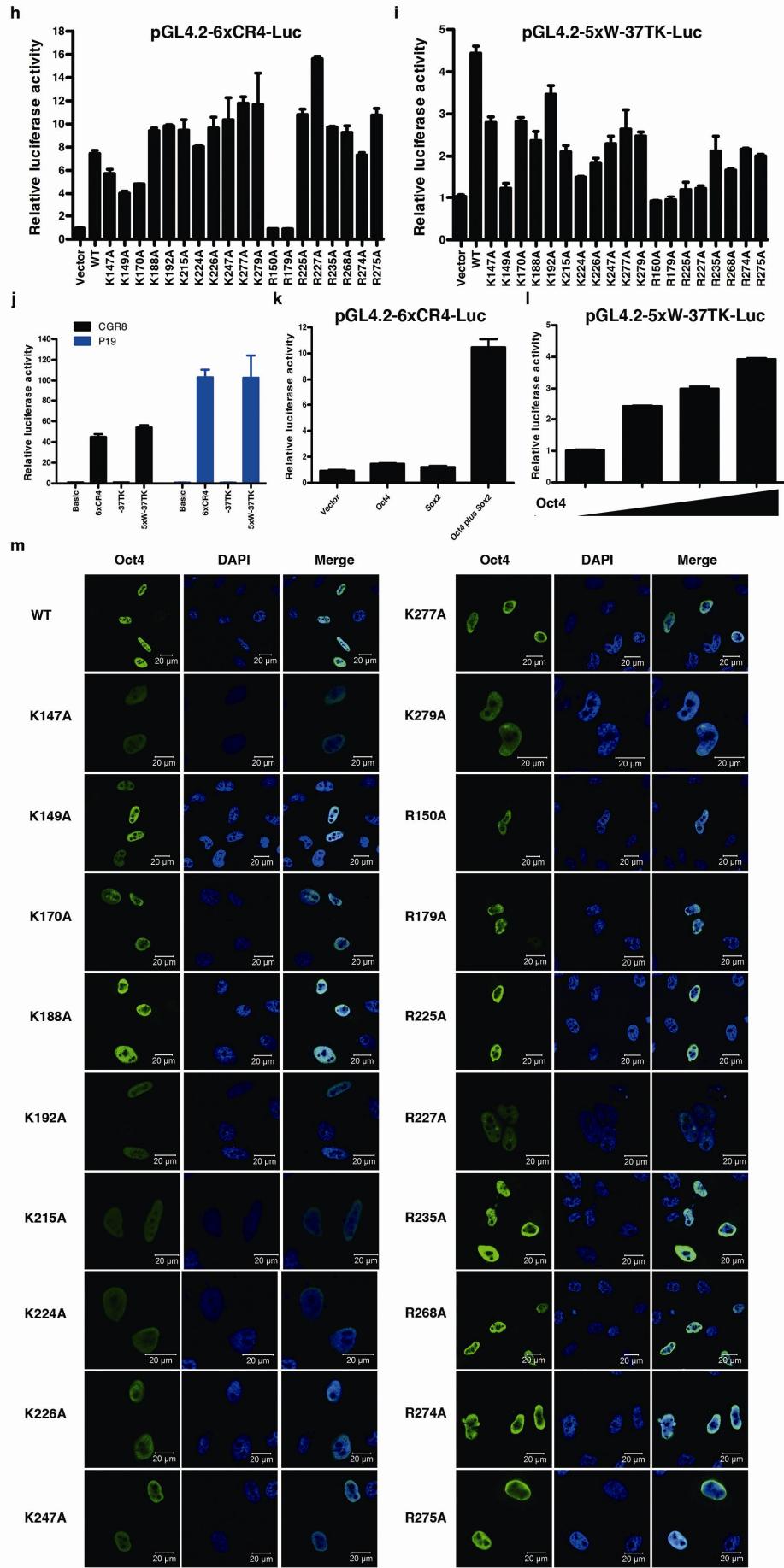
	mOct4	K224 R225 R226 R227 R228	R235	K247	R268 V269	N273 R274 R275 K277	K279	Y327	
pou5f1	ARKRKRTSIENRVWSLETMF L KCP K PSLQQITHIANQLGLEKD V RVWFC N RR R KGKRSS-----TTLY S -V								
pou5f1(n)GN.NL.Q.....T.....S...Q.....							A.....S.	
pou1f1	RKRKR...T.SVAAKDA.RH.GEHS...S.E.MRM.EE.N..E.....							RE.VK-----	
pou2f1	R.RK.....TNI.VA.KS.MENQ..TSED..L..E..NM..E.I.....							E.IN-----SAST.EA	
pou2f2	R.RK.....TN.FA.KS.ANQ..TSEE..LL..E..HM..E.I.....							E.IN-----GGGGGGG	
pou2f3	RKRK.....TNI.LT.KR.QDN...SEE.SM..E..SM..E.....							E.IN-----SG.HASS	
pou3f1	RKRK.....VG.KGA.SH.....AHE..GL.DS.Q...E.....							E.MT-----HHHHHTL	
pou3f2	RKRK.....VS.KGA.SH.....A.E..SL.DS.Q...E.....							E.MT-----HHG.	
pou3f3	RKRK.....VS.KGA.SH.....A.E..NL.DS.Q...E.....							E.MT-----HHGL	
pou3f4	RKRK.....VS.KGV..H.....AA.E.SSL.DS.Q...E.....							E.MT-----SCHD	
pou4f1	EK.....AAPEKR..AY.AVQ.R..SEK.AA..EK.D.K.N.....							Q.....MK-----	
pou4f2	EK.....AAPEKR..AY.AIQ.R..SEK.AA..EK.D.K.N.....							Q.....MKY-----	
pou4f3	E.....AAPEKR..AY.AIQ.R..SEK.AA..EK.D.K.N.....							Q.....MK-----	
pou5f2R.A.R.R.IGSN..KL.Q..E.TP...SY..GR.R.Q..L.Q..S..S.M.SWPT-----P...ST								
pou6f1	KKRKR...FTPQAIEA.NAY.E.N.L.TG.E..E..KE.NYDRE.....							TL.NT-----	
pou6f2	KKRKR...FTPQAIEI.NAH.E.NTH..G.EM.E..EK.NYDRE.....							K.AL.NTI-----	
pou2af1	.PTLSGPQFVQLPISIP.PVLQEMDD.RRAISSLTIDK.L..EEESNTYEL.HTL S VEGF-----								

f

H-bond \ AA	K147	K192	K247	K277	K279	R150	R179	R227	R268	R275
DNA										
backbone	✓	✓	✓	✓	✓	✓			✓	✓
base							✓	✓		

g



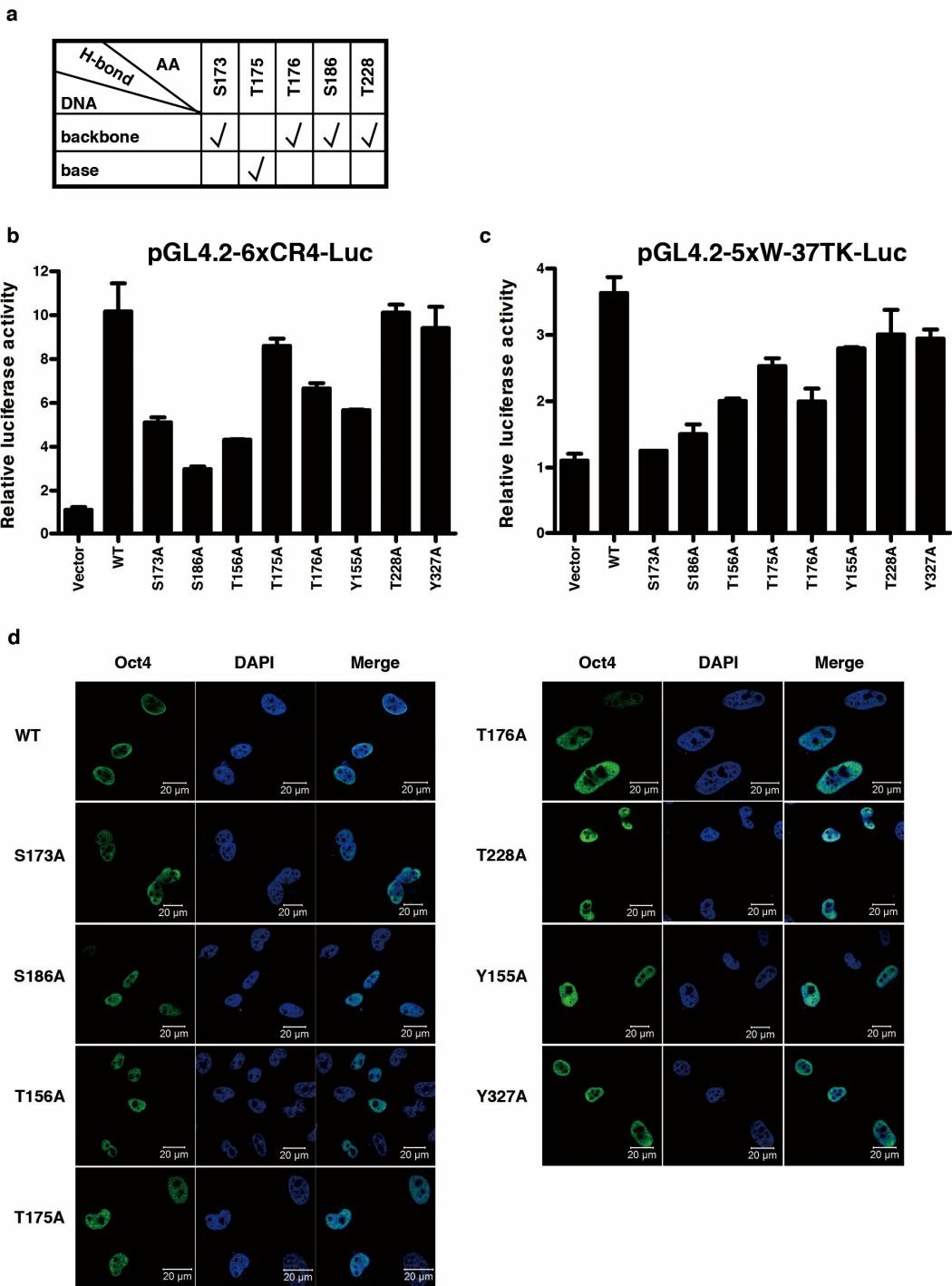


Supplementary Figure S1 Identification of critical Oct4 residues for somatic cell reprogramming by alanine scanning.

- (a) Schematic of experimental designs for library construction of mOct4 mutants and identification of the functional residues required for reprogramming activity.
- (b) GFP+ iPSC formation by different combination of reprogramming factors. Data are representative of two independent experiments. Statistics analysis was performed using a t-test (mean and s.d. of duplicate assays).
- (c) Percentage of GFP+ iPSC formation by the 90 mOct4 alanine mutants in the library. Data are representative of at least two independent reprogramming experiments for each mutant.
- (d) Sequence alignment of 12 different Oct4 POU proteins. Residues that are defective in reprogramming after mutating to alanine are highlighted in yellow.
- (e) Sequence alignment of 16 different murine POU proteins. Residues that are defective in reprogramming after mutating to alanine are highlighted in yellow.
- (f) Table to show the interaction between K/R residues and DNA in Figure 1b.
- (g) AP staining of OSKM-based reprogramming defective Oct4 K-to-A and R-to-A mutants.
- (h and i) The relative transcriptional activities of reprogramming defective K-to-A and R-to-A mutants were measured using 6xCR4-Luc and 5xW-Luc reporters, respectively. Data are representative of at least three independent experiments. Statistics analysis was performed using a t-test (mean and s.d. of duplicate assays).
- (j) Test of the 6xCR4-Luc or 5xW-Luc reporters in CGR8 ES cells and P19 EC cells. The Basic or -37TK luciferase enhancer-less vectors were served as negative controls.
- (k) Test of the 6xCR4-Luc in HEK293 cells. 6xCR4-Luc was transfected into HEK293 cells together with a mock vector, Oct4, Sox2, or Oct4 plus Sox2.

(l) Test of 5xW-Luc in HEK293 cells. 5xW-Luc was transfected into HEK293 cells together with a mock vector or increasing amounts of Oct4. A pRL-CMV renilla reporter was included in all transfection for normalization. Data (**j**, **k** and **l**) are representative of three independent experiments. Statistics analysis was performed using a t-test (mean and s.d. of duplicate assays).

(m) Nuclear localization of reprogramming defective mOct4 K-to-A and R-to-A mutants. K-to-A and R-to-A mutants were transfected into HeLa cells grown on coverslips. Immunostaining of Oct4 was performed 36 h after transfection with DAPI counterstaining.



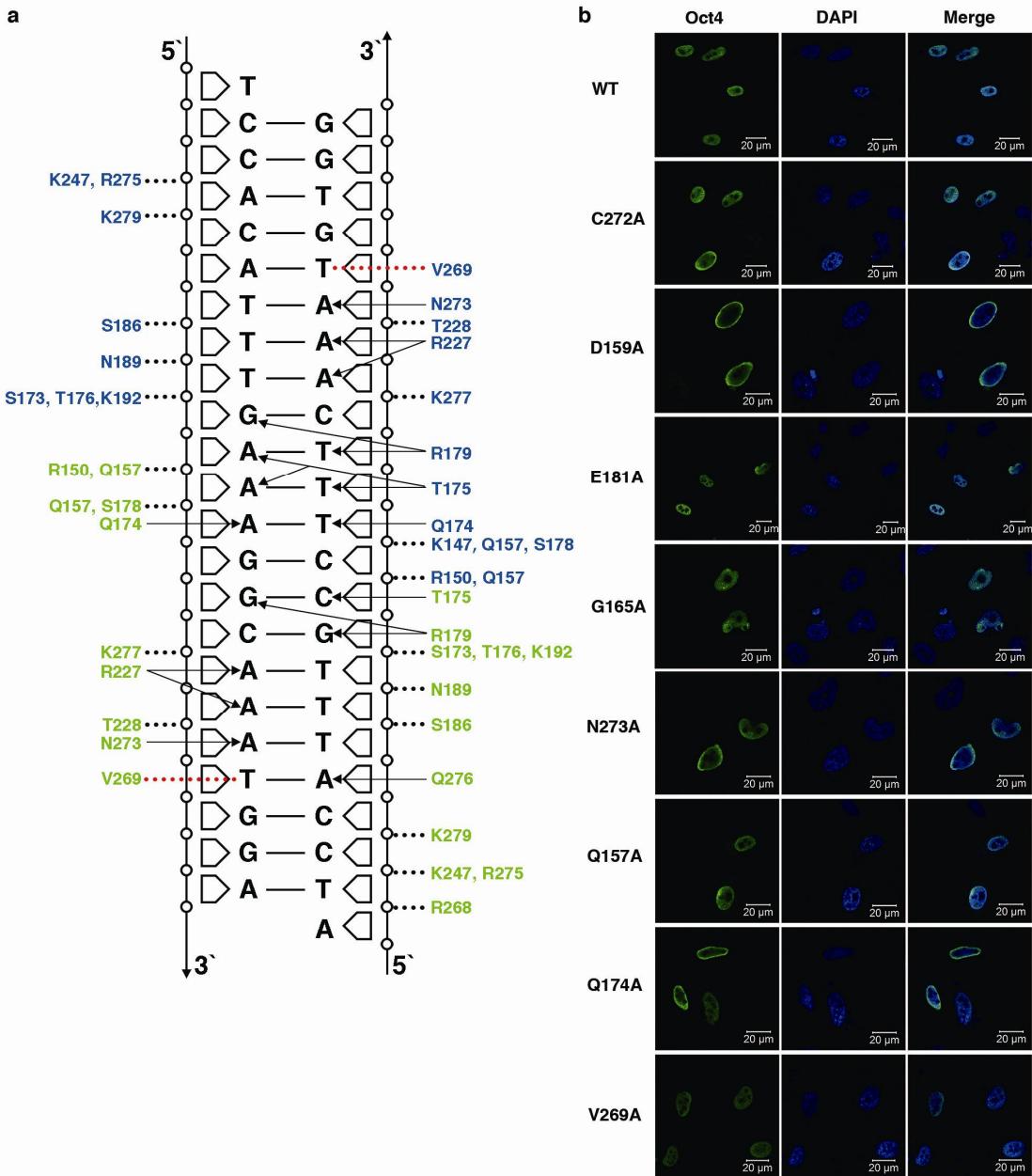
Supplementary Figure S2 Characterization of reprogramming defective mOct4 S-to-A, T-to-A and Y-to-A mutants.

- (a) Table to show the interaction between S/T/Y residues and DNA in Figure 2b.
- (b and c) The relative transcriptional activities of reprogramming defective S-to-A, T-to-A and Y-to-A mutants were measured using 6xCR4-Luc and 5xW-Luc reporters,

respectively. Data are representative of at least three independent experiments.

Statistics analysis was performed using a t-test (mean and s.d. of duplicate assays).

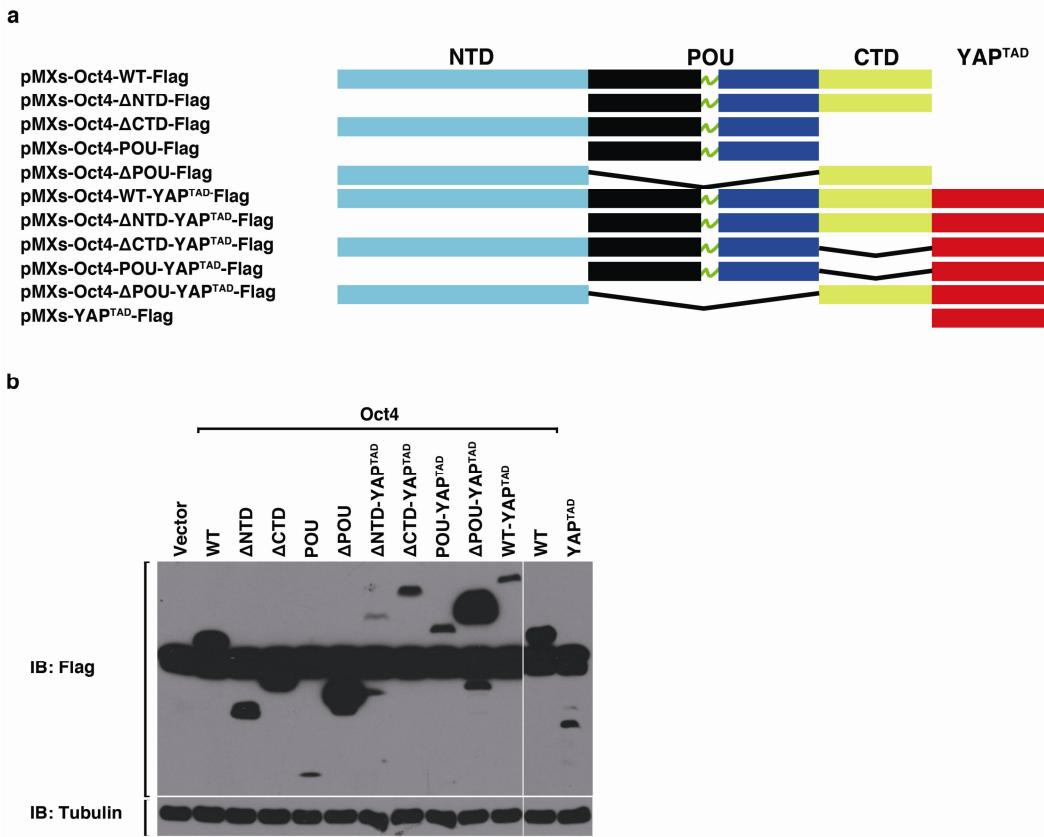
(d) Nuclear localization of reprogramming defective mOct4 S-to-A, T-to-A and Y-to-A mutants. The mutants were transfected into HeLa cells grown on coverslips. Immunostaining of Oct4 was performed 36 h after transfection with DAPI counterstaining.



Supplementary Figure S3 Characterization of mOct4 critical DNA binding residues.

(a) A sketch showing interactions or non-polar contacts between mOct4 residues and DNA in model of Oct4^{POU}:PORE complex. In the figure, interacting residues within the chain A (3L1P) are in blue, while interacting residues within the chain B are in green. Hydrogen bonds are shown by black dot lines for residues interacting with DNA phosphate and by solid arrows for those with DNA base. Non-polar contacts are shown by red dot lines.

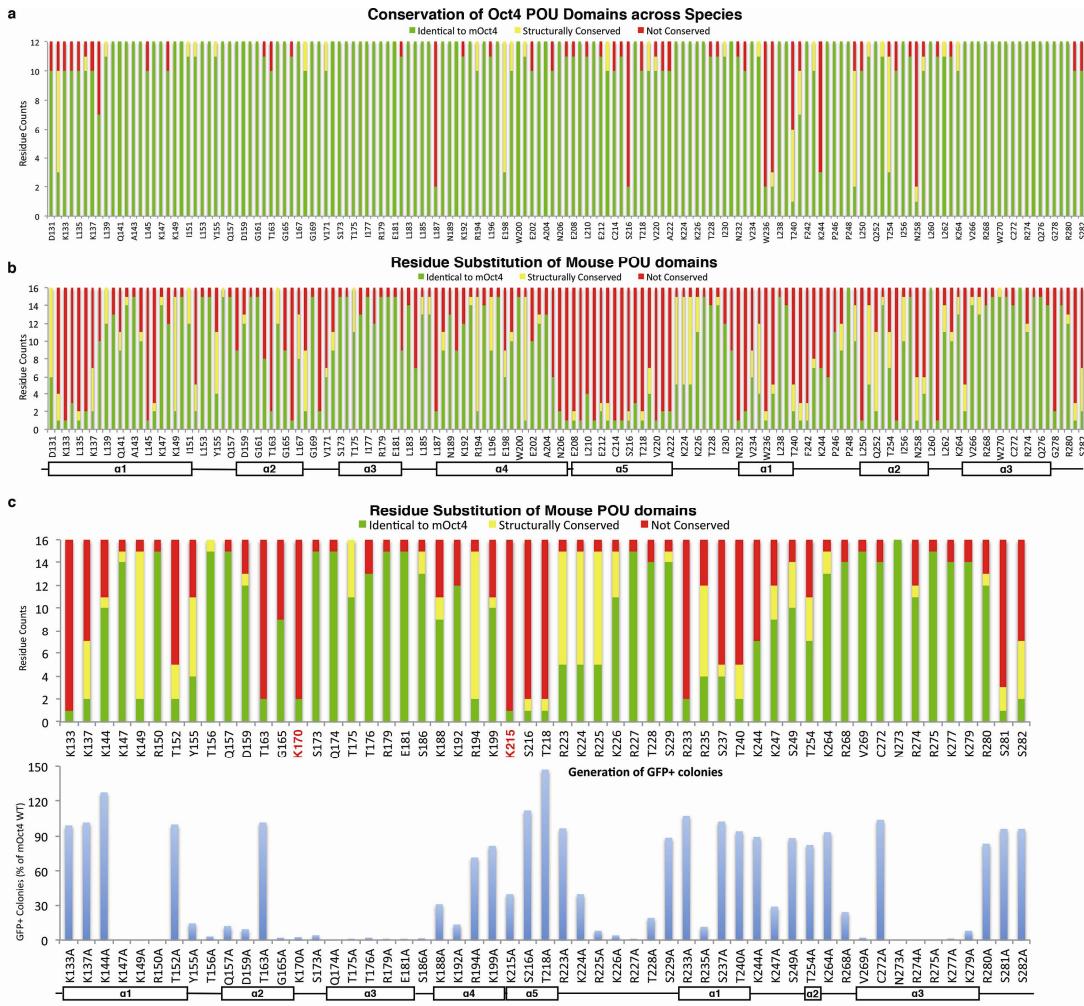
(b) Nuclear localization of reprogramming defective mOct4 mutants. The mutants were transfected into HeLa cells grown on coverslips. Immunostaining of Oct4 was performed 36 h after transfection with DAPI counterstaining.



Supplementary Figure S4 POU domain of Oct4 is the minimal requirement for generating iPS cells.

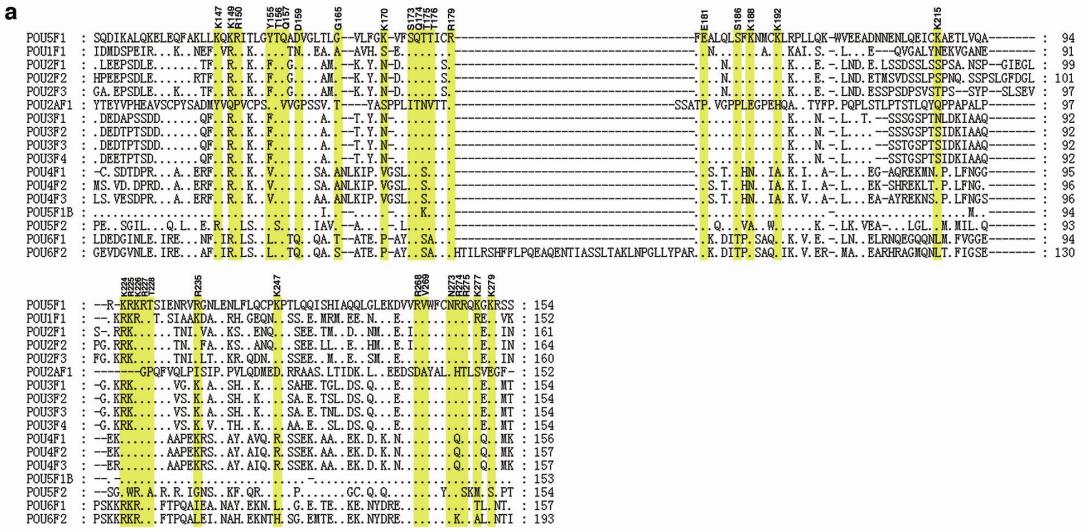
(a) Diagrams of Oct4 WT, WT-YAP^{TAD} and their deletion constructs.

(b) Protein stability of Oct4 WT, WT-YAP^{TAD} and their deletion constructs in MEFs.

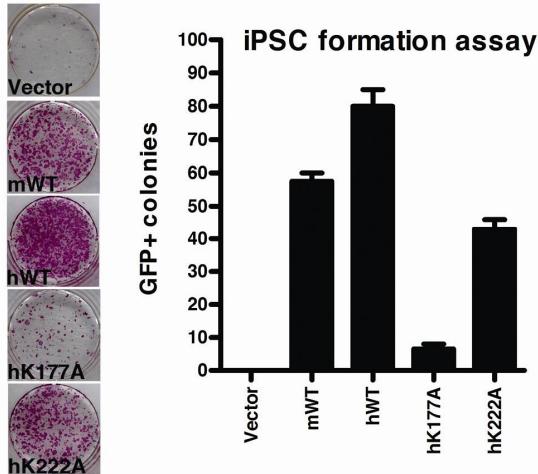


Supplementary Figure S5 Specific residues make Oct4 unique in reprogramming.

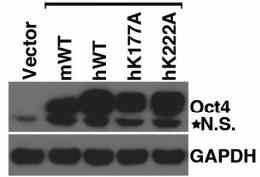
- (a) Conservation of Oct4 POU domains across 12 species. The numbering is in accord with mOct4.
- (b) Residue substitution among the 16 murine POU domain-containing members. The numbering is in accord with mOct4.
- (c) Substitution of residues tested in our alanine mutant library collection among the 16 murine POU domain-containing members. For easier viewing, the effects of the Oct4 mutants in iPSC assay are shown in the bottom panel. The numbering is in accord with mOct4.



b



c



Supplementary Figure S6 Specific residues make human Oct4 unique in reprogramming.

- (a) Sequence alignment of human POU domain-containing proteins. The numbering at the top is according to mouse Oct4 numbering.
- (b) AP staining of GFP+ colonies formed by human Oct4 WT, K177A and K222A mutants. Counting of GFP+ colonies is shown at the right. Data are representative of two independent experiments. Statistics analysis was performed using a t-test (mean and s.d. of duplicate assays).
- (c) Protein stability of hOct4 WT, the K177A and the K222A mutants in MEF cells.

Supplementary Table 1: Summary of key Oct4 residues

GFP+ colonies(% of WT)		Defective residues information (compared to WT)			Oct4 ^{POU} :PORE complex crystal structure (3L1P) was analyzed using Pymol software and analysis results are shown below.			putative or reported post translation modification sites	
		Reporter assay						PTM sites	Source
	Protein stability in MEF	6xCR4	5xW	Localization in nucleus	based on POU domain highly conserved between POU family members and available 1HF0 crystal structure, we repaired side chains of K188, K192,	Residues directly interact with DNA(backbone phosphate or base)	Residues directly interacting with functional residues are shown in black which were not tested in this study, and in blue which were already included in our iPSC screening and shown strong defects in reprogramming experiments. Five C to S mutants from crystal		

						K226, K277 and K279 in 3L1P.		structure are shown in red.	
Vector	0								
WT	100								
S7A	102.5 3								
S12A	84.18							putative phosphorylation site	PhosphoSite Plus
S21A	92.41								
R32A	152.9 4								
T33A	95.06								
S36A	92.41								
S50A	112.0 3								
S56A	110.1 3								
Y62A	79.75								
Y70A	94.94								
T87A	106.9 6							reported phosphorylation site	<i>Proc. Natl. Acad. Sci. U.</i>

									in human ES cells (human Oct4-T92)	S. A. 109 :7162- 7168.
R97A	134.5 7									
S100A	100								reported phosphorylation site in human ES cells (human Oct4-S105)	<i>Proc. Natl. Acad. Sci. U. S. A.</i> 109 :7162- 7168.
S102A	91.77								reported phosphorylation site in human ES cells (human Oct4-S107)	<i>Proc. Natl. Acad. Sci. U. S. A.</i> 109 :7162- 7168.
T105A	94.12									
S106- 107A	101.9								reported phosphorylation site in human ES cells (human Oct4-S111)	<i>Proc. Natl. Acad. Sci. U. S. A.</i> 109 :7162- 7168, <i>J. Biol. Chem.</i> 287 :38279- 38288.
R113A	97.5									
K118A	70.34								reported sumoylation site	<i>J. Bio. Chem.</i> 282 :21551- 21560; <i>FASEB J.</i> 21 :3042-51.
K121A	95.33									
T125A	82.35									
S129A	106.3 3									

K133A	99.17									putative ubiquitination site	PhosphoSite Plus
K137A	101.6 7									putative ubiquitination site	PhosphoSite Plus
K144A	127.5						H-bond to backbone phosphate (4.4 Å)			putative ubiquitination site	PhosphoSite Plus
K147A	0	low	low	low	YES			H-bond to A143, I151			
K149A	0	extremely low	low	extremely low	YES			H-bond to L-145, L146, L153			
R150A	0	low	extremely low	extremely low	YES		H-bond to backbone phosphate	H-bond to L146, G154, Y155 , E181			
T152A	100										
Y155A	14.36	slightly low	low	sightly low	YES			H-bond to R150 , E208, Q211			
T156A	3.08	slightly low	low	low	YES			H-bond to D159 , V160	putative phosphorylation site	PhosphoSite Plus	
Q157A	12	normal	extremely low	extremely low	YES		H-bond to backbone phosphate	H-bond to G161, E181			
D159A	9.33	normal	slightly low	slightly low	YES			H-bond to T156 , T163, K215			
T163A	101.6 7										
G165A	2	normal	slightly low	slightly low	YES			H-bond to L162, G169			
K170A	2.5	normal	slightly low	slightly low	YES	K side chain missin g					
S173A	4.1	slightly low	low	extremely low	YES		H-bond to backbone	H-bond to T176 , I177	putative phosphorylation site	PhosphoSite Plus	

								phosphate			
Q174A	0	normal	extremely low	normal	YES			H-bond to base	H-bond to S178		
T175A	1.03	normal	slightly low	sightly low	YES			H-bond to base	H-bond to R179		
T176A	2.05	slightly low	low	low	YES			H-bond to backbone phosphate	H-bond to S173, R179, F180		
								H-bond to backbone phosphate(C178 was not analyzed in this study)			
R179A	0.83	slightly low	extremely low	extremely low	YES			H-bond to base	H-bond to T175, T176, L183, Q184		
E181A	0.67	low	extremely low	extremely low	YES				H-bond to R150, Q157, I177, S178		
S186A	1.54	extremely low	extremely low	extremely low	YES			H-bond to backbone phosphate	H-bond to N189, M190		
K188A	30.67	normal	normal	slightly low	YES	K side chain missing	side chain was repaired		H-bond to K192		
								H-bond to backbone phosphate(N189 was not analyzed in this study)			
K192A	13.33	normal	normal	slightly low	YES	K side chain missing	side chain was repaired	H-bond to backbone phosphate	H-bond to K188, N189		
R194A	71.67										

K199A	81.67									putative ubiquitination site	PhosphoSite Plus
K215A	39.33	normal	normal	low	YES	K side chain missing			H-bond to D159, Q211	putative ubiquitination site	PhosphoSite Plus
S216A	112.03										
T218A	147.06										
R223A	96.67										
K224A	39.33	normal	normal	extremely low	YES	K side chain missing					
R225A	8	normal	normal	extremely low	YES	R side chain missing					
K226A	4	normal	normal	low	YES	K side chain missing	side chain was repaired				
R227A	0.67	normal	normal	extremely low	YES			H-bond to base			
T228A	18.97	normal	normal	sightly low	YES			H-bond to backbone phosphate		reported phosphorylation site in human ES and EC cells (human Oct4-T235) and reported glycosylation site	Proc. Natl. Acad. Sci. U. S. A. 109 :7162-7168, Mol. Cell 48 :1-14, Cell Stem Cell 11 :62-74.
S229A	88.61									reported	Proc. Natl.

									phosphorylation site in human ES cells (human Oct4-S236)	Acad. Sci. U. S. A. 109 :7162- 7168.
R233A	107.3 3									
R235A	11.33	normal	normal	low	YES			H-bond to E239, I230		
S237A	102.5 3									
T240A	94.12									
K244A	89.33									
K247A	28.67	normal	normal	low	YES			H-bond to backbone phosphate		
S249A	88.26									
T254A	82.35									
K264A	93.33									
R268A	24	normal	normal	low	YES			H-bond to backbone phosphate	H-bond to K264, D265, S272	
V269A	2	normal	normal	sightly low	YES			non polar contacts with base	H-bond to D265, S272 , N273	
C272A	104	normal	normal	normal	YES					
N273A	0	normal	normal	extremel y low	YES			H-bond to base	H-bond to V269 , Q276, K277	
R274A	0	normal	normal	low	YES				H-bond to E239, W270, G278	
R275A	0	normal	normal	low	YES			H-bond to backbone phosphate	H-bond to P246, F271, S272 , K279	
								H-bond to base (Q276 was not		

								analyzed in this study)			
K277A	1.33	normal	normal	low	YES	K side chain missing	side chain was repaired	H-bond to backbone phosphate	H-bond to N273		
K279A	8	normal	normal	low	YES	K side chain missing	side chain was repaired	H-bond to backbone phosphate	H-bond to R275		
R280A	83.54										
S281-282A	96.2									reported phosphorylation site in human ES cells (human Oct4-S288)	Proc. Natl. Acad. Sci. U. S. A. 109:7162-7168.
Y285A	96.2										
S286A	89.87										
R288A	109.33										
Y291A	91.18										
T294-296A	91.18										
S304A	88.89										
T315A	87.97										
Y318A	96.84										
S320A	98.77										
T324-325A	85.29										
Y327A	41.14	normal	normal	sightly low	YES					putative phosphorylation site	Plos One 4:e4467.
S328A	99.37										
S339A	85.19										

T343A	101.9												
S347-351A	92.5											reported phosphorylation site in human ES cells (human Oct4-S355 and human Oct4-S359)	<i>Proc. Natl. Acad. Sci. U. S. A.</i> 109 :7162-7168.

Supplementary Table 2: Vectors, Primers used for library construction

Entry clone	mutagenesis primer pairs		LR reaction with pMXs-CHR-ccdB (destination vector)
	Sense primer	Antisense primer	
pDONR223-mOct4-WT			pMXs-mOct4-WT
pDONR223-mOct4-S7A	GGACACCTGGCTGCAGACTCGCCTCTCAC	GTGAGAACGCCGAAGTCTGCAGCCAGGTGTCC	pMXs-mOct4-S7A
pDONR223-mOct4-S12A	GACTTCGCCTTCGCACCCCCCACCAGGTG	CACCTGGTGGGGGTGCGAACGGCGAACAGTC	pMXs-mOct4-S12A
pDONR223-mOct4-S21A	GGGGGTGATGGGGCAGCAGGGCTGGAG	CTCCAGCCCTGCTGCCCATCACCCCC	pMXs-mOct4-S21A
pDONR223-mOct4-S36A	CGAACCTGGCTAGCCTCCAAGGGCCTCC	GGAGGCCCTTGGAAAGGCTAGCCAGGTTCG	pMXs-mOct4-S36A
pDONR223-mOct4-S50A	GGAATCGGACCAGGCCAGAGGTATTGGGGATC	GATCCCCAATACCTCTGCGCCTGGTCCGATTCC	pMXs-mOct4-S50A
pDONR223-mOct4-S56A	GTATTGGGGATCGCCCCATGTCCGCC	GGGCGGACATGGGGCGATCCCCAATAC	pMXs-mOct4-S56A
pDONR223-mOct4-S100A	GGAGCACGAGTGGAAAGCGAACATTAGAGGAAACC	GGTCCCTCTGAATTGCTTCACTCGTGCTCC	pMXs-mOct4-S100A
pDONR223-mOct4-S102A	CGAGTGGAAAGCAACGCAGAGGGAACCTCCTC	GAGGAGGTTCCCTCTGCGTTGCTTCACTCG	pMXs-mOct4-S102A
pDONR223-mOct4-S106-107A	CTCAGAGGGAACCGCCGCTGAGCCCTGTGCCGAC	GTCGGCACAGGGCTCAGCGCGGTTCCCTGAG	pMXs-mOct4-S106-107A
pDONR223-mOct4-S129A	CCAACTCCCGAGGAGGCCAGGACATGAAAGCC	GGCTTCATGTCCTGGGCCTCCTCGGGAGTTGG	pMXs-mOct4-S129A
pDONR223-mOct4-S173A	GGAAAGGTGTTGCCAGACCACCATCTGTCG	CGACAGATGGTGGTCTGGCGAACACCTTCC	pMXs-mOct4-S173A
pDONR223-mOct4-S186A	GAGGCCTTGCAGCTGCCCTTAAGAACATGTG	CACATGTTCTAACGGCGAGCTGCAAGGCCTC	pMXs-mOct4-S186A
pDONR223-mOct4-S216A	CAGGAGATATGCAAAGCGGAGACCCTGGTCAGG	CCTGCACCAGGGCTCCGCTTGCATATCTCCTG	pMXs-mOct4-S216A
pDONR223-mOct4-S229A	GAGAAAGCGAACTGCCATTGAGAACCGTGTGAGG	CCTCACACGGTTCTCAATGGCAGTTCGCTTCTC	pMXs-mOct4-S229A
pDONR223-mOct4-S237A	CCGTGTGAGGTGGCTCTGGAGACCATGTTTC	GAAACATGGTCTCCAGAGGCCACCTCACACGG	pMXs-mOct4-S237A
pDONR223-mOct4-S249A	GTGCCCGAACCCGCCCTACAGCAGATC	GATCTGCTGTAGGGCGGGCTTCGGGCAC	pMXs-mOct4-S249A
pDONR223-mOct4-S281-282A	CCAGAAGGGAAAAGAGCAGCTATTGAGTATTCCCAAC	GTTGGGAATACTCAATAGCTGCTTTGCCCTCTGG	pMXs-mOct4-S281-282A
pDONR223-mOct4-S286A	GATCAAGTATTGAGTATGCCAACGAGAAGAG	CTCTCTCGTTGGGCATACTCAATACTTGATC	pMXs-mOct4-S286A

pDONR223-mOct4-S304A	GGGGGGGCTGTAGCCTTCCTGC	GGGGGCAGAGGAAGGCTACAGCCCCCCC	pMXs-mOct4-S304A
pDONR223-mOct4-S320A	CCAGGCTATGGAGCCCCCACTCACAC	GTGTGGTGAAGTGGGGGCTCATAGCCTGG	pMXs-mOct4-S320A
pDONR223-mOct4-S328A	CACCACACTCTACGCAGTCCCTTCTGAGGGC	GCCCTCAGGAAAAGGGACTGCGTAGAGTGTGGTG	pMXs-mOct4-S328A
pDONR223-mOct4-S339A	GAGGCCTTCCCGCTGTTCCCGTCAGTGC	GAGCAGTGACGGAACAGCGGGAAAGGCC	pMXs-mOct4-S339A
pDONR223-mOct4-S347-351A	CACTGCTCTGGCGCTCCATGCATGCAAACGA	CTTTGTAGTCGTTGCATGCATGGAGCGCCCAG	pMXs-mOct4-S347-351A
pDONR223-mOct4-T33A	CTGGGTGGATTCTCGAGCCTGGCTAAGCTTCC	GGAAGCTTAGCCAGGCTCGAGAACCTCACCCAG	pMXs-mOct4-T33A
pDONR223-mOct4-T87A	GTTGGCGTGGAGGCTTGCAGCCTGAGGGC	GCCCTCAGGCTGCAAAGCCTCACGCCAAC	pMXs-mOct4-T87A
pDONR223-mOct4-T105A	GCAACTCAGAGGGAGCCTCCTGAGCCCTG	CAGGGCTCAGAGGAGGCTCCCTCTGAGTTGC	pMXs-mOct4-T105A
pDONR223-mOct4-T125A	GAGAAGGTGGAACCAGCTCCGAGGAGTCCC	GGGACTCCTCGGGAGCTGGTCCACCTTCTC	pMXs-mOct4-T125A
pDONR223-mOct4-T152A	GCAGAAGAGGATCGCCTGGGTACACCCAG	CTGGGTGTACCCAAGGCGATCCTCTTCTGC	pMXs-mOct4-T152A
pDONR223-mOct4-T156A	CACCTTGGGTACGCCAGGCCAGGTG	CACGTGGCCTGGCGTACCCAAGGTG	pMXs-mOct4-T156A
pDONR223-mOct4-T163A	GACGTGGGCTGCCCTGGCGTTCT	GAGAACGCCAGGGCGAGCCCCACGTC	pMXs-mOct4-T163A
pDONR223-mOct4-T175A	GGTGTTCAGCCAGGCCACCATCTGTCGCTTCGAGG	CCTCGAACGCGACAGATGGTGGCTGGCTAACACAC	pMXs-mOct4-T175A
pDONR223-mOct4-T176A	GGTGTTCAGCCAGACGCCATCTGTCGCTTCGAGG	CCTCGAACGCGACAGATGGCGGTCTGGCTAACACAC	pMXs-mOct4-T176A
pDONR223-mOct4-T218A	GATATGCAAATCGGAGGCCCTGGTGCAGGCC	GGCCTGCACCAGGGCCTCCGATTGCATATC	pMXs-mOct4-T218A
pDONR223-mOct4-T228A	CGGAAGAGAAAGCGAGCTAGCATTGAGAACCGTG	CACGGTTCTCAATGCTAGCTCGCTTCTCTCCG	pMXs-mOct4-T228A
pDONR223-mOct4-T240A	GGTGGAGTCTGGAGGCCATGTTCTGAAGTGC	GGCACTTCAGAAACATGGCCTCCAGACTCCACC	pMXs-mOct4-T240A
pDONR223-mOct4-T254A	CCCTACAGCAGATCGCTCACATGCCAACATCAG	CTGATTGGCGATGTGAGCGATCTGCTGTAGGG	pMXs-mOct4-T254A
pDONR223-mOct4-T294-296A	GAAGAGTATGAGGCTGCAGGGCACCTTCCCGGGGG	CCCCCTGGAAAGGTGCCCTGCAGCCTCATAC	pMXs-mOct4-T294-296A
pDONR223-mOct4-T315A	CCCCACTTGGCGCCCCAGGCTATGGAAG	CTTCCATAGCCTGGGGGCCAAAGTGGGG	pMXs-mOct4-T315A
pDONR223-mOct4-T324-325A	GGAAGCCCCCACTCGCCGCACTCTACTCAGTC	GGAAAAGGGACTGAGTAGAGTGCAGCGAAGTGG	pMXs-mOct4-T324-325A
pDONR223-mOct4-T343A	CTCTGTTCCCGTCGCTGCTCTGGCTCTC	GAGAGCCCAGAGCAGCGACGGGAACAGAG	pMXs-mOct4-T343A
pDONR223-mOct4-Y62A	CCATGTCCGCCCGCAGCCGAGTTCTGCGGAG	CTCCGCAGAACACTGGCTGCAGGGCGACATGG	pMXs-mOct4-Y62A
pDONR223-mOct4-Y70A	GGAGGGATGGCAGCCTGTGGACCTCAGG	CCTGAGGTCCACAGGCTGCCATCCCTCC	pMXs-mOct4-Y70A
pDONR223-mOct4-Y155A	GGATCACCTGGGGGCCACCCAGGCCAGGTG	CACGTGGCCTGGTGGCCCCAAGGTGATCC	pMXs-mOct4-Y155A
pDONR223-mOct4-Y285A	GCAAAAGATCAAGTATTGAGGCTCCAAACGAGA	CATACTCTCTCGTTGGGAAGCCTCAATACTTGAT	pMXs-mOct4-Y285A
	AGAGTATG	CTTTTGC	

pDONR223-mOct4-Y291A	CCCAACGAGAAAGAGGCTGAGGCTACAGGGACAC	GTGTCCCTGTAGCCTCAGCCTTCTCGTTGGG	pMXs-mOct4-Y291A
pDONR223-mOct4-Y318A	GGCACCCCAGGCGCTGGAAGCCCCCAC	GTGGGGCTTCAGCGCTGGGTGCC	pMXs-mOct4-Y318A
pDONR223-mOct4-Y327A	CCACTTCACCACACTCGCCTCAGTCCCTTCC	GGAAAAGGGACTGAGGCGAGTGTGGTGAAGTGG	pMXs-mOct4-Y327A
pDONR223-mOct4-K118A	CCCAATGCCGTGGCGTTGGAGAAGGTGGAACCA ACTCC	GGAGTTGGTCCACCTCTCCAACGCCACGGCATT GGG	pMXs-mOct4-K118A
pDONR223-mOct4-K121A	GCCGTGAAGTTGGAGGCAGGTGGAACCAACTCCC	GGGAGTTGGTCCACCGCCTCCAACTTCACGGC	pMXs-mOct4-K121A
pDONR223-mOct4-K133A	GTCCCAGGACATGGCAGCCCTGCAGAAGGAG	CTCCTTCTGCAGGGCTGCCATGTCTGGGAC	pMXs-mOct4-K133A
pDONR223-mOct4-K137A	CATGAAAGCCCTGCAGGCGGAGCTAGAACAG	CTGTTCTAGCTCCGCCTGCAGGGCTTCATG	pMXs-mOct4-K137A
pDONR223-mOct4-K144A	CTAGAACAGTTGCCCGCCTGCTGAAGCAGAAG	CTTCTGCTTCAGCAGCGCGGAAACTGTTCTAG	pMXs-mOct4-K144A
pDONR223-mOct4-K147A	GCCAAGCTGCTGGCGCAGAACAGGAGATCACC	GGTGATCCTCTTCTGCGCCAGCAGCTTGGC	pMXs-mOct4-K147A
pDONR223-mOct4-K149A	GCTGCTGAAGCAGGCGAGGATCACCTTGGGG	CCCCAAGGTGATCCTCGCCTGCTTCAGCAGC	pMXs-mOct4-K149A
pDONR223-mOct4-K170A	GGCGTTCTCTTGGAGCGGTGTTCAGCCAG	CTGGCTGAACACCGCTCCAAGAGAACGCC	pMXs-mOct4-K170A
pDONR223-mOct4-K188A	GCAGCTCAGCCTTGCAGAACATGTGAAGCTGC	GCAGCTTACACATGTTGCAAGGCTGAGCTGC	pMXs-mOct4-K188A
pDONR223-mOct4-K192A	CTTAAGAACATGTGTGCGCTGCCCTG	CAGGGGCCGCAGCGCACACATGTTCTTAAG	pMXs-mOct4-K192A
pDONR223-mOct4-K199A	CCCCTGCTGGAGGCAGGGTGGAGGAAG	CTTCCTCCACCCACGCCCTCAGCAGGGG	pMXs-mOct4-K199A
pDONR223-mOct4-K215A	CAGGAGATATGCGCATCGGAGACCCTGGTG	CACCAGGGCTCCGATGCGCATATCTCCTG	pMXs-mOct4-K215A
pDONR223-mOct4-K224A	GTGCAGGCCCGGGCGAGAACCGAAGTAGC	GCTAGTTCGCTTCTGCCCGGGCCTGCAC	pMXs-mOct4-K224A
pDONR223-mOct4-K226A	GCCCGGAAGAGAGCGCGAACTAGCATTGAG	CTCAATGCTAGTTGCGCTCTTCCGGC	pMXs-mOct4-K226A
pDONR223-mOct4-K244A	GAGACCATGTTCTGGCGTGCCGAAGCCCTCC	GGAGGGCTCGGGCACGCCAGAACATGGTCTC	pMXS-mOct4-K244A
pDONR223-mOct4-K247A	CTGAAGTGCCCGGCCCTCCCTACAG	CTGTAGGGAGGGCGCCGGCACTTCAG	pMXs-mOct4-K247A
pDONR223-mOct4-K264A	CAGCTGGGCTAGAGGCAGGATGTGGTTGAG	CTCGAACACATCCGCCTCTAGCCAAGCTG	pMXs-mOct4-K264A
pDONR223-mOct4-K277A	GTAACCGGCCAGGCGGGAAAAGATCAAG	CTTGATCTTGGCCCTGGCGCCGGTTAC	pMXS-mOct4-K277A
pDONR223-mOct4-K279A	CGCCAGAAGGGCGCAAGATCAAGTATTG	CAATACTGATCTGCGCCCTCTGGCG	pMXS-mOct4-K279A
pDONR223-mOct4-R32A	GGCTGGGTGATTCTGCAACCTGGCTAAGC	GCTTAGCCAGGTTGCAGAACATCACCAGCC	pMXs-mOct4-R32A
pDONR223-mOct4-R97A	CAGGCAGGAGCAGCAGTGGAAAGCAACTCAG	CTGAGTTGCTTCCACTGCTGCTCCTGCCTG	pMXS-mOct4-R97A
pDONR223-mOct4-R113A	CCCTGTGCCAGGCCCTTCAATGCCGTG	CACGGCATTGGGGCGTCGGCACAGGG	pMXs-mOct4-R113A
pDONR223-mOct4-R150A	CTGCTGAAGCAGAACGGCGATCACCTTGGGG	CCCCAAGGTGATCGCCTTCTGCTTCAGCAG	pMXS-mOct4-R150A
pDONR223-mOct4-R179A	CAGACCAACATCTGTGCCCTCGAGGCCTTGAG	CTGCAAGGCCTCGAAGGCACAGATGGTGGTCTG	pMXS-mOct4-R179A
pDONR223-mOct4-R194A	CATGTGAAGCTGGCGCCCTGCTGGAGAAG	CTTCTCAGCAGGGCGCCAGCTTACACATG	pMXs-mOct4-R194A
pDONR223-mOct4-R223A	CTGGTGCAGGCCCGAAGAGAAAGCGAAC	GTTCGCTTCTCTCGCGGCCTGCACCAAG	pMXS-mOct4-R223A
pDONR223-mOct4-R225A	CAGGCCCGGAAGGCAGAACCGAACATGATTGAG	CTCAATGCTAGTTGCGCTTGCCTCCGGCCTG	pMXs-mOct4-R225A
pDONR223-mOct4-R227A	GCCCGGAAGAGAAAGGCAACTAGCATTGAGAAC	CGGTTCTCAATGCTAGTTGCCTTCTCTCCGGGC	pMXs-mOct4-R227A

	CG		
pDONR223-mOct4-R233A	CTAGCATTGAGAACGCTGTGAGGTGGAGTCTG	CAGACTCCACCTCACAGCGTTCAATGCTAG	pMXs-mOct4-R233A
pDONR223-mOct4-R235A	GAGAACCGTGTGGCGTGGAGTCTGGAG	CTCCAGACTCCACGCCACACGGTTCTC	pMXs-mOct4-R235A
pDONR223-mOct4-R268A	GGGCTAGAGAAAGGATGTGGTTGCAGTATGGTTC TGTAAC	GTTACAGAACCATACTGCAACCACATCCTCTA GCC	pMXs-mOct4-R268A
pDONR223-mOct4-R274A	GTATGGTTCTGTAACCGCGCCAGAACGGC	GCCCTTCTGGCGCGTACAGAACCATAC	pMXs-mOct4-R274A
pDONR223-mOct4-R275A	GGTTCTGTAACCAGGGCCCAGAACGGCAAAGAT C	GATCTTTGCCCTCTGGGCCCGTTACAGAACACC	pMXs-mOct4-R275A
pDONR223-mOct4-R280A	CGCCAGAACGGCAAAGCATCAAGTATTGAGTATT CCC	GGGAATACTCAATACTTGATGCTTGCCCTCTGG CG	pMXs-mOct4-R280A
pDONR223-mOct4-R288A	CAAGTATTGAGTATTCCAAGCAGAACAGAGTATGA GGCTACAGGG	CCCTGTAGCCTCATACTCTCTGCTTGGAAACT CAATACTTG	pMXs-mOct4-R288A
pDONR223-mOct4-C272A	GTTCGAGTATGGTTCGCTAACCGGCCAG	CTGGCGCCGGTTAGCGAACCATACTCGAAC	pMXs-mOct4-C272A
pDONR223-mOct4-D159A	GGGTACACCCAGGCCCGTGGGCTCACCT G	CAGGGTGAGCCCCACGGCGGCCTGGGTGTACCC	pMXs-mOct4-D159A
pDONR223-mOct4-E181A	CCATCTGTCGCTTCGCGGCCCTGCAGCTCAGC	GCTGAGCTGCAAGGCCCGAACGACAGATGG	pMXs-mOct4-E181A
pDONR223-mOct4-G165A	GTGGGGCTCACCTGGCCGTTCTCTTGAAAG G	CCTTCCAAAGAGAACGCCAGGGTGAGCCCCAC	pMXs-mOct4-G165A
pDONR223-mOct4-N273A	CGAGTATGGTTCTGTGCCCGGCCAGAACGG	CCTTCTGGCGCCGGGACAGAACCATACTCG	pMXs-mOct4-N273A
pDONR223-mOct4-Q157A	CCTTGGGGTACACCGCGGCCGACGTGGGC	GCCCCACGTCGCCCGCGTGTACCCCAAGG	pMXs-mOct4-Q157A
pDONR223-mOct4-Q174A	GGAAAGGTGTTCAGCGCAGCACCATCTGCGC	GCGACAGATGGTGGTCGCGCTGAACACCTTCC	pMXs-mOct4-Q174A
pDONR223-mOct4-V269A	GGATGTGGTTCGAGCATGGTTCTGTAACCGG	CCGGTTACAGAACCATGCTCGAACACATCC	pMXs-mOct4-V269A
pDONR223-mOct4-S186D	GAGGCCTTGCAGCTGACCTTAAGAACATGTG	CACATGTTCTTAAGGTCGAGCTGCAAGGCCTC	pMXs-mOct4-S186D
pDONR223-mOct4-S186E	GAGGCCTTGCAGCTCGAACCTTAAGAACATGTG	CACATGTTCTTAAGGTCGAGCTGCAAGGCCTC	pMXs-mOct4-S186E
Other constructs	Primer pairs		Source
	Sense primer	Antisense primer	
pDONR223-mOct4	CGGTGTACAAAAAAGCAGGCTGGATGGCTGGAC ACCTGGCTTCAGACTTCG	CGGTGTACAAGAACGCTGGGTTCACTTGTCACTG TCGTCTTGTAGTCGTTGAATGCATGGAGAGCC CAGAGC	
pMXs-CHR-ccdB	CGGTTAATTAA ATAAGCAGAGCTCTGGCTAACTG	CGGCTCGAGCATTACTACGTAGAACATCGAGACCGA	

pUAST-Sapl-1xCR4	CGGGGTACC GCTCTCGAACTGGGTGTGGGGAGGTTGAGC	CGGTCTAGA GCTCTCGGTTAGCAGATTAAGGAAGGGC	
pUAST-Nhel-1xCR4	CGGGGTACC GCTCTCGAACTGGGTGTGGGGAGGTTGAGC	CGGTCTAGAGCTAGCGGTTAGCAGATTAAGGAA GGGC	
pUAST-6xCR4			subcloned from pUAST-Sapl-6XCR4
pGL4.2-6xCR4			subcloned from pUAST-6XCR4
pGL4.2-5xW	CCCTGAGCAAAACACCACCTGGTAATTGCAT TTCTAAAATAAGTCGA	GGGTGACTTATTAGAAATGCAAATTACCCAGG TGGTGTGGCTCAG	
pGL4.2-5xW-37TK	CGGCTGAGGGTCCACTTCGCATATTAAAGGTGA CG	CGGAGATCTCGGCACGCTGTTGACGCTGTTAACG GGGTGCGCTGC	
pMXs-mOct4-△NTD-Flag	CGGGGATCCATGGACATGAAAGCCCTGCAGAAG G	CGGGCGGCCGCTCACTTGTGTCATCGTCTTG AGTCGTTGAATGCATGGG	
pMXs-mOct4-△CTD-Flag	CGGGGATCCATGGCTGGACACCTGGCTTCAGAC	CGGGCGGCCGCTCACTTGTGTCATCGTCTTG AGTCACTTGATCTTGCC	
pMXs-mOct4-△POU(NTD+CTD)-Flag			
pMXs-mOct4-NTD	CGGGGATCCATGGCTGGACACCTGGCTTCAGAC	CGGGAAATTCCCTGGGACTCCTCGGGAGTTGGTTCC	
pMXs-mOct4-CTD-Flag	CGGGAATTCAATTGAGTATTCCCAACGAGAAGAG	CGGGCGGCCGCTCACTTGTGTCATCGTCTTG AGTCGTTGAATGCATGGG	
pMXs-mOct4-POU-Flag	CGGGGATCCATGGACATGAAAGCCCTGCAGAAG G	CGGGCGGCCGCTCACTTGTGTCATCGTCTTG AGTCACTTGATCTTGCC	
pMXs-mOct4-△NTD-YAP ^{TAD} -Flag	CGGGGATCCATGGACATGAAAGCCCTGCAGAAG G	CGGGCGGCCGCTTGAATGCATGGGAGAGCCC AGAG	
pMXs-mOct4-△CTD-YAP ^{TAD} -Flag	CGGGGATCCATGGCTGGACACCTGGCTTCAGAC	CGGGCGGCCGCACTTGATCTTGCCCTCTGGC GC	
pMXs-mOct4-△POU-YAP ^{TAD} -Flag	CGGGGATCCATGGCTGGACACCTGGCTTCAGAC	CGGGCGGCCGCTTGAATGCATGGGAGAGCCC AGAG	
pMXs-mOct4-POU-YAP ^{TAD} -Flag	CGGGGATCCATGGACATGAAAGCCCTGCAGAAG G	CGGGCGGCCGCACTTGATCTTGCCCTCTGGC GC	
pMXs-YAP ^{TAD} -Flag	CGGTTAATTAAATGCAGGGAGGCGCTGGGTG GAG	CGGGGATCCCTACTTGTGTCATCGTCATCCTTGTAA CTAACCC	

pMXs-EGFP-FLAG	CGGTTAATTAAATGGT GAGCAAGGGCGAGGAGC TGTTC	CGGCTCGAGTCAC TTGTCATCGT CCGTCTTTGAGT CCTTGTA CAGCTCGTCCATGCCGAGAGTG	
pMXs-mSox2-Myc	CGGAATT CATGTATAACATGATGGAGACGGAG CTGAAGC	CGGGCGGCCGCTCACAGATCCTCTTGAGATGA GTTTTGTTCCATGTGCGACAGGGG	
pMXs-mOct4-WT-YAP ^{TAD} -Flag			<i>Stem Cell Reports</i> 2:1-9.
pMXs-mKlf4			purchased from addgene
pMXs-mNanog			purchased from addgene
pMXs-m-cMyc			purchased from addgene
pMXs-hOct4-WT-Flag	CGGGGATCCATGGCGGGACACCTGGCTTCGGAT TTCG	CGGGCGGCCGCTCACTTGT CGTCATCGTCTTG AGTCGTTGAATGCATGGGAGAG	
pMXs-hOct4-K177A-Flag	GGTTCTATTGGGGCGGTATTCAGCCAAAC	GTTTGGCTGAATACCGCCCCAAATAGAACCC	
pMXs-hOct4-K222A-Flag	CAGGAGATATGCGCAGCAGAAACCCTC	GAGGGTTCTGCTGCGCATATCTCCTG	