

ARTD1 regulates osteoclastogenesis and bone homeostasis by dampening NF- κ B-dependent transcription of *IL-1 β*

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Supplementary Figures and Table

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Table S1. List of real-time PCR and ChIP primer sequences.

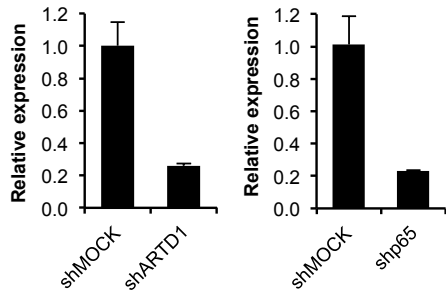
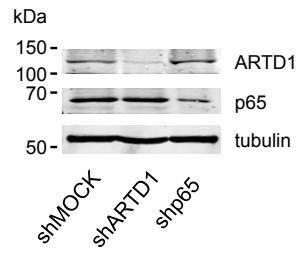
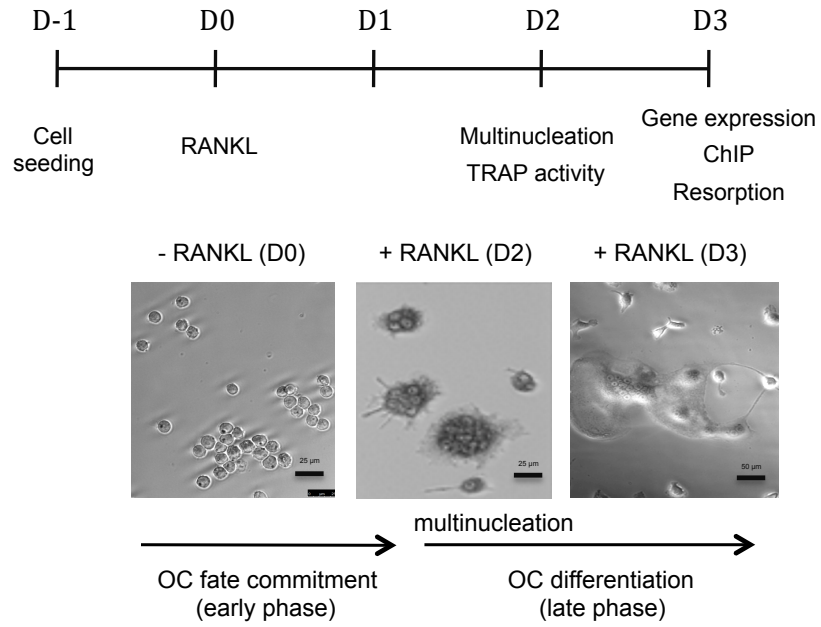
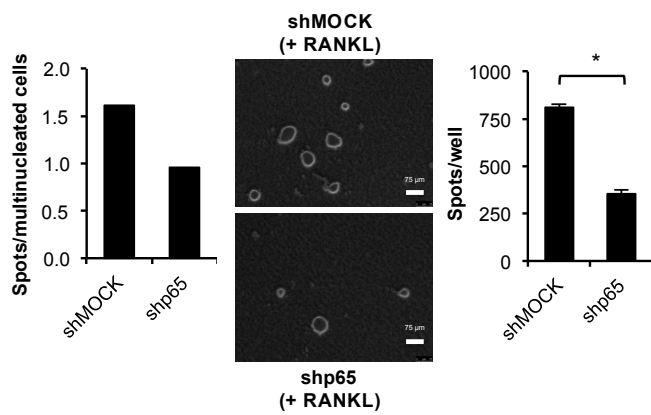
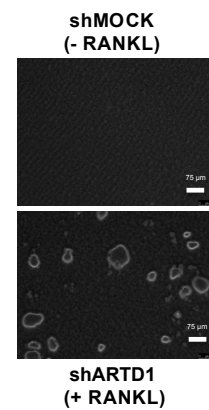
A**B****C****D****E**

Figure S1. Efficiencies of stable knock-downs, experimental design of in vitro differentiation model, and functional osteoclast activity in knock-down cells. (A) Efficiency of *Artd1* and *p65* silencing after RAW 264.7 macrophages transduction with retroviruses carrying shRNA was estimated using real-time PCR. Expression of *Gapdh* was used for normalization. (B) Efficiency of *Artd1* and *p65* silencing with shRNA was determined at the level of protein by Western blotting. Tubulin was used as an internal control. (C) Experimental design of the in vitro differentiation model (upper panel). Graphic representation of osteoclast formation from RAW 264.7 macrophages; from the left: macrophages without RANKL treatment (-RANKL, day 0 (D0)), 2-day-old (D2) and 3-day-old (D3) osteoclasts (lower panel). (D) The resorbing activity of 3-day-old osteoclasts derived from shMOCK and shp65 macrophages were compared based on their capacity to induce matrix dissolution (resorption of bone-mimicking mineral surface) (left panel). The number of matrix dissolution spots was normalized to the number of multinucleated cells (right panel). (E) The functional activity of osteoclasts derived from shARTD1 RAW 264.7 macrophages was verified based on their capacity to induce matrix dissolution.

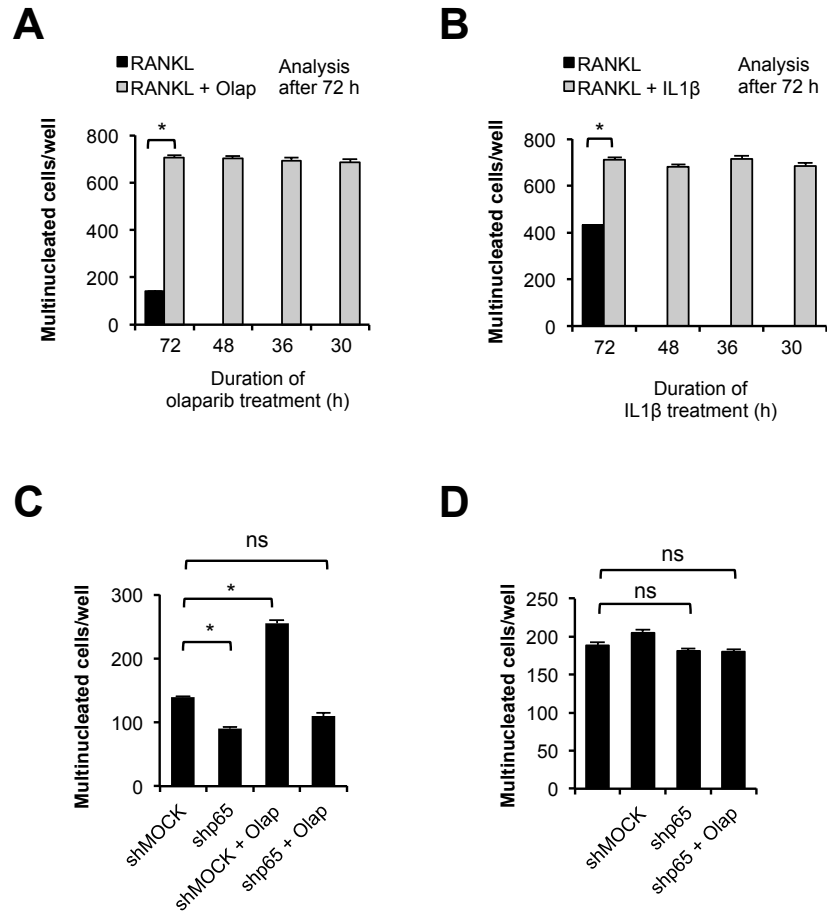


Figure S2. Timing of olaparib or IL-1 β effect on multinucleation, and contribution of p65 on olaparib effect on multinucleation. (A) Olaparib was administrated at selected time points after induction of osteoclastogenesis, and multinucleated cells were counted after 72 h. (B) IL-1 β (2 ng/ml) was added at the indicated time points to shMOCK RAW 264.7 macrophages cultured in the presence of RANKL, and multinucleated cells were quantified 72 h after RANKL administration. (C) The contribution of p65 in the ADP-ribosylation-mediated increase in osteoclast formation was studied in shMOCK and shp65 RAW 264.7 macrophages induced to differentiate in the presence or absence of olaparib. (D) The multinucleation of osteoclasts differentiated in the presence of medium conditioned by shMOCK and shp65 (\pm olaparib) was compared 48 h after RANKL administration.

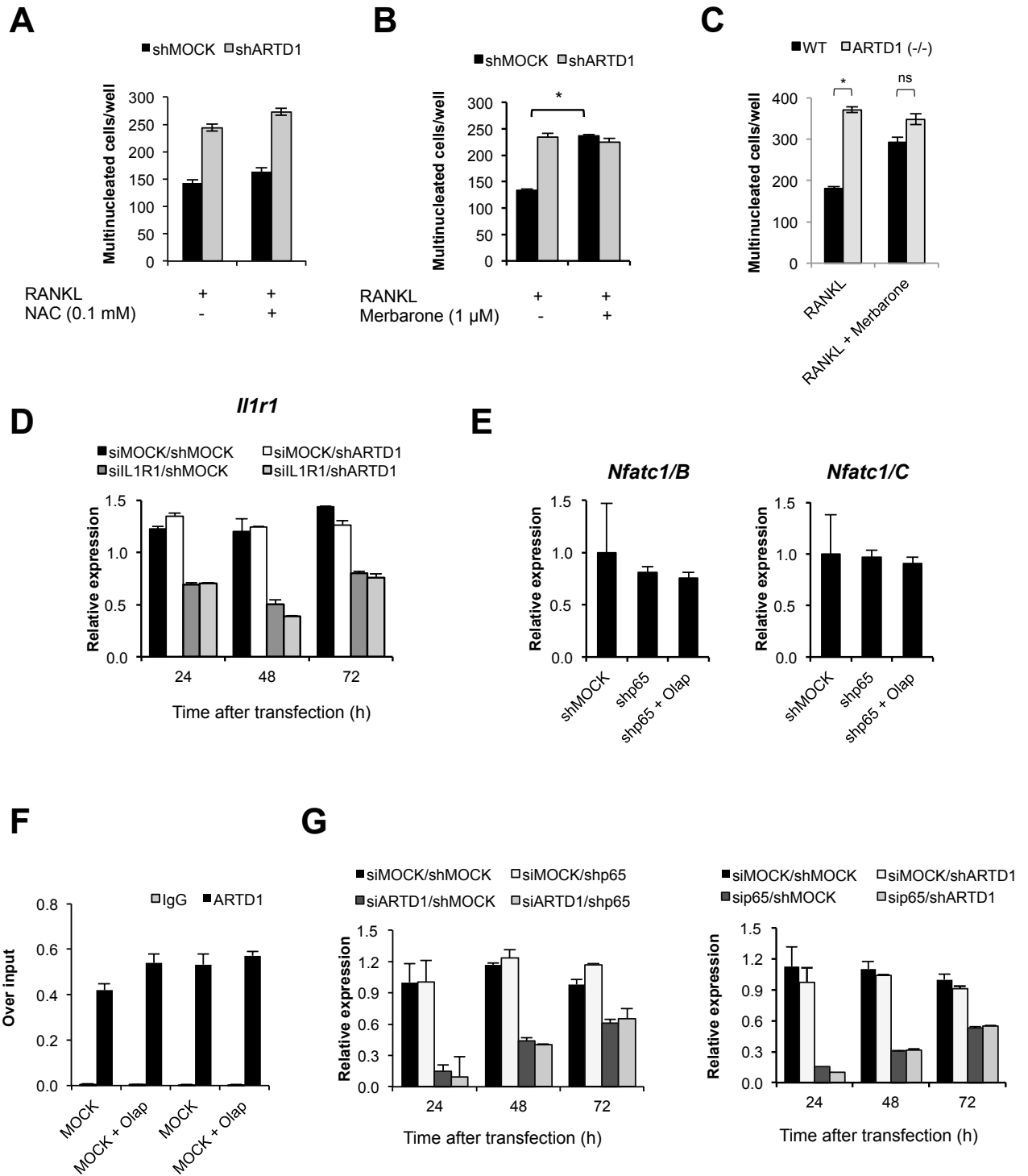


Figure S3. Effect of NAC or merbarone on enhanced multinucleation by ARTD depletion/deletion. (A) The ROS scavenger N-acetylcysteine or (B) the topoisomerase II inhibitor merbarone were added together with RANKL to shMOCK and shARTD1 RAW 264.7 macrophages and multinucleation was quantified 48 h later. (C) The topoisomerase II

inhibitor merbarone was added together with RANKL to the culture of WT and *Artd1* (-/-) osteoclast precursors and multinucleation was quantified 48 h later. **(D)** The recovery of IL-1 receptor 1 (*IL-1r1*) mRNA after silencing with siRNA was determined using real-time PCR. **(E)** The expression of *NAFTc1/B* and *NAFTc1/C* was compared between 3-day-old osteoclasts derived from shMOCK and shp65 (\pm olaparib) RAW 264.7 macrophages using real-time PCR. **(F)** The association of ARTD1 with chromatin in 3-day-old shMOCK (\pm olaparib) osteoclasts was determined by ChIP. **(G)** The double *Artd1/p65* silencing with siRNA/shRNA over time was monitored with real-time PCR.

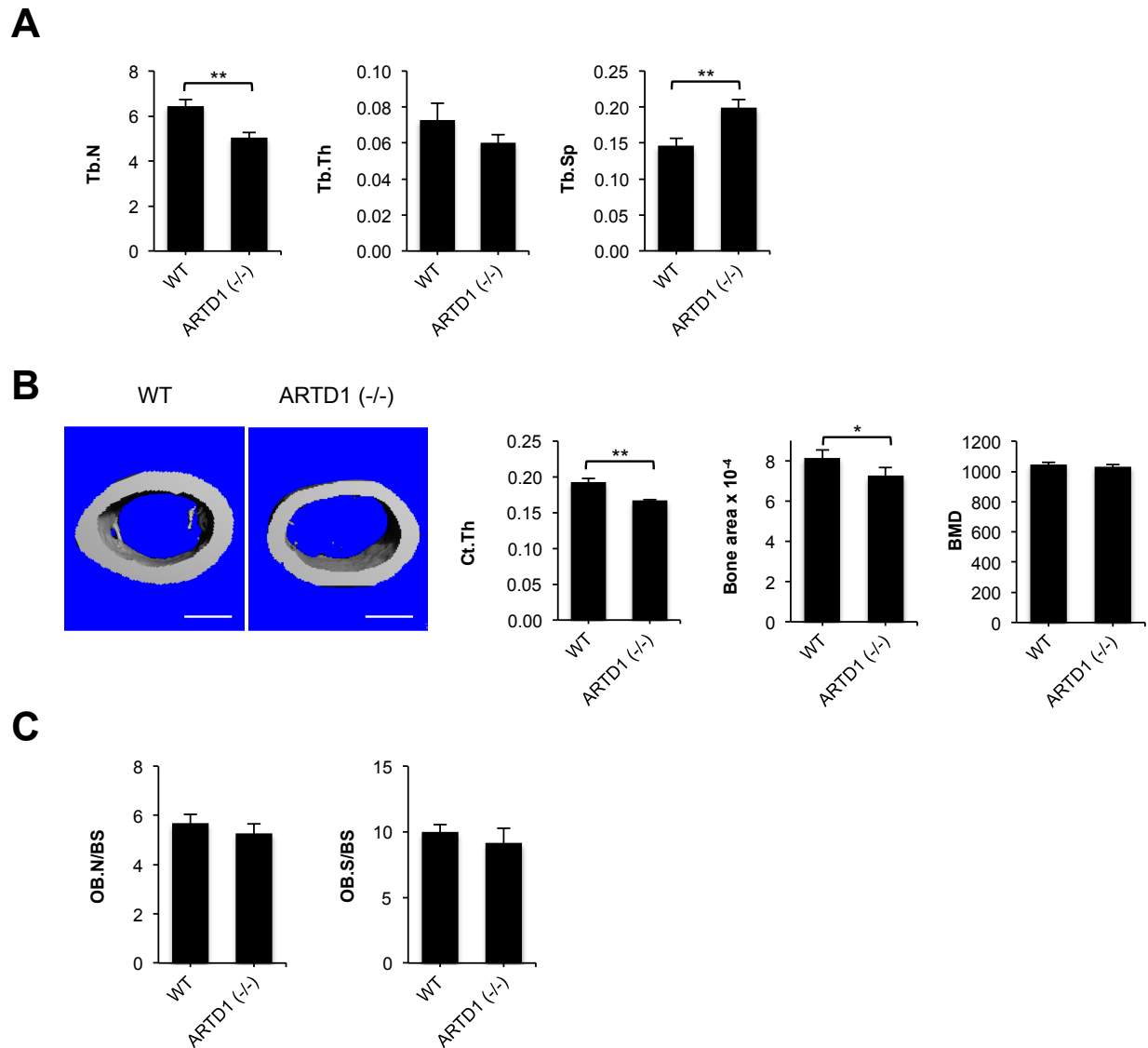


Figure S4. ARTD1 deficiency causes osteopenia without changes in osteoblast parameters. Femurs of 8-week-old *Artd1*-deficient mice (n=4) and their wild-type littermates (n=4) were analysed by μ CT. **(A)** Trabecular number (Tb.N), thickness (Tb.Th) or spacing (Tb.Sp). **(B)** Representative 3D reconstruction of mid-shaft cortical bone (left panels, size bars = 500 μ m); cortical thickness (Ct.Th), cortical bone area and BMD were measured. **(C)** Osteoblast number/bone surface (OB.N/BS) or surface/bone/surface (OB.S/BS) was quantified from H&E stainings of tibial sections. Values are mean \pm SD. * p<0.05, ** p<0.01.

Supplementary Table

Table S1. List of real-time PCR and ChIP primer sequences

Primers	Sequence
Used for shRNA constructs	
<i>Ard1</i> Fwd	GATCCCCGGTGATCGGTAGCAACAAATTC AAGAGATTTGTTGCTACCGATCACCTTTTG GAAA
<i>Ard1</i> Rev	AGCTTTTCCAAAAAGGTGATCGGTAGCAACAAATCTCTTGAATTTGTTGCTACCGATCAC CGGG
<i>p65/RelA</i> Fwd	GATCCCCGAGTTTCAGCAGCTCCTGAACTTCAAGAGAGTTCAGGAGCTGCTGAAACTCT TTTTGGAAA
<i>p65/RelA</i> Rev	AGCTTTTCCAAAAAGAGTTTCAGCAGCTCCTGAACTCTCTTGAAGTTCAGGAGCTGCTAA ACTCGGG
Used for real-time PCR	
<i>Nfat1/A</i> Fwd	GGTAACTCTGTCTTTCTAACCTTAAGCTC
<i>Nfat1/A</i> Rev	GTGATGACCCAGCATGCACCAGTCACAG
<i>Nfat1/B</i> Fwd	CCCATCCGCCAGGCTACAGCCGCAGTAA
<i>Nfat1/B</i> Rev	TTCGGTAAGTTGGGATTTCTGAGTGGTACC
<i>Nfat1/C</i> Fwd	CCCATCCGCCAGGCTACAGCCGCAGTAA
<i>Nfat1/C</i> Rev	TGAGTGGTACCAGATGTGGGTCCAGTTTAT
<i>Tracp</i> Fwd	AGATTTGTGGCTGTGGGCGA
<i>Tracp</i> Rev	CTGCACGGTTCTGGCGATCT
<i>Ctsk</i> Fwd	AAGTGGTTCAGAAGATGACGGGAC
<i>Ctsk</i> Rev	TCTTCAGAGTCAATGCCTCCGTTT
<i>Gapdh</i> Fwd	ACTCCACTCACGGCAAATTCA
<i>Gapdh</i> Rev	GGTCTCGCTCCTGGAAGATG
<i>p65/RelA</i> Fwd	GCGAGACCTGGAGCAAGCCATT
<i>p65/RelA</i> Rev	GTGTTGGGGGCCCGGTTATCAA
<i>Ard1</i> Fwd	GAAGGAAAGAGAAAAGGTGACG
<i>Ard1</i> Rev	GCAACTCTGTCCAAGATCGCTG
<i>IL-1b</i> Fwd	AAGGAGAACCAAGCAACGACAAAA
<i>IL-1b</i> Rev	TGGGGA ACTCTGCAGACTCAA ACT
<i>IL-1r1</i> Fwd	TGGAACAGAGCCAGTGT CAG
<i>IL-1r1</i> Rev	CAGGAGAAGTCGCAGGAAGT
<i>Rankl</i> Fwd	TGTACTTTCGAGCGCAGATG
<i>Rankl</i> Rev	AGGCTTGTTTCATCCTCCTG

<i>Dspp</i> Fwd	CCTCGGAGGCTTTGAAGACAT
<i>Dspp</i> Rev	CCTCTCGCATGTACCCCATC
<i>Opn</i> Fwd	TCCCTCGATGTCATCCCTGT
<i>Opn</i> Rev	ATCACATCCGACTGATCGGC
<i>Rank</i> Fwd	GCATCCCTTGCAGCTCAACA
<i>Rank</i> Rev	ATGGAAGAGCTGCAGACCAC
<i>Coll1a1</i> Fwd	CGATGGATTCCCGTTCGAGT
<i>Coll1a1</i> Rev	CGATCTCGTTGGATCCCTGG
Used for ChIP	
<i>IL-1b</i> promoter Fwd	CCCCTAAGAATTCCCATCAAGC
<i>IL-1b</i> promoter Rev	GAGCTGTGAAATTTCCCTTGG
<i>Il6</i> body Fwd	AAGTGGATTCCCCTGCCACT
<i>Il6</i> body Rev	ACAACAGGCAGCATCAGCTGACC