

SUPPLEMENTAL INFORMATION

Collaborative Enhancement of Endothelial Targeting of Nanocarriers by Modulating Platelet Endothelial Cell Adhesion Molecule-1/CD31 Epitope Engagement

Ann-Marie Chacko,^{1,2,§} Jingyan Han,^{3,§} Colin F. Greineder,³ Blaine J. Zern,³ John L. Mikitsh,¹ Madhura Nayak,¹ Divya Menon,³ Ian H. Johnston,³ Mortimer Poncz,⁶ David M. Eckmann,⁴ Peter F. Davies⁵ and Vladimir R. Muzykantov^{2,3*}

¹Department of Radiology, Division of Nuclear Medicine and Clinical Molecular Imaging, ²Center for Targeted Therapeutics and Translational Nanomedicine, Institute for Translational Medicine and Therapeutics, ³Department of Systems Pharmacology and Translational Therapeutics, ⁴Department of Anesthesiology & Critical Care, ⁵Department of Pathology and Institute for Medicine and Engineering, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, United States, and ⁶Department of Pediatrics, Division of Hematology, The Children's Hospital of Philadelphia, Philadelphia, PA, United States

[§]Authors contributed equally to this work; *Address correspondence to muzykant@mail.med.upenn.edu

Table S1.^a Binding Epitopes and Parameters of Radioiodinated anti-PECAM Ab to Live Endothelial Cells (EC)

Ab	Species Reactivity	clone	Amino Acid (AA) Epitope Mapping, Ig Domain (IgD)	PECAM		Effect on PECAM Homophilic Adhesion
				K _d (nM)	B _{max} (10 ⁵ Ab/cell)	
Ab _{1h}	Human	62	AA26-AA30, IgD1	4.32 ± 0.30 ^b	2.62 ± 0.26 ^b	Inhibit
Ab _{2h}	Human	37	AA42-AA45, IgD1	0.24 ± 0.02 ^b	1.51 ± 0.12 ^b	No effect
Ab _{1m}	Mouse	MEC13	AA164-AA177, IgD2	2.81 ± 0.13 ^c	5.81 ± 0.2 ^c	No effect
Ab _{2m}	Mouse	390	AA194-AA205, IgD2	0.25 ± 0.01 ^c	2.60 ± 0.06 ^c	Inhibit

^aRef.¹

^bAb binding to native huPECAM on HUVEC

^cAb binding to native muPECAM on murine MS1 cells

Table S2.^a Lung:Blood Tissue Selectivity Summary

Formulation	Localization Ratio		
	IgG	Paired Ab	Specificity Index
¹²⁵ I-Ab _{1m}	2.98	10.22	3.4
¹²⁵ I-Ab _{2m}	2.40	7.75	3.2
Ab _{1m} / ¹²⁵ I-NC	17.55	183.58	10.5
Ab _{2m} / ¹²⁵ I-NC	62.42	238.75	3.8

^aMice were injected intravenously with IgG or paired Ab (at t=-30 min), followed by radiolabeled probe (anti-PECAM ¹²⁵I-Ab or ¹²⁵I-NC) (at t=0 min). Tissues were harvested at t=30 min. Specificity Index denotes the relative lung: blood localization ratio of paired Ab group to IgG control group.

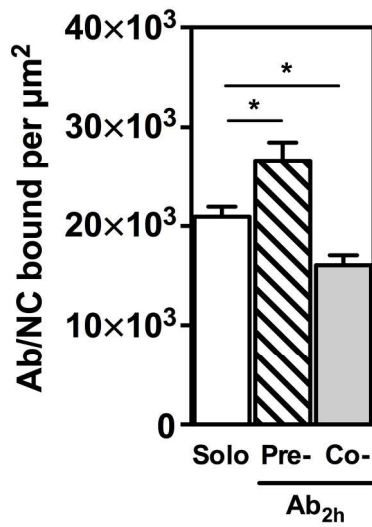


Figure S1. Free paired Ab_{2h} pre-incubation specifically enhances EC binding of $\text{Ab}_{1h}/\text{NCs}$. EC binding of Ab_{1h}/NC at high Ab density over particle surface ($200 \text{ Ab}/\text{NC}$) was enhanced by paired free Ab_{2h} when pre-incubated with cells (hashed bar) than when cells were co-incubated with paired Ab and Ab_{1h}/NC (grey bar) when compared to solo binding (open bar). The number of total EC bound fluorescent particles in each image field as quantified by fluorescence microscopy. Data are mean \pm SE (n=8).

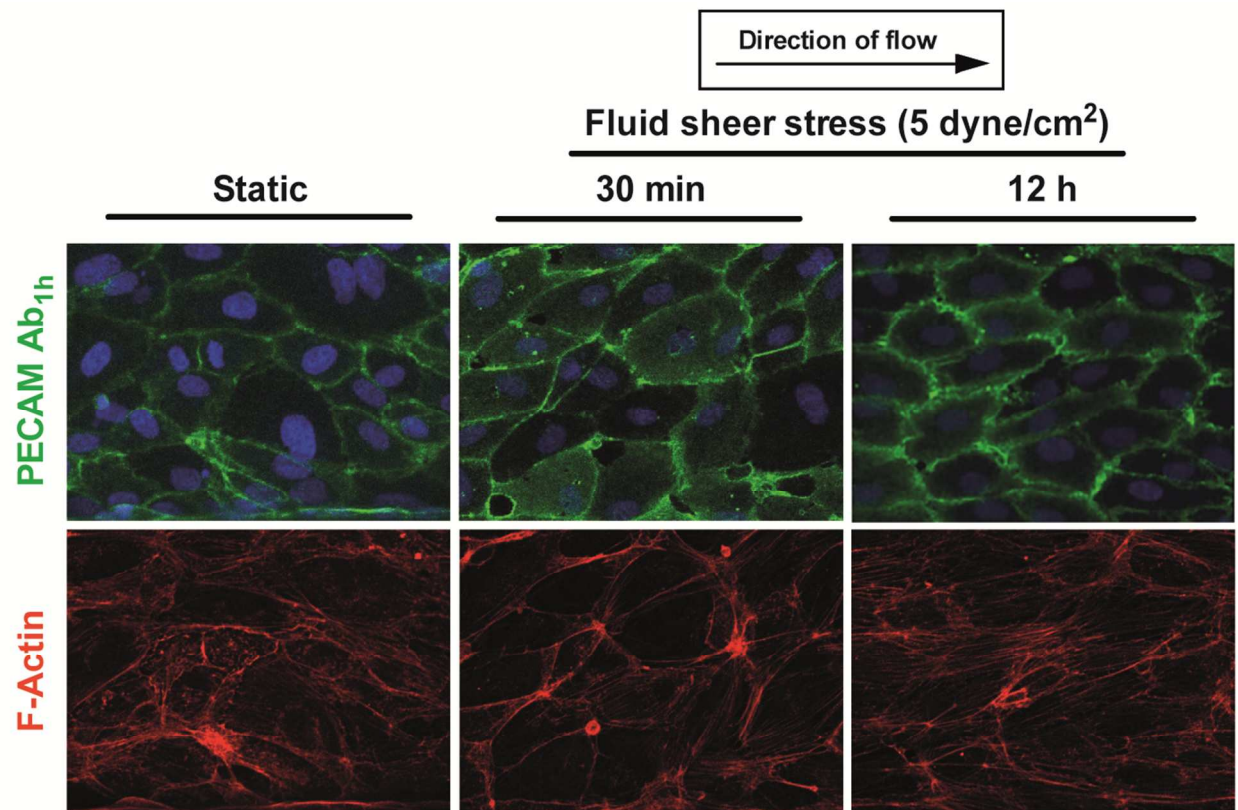


Figure S2. Distribution of PECAM and F-actin with and without exposure to flow. Confluent monolayers of HUVEC were exposed to static conditions, short-term (30 min) acute flow, or overnight (12 h) flow adaption and subsequently fixed, permeabilized, and stained for PECAM (green) and F-actin (red). In contrast to F-actin, PECAM staining demonstrates neither a change in intensity nor distribution, with PEMCAM remaining predominantly at the cell-cell junctions.

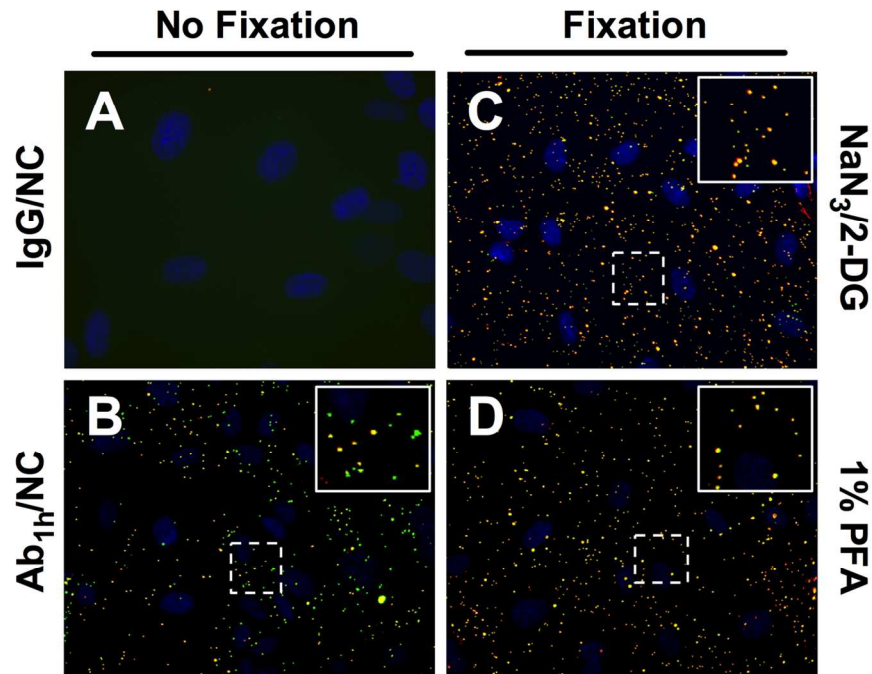


Figure S3. Microscopy studies reveal no internalization of fully coated Ab_{1h}/NC in fixed ECs *versus* live ATP-depleted ECs under fluid shear stress (4 dyne/cm²) following counterstain with secondary red anti-IgG. Merged fluorescence images show internalized (green) and surface-bound (yellow) Ab/NC, with DAPI-stained nuclei (blue). (A) Control IgG/NC have negligible surface binding, whereas Ab_{1h}/NC undergo endocytosis under flow (B). (C) Endocytosis of Ab_{1h}/NC was blocked in ATP-depleted EC treated with NaN₃/2-DG (5 mM, 30 min) as well as with (D) EC pre-fixed with PFA (1%, 10 min). Inset in each panel is close up of selected field.

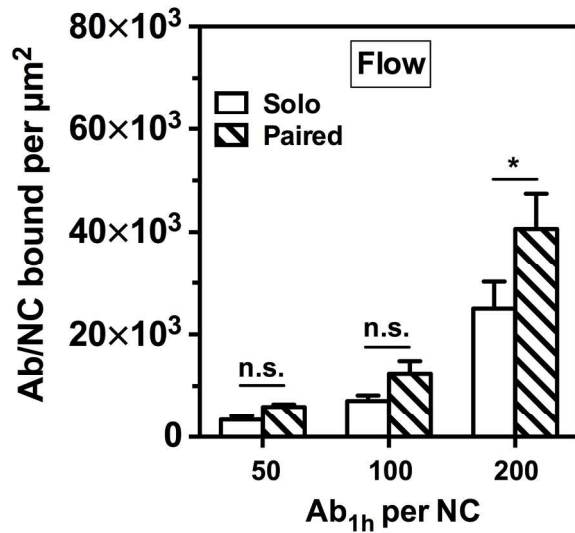


Figure S4. Under flow conditions (4 dyne/cm²), pre-incubation of paired Ab_{2h} (20 nM) stimulated endothelial binding of Ab_{1h}/NCs with different targeted antibody density on particle surface (*i.e.*, # Ab per particle) as compared to pre-incubation with solo control IgG. The number of total EC bound fluorescent particles in each image field as quantified by fluorescence microscopy. Data are shown as mean ± SE (n=8). *, $P < 0.05$; ***, $P < 0.001$; n.s., $P > 0.05$. Open bars show NC binding with solo treatment; hashed bars show NC binding with paired free Ab treatment.

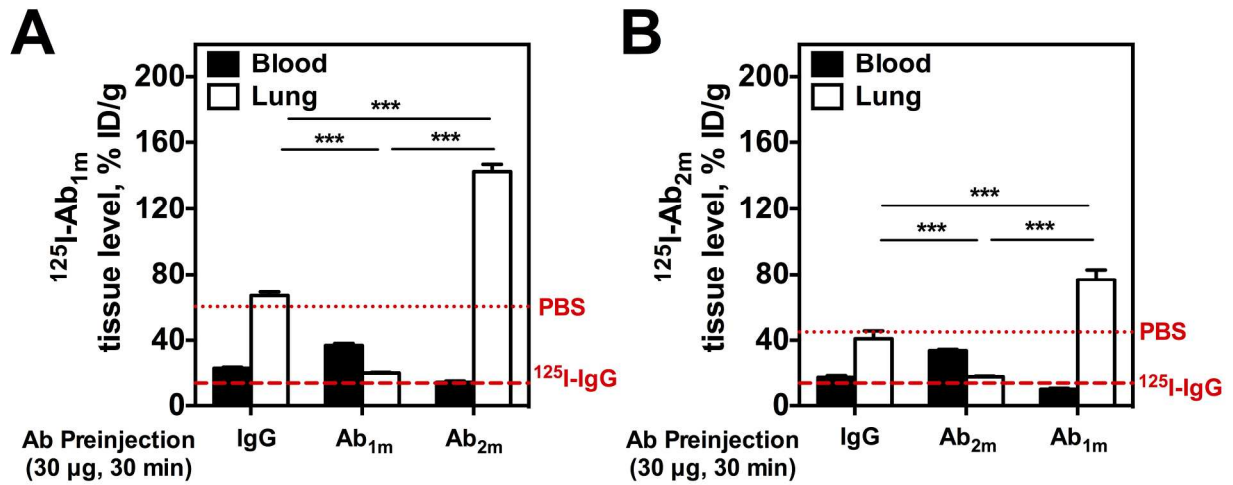


Figure S5. *In vivo* endothelial targeting of $\text{Ab}/^{125}\text{I-NC}$ to μPECAM is enhanced by paired μPECAM Ab. (A) Lung tissue radioactivity levels at 30 min p.i. of anti- μPECAM $^{125}\text{I-Ab}_{1\text{m}}$ is enhanced when mice are preinjected with paired $\text{Ab}_{2\text{m}}$ and uptake blocked with self-paired $\text{mAb}_{1\text{m}}$. Paired enhancement of lung activity levels results in significantly decreased blood levels. The dashed line indicates non-specific lung activity levels of control $^{125}\text{I-IgG}$ and dotted line indicates lung activity levels of probe with vehicle control PBS pre-injection. (B) Lung tissue activity of $\text{Ab}_{1\text{m}}/^{125}\text{I-NC}$ (200 μg NC per mouse) at 30 min p.i. is enhanced significantly with paired $\text{Ab}_{2\text{m}}$ (30 μg for 30 minutes) as compared to IgG pre-injection. Blood levels of $\text{Ab}_{1\text{m}}/^{125}\text{I-NC}$ following paired Ab treatment is lower than blood in IgG pre-treated mice. Dashed line indicates lung uptake of non-specific IgG/ $^{125}\text{I-NC}$ and dotted line is $\text{Ab}_{1\text{m}}/^{125}\text{I-NC}$ lung uptake blocked with $\text{Ab}_{1\text{m}}$ pretreatment. Data represented as mean \pm SE (n = 5 mice per group). *, $P < 0.05$, ***, $P < 0.001$.

REFERENCES

1. Chacko, A. M.; Nayak, M.; Greineder, C. F.; Delisser, H. M.; Muzykantov, V. R. Collaborative enhancement of antibody binding to distinct PECAM-1 epitopes modulates endothelial targeting. *PLoS One* **2012**, *7*, e34958.