Supplementary Information

Truncated yet functional viral protein produced *via* **RNA polymerase slippage implies underestimated coding capacity of RNA viruses**

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Supplementary Figure 1. Schematic model of the production of viral protein truncations *via* **transcriptional slippage (TS) by viral RNA polymerases.** (**a**) A small ORF exists in the $P3$ cistron at the -1 reading frame in the large ORF that encodes the potyvirus polyprotein. Potyviruses produce the P3N-PIPO protein *via* TS by viral RNA polymerase NIb at the $G_{1-2}A_{6-7}$ motif. This study revealed that TS also causes frameshift to the +1 reading frame to produce P3N-ALT. Although P3N-ALT is considered a truncation of P3 because there is no substantial ORF at the +1 reading frame following to the motif, it contributes to the cell-to-cell movement of ClYVV in infected plants. (**b**) The findings in this study imply that truncated, yet functional, proteins from a certain gene *X* can be produced *via* TS, even without a substantial ORF in alternative reading frames of the RNA genomes of other viruses. In fact, as unveiled in this study, a number of $G_{1-2}A_{6-7}$ and G_0A_{6-7} motifs with no substantial ORFs in one or two alternative frames are found in a diverse range of RNA viruses. Our study implied that not only a fusion protein such as P3N-PIPO shown in panel (**a**), but also a truncated protein as P3N-ALT could be produced *via* TS that causes a frameshift to an alternative frame where no substantial ORF exists.

RB-P3(PIPO:FLAG-1)

P3N' (34.3 kDa)

MGKSLTGQVIQFDTKMLISSIYRPRQMEMIINEEPFVLVLAMQSPSVLLALFNSASLEKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQMEIIEA SASHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLMEKIWAEDLEQQWLGLRLSQKFYLIRQSWKQRAKYSKILAQRDELG ASDKFSASLRLSATSIKNQAISCRKRMVATRLVVEVEVVVEVEVVWTTKTMTVIIKIMTSITRIMMGKLDDPGLPAFVRIIGFDNVRQVQCISFIAHTPE

SY* **P3N-PIPO (29.8 kDa)**

MGKSLTGQVIQFDTKMLISSIYRPRQMEMIINEEPFVLVLAMQSPSVLLALFNSASL<mark>EKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQME</mark>IIEA
SASHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLM<mark>EKNLGRGFRAAMARFKIVTKVLFNKAVMEAAGKVFQNISPERRAR</mark> CQRQVQRITQIVSNKHQKPSNQLQEENGRNKTSGGSGGSGGSGGSM**DYKDHDGDYKDHDIDYKDHDG***

P3N-ALTy (18.2 kDa)

MGKSLTGQVIQFDTKMLISSIYRPRQMEMIINEEPFVLVLAMQSPSVLLALFNSASLEKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQMEIIEA
SASHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLMEKFGQRI*

Cl30-P3(PIPO:FLAG-1)

P3N' (32.5 kDa)

MGKSLTGQVIQFDTKMLISSIYRPRQMEKIINEEPFVLVLAMQSPSVLLALFNSASLEKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQMEIIEA SAGHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLMEKIWAEDLEQQWLGLRLSQKFYLIKQSWKQRAKYSKILAQRDELG ASDKFSASLRLSVTSILVVEVEVVVEVEVVWTTKTMTVIIKIMTSITRIMMGKLDDPGLPAFVRIIGFDNVRQVQCISFIAHTPESY*

P3N-PIPO (28.0 kDa) MGKSLTGQVIQFDTKMLISSIYRPRQMEKIINEEPFVLVLAMQSPSVLLALFNSASLEKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQMEIIEA SAGHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLMEKNLGRGFRAAMARFKIVTKVLFNKAVMEAAGKVFQNTSPERRAR CQRQVQRITQIVSNKHTSGGSGGSGGSGGSM**DYKDHDGDYKDHDIDYKDHDG***

P3N-ALT (18.1 kDa)

MGKSLTGQVIQFDTKMLISSIYRPRQMEKIINEEPFVLVLAMQSPSVLLALFNSASLEKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQMEIIEA SAGHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLMEKSGQRI*

CS-P3(PIPO:FLAG-1)

P3N' (35.3 kDa)
MGNQSVGSHIRIDTELLVKSVYKPELMAHIIEQEPFLLVLAMQSPATLMALFNSSSLEKAVQYWLHKDMQVSHIMTMLAVLASNVSASKLLTTQFEVIEA
SAPQILAAMDHVYKPMHSINTANTFLMNLNESRETDKTIDELGFYSFKKSTRILMEKTLMADLDQQWQELGLLERFSLIKRSWQVRAKYSSFAIQREEKG ITDKFTTSLRLSGAQIKHQALAKRDQIVRFAERRVERTLVVEVEVVVEVEVVWTTKTMTVIIKIMTSITRIMMGKLDDPGLPAFVRIIGFDNVRQVQCIS FIAHTPESY*

P3N-PIPO (30.6 kDa)

MGNQSVGSHIRIDTELLVKSVYKPELMAHIIEQEPFLLVLAMQSPATLMALFNSSSLEKAVQYWLHKDMQVSHIMTMLAVLASNVSASKLLTTQFEVIEA SAPQILAAMDHVYKPMHSINTANTFLMNLNESRETDKTIDELGFYSFKKSTRILMEKNLDGGLRSTMARIRIVGKILFNKAIVASASKVFKFCNSERRER
YHRQVHNLTQVIRGTNKAPGPCQERSNCAICREKSREDTSGGSGGGGGGGGGM**DYKDHDGDYKDHDIDYKDHDG***

P3N-ALT (17.8 kDa)

MGNQSVGSHIRIDTELLVKSVYKPELMAHIIEQEPFLLVLAMQSPATLMALFNSSSLEKAVQYWLHKDMQVSHIMTMLAVLASNVSASKLLTTQFEVIEA SAPQILAAMDHVYKPMHSINTANTFLMNLNESRETDKTIDELGFYSFKKSTRILMEKP*

RB-P3(ALT:FLAG+1)

P3N' (18.4 kDa)

MGKSLTGQVIQFDTKMLISSIYRPRQMEMIINEEPFVLVLAMQSPSVLLALFNSASLEKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQMEIIEA SASHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLMEKIWAEDY*

P3N-PIPO' (26.2 kDa)

MGKSLTGQVIQFDTKMLISSIYRPRQMEMIINEEPFVLVLAMQSPSVLLALFNSASLEKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQMEIIEA SASHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLMEKNLGRGLLVVEVEVVVVEVEVVVWTTKTMTVIIKIMTSITRIMMGK
LDDPGLPAFVRIIGFDNVRQVQCISFIAHTPESY*

P3N-ALT:FLAG (21.9 kDa)

MGKSLTGQVIQFDTKMLISSIYRPRQMEMIINEEPFVLVLAMQSPSVLLALFNSASLEKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQMEIIEA
SASHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSOVLMEKFGORITSGGSGGSGGSGGSM**DYKDHDGDYKDHDIDYKDHDG*** SASHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLMEKFG **Cl30-P3(ALT:FLAG+1)**

P3N' (18.4 kDa)

MGKSLTGQVIQFDTKMLISSIYRPRQMEKIINEEPFVLVLAMQSPSVLLALFNSASLEKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQMEIIEA
SAGHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLMEKIWAEDY*

P3N-PIPO' (26.2 kDa)

MGKSLTGQVIQFDTKMLISSIYRPRQMEKIINEEPFVLVLAMQSPSVLLALFNSASLEKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQMEIIEA
SAGHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLMEK<mark>NLGRGLLVVEVEVVVVEVEVVWTTKTMTVIIKIMTSITRIMMGK</mark>
<u>LDDPGLPAFVRIIGFD</u>

P3N-ALT:FLAG (21.8 kDa)

MGKSLTGQVIQFDTKMLISSIYRPRQMEKIINEEPFVLVLAMQSPSVLLALFNSASLEKAVEVWLHKDMRVSHVMTMLALLAAKVSAAKMVNLQMEIIEA
SAGHELAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGEHSLKKSSOVLMEKSGORITSGGSGGSGGSGGSMO**VKDHDGDVKDHDIDYKDHDG*** SAGHFLAAMDTIHKPMHSINTANIFLMNLEEGRSTDRTIDELGFHSLKKSSQVLMEKS **CS-P3(ALT:FLAG+1)**

P3N' (18.0 kDa)

MGNQSVGSHIRİDTELLVKSVYKPELMAHIIEQEPFLLVLAMQSPATLMALFNSSSLEKAVQYWLHKDMQVSHIMTMLAVLASNVSASKLLTTQFEVIEA
SAPQILAAMDHVYKPMHSINTANTFLMNLNESRETDKTIDELGFYSFKKSTRILMEKTY*

P3N-PIPO' (25.9 kDa)

MGNQSVGSHIRIDTELLVKSVYKPELMAHIIEQEPFLLVLAMQSPATLMALFNSSSLEKAVQYWLHKDMQVSHIMTMLAVLASNVSASKLLTTQFEVIEA SAPQILAAMDHVYKPMHSINTANTFLMNLNESRETDKTIDELGFYSFKKSTRILMEK<u>NLLVVEVEVVVEVEVVWTTKTMTVIIKIMTSITRIMMGKLDDP</u>
GLPAFVRIIGFDNVRQVOCISFIAHTPESY* **P3N-ALT:FLAG (21.5 kDa)**

MGNQSVGSHIRIDTELLVKSVYKPELMAHIIEQEPFLLVLAMQSPATLMALFNSSSLEKAVQYWLHKDMQVSHIMTMLAVLASNVSASKLLTTQFEVIEA
SAPQILAAMDHVYKPMHSINTANTFLMNLNESRETDKTIDELGFYSFKKSTRILMEKPTSGGSGGSGGSGGSM**DYKDHDGDYKDHDIDYKDHDG***

Supplementary Figure 2. Amino acid sequence of the constructs used in Fig. 2. Amino acid sequences of the three alternative reading frame products are shown for each of *RB-P3(PIPO:FLAG–1)*, *Cl30-P3(PIPO:FLAG–1)*, *CS-P3(PIPO:FLAG–1)*, *RB-P3(ALT:FLAG+1)*, $RB-P3(ALT: FLAG^{+1})$ and $CS-P3(ALT: FLAG^{+1})$ RNAs. For P3N' and P3N-PIPO', the Cterminal regions are different among the virus constructs. P3 and P3N sequences are shown in green, PIPO sequences are in blue (underlined), ALT sequences are in magenta (underlined), and the FLAG-tag sequence is shown in black (bold).

Supplementary Figure 3. Detection of P3N-ALT accumulation in ClYVV-infected broad bean stems. The stem tissues systemically infected with Cl30 and RB as in Fig. 3 were harvested at 10 days post-inoculation. The infected areas were confirmed by virus-derived GFP fluorescence (left panel). P3N-ALT was detected by western blotting using a polyclonal antibody raised against the N-terminal region of P3. CBB-stained gel image is shown at the bottom of the western blot panel as a loading control. The position of P3N-ALT is marked with an arrowhead. Positions of molecular mass markers (kDa) are indicated on the left side of the panel.

Supplementary Figure 4. Systemic infection of broad bean plants with ClYVV requires P3N-ALT and P3N-PIPO. Broad bean leaves were biolistically co-inoculated with Cl-P3ΔARFPs (Fig.4a) and WClMV vectors, WCl/P3N-ALT expressing P3N-ALT, WCl/P3N-PIPO expressing P3N-PIPO, WCl/P3N-PIPO(ALT^{-1} , $P3^{-1}$) expressing both P3N-PIPO and P3N-ALT, or an empty vector WCl/empty as indicated. Images of a healthy plant (**a**, **e**) are shown for comparison. Systemic movement of Cl-P3ΔARFPs was examined by monitoring GFP fluorescence derived from Cl-P3ΔARFPs. Visible images (**a**–**d**) and GFP fluorescence images in stems at 7 or 8 days post-inoculation (**e**–**j**) are shown. When co-inoculated with Cl-P3ΔARFPs and WCl/empty, no GFP signal was detected in any of the non-inoculated parts of the plants (**f**), whereas sporadic GFP signals (arrows) were detected in systemic stems of plants that were co-inoculated with WCl/P3N-ALT plus WCl/P3N-PIPO (**g**, **i**) or WCl/P3N-PIPO(ALT–1, P3+1) (**h**, **j**). This suggests that expression of P3N-ALT and P3N-PIPO *in trans* using WClMV vectors at least partly compensated for the defect of Cl-P3ΔARFPs to enable systemic spread of the virus in the inoculated broad bean plants. The test was repeated twice, and representative images are shown. Scale bar = 5 mm.

Supplementary Figure 5. Alternative reading frame proteins are produced regardless of the original reading frame *in vitro***.** (**a**) Schematic diagram of RNA used for *in vitro* translation analysis. The nucleotide sequences around the G_2A_6 motif derived from RB were replaced with *GFP* RNA sequence (red italic letters) in three different reading frames to generate *GFP-G1A6(f1)*, *GFP-G1A6(f2)* and *GFP-G1A6(f3)* RNAs. The replaced sequences are marked in red italic letters. Spaces denote the zero reading frame. The G_2A_6 motif is underlined. (**b**) *In vitro* translation analysis using WGE. RNA prepared *in vitro* were translated in the presence of $\int^{35}S$]methionine. The translation products were separated by SDS-PAGE, and the signals were visualized by autoradiography. The positions of zero frame, +1 frame, and –1 frame products are marked on the right side of the panel. Positions of molecular mass markers (kDa) are indicated on the left. (**c**) The accumulated levels of –1 and +1 frame proteins produced in each construct. In the calculation, the number of methionine codons present in each product was taken into account. The value for each product was obtained from at least two independent translation assays. The means and standard deviations are shown.

Supplementary Figure 6. Results of statistical tests for enrichment of indels in systemically infected viral cDNA. (**a**–**d**) Solid lines indicate *q*-values for enrichment of one base insertion (**a**,**b**) and one base deletion (**c**,**d**) in viral cDNAs from Cl30 (**a**,**c**) and RB (**b**,**d**), which are shown in Fig. 5b,c. Fisher's exact test was performed for the indel counts observed at each position in the amplicons from systemically infected viral RNA and control plasmids used for inoculation (Supplementary Methods). The *q*-values were obtained by multiple test correction of *p*-values calculated by Fisher's exact test using Storey's method. The vertical axis represents negative log_{10} -transformed *q*-values. The red dashed lines indicate a statistical cut-off (*q*-value = 0.05).

Supplementary Figure 7. Frequency of transcriptional slippage at ClYVV P3 region estimated using a negative binomial regression. Frequencies of one base insertions in Cl30 (**a**) and RB (**b**), and that of one base deletions in Cl30 (**c**) and RB (**d**), are shown. Sites and distribution of indels at the ClYVV P3 region were measured by amplicon sequencing of cDNA prepared from total RNA of Cl30 or RB-infected broad bean leaves. As controls, plasmids used for inoculation were subjected to sequencing. The frequency of insertions and deletions per base was estimated using a negative binomial regression as a ratio to the total number of reads obtained from respective samples. Solid and dashed lines indicate estimated indel frequencies in systemically propagated viral sample and inoculated plasmid sample, respectively. The indicated frequency was estimated in a window size of seven bases (Supplementary Table 2), with a step size of one base. Standard errors of the estimated frequencies are shown as shaded area. Numbers at the bottom indicate the position in the amplicons (numbered from the 5′ end). Seven base-windows in which indel frequencies are statistically significantly different between viral RNA and control plasmid are shown with red bars under the plot (adjusted $p < 0.05$). The adjusted *p*-values were obtained by multiple test correction using Holm's method. The $33rd$ A, which is the common site among all the significant windows is highlighted by orange bars. Estimated indel frequencies obtained using different window sizes are shown in Supplementary Table 2.

Supplementary Figure 8. Alignment of the P3N-ALT amino acid sequences of potyviruses. Potyviral ORF sequences were retrieved from GenBank using the queries "potyvirus" and "complete genome". These entries were further filtered by selecting for those with annotation for the *P3* cistron. The *P3* cistron sequences of the resultant 197 potyvirus genomes were obtained and one base deletion at the $G_{1-2}A_{6+}$ or G_0A_{6+} motif was simulated using a custom Perl script. The simulated sequences were aligned using Muscle v3.8.31 software. For those entries that gave identical P3N-ALT amino acid sequence, only one entry is shown. These include (the omitted Accession Nos. are listed in the parentheses), plum pox virus: AB576046 (AB576047, AB576048, AB576049, AB576050, AB576051, AB576052, AB576054, AB576055, AB576056, AB576057, AB576058, AB576059, AB576061, AB576062, AB576063, AB576064, AB576065, AB576066, AB576067, AB576068, AB576069, AB576070, AB576073, AB576074, AB576075, AB576076, AB576077, AB576078, AB576079, AB576080); potato virus Y: AB711153 (AB711154, AB711155, AB714134); AB711143 (AB711152); AB711147 (AB711148); AB711145 (AB711146); bean yellow mosaic virus: AB079886 (AB079887); konjac mosaic virus: NC_007913 (AB219545); papaya leaf distortion mosaic virus: AB088221 (NC_005028); soybean mosaic virus: AY294044 (AJ312439); zucchini yellow mosaic virus: AB188115 (AB188116); Algerian watermelon mosaic virus: EU410442 (NC_010736); Moroccan watermelon mosaic virus: EF579955 (NC_009995); sugarcane mosaic virus: JX237862 (JX237863); peanut mottle virus: AF023848 (NC_002600); turnip mosaic virus: AB701740 (AB701741); AB701702 (AB701728, AB701731, AB701732); AB701700 (AB701719); AB701712 (AB701714); AB194790 (AB194791); AB194785 (AB194786, AB194787, AB194788, AB194789, AB194792, AB194793, AB194794, AB194795, AB194796, AB194797, AB194798, AB194799, AB194800, AB194801, AB194802); AB701733 (AB701738); AB701734 (AB701735); AB701690 (AB701691); Habenaria mosaic virus: AB818538 (NC_021786); chilli veinal mottle virus: AJ972878 (AM909717). As a result, 125 entries are shown. Lupine mosaic virus (LuMV; Accession No. NC 014898) was recently isolated 68 and is distantly related to other potyviruses⁴⁹. LuMV does not carry the $G_{1-2}A_{6-7}$ motif⁶⁷, but carries G_0A_6 within the *P3* cistron at a similar position to other potyviruses. Amino acid sequences of the ALT region were aligned manually. LuMV was placed at the bottom of the figure. Number of amino acid residues from the N-terminus of P3N is indicated. The amino acid sequence EK, which is highlighted in yellow, is of the expected slippage site of $G_{1-2}A_{6-7}$ motif in P3. P3N and ALT regions are marked in green and magenta, respectively.

Supplementary Figure 8 (part 1/3)

Supplementary Figure 8 (part 2/3)

Supplementary Figure 8 (part 3/3)

Supplementary Figure 9. Western blotting to confirm specific binding of the anti-P3N peptide antibody to the FLAG-tagged P3 products of Cl30, P3, P3N-PIPO and P3N-ALT. RNAs that produce FLAG:P3, P3N-PIPO:FLAG and P3N-ALT proteins (Supplementary Methods) were transiently expressed in *N. benthamiana* leaves by agroinfiltration, as in Fig. 2. The products were detected by western blotting using an anti-FLAG monoclonal antibody (**a**) or the anti-P3N peptide polyclonal antibody (**b**). Loading controls are the part of the PVDF membranes covering the RubisCO large subunit band that were stained with Amido Black after western transfer. The membranes were stained with 0.1% Amido Black in 45% methanol and 10% acetic acid followed by destaining in 90% methanol and 2% acetic acid.

Supplementary Table 1. Occurrence of G_0A_{6+} and $G_{1\cdot 2}A_{6+}$ motifs in GenBank/EMBL/DDBJ accessions.

^a Number of accessions in GenBank/EMBL/DDBJ.
^b Number of accessions containing the indicated motif at least once.

^a Indel frequencies were estimated by fitting the negative binominal model to observed counts of the reads with indels obtained by amplicon sequenceing. SE, standard error of estimated mean.

b *p*-values for differences between the estimated indel frequencies between inoculated plasmid and systemically propagated virus were obtained by the model fitting and were corrected for multiple tests by Holm's method.

Supplementary Table 3. Primers used in this study.

Supplementary Methods

Plasmids used for Supplementary Fig. 5

To construct pSP/GFP-f1 [to prepare *GFP-G1A6(f1)* RNA], pSP/GFP-f2 [to prepare *GFP-G1A6(f2)* RNA] and pSP/GFP-f3 [to prepare *GFP-G1A6(f3)* RNA], a template plasmid pSP/GFPstp was generated. The sGFP(S65T) sequence present in the 35Somega-sGFP(S65T) plasmid69 was PCR amplified using primers BamHI-kzk-GFP-F and GFP-SmaI-SacI-R (Supplementary Table 3). The PCR product was inserted into pSP64 Poly(A) vector using the *Bam*HI and *Sac*I sites. The resulting plasmid was used as a template for inverse PCR with primers sGFP642-stop-F and sGFP6421-R (Supplementary Table 3) to introduce a stop codon with the $+1$ reading frame. The PCR product was phosphorylated and circularized to produce pSP-GFPstp. Inverse PCR was further conducted using pSP/GFPstp as a template, reverse primer sGFP-387-R (Supplementary Table 3) and one of the following forward primers; TTTf1 for pSP/GFP-f1; TTTf2 for pSP/GFP-f2; and TTTf3 for pSP/GFP-f3 (Supplementary Table 3). The PCR product was phosphorylated and circularized.

Indel enrichment analysis for Supplementary Fig. 6

Enrichment of indels at each position in the amplicons was analysed using Fisher's exact test. Briefly, a 2×2 contingency table comprising counts of reads with or without indels in viral or plasmid samples was prepared for each position and each genotype. The ratios of counts of reads with or without indels for each position were used as null hypotheses and *p*-values for enrichment of indels in viral samples were obtained. Storey's *q*-values were calculated from the *p*-values. These were performed using R software (version 3.1.2) with the qvalue package (version 1.43).

Plasmids used for Supplementary Fig. 9

P3 of RB and Cl30 were amplified with primers 1009/1049 and 1009/1007 (Supplementary Table 3) from P3+P3N-PIPO and pCl30, respectively. Each fragment was digested with *Xho*I and *Spe*I, and introduced into the pTA/3FLAG-XhSp vector to obtain pTA/FLAG-P3-RB and pTA/FLAG-P3-Cl30, which produce P3 proteins tagged with 3×FLAG at the N-terminus. The pTA/3FLAG-XhSp vector was created by introducing 3×FLAG tag *+* Gly/Ser linker + *Xho*I + *Spe*I fragment with *Sal*I site at the 5′ terminus (gtcgacATGGACTACAAAGACCATGACGGTGATTATAAAGATCATGACATCGATTACA AGGATCATGATGGGggtggaagtggaggtagtggtggaagtggaggtagtCTCGAGatggccACTAGT) into the cloning site using the *Xho*I and *Spe*I sites of pTA7001.

Supplementary References

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