SUPPORTING INFORMATION

Quantitative Profiling of Protein S-Glutathionylation Reveals Redox-Dependent

Regulation of Macrophage Function During Nanoparticle-Induced Oxidative Stress

Jicheng Duan¹, Vamsi K. Kodali¹, Matthew J. Gaffrey¹, Jia Guo^{1,2}, Rosalie K. Chu,³ David G.

Camp,¹ Richard D. Smith,^{1,3} Brian D. Thrall,^{1*} and Wei-Jun Qian^{1*}

¹Biological Sciences Division, ³Environmental Molecular Sciences Laboratory, Pacific

Northwest National Laboratory, Richland, WA 99352

²Current address: BioAnalytical Sciences, BioMarin Pharmaceutical Inc., Novato, CA 94949

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*Address correspondence to:

Dr. Wei-Jun Qian Biological Science Division, Pacifica Northwest National Laboratory P.O. BOX 999, MSIN: K8-98, Richland WA, 99352 Phone: (509) 371-6572; Fax: (509) 371-6546 Email: weijun.gian@pnnl.gov

Dr. Brian D. Thrall Biological Science Division, Center for Nanotoxicology, Pacifica Northwest National Laboratory P.O. BOX 999, MSIN: J4-02, Richland WA, 99352 Phone: (509) 371-7307; Fax: (509) 371-7304 Email: brian.thrall@pnnl.gov



Figure S1. Western blot analysis of time-dependent increase of SSG in RAW 264.7 cells after treatment by CoO nanoparticles. Cells were treated by 25 μ g/mL CoO nanoparticles and cells treated by 500 μ M diamide for 30 min were used as positive control. The overall level of SSG in cells was detected on the blot using anti-glutathione monoclonal antibody (Virogen, Watertown, MA).



Figure S2. Venn diagram summarized identification of SSG-modified proteins. (a) The overlaps of identified SSG-modified sites, and (b) SSG modified proteins between different ENP-treatment conditions. (c) The overlap of identified proteins between SSG enrichment and total Cys enrichment after CoO treatment. (d) The overlap of substantial proteins between SSG enrichment SSG enrichment and total Cys enrichment after CoO treatment. Threshold of significance: p-value ≤ 0.05 and $|\log 2$ ratio $| \geq 0.3$ versus controls.



Figure S3. Summary of folder change for proteins with substantial change in SSG modification and total expression after CoO treatment. Threshold of significance: p-value ≤ 0.05 and $|\log 2$ ratio $|\geq 0.3$ versus controls.



Figure S4. Distribution of molecular types of identified SSG-modified proteins with substantial SSG alteration after Fe₃O₄ and CoO treatments. Threshold of significance: p-value ≤ 0.05 and log2 ratio ≥ 0.3 versus controls.



Figure S5. TEM characterization for Fe₃O₄ ENPs. The SEM pictures for SiO₂ and CoO ENPs can be found in manufacturers' websites (SiO₂: http://www.nanoamor.com/inc/sdetail/260; CoO: http://www.ssnano.com/inc/sdetail/cobalt_oxide_nanoparticles/238).