

22 **Supplementary Information**

23

24 **Figure S1**

25 FTIR analysis of the EPS from *Acidovorax avenae* subsp. *avenae* strain RS-1. (a) The
26 *hcp*-mutant strain; (b) The wild type strain under exposure to Amp; (c) The wild type
27 strain. Compared to the wild type, both the mutation of *hcp* gene and exposure to
28 Amp caused the disappearance of one peak at 859.16 cm^{-1} representing phenyl ring
29 substitution bands/alkenes. Each experiment was repeated three times independently
30 with similar results.

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33 **Figure S2**

34 Gene ontology categories for the differentially expressed genes under Amp (+) vs.
35 Amp (-) condition of *Acidovorax avenae* subsp. *avenae* strain RS-1 transcriptome. All
36 genes were classified into Biological process, Molecular function and Cellular
37 component catalogs based on their gene ontology annotations.

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40 **Figure S3**

41 Expression changes of T6SS-related genes in *Acidovorax avenae* subsp. *avenae* strain
42 RS-1. (a) Mutation of *hcp* gene; (b) Exposure to Amp. Each result represents the
43 average of three independent determinations with similar results.

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48 **Table S1**

49 Primers used in this study.

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52 **Table S2.**

53 Effect of three kinds of β -lactam antibiotics on the growth and pathogenicity of
54 *Acidovorax avenae* subsp. *avenae* strain RS-1 to rice seedlings.

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57 **Table S3.**

58 Summary of *Acidovorax avenae* subsp. *avenae* strain RS-1 cDNA samples sequenced
59 using the Illumina genome analyzer.

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62 **Table S4**

63 RNA-Seq data of *Acidovorax avenae* subsp. *avenae* strain RS-1 under Amp (-) and
64 Amp (+) conditions. The data of Amp (-) obtained from Li *et al.* (2014)¹⁰ were used as
65 the control in this study.

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68 **Table S5**

69 List of specifically expressed genes under Amp (+) condition of *Acidovorax avenae*
70 subsp. *avenae* strain RS-1 transcriptome.

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73 **Table S6**

74 List of specifically expressed genes under Amp (-) condition of *Acidovorax avenae*
75 subsp. *avenae* strain RS-1 transcriptome.

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78 **Table S7**

79 List of the common expressed genes under both Amp (-) and Amp (+) conditions of
80 *Acidovorax avenae* subsp. *avenae* strain RS-1 transcriptome.

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83 **Table S8**

84 List of up-regulated expressed genes under Amp (+) vs. Amp (-) condition of
85 *Acidovorax avenae* subsp. *avenae* strain RS-1 transcriptome.

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88 **Table S9**

89 List of down-regulated expressed genes under Amp (+) vs. Amp (-) condition of
90 *Acidovorax avenae* subsp. *avenae* strain RS-1 transcriptome.

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93 **Table S10**

94 List of differentially expressed β -lactamase-related genes under Amp (+) vs. Amp (-)
95 condition of *Acidovorax avenae* subsp. *avenae* strain RS-1 transcriptome.

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98 **Table S11**

99 qRT-PCR analysis of T6SS genes expression in *Acidovorax avenae* subsp. *avenae*
100 strain RS-1 under Amp (+) vs. Amp (-) condition.

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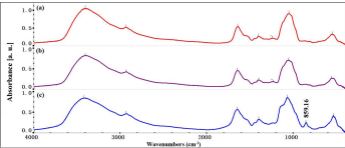
102

103 **Table S12**

104 Gene ontology enrichment analysis for differentially expressed genes under Amp (+)
105 vs. Amp (-) condition of *Acidovorax avenae* subsp. *avenae* strain RS-1 transcriptome.

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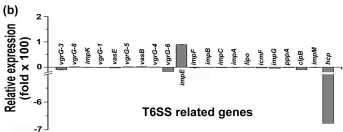
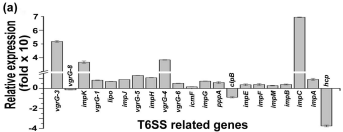


Table S1 Primers used in this study

Name	Sequence (5'- 3')	Usage description	Sources or References
Pt-hcp	F-CCTCGGATCCATGTCCGTCGATATGTTTCATG R-GCGCGAATTCTTACATTTTCCTTGTGGCCCTTG	Pull-Down and Bacterial Two-hybrid	This study
Pt-vgrG	F-CCGAATTCATGACACGCAGCGTCACCATAC R-CATCTCGAGGTGCACGATGAGCCCGTTCTTG	Pull-Down	This study
Pt-lipo	F-GCGGATCCATGCATCGGGAAGCGATGGC R-CCGGAATTCGCGGTCTGGATCTTCACCGC	Pull-Down	This study
Th-hcp	F-CCTCGGATCCATGTCCGTCGATATGTTTCATG R-GCGCGAATTCTTACATTTTCCTTGTGGCCCTTG	Bacterial Two-hybrid	This study
Th-lipo	F-GCGGATCCATGCATCGGGAAGCGATGGC R-CCGGAATTCGCGGTCTGGATCTTCACCGC	Bacterial Two-hybrid	This study
Th-vgrG	F-CCTCTAGAATGACACGCAGCGTCACCATAC R-CCGAATTCGTGCACGATGAGCCCGTTCTTGCC	Bacterial Two-hybrid	This study
GK-hcp-F	F-TATGGATCCATTTTCCTTGTGGCCCTTG R-GCGGAATTCGACATCCGTTTCCTTCAC	Gene Knockout	This study
16S	F-TTGCGGTCCCCTGCTTTCAT	Quantitative	Li et al.
rRNA	R-CGGTAACAGGTCTTCGGATGCT	Real-time PCR	(2014) ¹⁰
clpB	F-GCAGGGCGAGAAGGACAAG R-GCCGAGGAACAGGAACGAG	Quantitative Real-time PCR	Li et al. (2014) ¹⁰
hcp	F-CTGGGTCAGGTGGAGATCTC R-TGGGTCTTGATTTGGCTGC	Quantitative Real-time PCR	Li et al. (2014) ¹⁰
dotU	F-CCAGCATTACCTGCTCGAAT R-CCAGGTCTCGTTGTGCAGT	Quantitative Real-time PCR	Li et al. (2014) ¹⁰
icmF	F-ACCGTGGGCAGCAATCTCA R-GCGAAGTCATCGCTCGTCA	Quantitative Real-time PCR	Li et al. (2014) ¹⁰
impA	F-CTTGAACCTGCGGCGGACAC R-GCTCGGCGGGAATCACCAT	Quantitative Real-time PCR	Li et al. (2014) ¹⁰
impB	F-ATCTCCCTCATCCTGCTCA R-TCAGATGCGTCCCATCAG	Quantitative Real-time PCR	Li et al. (2014) ¹⁰
impC	F-GCACCACCTGGTCCACAACA R-CGAACTGGCCGTATTCTCT	Quantitative Real-time PCR	Li et al. (2014) ¹⁰
impE	F-TGATCGGCTCGCTGTTCG R-TGCTTGTA CTGCCCCTTGTT	Quantitative Real-time PCR	Li et al. (2014) ¹⁰
impF	F-TGGACTGGAAGGACGTGGAA R-AGGGTGTGTGGTGGTTGAA	Quantitative Real-time PCR	Li et al. (2014) ¹⁰
impG	F-TGGAACCTCGGCCTCTATGG R-TGGTGGAAAGATGTCCGAGAA	Quantitative Real-time PCR	Li et al. (2014) ¹⁰
impH	F-GCCACAAGTTCCTTTTGCA R-AAGAACGGCACGAAATCC	Quantitative Real-time PCR	Li et al. (2014) ¹⁰
impJ	F-TCCAGGATGCCAACGACA R-GACCACGGTGGGAATGAA	Quantitative Real-time PCR	Li et al. (2014) ¹⁰

impM	F-GCAATGGCGTCGTCCTCT	Quantitative	Li et al.
	R-CGGTCGTGCCGATCTTCT	Real-time PCR	(2014) ¹⁰
lipo	F-GCAGTGGGATGTCCGTACCTT	Quantitative	Li et al.
	R-TCCTTGCCACCGTGATGCT	Real-time PCR	(2014) ¹⁰
pppA	F-AGATCACGCGGGACCATT	Quantitative	Li et al.
	R-TTCCTCGTCGTCGAGCAT	Real-time PCR	(2014) ¹⁰
vgrG-1	F-ATCCGATGGAAAAGAACTC	Quantitative	Li et al.
	R-AATAGATGCCCTCGTGCT	Real-time PCR	(2014) ¹⁰
vgrG-2	F-GCGTGCAATATGACGAGAGC	Quantitative	Li et al.
	R-CCGGCGGATAGAAGGGAATC	Real-time PCR	(2014) ¹⁰
vgrG-4	F-CTGACGCAGAGCACGAAT	Quantitative	Li et al.
	R-CCGAAGCACCATACCA	Real-time PCR	(2014) ¹⁰
vgrG-5	F-CATCAAGACCAAGTCCAGC	Quantitative	Li et al.
	R-CAGCCATAATTGCTCTGC	Real-time PCR	(2014) ¹⁰
vgrG-7	F-CCGATGGAAAAGAACTCAG	Quantitative	Li et al.
	R-AATAGATGCCCTCGTGCT	Real-time PCR	(2014) ¹⁰
vgrG-8	F-TCCTTCCAGAAGTTCAGCC	Quantitative	Li et al.
	R-GGTATTCGTCGGTCCAGATT	Real-time PCR	(2014) ¹⁰

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3 **Table S2.**4 Effect of three kinds of β -lactam antibiotics on the growth and pathogenicity of5 *Acidovorax avenae* subsp. *avenae* strain RS-1 to rice seedlings.

Treatments	Bacterial growth (OD600)			Plant parameters		
	12 h	24 h	48 h	Root length	Height	Emergence
Strain RS-1	0.15 b	0.82 c	1.24 b	15.94 b	9.16 b	23.33 c
+ Amp	0.03 d	0.79 d	1.20 b	28.97 a	21.06 a	67.74 a
+ Amo	0.03 c	0.92 b	0.97 c	38.47 a	36.63 a	40.00 b
+ Pen	0.34 a	1.12 a	1.33 a	49.10 a	43.60 a	36.67 b
None	-	-	-	177.40 a	132.47 a	100.0 a

6 In absence of pathogen, exposure to Amo caused a 20.05%, 40.49% and 20.00%

7 reduction, while exposure to Pen resulted in a 48.93%, 32.79% and 20.00% reduction

8 in root length, plant height and emergence compared to the control. However, there

9 was no significant ($P < 0.05$) difference in plant parameter between Amp (+) and10 Amp (-). The same letters indicate no significant differences ($P < 0.05$) among

11 treatments. Each treatment has 25 replicates and this experiment was repeated four

12 times independently with similar results.

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3 **Table S3.**

4 Summary of *Acidovorax avenae* subsp. *avenae* strain RS-1 cDNA samples sequenced
5 using the Illumina genome analyzer

Sample name	Amp (+, 1)	Amp (+, 2)
Number of reads	14,537,291	16,443,928
Mapped reads	12,679,967	15,732,263
Unique mapped	4,755,618	6,149,831
mRNA percent	35.1%	39.1%
Number and percentage at different transcript levels		
High-	809 (16.68%)	
Medium-	1088 (22.43%)	
Low-	2014 (41.52%)	
None-	940 (19.38%)	

6 Classification of transcription profile for various categories of transcription level was
7 carried out as described by Nagalakshimi et al. (2008)²⁵. The data of Amp (-) used as
8 the control in this study were obtained from Li et al. (2014)¹⁰.

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