SUPPLEMENTARY DATA





Figure S1 | Occurrence of mRNA codons for each amino acid in the A-, P- and E-site.

(A-C) Ribosomal profiling was performed on the ribosomes from the *in vivo* pullout using the affinity-tagged Ski complex and the normalized occurrence of the mRNA codons for each amino acid in the (A) A-site, (B) P-site, and (C) E-site was calculated.



Figure S2 | Resolution of the eIF-5A-80S complex cryo-EM reconstruction. (A) Resolution curve for the final cryo-EM map of the eIF-5A-80S complex as obtained from RELION (28). The resolution was determined to 3.9 Å at a FSC cut-off at 0.143 according to the gold-standard criterion. (B) eIF-5A-80S map and the isolated density for eIF-5A colored according to the local resolution as obtained by ResMap (30). (C-E) Examples densities (grey mesh) and models for (C) uL16 (orange), (D) CCA-end of the P-tRNA (green) and (E) H93 of the 23S rRNA (blue). (F) Density for cycloheximide (CHX) in the E-site and the potential clash with the CCA end of an E-site tRNA.



Figure S3 | Comparison of eIF-5A, aIF-5A and EF-P structures.

(A-F) Comparison of the conformation of yeast eIF-5A in complex with the 80S ribosome (bound, red) with (A) crystal structure of free yeast eIF-5A (unbound, green, PDB ID 1ER0), (B) with crystal structures of free eukaryotic eIF-5As from yeast (green, 1ER0), human (49) (green, 3CPF), *Leishmania mexicana* (light green, 1X60) and *L. braziliana* (bright green, 1XTD), (C) with crystal structures of free archaeal aIF-5As from *Methanococcus jannaschii* (59) (cyan, 1EIF), *Pyrococcus horikoshii* (60) (blue, 1I26), and *Pyrobaculum aerophilum* (61) (grape, 1BKB), (D,E) crystal structure of bound *Thermus thermophilus* EF-P (16) (pink, 4U6A) relative P-tRNA (green), and (F) with crystal structures of free forms of bacterial EF-Ps from *Coxiella burnetii* (62) (yellow, 3TRE), *Pseudomonas aeruginosa* (63) (bright green, 3OYY), *Thermus thermophiles* (64) (khaki, 1UEB) and *Clostridium thermocellum* (brown, 1YBY).



Figure S4 | Comparison of L1 conformations.

(A) Comparison of the L1 stalk conformation in the presence of yeast eIF-5A (upper panel), eEF2 (50) (second row), E-tRNA (48) (third row) and with the uL1 out position observed in stalled wheat germ 80S ribosome (65,66) (lower panel). (B) Comparison of uL1 (yellow) in presence eIF-5A (upper panel) compared with uL1 (orange) in presence of EF-P (16) (lower panel). (C) Comparison of uL1 (yellow) stalk in presence eIF-5A (top left) compared with uL1 (orange) in presence of EF-P (16) (lower panel). (C) Comparison of uL1 (yellow) stalk in

right), or with uL1 (bright orange) in presence of E-tRNA (48) (bottom left). An overlay of uL1 positions is shown in the bottom right panel.



Figure S5 | Interactions between eIF-5A and the 25S rRNA.

(A) Schematic and (B) molecular representation of the interactions between eIF-5A (red), P-tRNA (green) and 25S rRNA nucleotides (slate). The interactions with P-tRNA are omitted for clarity in (B).



Figure S6 | Hydroxyl-radical probing of eIF-5A.

(A-B) Hydroxyl-radical probing cleavages (7) are mapped onto the (A) 25S rRNA (light blue ribbons) and (B) P-tRNA (green ribbon) from iron tethers located at positions S36 (green), K48 (magenta), M105 (blue) and T126 (red) of eIF-5A (pink ribbons).

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