

Supplementary Information

Identification of a Conserved 5'-dRP Lyase Activity in Bacterial DNA Repair Ligase D and Its Potential Role in Base Excision Repair

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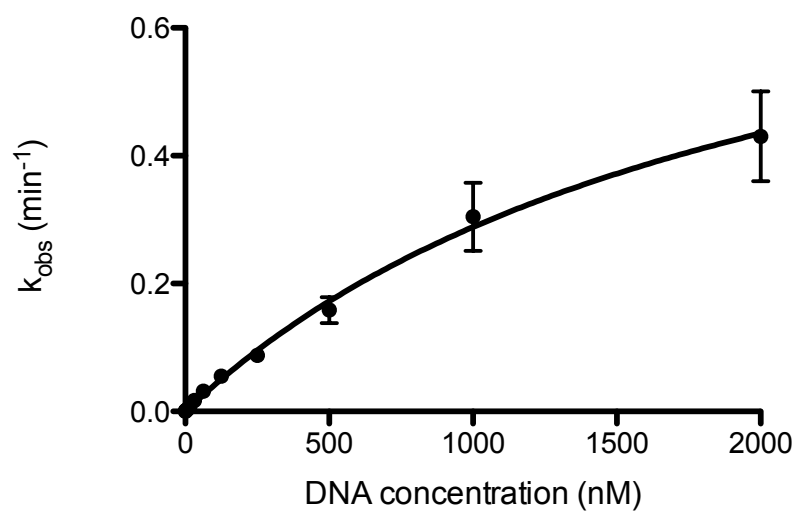
Table S1. Bacterial strains used in this study

Strain	Gene/mechanism	Genotype	Source/reference
168	wt	<i>trpC2</i>	Laboratory strain
BFS184	LigD [NHEJ]	+ <i>ykoU:erm^a</i>	(1)
GP1502	Nfo [EndoIV]	+ <i>Δnfo:cat^b</i>	(2)
GP1502L	LigD Nfo	+ <i>ykoU:erm Δnfo:cat^c</i>	GP1502 DNA transform BFS1845
BG214	wt	<i>trpCE metA5 amyE1 ytsJ1 rsbV37 xre1 xkdA1 att^{SPB} att^{ICEBs1}</i>	Laboratory strain
BC1000	wt	+ <i>neo^d</i>	This work
BC1001	LigDE184A	+ <i>ykoUE184A neo^d</i>	This work
BC1002	Nfo	+ <i>Δnfo:cat^b neo^d</i>	This work
BC1003	LigDE184A Nfo	+ <i>ykoUE184A neo^d Δnfo:cat^b</i>	This work

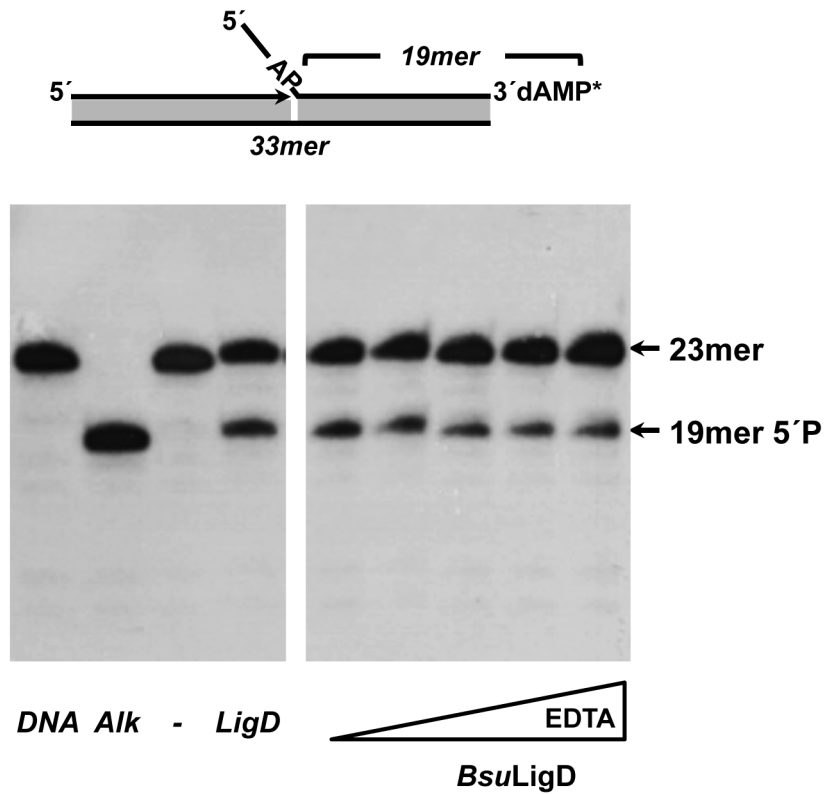
^aResistance to erythromycin (Erm^R, 2 μg/ml), ^bResistant to chloramphenicol (Cam^R, 5 μg/ml), ^cErm^R (2 μg/ml) and Cam^R (5 μg/ml), ^dResistant to neomycin^R (Neo^R, 5 μg/ml).

References

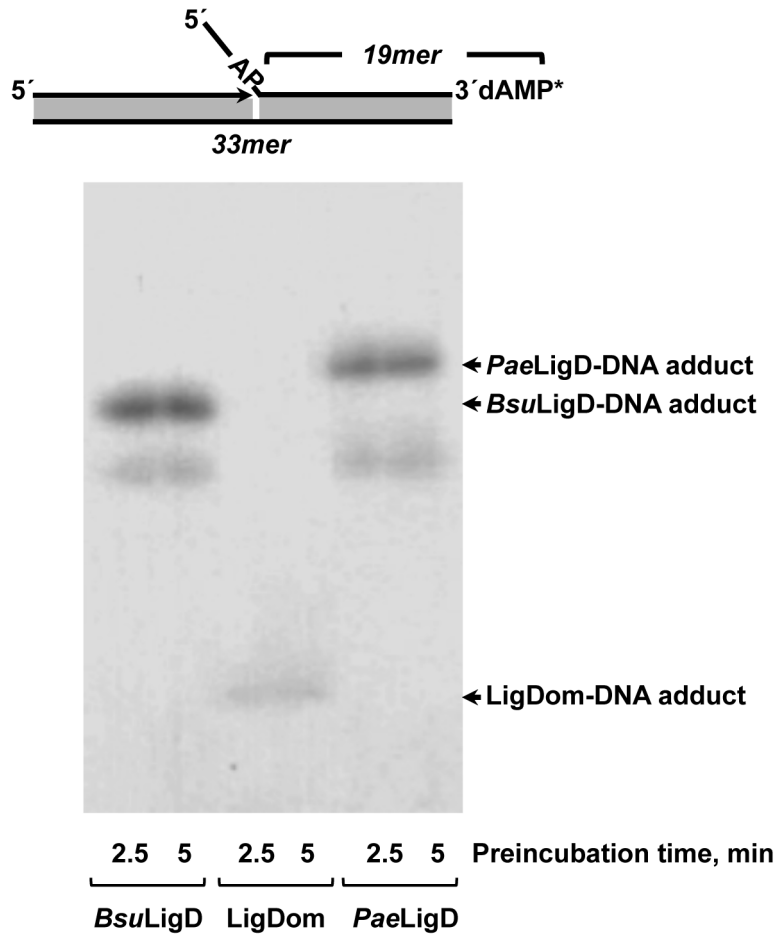
1. Weller GR, Kysela B, Roy R, Tonkin LM, Scanlan E, *et al.* (2002) Identification of a DNA nonhomologous end-joining complex in bacteria. *Science* **297**(5587):1686-1689.
2. Gunka K, Tholen S, Gerwig J, Herzberg C, Stulke J, *et al.* (2012) A high-frequency mutation in *Bacillus subtilis*: requirements for the decyptification of the *gudB* glutamate dehydrogenase gene. *J Bacteriol* **194**(5):1036-1044.



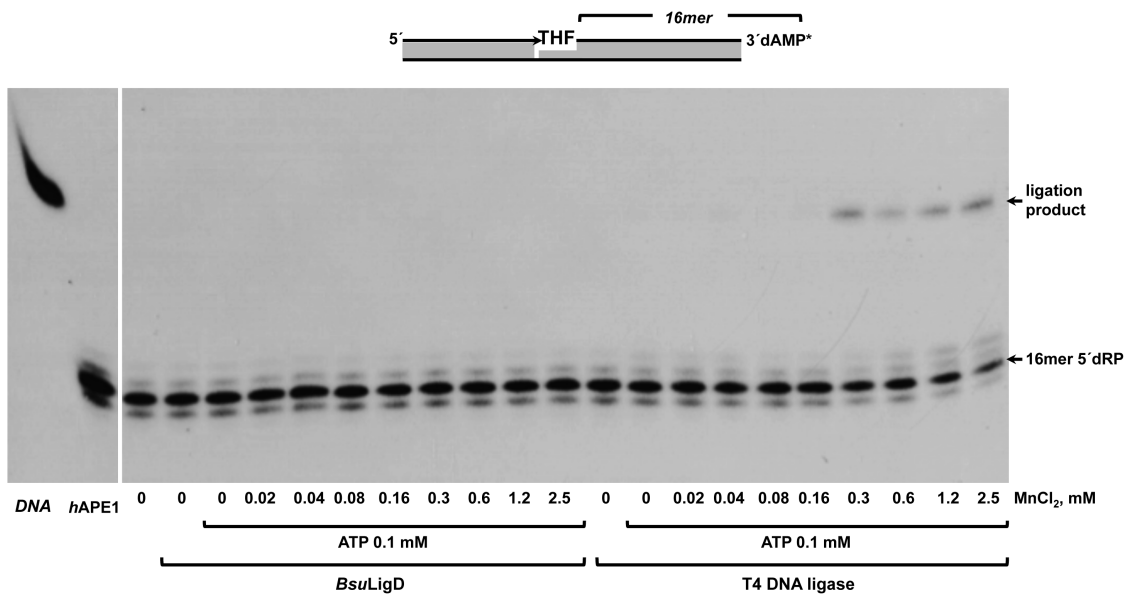
Supplementary Figure S1. Steady-state kinetic analysis of the 5'-dRP lyase activity of *BsuLigD*. The release of dRP from a 3' labeled 5'-dRP-containing DNA substrate was examined as a function of the indicated substrate concentration as described in Materials and Methods. Reactions contained 50 nM *BsuLigD* and were incubated at 30 °C for 20 min. Apparent activities were fitted to the Michaelis equation by least-squares nonlinear regression. The values plotted are the mean of three independent experiments.



Supplementary Figure S2. *BsuLigD* AP-lyase activity in the presence of EDTA. 0.83 nM of the [³²P]3'-labeled uracil-containing hybrid E depicted on top of the figure was treated with *E. coli* UDG, leaving an intact AP site (*DNA*). 57 nM *BsuLigD* was preincubated with increasing concentrations of EDTA (0, 12.5, 25, 50 and 100 mM). After overnight incubation at 4°C, both the EDTA-treated *BsuLigD* and 57 nM of fresh protein were incubated with the DNA substrate during 30 min or 30°C, as described in Materials and Methods. After incubation samples were analyzed by 8 M urea-20% PAGE and autoradiography. Position of products is indicated. *Alk*, alkaline hydrolysis of the AP site.

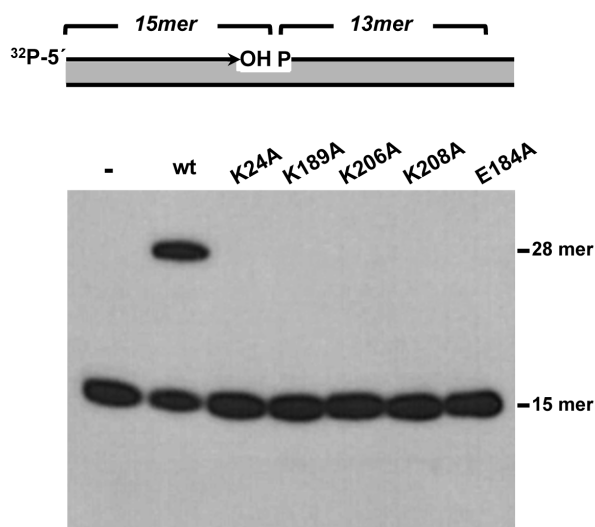


Supplementary Figure S3. Formation of protein-DNA adducts. The assay was performed as described in Materials and Methods by incubating 4 nM of the [³²P]3'-labeled uracil-containing oligonucleotide (depicted on top of the figure) pretreated with *E. coli* UDG to leave an intact AP site and further incubated with the indicated protein (100 nM *BsuLigD* or *PaeLigD*, 147 nM *LigDom*) for the indicated times in the presence of 0.64 mM MnCl₂. After incubation samples were reduced by adding 100 mM NaBH₄ and further subjected to SDS-PAGE and autoradiography. The mobility of the different protein-DNA adducts is indicated.

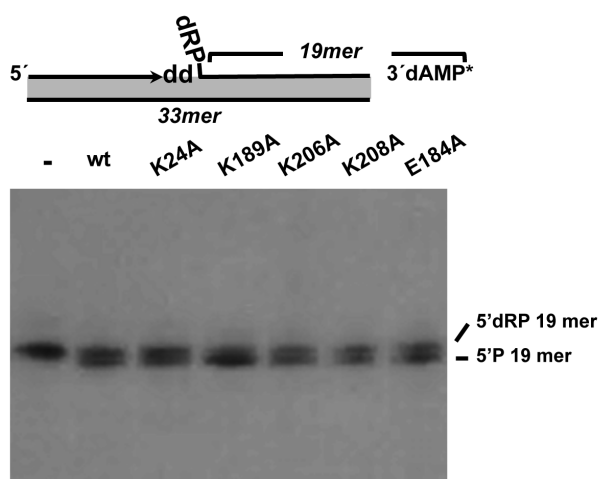


Supplementary Figure S4. *BsuLigD* does not seal 3'-OH and 5'-dRP ends. A 3'-end labeled (*) oligonucleotide containing a single synthetic AP (THF) site (lane *DNA*) was treated with *hAPE1* to insert a nick at the 5' side of the molecule (lane *hAPE1*). The assay was performed as described in Materials and Methods. Briefly, 0.53 nM of the nicked substrate was incubated with either 57 nM *BsuLigD* or 40 U of T4 DNA ligase with the indicated MnCl₂ concentrations and in the absence or presence of 0.1 mM ATP. After incubation for 30 min at 30°C, samples were analyzed by 8 M urea-20% PAGE and autoradiography. Position of products is indicated.

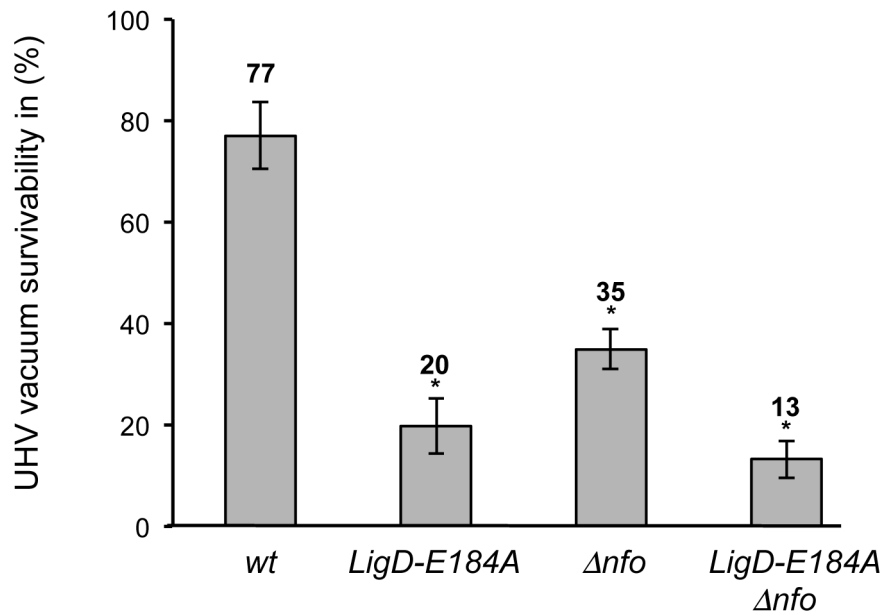
A Ligation activity



B 5'dRP lyase activity



Supplementary Figure S5. Analysis of dRP lyase and ligase activity of *BsuLigD* mutants K24A, K189A, K206A, K208A and E184A. (A) Ligase activity. The assay was performed by incubating 23 nM of the indicated protein with 1.2 nM of the nicked DNA hybrid F (depicted in top of the panel) in the presence of 5 μM MnCl_2 . After incubation for 30 min at 30°C, samples were analyzed by 8 M urea-20% PAGE and autoradiography. (B) dRP lyase activity. The assay was performed as described in Materials and Methods. Briefly, 50 nM of the DNA hybrid B was preincubated with *E. coli* UDG as described above, to render a 5'-dRP end (see scheme of the substrate on top of the panel). Reactions were initiated by adding 28 nM of the indicated protein in the presence of 0.64 mM MnCl_2 . After incubation for 20 min at 30 °C, reactions were stopped by addition of freshly prepared NaBH_4 . Samples were analyzed by 8 M urea-20% PAGE and autoradiography.



Supplementary Figure S6. Survival of *B. subtilis* spores expressing the *BsuLigD* ligase deficient mutant E184A and/or Nfo AP endonuclease. The assay was performed as described in Materials and Methods. The CFUs of UHV-treated spores were divided with the average CFU-value of untreated spore samples in order to obtain the survival after UHV. The data presented are expressed as average values \pm SD, $N=4$. Asterisks indicate UHV survival values that were significantly different (P -values ≤ 0.05) from values for wild-type (*wt*) spores.