

## Supplemental Legends

**FIG S1** Nucleotide sequence alignment of the *iglA-D* containing region of *F.n.n.* FSC769 and other selected *Francisella* sp.: *F.n.o.* LADL--07-285A (GI: 564747871), *F. philomiragia* ATCC 25017 (GI: 167596226), *F. novicida* U112 (GI: 754269614), *F. holarctica* LVS (GI: 754265763) and *F.t.t.* Schu S4 (GI: 754282044). Pink background indicates nucleotide differences and dots represent identical residues between strains. Genes are labeled with yellow arrows behind the nucleotide sequence pointing in the coding direction and the gene order is *iglA*, *iglB*, *iglC* and *iglD*. *F.n.o.* has a premature stop codon 21 bp before the end of *iglA* compared to the other *Francisella* sp. analyzed.

**FIG S2** Representative dotplots of *Dictyostelium* wt cells infected with *F.n.n.* wt and  $\Delta iglC$  (upper and middle panel) and *Dictyostelium*  $\Delta atg1$  cells infected with *F.n.n.* wt (lower panel) at 1, 4, 24 and 48 hpi. Displayed is the sideward scatter (SSC) plotted as a function of green fluorescence (FL-1). After enhancement of the endogenous *F.n.n.* GFP signal with an AlexaFluor488-coupled antibody we observe two separate populations of weakly (non-infected) and strongly fluorescent (infected) cells.

**FIG S3** Replication of *F.n.n.* in zebrafish is dependent on IgIC

(A) Site of microinjection (white circle) and observation area of bacterial growth using fluorescence microscopy (black rectangle). (B-G) Representative micrographs show the fluorescent signal of *F.n.n.* wt (B, C), *F.n.n.*  $\Delta iglC$  (D, E) and PBS as a negative control (F, G) at 6 dpi. Displayed are the green channel (GFP) and an overlay with the corresponding bright field image of the zebrafish tail region. (H) Bacterial

replication of *F.n.n.* wt (black), *F.n.n.*  $\Delta$ *ig/C* (white) and *F.n.n.*  $ig/C^+$  (grey) measured by qPCR at 3 and 7 dpi (n=3). In the *F.n.n.*  $\Delta$ *ig/C* infected group (6 dpi), a few of the embryos displayed a substantial amount of green fluorescent granulas in parts of the yolk sac. This could suggest that the *F.n.n.*  $\Delta$ *ig/C* mutant succeeds in some individuals, however selected individual responses of zebrafish embryos to bacterial infections were observed before (91). This different degree of success is reflected in the gDNA samples, where one  $\Delta$ *ig/C* sample (consisting of 3 embryos each) contained a much higher quantity of *F.n.n.* genomes than the others 7 dpi.

**FIG S4** Additional ultrastructural analysis of intracellular *F.n.n.* wt samples processed by chemical fixation and room temperature dehydration (A, B) or HPF and freeze substitution (C, D). (A, B) Ultrastructural analysis of phagosomal (A) and cytosolic (B) *F.n.n.* after chemical fixation at 2 and 4 hpi, respectively. Cytosolic as well as phagosomal bacteria are surrounded with the electron-translucent area, which indicates fixation artefacts. (C, D) Overview (C) and high magnification (D) showing formerly cytosolic *F.n.n.* wt at 1 hpi possibly contained within an autophagosome as indicated by the limiting autophagic-like membrane and the ribosomes in the lumen. Bacteria are surrounded with an electron pale area indicating the presence of weakly contrasted material, which was a characteristic appearance of cytosolic bacteria after HPF and freeze substitution. No binding of various lectins (wheat germ agglutinin, concanavalin A, ricinus agglutinin I, *Erythrina cristagalli* lectin, *Maackia amurensis* Lectin I or *Maclura pomifera* lectin) was observed indicating no enrichment in mannose, glucose, N-acetylglucosamine, galactose, N-acetylgalactoseamine or sialic acid components. Asterisks: *F.n.n.*, arrows: *F.n.n.*-enclosing membrane, white arrowheads: electron-translucent area characteristic of chemically fixed bacteria,

black arrowheads: electron-pale area characteristic for HPF. Scale bars: 500 nm (A, B, D), 2  $\mu$ m (C).

**Fig S5** Visualization of the rupture of the phagosomal membrane by electron and fluorescence microscopy. (A, B) Electron microscopy following HPF and freeze substitution reveals rupturing phagosomal membrane in *Dictyostelium* cells infected with *F.n.n.* wt at 1 hpi. Asterisks: *F.n.n.*, arrowheads: broken membrane. (C, D) Confocal fluorescence microscopy shows the broken p80-positive compartment (green) and escaping *F.n.n.* wt (red) at 4 hpi. The inlet displays the green channel (p80) of the region of interest (white square). Scale bars: 500nm (A, B), 5  $\mu$ m (C, D).

**FIG S6** Phagosomal maturation and ubiquitination in *Dictyostelium Δatg1* cells  
(A-C) Association rates of *F.n.n.* wt with p80 (A), VatA (B) and ubiquitin (C) in *Dictyostelium Δatg1* cells over 48 hpi. Data of 3 independent experiments (mean $\pm$ SEM) was collected for each association study and a minimum of 100 bacteria was counted at each timepoint. (D) Representative micrographs of accumulated, ubiquitinated (green) *F.n.n.* wt (red) in *Dictyostelium Δatg1* cells at 48 hpi. The margin of the cell is marked with a dashed line and the inlet shows only ubiquitin in the green channel. Scale bar: 5  $\mu$ m.

**Movie S1** Live cell microscopy of *Dictyostelium* Lifeact-RFP (red) infected with *F.n.n.* wt expressing GFP (green) at 19 hpi. The inlet displays the red channel (actin) of the region of interest (white square). Scale bar: 5  $\mu$ m.

**Movie S2** Live cell microscopy of *Dictyostelium* Lifeact-RFP (red) infected with *F.n.n.*  $\Delta ig/C$  expressing GFP (green) at 2 hpi. The inlet displays the red channel (actin) of the region of interest (white square). Scale bar: 5  $\mu\text{m}$ .

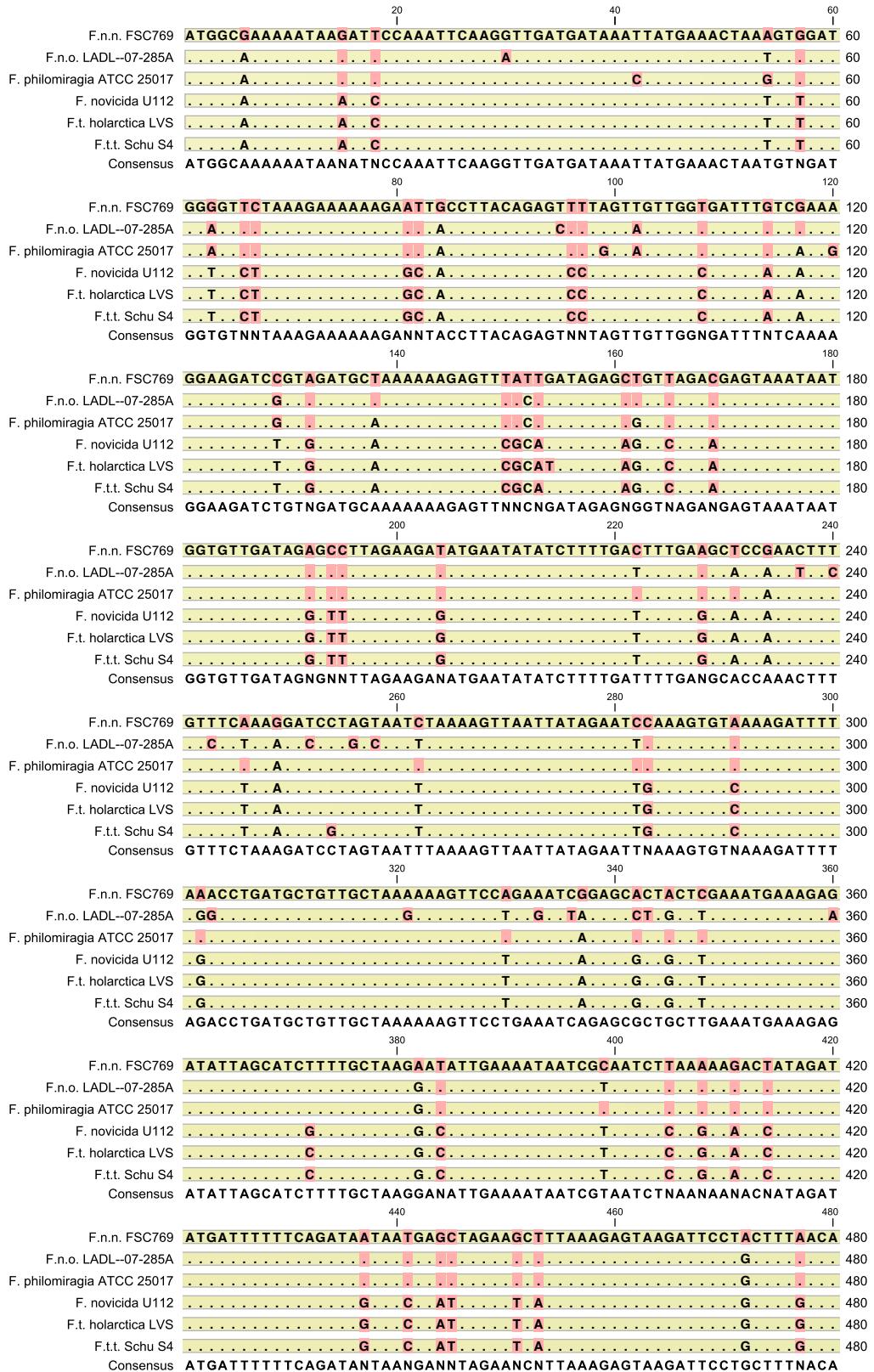
### **Cryofixation, chemical fixation and Transmission Electron Microscopy**

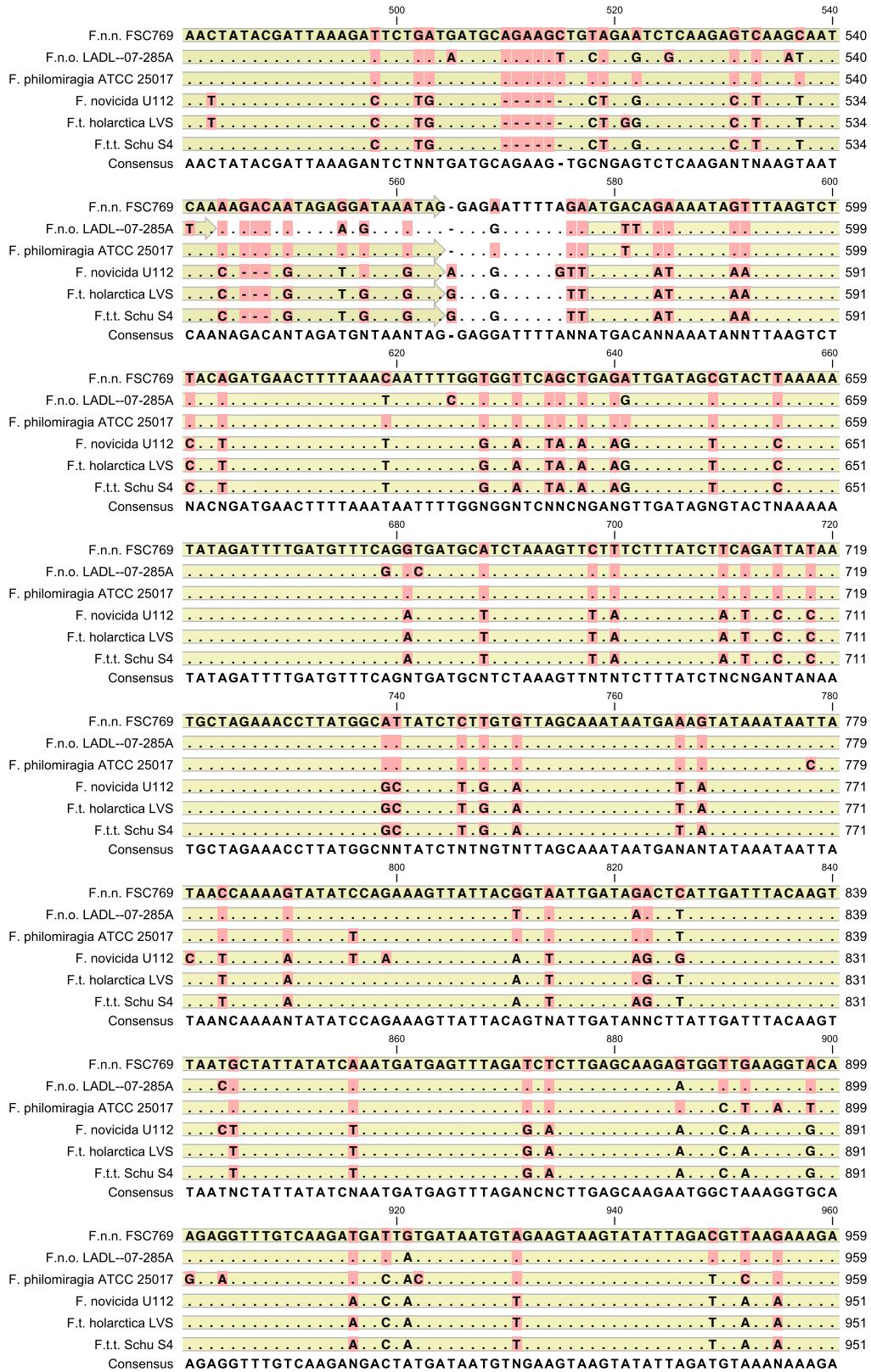
Cellulose capillary tubes (a kind gift by Heinz Schwarz) with an inner diameter of 200 µm were filled with a concentrated suspension of infected cells by capillary action (Hohenberg *et al.*, 1994), cut into short segments and transferred into aluminium planchettes (with a 6 mm diameter and a 150 µm recess), filled with 1-hexadecene (Merck Millipore, Darmstadt, Germany). Samples were cryofixed in an EM HPM100 high-pressure freezing machine (Leica Microsystems, Vienna, Austria) (Kaech *et al.*, 2014). Freeze substitution (Balzers FSU 010; Bal-Tec, Balzers, Liechtenstein) was performed as described (Leonidova *et al.*, 2014) in acetone containing 1% osmium tetroxide for 8 h at -90°C, 6 h at -60°C and 4 h at -40°C. Afterwards samples were incubated for 1 h on ice, washed with acetone and incubated with 1% uranyl acetate in acetone for 1 h on ice. After washing samples were then progressively infiltrated with epon resin (Fluka, St. Louis, Missouri, USA) (15, 33, 50, 80, 100%) over two days at room temperature or they were first incubated with 0.5% glutaraldehyde (GA) for 30 min at 4°C, washed and then infiltrated with LR White resin (Electron Microscopic Sciences, Hatfield, PA, USA) (50, 100%) over 10 hours. Eventually samples were polymerized overnight at 70°C. Around 70 nm sections were cut using an ultra 45° diamond knife (Diatome, Biel, Switzerland). Sections were examined without additional contrasting using a CM100 transmission electron microscope (Philips, Eindhoven, Netherlands). The images were recorded digitally with a Quemesa TEM CCD camera and iTEM software v 5.1 (both Olympus Soft Imaging Solutions, Münster, Germany).

The embedding of chemically fixed samples was performed by the EM facility at the department of Biosciences, University of Oslo. Infected, attached cells were fixed in a solution of 2% GA and 1% PFA in HL5c for 2 h at RT and washed with sterile PBS. Subsequently, 2% GA/sodium cacodylate (0.1 M) was added before the fixed cells

were scraped off the well bottom with a cell scraper and stored at 4°C. The cell pellets were subsequently washed 2 x 10 min with 0.1 M sodium cacodylate buffer, then incubated for 1 h in darkness with the aqueous solution of 2% OsO<sub>4</sub> and 1.5% K<sub>3</sub>Fe(CN)<sub>6</sub>, washed 2 x 10 min in dH<sub>2</sub>O and stained with 1.5% uranylacetate for 30 min. Samples were dehydrated in 70, 80, 90, 96% EtOH for 10 min in each concentration before 4 x 15 min in absolute EtOH. The pellets were resuspended in 1:1 epon/propylene oxide and rotated O/N with open lids before Epon polymerization at 60°C for 3 days. Sections were prepared and examined as described above for cryofixed samples.

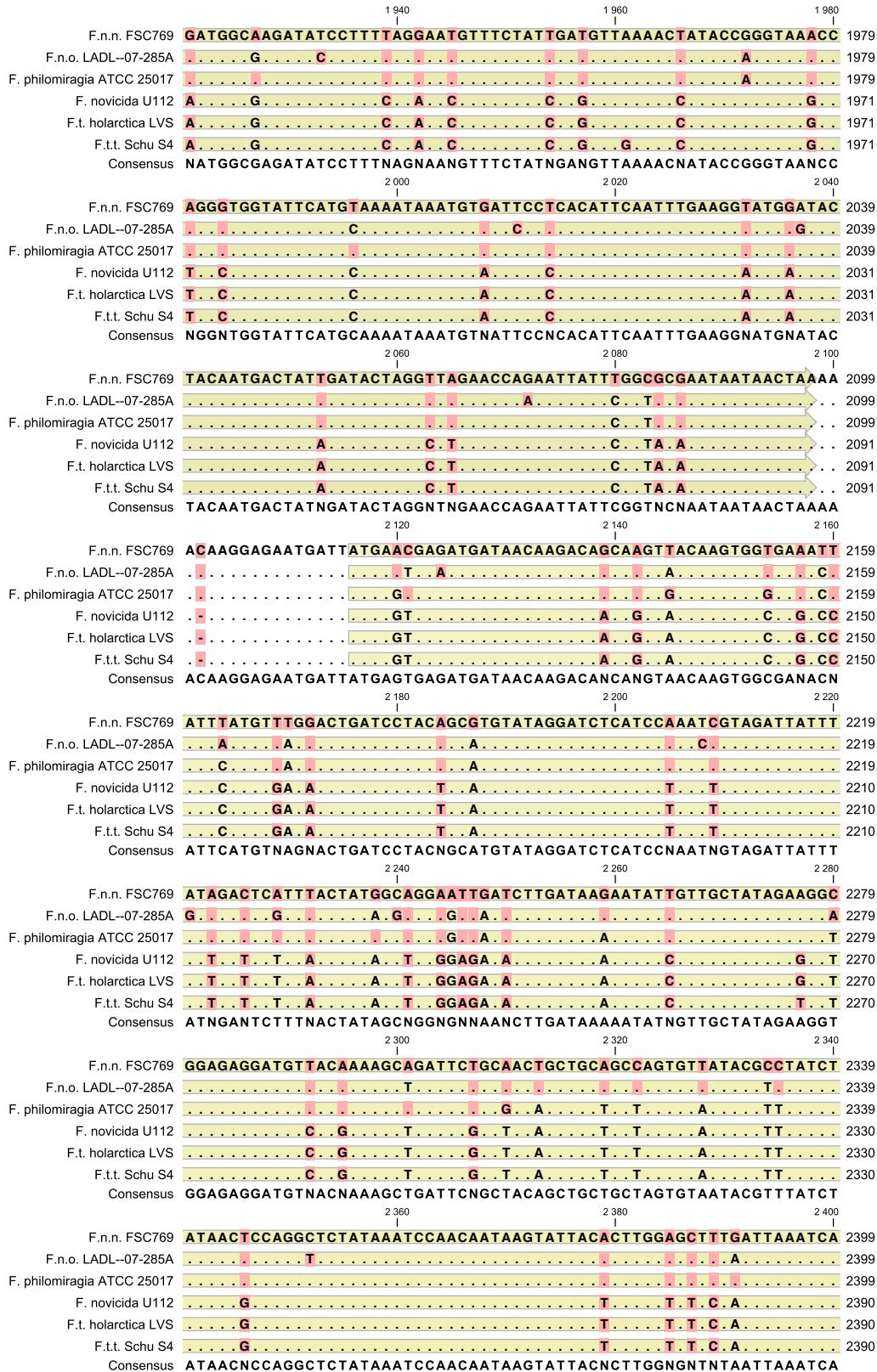
- Hohenberg, H., Mannweiler, K. and Muller, M. (1994). High-pressure freezing of cell suspensions in cellulose capillary tubes. *Journal of microscopy* **175**, 34-43.
- Kaech, A. and Ziegler, U. (2014). High-pressure freezing: current state and future prospects. *Methods in molecular biology* **1117**, 151-171.
- Leonidova, A., Pierroz, V., Rubbiani, R., Lan, Y.J., Schmitz, A.G., Kaech, A., et al. (2014). Photo-induced uncaging of a specific Re(I) organometallic complex in living cells. *Chem Sci* **5**, 4044-4056.





F.n.n. FSC769 **AGAATTACAGTATGACTTGGAGAGAAATCTATATGACATATCTAGTAGCGATTCTTAA** 1019  
 F.n.o. LADL-07-285A ...**G**... 1019  
 F. philomiragia ATCC 25017 ...**T**... 1019  
 F. novicida U112 ...**GC**...**A**...**T**...**C**...**T**...**T**...**T**...**C**...**T**...**C**... 1011  
 F.t. holarctica LVS ...**GC**...**A**...**T**...**C**...**T**...**T**...**T**...**C**...**T**...**C**... 1011  
 F.t. Schu S4 ...**GC**...**A**...**T**...**C**...**T**...**T**...**T**...**C**...**T**...**C**... 1011  
 Consensus **AGAGNTACANTATGATTNGAGAGAAATNTATATGANATATCTAGTAGNGACTTTNAA**  
 980 1 000 1 020  
 F.n.n. FSC769 **GAAAGTGTATGTTGCAGAATTGATCAATATGGTGGTAACCATATGGCGCAATACTCGG** 1079  
 F.n.o. LADL-07-285A ...**A**...**CT**... 1079  
 F. philomiragia ATCC 25017 ...**T**...**G**... 1079  
 F. novicida U112 **A**...**A**...**T**...**C**...**T**...**T**...**A**... 1071  
 F.t. holarctica LVS **A**...**T**...**C**...**T**...**A**...**T**...**T**...**A**... 1071  
 F.t. Schu S4 **A**...**T**...**C**...**T**...**C**...**T**...**T**...**A**... 1071  
 Consensus **NAAAGTNTATGTTTCAGAATTGATCAATATGGTGGTAACCTATGGCGCAATANTAGG**  
 1 040 1 060 1 080  
 F.n.n. FSC769 **CTTGATAATTGAAAACACTACAAATGATATAATTGGTGGACAGGGATGGGCATGGT** 1139  
 F.n.o. LADL-07-285A **AC**...**C**...**G**...**G**... 1139  
 F. philomiragia ATCC 25017 ...**T**... 1139  
 F. novicida U112 **A**...**T**...**C**...**T**...**T**...**A**...**T**... 1131  
 F.t. holarctica LVS **A**...**T**...**C**...**T**...**T**...**A**...**T**... 1131  
 F.t. Schu S4 **A**...**T**...**C**...**T**...**T**...**A**...**T**... 1131  
 Consensus **ATTGATAATTGAAAANACNACAAATGATATAATTGGTGGACTGGNATGGGNATGGT**  
 1 100 1 120 1 140  
 F.n.n. FSC769 **GGCAAAGAATTCTCATGCCATTAGCATCAATAGACAAGTCATTGGTGTAA** 1199  
 F.n.o. LADL-07-285A ...**A**...**C**...**AT**...**T**... 1199  
 F. philomiragia ATCC 25017 ...**T**... 1199  
 F. novicida U112 ...**A**...**T**...**T**...**T**...**C**... 1191  
 F.t. holarctica LVS ...**A**...**T**...**T**...**T**...**C**... 1191  
 F.t. Schu S4 ...**A**...**T**...**T**...**T**...**C**... 1191  
 Consensus **GGCAAAGAATTCTCATGCCATTAGCATCAATTGANAAGTCATTGGTGTAA**  
 1 160 1 180 1 200  
 F.n.n. FSC769 **AGATCTATCAGAAATACTCACATAAGAGTTTGATCCTTGTAGAACATCCTAGATA** 1259  
 F.n.o. LADL-07-285A ...**T**...**T**...**T**...**C**...**G**...**T**...**A**...**C**... 1259  
 F. philomiragia ATCC 25017 ...**T**... 1259  
 F. novicida U112 **G**...**T**...**C**...**T**...**A**...**G**...**T**...**C**...**T**...**G**... 1251  
 F.t. holarctica LVS **G**...**T**...**C**...**T**...**A**...**G**...**T**...**C**...**T**...**G**... 1251  
 F.t. Schu S4 **G**...**T**...**C**...**T**...**A**...**G**...**T**...**C**...**T**...**G**... 1251  
 Consensus **NGATTATCAGAAATNACTATATAANAGTTGAANCNTTGTGANCATCCTAGATA**  
 1 220 1 240 1 260  
 F.n.n. FSC769 **TAAAGAGTGGAAATGACTTGGAAACCTAGATGTAGCTGCATATATCGGTTGACTATCGG** 1319  
 F.n.o. LADL-07-285A ...**A**...**T**...**G**...**T**...**T**...**C**... 1319  
 F. philomiragia ATCC 25017 ...**T**...**G**...**T**...**T**...**T**...**C**... 1319  
 F. novicida U112 ...**T**...**T**...**T**...**T**...**A**...**C**...**G**...**A**... 1311  
 F.t. holarctica LVS ...**T**...**T**...**T**...**T**...**A**...**C**...**G**...**A**... 1311  
 F.t. Schu S4 ...**T**...**T**...**T**...**T**...**A**...**C**...**G**...**A**... 1311  
 Consensus **TAAAGAGTGGAAATGATTAGAAACCTTGTAGTGTGCTGCATATATNGTTGACNGTNGG**  
 1 280 1 300 1 320  
 F.n.n. FSC769 **AGATTTATGTTACGTCAGCCATATAATCCTGAGAATAACCCAGTCAGTATAAGCTTAT** 1379  
 F.n.o. LADL-07-285A ...**A**...**T**...**A**...**C**... 1379  
 F. philomiragia ATCC 25017 ...**A**...**T**...**A**...**C**... 1379  
 F. novicida U112 **T**...**G**...**G**...**A**...**T**...**A**...**A**... 1371  
 F.t. holarctica LVS **T**...**G**...**G**...**A**...**T**...**A**...**A**... 1371  
 F.t. Schu S4 **T**...**G**...**G**...**A**...**T**...**A**...**A**... 1371  
 Consensus **NGATTATGTTNCNCATATAATCCTGAGAATAATCCAGTCAGTATAANCTTAT**  
 1 340 1 360 1 380  
 F.n.n. FSC769 **GGAAGGCTTTAACGAGTTGGATCATAGAATAATGATAGCTACTATGGGGACCTC** 1439  
 F.n.o. LADL-07-285A ...**T**...**A**...**T**...**T**...**G**...**G**...**T**...**T**...**G**...**C**...**T**... 1439  
 F. philomiragia ATCC 25017 ...**C**...**T**...**A**...**T**...**T**...**G**...**G**...**T**...**T**...**G**...**G**...**T**... 1439  
 F. novicida U112 ...**T**...**T**...**T**...**G**...**T**...**G**...**A**...**T**...**T**...**C**...**T**...**G**... 1431  
 F.t. holarctica LVS ...**T**...**T**...**T**...**G**...**T**...**G**...**A**...**T**...**T**...**C**...**T**...**G**... 1431  
 F.t. Schu S4 ...**T**...**T**...**T**...**G**...**T**...**G**...**A**...**T**...**T**...**C**...**T**...**G**... 1431  
 Consensus **GGAAGGCTTTATGAGTTGATTATGANAANAATGANAGTTATNTATGGGACCTNC**

F.n.n. FSC769 **ATCTATTCAAGTTAGTTAAAATATGGTGAGATCATATGATAAAACTAGATGGTTCAATA** 1499  
 F.n.o. LADL-07-285A ...C.....A.....G..... 1499  
 F. philomiragia ATCC 25017 ...A.....A..... 1499  
 F. novicida U112 T...A.....TC.....G.....A.....T.....C..... 1491  
 F.t. holarktica LVS T...A.....TC.....G.....A.....T.....C..... 1491  
 F.t. Schu S4 T...A.....TC.....G.....A.....T.....C..... 1491  
 Consensus NTCNATTCACTAGTTAANAAATGATGAGATCTTATGATAAAACTAGATGGTTCAATA  
 1460 1480 1500  
 F.n.n. FSC769 **CATAAGAGGTGTTGAAAGTGGTTATGTAAGAAACCTAGTGTCAATGTTATGATAA** 1559  
 F.n.o. LADL-07-285A ..... 1559  
 F. philomiragia ATCC 25017 T.....T.....A.....G..... 1559  
 F. novicida U112 T.....A.....G.....A.....G.....AG.T.C.A..... 1551  
 F.t. holarktica LVS T.....A.....G.....AG.....G.....AG.T.C.A..... 1551  
 F.t. Schu S4 T.....A.....G.....AG.....G.....AG.T.C.A..... 1551  
 Consensus TATAAGAGGNNTGAGTGGTTATGTAANAAACCTNGTANCTTGNNTATGATAA  
 1520 1540 1560  
 F.n.n. FSC769 **TAAAGGTGTCTTAGAACAAAGCCGCCACTGAATGCTTTATTTGCAGATTATATGGAGCT** 1619  
 F.n.o. LADL-07-285A ..... 1619  
 F. philomiragia ATCC 25017 ..... 1619  
 F. novicida U112 .....CA.TC.....T.....T.A.....T.....T.....T..... 1611  
 F.t. holarktica LVS .....CA.TC.....T.....T.A.....TT.A.....T.....T.....T..... 1611  
 F.t. Schu S4 .....CA.TC.....T.....T.A.....TT.A.....T.....C.T.....T..... 1611  
 Consensus TAAAGGNNTNTAGAACNAAGTCNNCTAAATGTNTTATTCNGNGATTATATGGAGNT  
 1580 1600 1620  
 F.n.n. FSC769 **ATCACTTACAATATTGGTTAATACCATTGTAAGTGAAAAGGTACAAGTAATGCTTG** 1679  
 F.n.o. LADL-07-285A .....G..... 1679  
 F. philomiragia ATCC 25017 .....G..... 1679  
 F. novicida U112 .....G..... 1671  
 F.t. holarktica LVS .....G..... 1671  
 F.t. Schu S4 .....G..... 1671  
 Consensus ATCACTTGCAATATTGGTTAATACCATTGTAAGTGAAAAGGTACAAGTAATGCTTG  
 1640 1660 1680  
 F.n.n. FSC769 **TTTTTTTAGTGTGAACTCAGCTAAGAAAGTTGAAGAGTTTAGATAGTTTGATTCTGC** 1739  
 F.n.o. LADL-07-285A ...C..... 1739  
 F. philomiragia ATCC 25017 ...C..... 1739  
 F. novicida U112 ...C.....A.T.T.....A.....C.....A.....G.A.....C.A..... 1731  
 F.t. holarktica LVS ...C.....A.T.T.....A.....C.....A.....G.A.....C.A..... 1731  
 F.t. Schu S4 ...C.....A.T.T.....A.....C.....A.....G.A.....C.A..... 1731  
 Consensus TTTCTTTAGTGTNAANTCNGCTAANAAAGTCGAAGANTTTGTAGATNGNTTGANCN  
 1700 1720 1740  
 F.n.n. FSC769 **TAATTCAATGTTGATTGTAATCTTCTTAACTATGTTATCCAGAATTTCACATTA** 1799  
 F.n.o. LADL-07-285A ...C.....C.A..... 1799  
 F. philomiragia ATCC 25017 ...C..... 1799  
 F. novicida U112 A...C.....GA.....A.....C.....C.....G.....A.....T..... 1791  
 F.t. holarktica LVS A...C.....GA.....A.....C.....C.....G.....A.....T..... 1791  
 F.t. Schu S4 A...C.....GA.....A.....C.....C.....G.....A.....T..... 1791  
 Consensus NAACTCANNTTAATTGCTAANCTTCTTANACTATGTATATCNGAATNTCN  
 1760 1780 1800  
 F.n.n. FSC769 **TATTAAGTGTATTCAAGAGATAAGAGTTGAGCTGAGCAAATTCAAAA** 1859  
 F.n.o. LADL-07-285A .....A..... 1859  
 F. philomiragia ATCC 25017 ..... 1859  
 F. novicida U112 .....A.....A.T.....T.T.....A.....G.A.....TC.....TCG..... 1851  
 F.t. holarktica LVS .....A.....A.T.....T.T.....A.....G.A.....TC.....TCG..... 1851  
 F.t. Schu S4 .....A.....A.T.....T.T.....A.....G.A.....TC.....TCG..... 1851  
 Consensus TATTAANTGTGTNATNAGAGATAAGATNGGNAGNTTGTGGNTNNNGAGNNNATT  
 1880 1900 1920  
 F.n.n. FSC769 **TATTCTTCAGATTGGATATCAGAGTTGTTACTACTGTATATCAACCAACGCCCTT** 1919  
 F.n.o. LADL-07-285A .....G.....A.C..... 1919  
 F. philomiragia ATCC 25017 .....C..... 1919  
 F. novicida U112 A.....T.....A.....C.....C.....A.....C.....C..... 1911  
 F.t. holarktica LVS A.....T.....A.....C.....C.....A.....C.....C..... 1911  
 F.t. Schu S4 A.....T.....A.....C.....C.....C.....A.....C..... 1911  
 Consensus NATTCTTCAGATTGGATATCAGANNTTGTACNACAGTNTATCAACCAACCC  
 CTTAGA



	2 420	2 440	2 460
F.n.n. FSC769	<b>AATACTAGGACTCTACTTGAAAGTGCTGTCAAGTATATTACAAGCAGGGCACTGAT</b>		2459
F.n.o. LADL-07-285A	.G.GT.....CT.G.....G.A.....T.T.A.....G.T..A..		2459
F. philomiragia ATCC 25017	.....G.....G.....G.....G.....		2459
F. novicida U112	...GTC..A...AAA...GAGAAA..T..G..C..A.T..T..A..		2450
F.t. holarktica LVS	...GT..A...AAA...GAGAAA..T..G.....A.T..T..A..		2450
F.t. Schu S4	...GT..A...AAA...GAGAAA..T..G.....A.T..T..A..		2450
Consensus	<b>AATGTTAGNACTANANTGGAAGNGNNNGNTTCGAGTATATTACAAGCANGTGCNACAGT</b>		
	2 480	2 500	2 520
F.n.n. FSC769	<b>ATGAAAATAAAATTGGGTAACTCGAACAAAAACAAAGAGTACAAAACAGATGATGCTTGG</b>		2519
F.n.o. LADL-07-285A	.....T.GC.T.G..T..T..G.....T.....T..A..		2519
F. philomiragia ATCC 25017	.....T.....T.....T.....T.....T.....A..A..		2519
F. novicida U112	...T.G..A...T..T..T.....T.....T..A..A..		2510
F.t. holarktica LVS	...T.G..A...T..T..T.....T.....T..A..A..		2510
F.t. Schu S4	...T.G..A...T..T..T.....T.....T..A..A..		2510
Consensus	<b>ATGAAAATTAAGTTAGGTAANTCTAATAAAAAACAAAGAGTANAACACTGATGAAGCNTGG</b>		
	2 540	2 560	2 580
F.n.n. FSC769	<b>GCGATTATGATTGATATCTAATTTAGAATTATATCCTATTAGTCAGAACGATTTAGT</b>		2579
F.n.o. LADL-07-285A	...T.C.....C.....G.....C.....T.T.C..		2579
F. philomiragia ATCC 25017	.....T.....T.....T.....T.....T.....T.....		2579
F. novicida U112	...T.....A..C.....G.....A..A.....T.A.G..T..		2570
F.t. holarktica LVS	...T.....A..C.....G.....A..A.....T.A.G..T..		2570
F.t. Schu S4	...T.....A..C.....G.....A..A.....T.A.G..T..		2570
Consensus	<b>GGTATTATGATNGATNTCTAATTTAGAGTTATATCCNATAAGTGCNTAGGCNTTTAGT</b>		
	2 600	2 620	2 640
F.n.n. FSC769	<b>ATAAAAATAGAACCAACTGAACTTATGGGGTTGCGAAAGATGGTATGAGATATCATGTA</b>		2639
F.n.o. LADL-07-285A	.....A..G.....C..AT..T..G.....A..		2639
F. philomiragia ATCC 25017	.....A.....A.....A.....A.....A.....A..		2639
F. novicida U112	...T.GT..G..A.....T..T..A.....A..T..A..T..		2630
F.t. holarktica LVS	...T.GT..G..A.....T..T..A.....A..T..A..T..		2630
F.t. Schu S4	...T.GT..G..A.....T..T..A.....A..T..C..A..T..		2630
Consensus	<b>ATNANNATAGANCCAACAGAACTTATGGGTGTTCAAAAGATGGNATGAGATATCATATN</b>		
	2 660	2 680	2 700
F.n.n. FSC769	<b>GTTATCTATTGATGGACTTACAACCTTCACAAGGTAGCTTGCCTGTATGCTGTGCTGCAAGT</b>		2699
F.n.o. LADL-07-285A	.....A..G..C.....A.....A.....G..		2699
F. philomiragia ATCC 25017	.....A.....C.....A.....A.....G.....A..		2699
F. novicida U112	.....A.....A..T..A..T..A.....A..T..C..A..T..C..		2690
F.t. holarktica LVS	.....A.....A..T..A..T..A.....A..T..C..A..T..C..		2690
F.t. Schu S4	.....A.....A..T..A..T..A.....A..T..C..A..T..C..		2690
Consensus	<b>ATATCTATNGATGGTCTTACAACNTCNCAAGNAGCTTGCCNGTATGNTGNGCNGCAGN</b>		
	2 720	2 740	2 760
F.n.n. FSC769	<b>ACTGATAAAGGAGTTGCTAGAATAGGGTACATAGCAGCTGTATAGTAA-ATAGGAGTTA</b>		2758
F.n.o. LADL-07-285A	.....A.....A.....T..C.....A.....A..		2758
F. philomiragia ATCC 25017	.....A.....A.....A.....C.....A.....A..		2758
F. novicida U112	.....A.....A..A..T..T..C.....G..C.....G..		2749
F.t. holarktica LVS	.....A.....A..A..T..T..C.....G..C.....G..		2749
F.t. Schu S4	.....A.....A..A..T..T..C.....G..C.....G..		2749
Consensus	<b>ACNGATAAAGGAGTTGCTAAAATAGGNATANNGCAGCTGCATAGTAAGATNGGAGTTN</b>		
	2 780	2 800	2 820
F.n.n. FSC769	<b>ATTATTAAAATGTTCTCGAAAGGATATGTTGGAAAGATGGCTAAGATTAGACAAAGT</b>		2818
F.n.o. LADL-07-285A	.....A.....A.....A.....A.....A..		2818
F. philomiragia ATCC 25017	.....A.....A.....A.....A.....A..		2818
F. novicida U112	...-..CT..A..T..A.....T.....T..T.GC..A..		2806
F.t. holarktica LVS	...-..CT..A..T..A.....T.....T..T.GC..A..		2806
F.t. Schu S4	...-..CT..A..T..A.....T.....T..T.GC..A..		2806
Consensus	<b>ATTATTNAATGTTCTNGAAAGGATNTATGGGAAGATGGNTAAGATTAGANAGNGAT</b>		
	2 840	2 860	2 880
F.n.n. FSC769	<b>ATCTTAGACAAATCAAATTGTCTATTTAGAAAGGTCAAAGTCTGCAAATTATTTGCCA</b>		2878
F.n.o. LADL-07-285A	.....T..TC..G.....A.....G..		2878
F. philomiragia ATCC 25017	.....T..TC..G.....A.....A..		2878
F. novicida U112	.....T..T..G..C..A..G..T..G..C..A..G..		2866
F.t. holarktica LVS	.....T..T..G..C..A..G..T..G..C..A..G..G..G..		2866
F.t. Schu S4	.....T..T..G..C..A..G..T..G..C..A..G..G..		2866
Consensus	<b>ATTTAGATAANTCAAANTNTCTNTTAAAGGTNAANNNCNGCAAGNTATTTGCCA</b>		

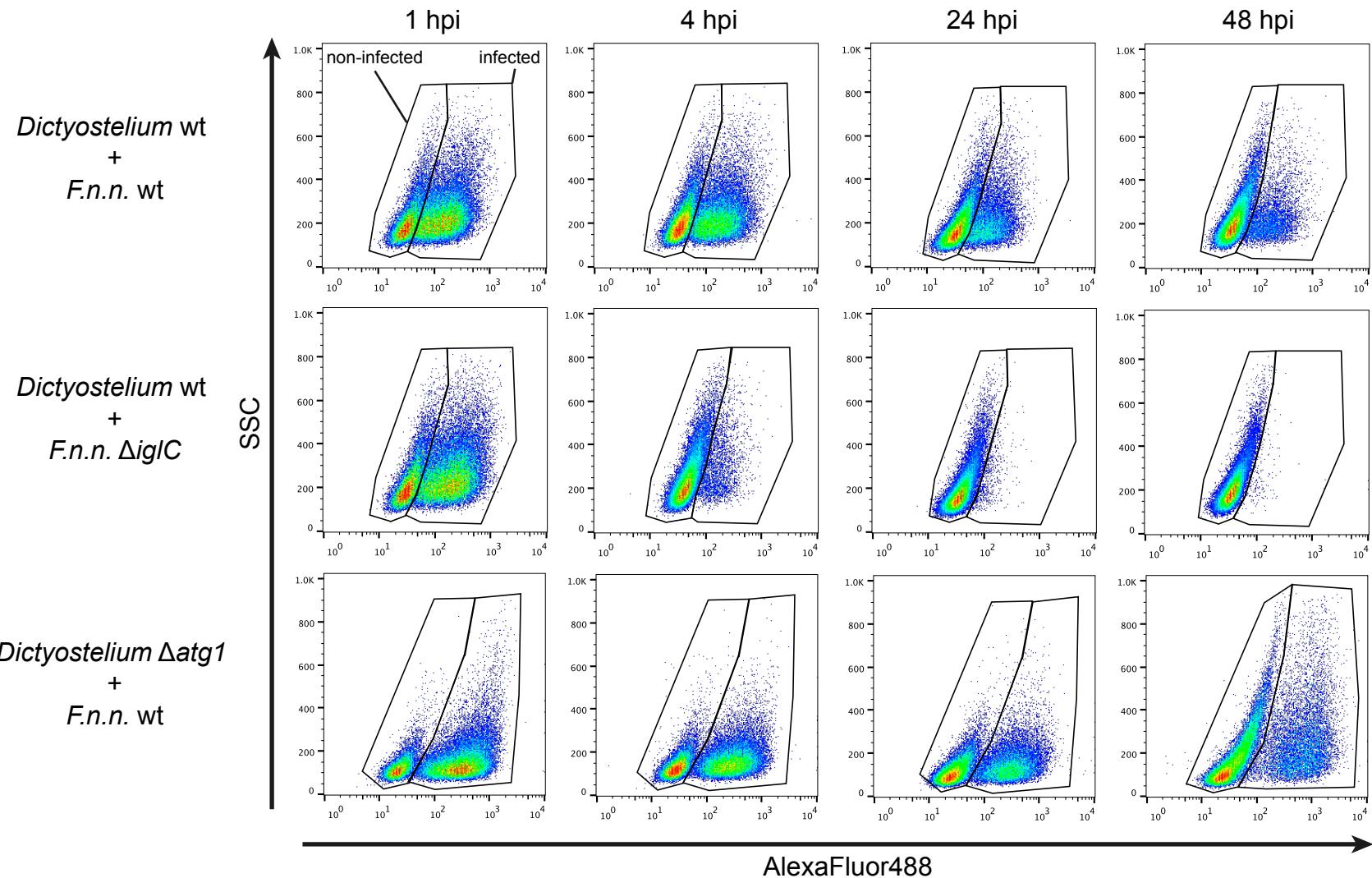
F.n.n. FSC769 **GCTAATCTTAATAAAAGGTATCGTTAGTTCGATTAGATATGGAGAGTTGCAGACGGGA** 2938  
 F.n.o. LADL-07-285A .A.....TC.....A.....T..... 2938  
 F. philomiragia ATCC 25017 .....G.A.....C.T.....G.T.A.....A.T 2926  
 F. novicida U112 .....G.A.....C.T.....G.T.A.....A.T 2926  
 F.t. holarctica LVS .....G.A.....C.T.....G.T.A.....A.T 2926  
 F.t. Schu S4 .....G.A.....C.T.....G.T.A.....A.T 2926  
 Consensus **GCTAATCTTAATAANGGNATCGTTAGNTTNGATTTAGATNTTGANAGTTGCAGACAGGN**  
 2 900 2 920 2 940  
 F.n.n. FSC769 **CTTATTCTAATAAAAGACTTGTATTGACTTAGATGAAAAAACATTTATTTTTATGAT** 2998  
 F.n.o. LADL-07-285A .....A.....A.....A..... 2998  
 F. philomiragia ATCC 25017 .....G.....A..... 2998  
 F. novicida U112 .....C.T.....AA.....AT.G..... 2986  
 F.t. holarctica LVS .....C.T.....AA.....AT.G..... 2986  
 F.t. Schu S4 .....C.T.....AA.....AT.G..... 2986  
 Consensus **CTTATNCTNATAAAAGACTTANATTGACTTAGATGAAAAAANNTNTTTTTATGAT**  
 2 960 2 980 3 000  
 F.n.n. FSC769 **AAATGCTATCCTCTGTCTTGCAGGTAATGACAGATGAGTTAACATATGATATCCCCTCA** 3058  
 F.n.o. LADL-07-285A .....A.....A.G..... 3058  
 F. philomiragia ATCC 25017 .....A..... 3058  
 F. novicida U112 .....G.CT.....GT.A.....A.AA.....T.A.....G.G.....A.A.CT..... 3046  
 F.t. holarctica LVS .....G.CT.....GT.A.....A.AA.....T.A.....G.G.....A.CT..... 3046  
 F.t. Schu S4 .....G.CT.....GT.A.....A.AA.....T.A.....G.G.....A.CT..... 3046  
 Consensus **AAANTNNATCCNNATCTTNCANATAATGACNGATNAGTTAANTNATGANATCCNNTA**  
 3 020 3 040 3 060  
 F.n.n. FSC769 **TTCTTGAATGTTAAGGAGAAGATAGTTGAAAAAGAAGGTGTTAAGTACATATATAATCAA** 3118  
 F.n.o. LADL-07-285A .....T..... 3118  
 F. philomiragia ATCC 25017 .....A..... 3118  
 F. novicida U112 .....TC.....A.C.GA.....AG.A.....AT.G.....A.T.C..... 3106  
 F.t. holarctica LVS .....TC.....A.C.GA.....AG.A.....AT.GA.....A.T.C..... 3106  
 F.t. Schu S4 .....TC.....A.C.GA.....AG.A.....AT.GA.....A.T.C..... 3106  
 Consensus **TTNNNTGAATNTNANAGAGAANNTANTTGAAAAANANGGNGTTAANTANATNTATAATCAA**  
 3 080 3 100 3 120  
 F.n.n. FSC769 **CTATCACTTCATTAGACTATGATTATAGTGTAAAGTATAGTACGCAAATTGCATTATTT** 3178  
 F.n.o. LADL-07-285A .....C..... 3178  
 F. philomiragia ATCC 25017 ..... 3178  
 F. novicida U112 .....T.G.....T.A.....C..AGC..G.T.....AC.....C.TC..... 3166  
 F.t. holarctica LVS .....T.G.....T.A.....C..AGC..G.T.....AC.....C.TC..... 3166  
 F.t. Schu S4 .....T.G.....T.A.....C..AGC..G.T.....AC.....C.TC..... 3166  
 Consensus **NTNTCANTNTCATTAGACCATNNNTATGNTNTAANNATAGNNANCAAATTGCATTATTT**  
 3 140 3 160 3 180  
 F.n.n. FSC769 **AAGTTAGATAGAGGCAAGTAGTATCAGATACATATGATTTCTTACTAACTCTTAAT** 3238  
 F.n.o. LADL-07-285A .....T..... 3238  
 F. philomiragia ATCC 25017 ..... 3238  
 F. novicida U112 .....G.C.....GCGA.....C.....A.TT.....C.....GC.T..... 3226  
 F.t. holarctica LVS .....G.....GCGA.....C.....A.TT.....C.C.GC.T..... 3226  
 F.t. Schu S4 .....G.....GCGA.....C.....A.TT.....C.C.GC.T..... 3226  
 Consensus **ANGTTAGATAGAGGGNNNTTAGTANCAGANATNTATGANNTCCNNNTANTAACTCTTAAT**  
 3 200 3 220 3 240  
 F.n.n. FSC769 **CACTATTTGATGCATGATACTTTATAAAGCTCAATAGAATGGTTCTGAATTAAAATCT** 3298  
 F.n.o. LADL-07-285A .....C.....T.....A..... 3298  
 F. philomiragia ATCC 25017 .....T..... 3298  
 F. novicida U112 .....T.....ATT.AGG.....T.....G.....A.T.....G.CT.....C.G 3286  
 F.t. holarctica LVS .....T.....ATT.AGG.....T.....G.....A.T.....G.CT.....C.G 3286  
 F.t. Schu S4 .....T.....ATT.AGG.....T.....G.....A.T.....G.CT.....C.G 3286  
 Consensus **CANTATTANNTNGNTGATTTNTAAANCTNAATAGNANTGTTCTGAANTAAANTCT**  
 3 260 3 280 3 300  
 F.n.n. FSC769 **TTCACATCGTTTGTCTCAACCTCAAGATCATATGCTGCAATTACTTGTATTCTTA** 3358  
 F.n.o. LADL-07-285A ..... 3358  
 F. philomiragia ATCC 25017 ..... 3358  
 F. novicida U112 .....T.....C.....T.....G.T.....T.....GT.....A.....T.G 3346  
 F.t. holarctica LVS .....T.....C.....T.....G.T.....T.....GT.....A.....T.G 3346  
 F.t. Schu S4 .....T.....C.....T.....G.T.....T.....GT.....A.....T.G 3346  
 Consensus **TTNAATCGNTTGTNTCAAGATCNTATGCNNCAATNTTACTTGTATTNTT**

		3 380	3 400	3 420
F.n.n. FSC769	<b>ATTAATAAATTAGAAAGAGAACTTAAATTGCTGAATCTAATAAGTTAAATAGTTCTCT</b>			3418
F.n.o. LADL-07-285A	.....	.....	.....	3418
F. philomiragia ATCC 25017	.....	.....	.....	3418
F. novicida U112	.....	T G G	G	3406
F.t. holartica LVS	.....	T G G C A	G GC	3406
F.t.t. Schu S4	.....	T G G C A	G GC	3406
Consensus	<b>ATTAATAAATTAGAAAGAGAACTTAAATTGCTGAATCTAATANGNNAAATAGTTCCNCCN</b>	3 440	3 460	3 480
F.n.n. FSC769	<b>AAGCAAATATTTGATCTAATTCTATGATATTATAGTTAACTCCAGCTTAACCTTAGATAAG</b>	I	I	3478
F.n.o. LADL-07-285A	.....	A.....	G.....	3478
F. philomiragia ATCC 25017	.....	A.....	G.....	3478
F. novicida U112	.....	A.....	T G C C T A C	3466
F.t. holartica LVS	.....	A.....	T G C C T A C	3466
F.t.t. Schu S4	.....	A.....	T G C C T A C	3466
Consensus	<b>AAACAAATATTTGATNTAATTNTGATATTANAGNTTAATNCNAACTTAACNTAGATAAN</b>			
		3 500	3 520	3 540
F.n.n. FSC769	<b>GTTGGGGATTTGATAATATTGAGTTGATTTTATAAACTCTAAAGGAAGATAAACTTAA</b>	I	I	I
F.n.o. LADL-07-285A	.....	A.....	G.....	3538
F. philomiragia ATCC 25017	.....	A.....	G.....	3538
F. novicida U112	.....	A A G C G C A C C A G C T G C T A T C	A.....	3526
F.t. holartica LVS	.....	A A G C G C A C C A G C T G C T A T C	A.....	3526
F.t.t. Schu S4	.....	A A G C G C A C C A G C T G C T A T C	A.....	3526
Consensus	<b>GTTGANGANTTGATAGNATTGANTTTGATTNNANAANCNTNACTAANNTAAANNTA</b>			
		3 560	3 580	3 600
F.n.n. FSC769	<b>TTGGCTGATAGATTATTGACTCTTGTGAAATAGAAAAATTAAACTTTATTAAATT</b>	I	I	I
F.n.o. LADL-07-285A	.....	C.....	C.....	3598
F. philomiragia ATCC 25017	.....	C.....	C.....	3598
F. novicida U112	.....	C T G A G G	G.....	3586
F.t. holartica LVS	.....	C T G A G G	G.....	3586
F.t.t. Schu S4	.....	C T G A G G	G.....	3586
Consensus	<b>NTNGCTGATAGATTATTNACTCTTGTGANTATAGAAANNTAAACTTTATTANATT</b>			
		3 620	3 640	3 660
F.n.n. FSC769	<b>GAAC TGCAAGGAAAAAAATTTATGTGAAAACCTTCCCCGAAGAGTTTTCGTTGCTACT</b>	I		3658
F.n.o. LADL-07-285A	.....	.....	.....	3658
F. philomiragia ATCC 25017	.....	.....	.....	3658
F. novicida U112	.....	T T A G G	T T T T	3646
F.t. holartica LVS	.....	T T A G G	T T T T	3646
F.t.t. Schu S4	.....	T T A G G	T T T T	3646
Consensus	<b>GAANTGCANGGAAAAAAATATNTATGTGAAANCTTNCCNGAACAGGTTTTNGTCTACT</b>			
		3 680	3 700	3 720
F.n.n. FSC769	<b>AGGTATTATCTTTCATCAAAAGAAAGGCAATAGCTCCAGCTAATGTAATTTGAAAAT</b>	I	I	I
F.n.o. LADL-07-285A	.....	A.....	A.....	3718
F. philomiragia ATCC 25017	.....	A.....	A.....	3718
F. novicida U112	.....	A C C T G A C	C.....	3706
F.t. holartica LVS	.....	A C C T G A C	C.....	3706
F.t.t. Schu S4	.....	A C C T G A C	C.....	3706
Consensus	<b>AGATANTATCTTTCNNTTAAAGNAANGAACAGCTCCAGCNAATGTAANNNTTGAAAAT</b>			
		3 740	3 760	3 780
F.n.n. FSC769	<b>AAAAAAGCTATGAGAAATAACAAGTATAAGTAGAAATAAAACGTTGTAACCTCTCTCT</b>			3778
F.n.o. LADL-07-285A	.....	.....	.....	3778
F. philomiragia ATCC 25017	.....	.....	.....	3778
F. novicida U112	.....	G T C T T	G TA T G	3766
F.t. holartica LVS	.....	G T C T T	G TA T G	3766
F.t.t. Schu S4	.....	G T C T T	G TA T G	3766
Consensus	<b>AANAANGCTNTGAGAATNACNAGTATAAGTAGAAATAANAANNTGTAACCTNTCTCT</b>			
		3 800	3 820	3 840
F.n.n. FSC769	<b>TCTGGAGTGAACACTGTTGAAGTTGAGTATTCTATGATGAATTTCACACGGGTTGAT</b>			3838
F.n.o. LADL-07-285A	.....	.....	.....	3838
F. philomiragia ATCC 25017	.....	.....	.....	3838
F. novicida U112	.....	A A G C A G	T A T	3826
F.t. holartica LVS	.....	A A G C A G	T A T	3826
F.t.t. Schu S4	.....	A A G C A G	T A T	3826
Consensus	<b>TCNGAGTAAACACTGGTTGANGTTGANTTCTATGATNAATTACACNNGNTTGT</b>			

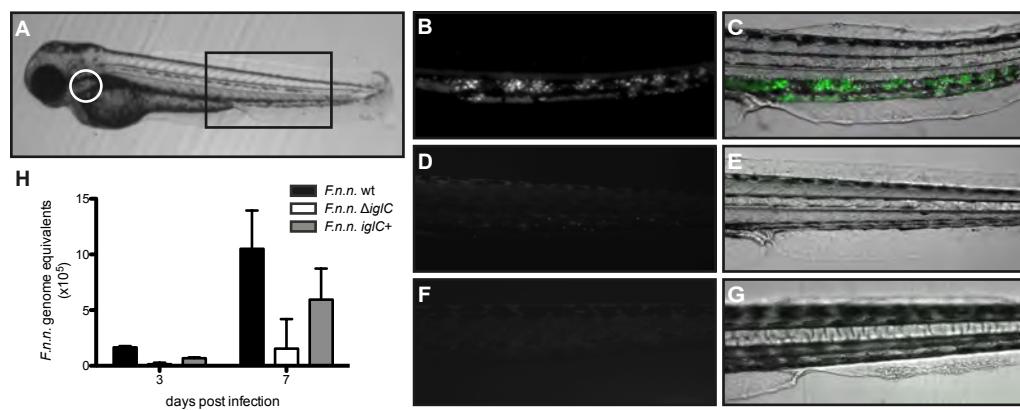
The figure displays sequence alignments for two sets of bacterial strains. The top set includes F.n.n. FSC769, F.n.o. LADL-07-285A, F. philomiragia ATCC 25017, F. novicida U112, F.t. holartica LVS, F.t.t. Schu S4, and a Consensus sequence. The bottom set includes F.n.n. FSC769, F.n.o. LADL-07-285A, F. philomiragia ATCC 25017, F. novicida U112, F.t. holartica LVS, F.t.t. Schu S4, and a Consensus sequence. Each strain's sequence is shown as a horizontal line with colored boxes indicating specific nucleotides at positions 3860, 3880, and 3900. The bottom set shows additional mutations at positions 3920, 3940, and 3960. A yellow arrow points to the F.n.n. FSC769 sequence in the bottom set.

Strain	Sequence	Position 3860	Position 3880	Position 3900	Position 3920	Position 3940	Position 3960	
F.n.n. FSC769	AATATTGATGCAATATATGAAATTCAAAAAGGCTCGGAATGGGACTTATACTTGCGGAT							3989
F.n.o. LADL-07-285A	.....						A	3989
F. philomiragia ATCC 25017	.....						A	3989
F. novicida U112	.....C.....	T	T	G	T	T	A	3886
F.t. holartica LVS	.....	T	T	G	T	T	A	3886
F.t.t. Schu S4	.....C.....	T	T	G	T	T	A	3886
Consensus	AATATTGATGCAATATATGAAATTCAAAAAGGNTCNGANTGGGANNTTATANTNGCGGAT							
F.n.n. FSC769	AGTAGTGC CGTTTACAGCCTTGAAGGTAGTGAAAACCTTGATTTCTTATAGCATT	I	I					3958
F.n.o. LADL-07-285A	.....T.....					G		3958
F. philomiragia ATCC 25017	.....T.....							3958
F. novicida U112	.....G.T.....			G	T		C	3946
F.t. holartica LVS	.....G.T.....			G	T		C	3946
F.t.t. Schu S4	.....G.T.....			G	T		C	3946
Consensus	AGTAGTGC CGTTTACNGCTTTGAAGGTAGTGANAANTTGATTTCTTATAGCNTT							

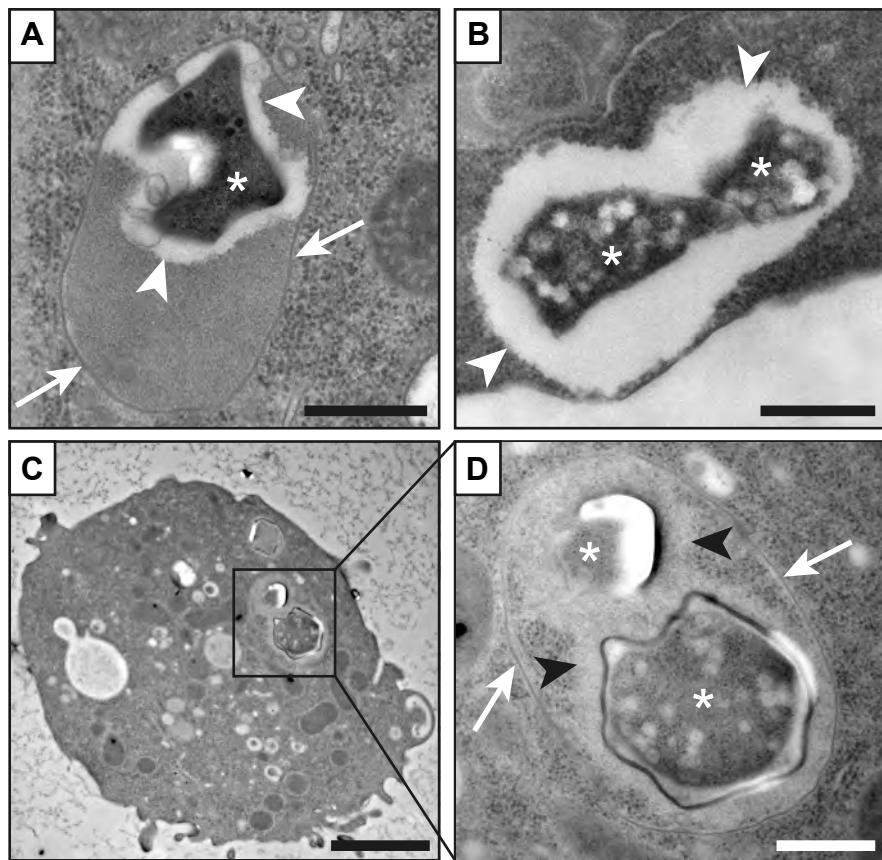
**FIG S2**



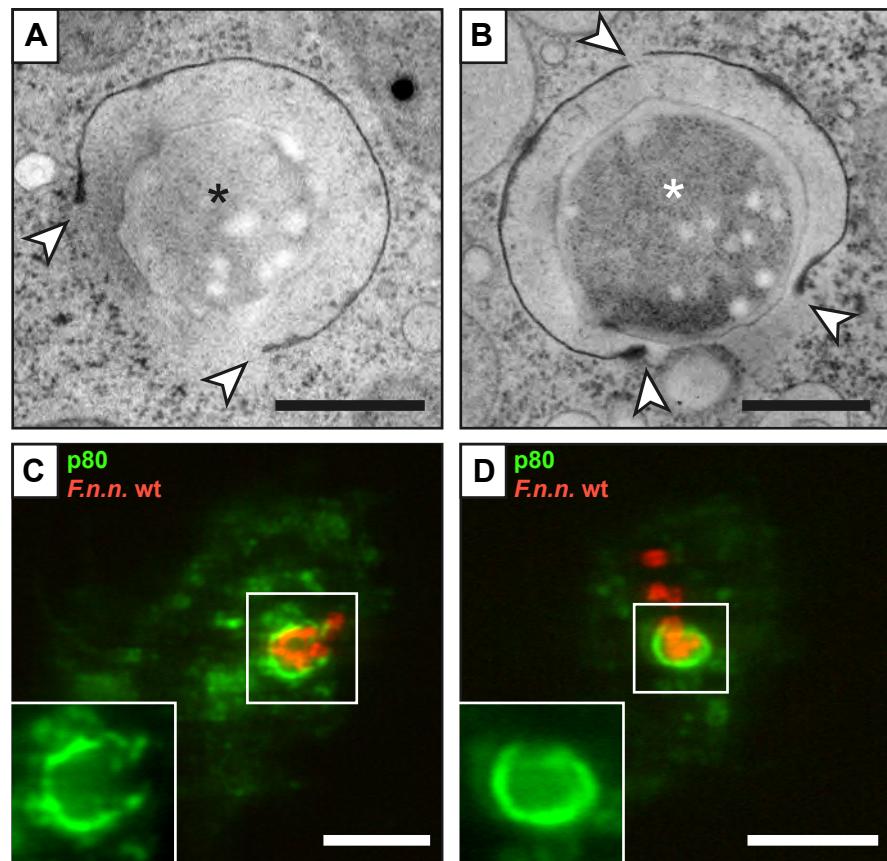
**FIG S3**



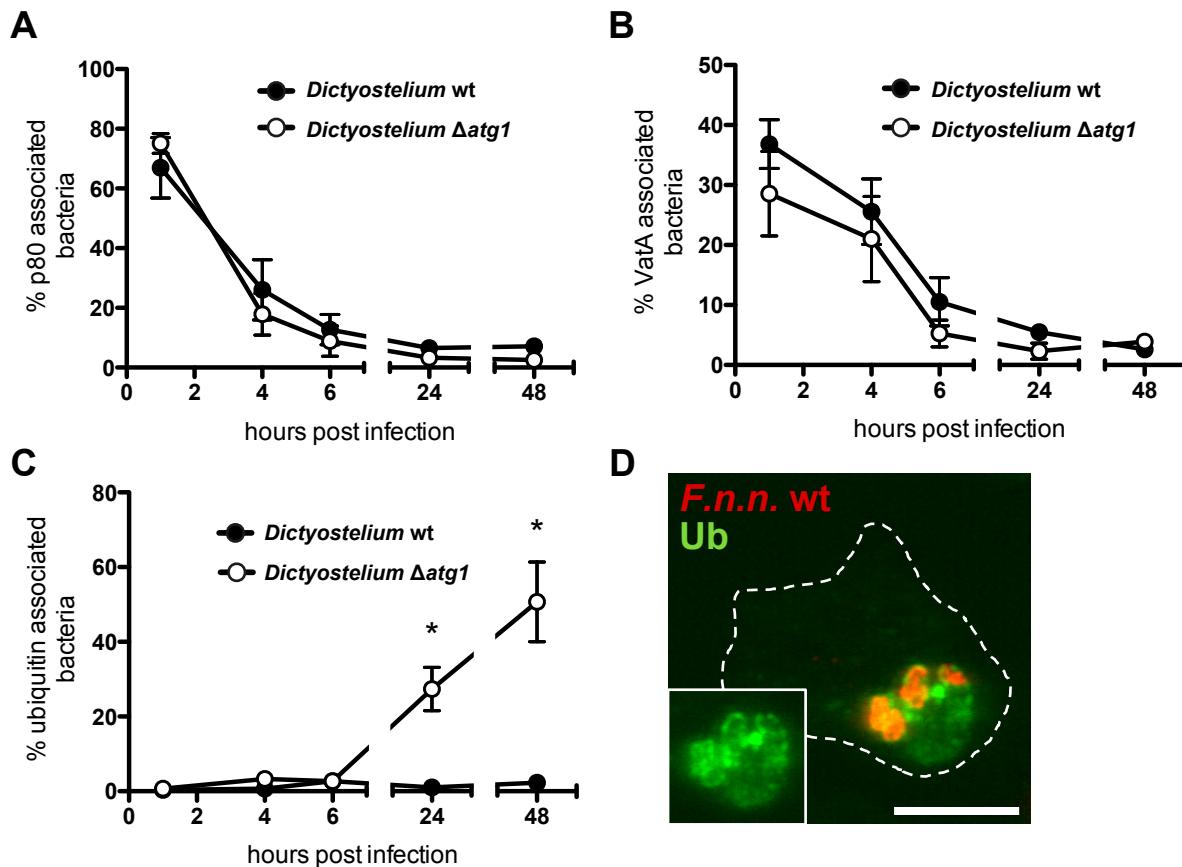
**FIG S4**



**FIG S5**



**FIG S6**



Primers used for cloning				
	Name	5'-3' sequence	Product size	Source
<b>P1</b>	Fnn_igIC_OF	GGTGGTGAACCATATGGCGC	P1+2: 2271 bp in mutant, 2837 bp in wt <i>F.n.n.</i>	This study
<b>P2</b>	Fnn_igIC_OR	TAAAGTCCCATTCCGAGCCTT		
<b>P3</b>	Fnn_igIC_IF	CACAGCAAGTTGCTAGAATAGGGTACATAGCAGCT	P1+4: 1184 bp	This study
<b>P4</b>	Fnn_igIC_IR	GCTATTCTAGCAACTGCTGTCTGTTATCATCTC	P2+3: 1105 bp	This study
<b>P5</b>	Fnn_igIC_C1F	GTGTATGTTGCAGAATTGATCA	P2+5: 2298 bp in mutant, 2864 bp in wt <i>F.n.n.</i>	This study
<b>P6</b>	Fnn_igIC_C2R	AACCGCACTACTATCCGCAAG	P1+6: 2294 bp in mutant, 2860 bp in wt <i>F.n.n.</i>	This study
<b>P7</b>	Fnn_igID_IR	TC AAA GGC TGT TCT TAG ACC ATC TTC CCA ACA	P5+7: 1228 bp in mutant, 1794 bp in wt <i>F.n.n.</i>	This study
<b>P8</b>	Fn_groEL F	CATAAGACCAGCGATTGATGC	367 bp	This study
<b>P9</b>	Fn_groEL R	AAAAGATCGTGTGGATGATGC		
<b>P10</b>	Fnn_compl_tr_igIC_F(Ndel)	TCT GCA CAT ATG ATT ATG AAC GAG ATG ATA ACA AGA CAG	635 bp	This study
<b>P11</b>	Fnn_compl_tr_igIC_R(EcoRI)	CTGA GAA TTC CTA TAC AGC TGC TAT GTA CCC TAT TCT		
Primers used for qPCR				
	Name	5'-3' sequence	Product size	Source
<b>Q1</b>	Dd_rnIA_F	GGCGCTGGTGAAATAGTAAGTATATTAGA	85 bp	(1)
<b>Q2</b>	Dd_rnIA_R	GGTTACCGCCCCAGTCAAA		
<b>Q3</b>	Dd_gapdh_F	GGTTGTCCAATTGGTATTAATGG	247 bp with gDNA, 156 bp with cDNA	(2)
<b>Q4</b>	Dd_gapdh_R	CCGTGGGTTGAATCATATTGAAC		
<b>Q5</b>	Dd_atg8_F	CTCCAAGATCAGATGCACCA	163 bp	Frauke Bach (GI: 8625555)
<b>Q6</b>	Dd_atg8_R	GCAGCAGTTGGTGGGATAGT		
<b>Q7</b>	Dd_p62_F	TTGAAAATCGCACAAACCAAC	172 bp	Frauke Bach (GI: 8617503)
<b>Q8</b>	Dd_p62_R	AGGAACCCTTGGAATGACA		
<b>Q9</b>	Fnn spec_F	TGAGTTGGTAACCATTGATTGTACATAGT	97 bp	(3)
<b>Q10</b>	Fnn spec_R	CGAGTACCTGGTGGGAGAAAGA		
<b>Q11</b>	Fnn_igIB_F	GGTGCTGAGCAAATTCAAAA	85 bp	This study (GI:169589433)
<b>Q12</b>	Fnn_igIB_R	CCATCTCTAACAGGCCTTGGT		
<b>Q13</b>	Fnn_igIC_F	TAGGCGTATAACACTGGCTGC	70 bp	(4)
<b>Q14</b>	Fnn_igIC_R	TGCTATAGAAGGCCGGAGAGG		
<b>Q15</b>	Fnn_igID_F	TGTAAAACCTCCCCGAAGA	77 bp	This study (GI:169589431)
<b>Q16</b>	Fnn_igID_R	GCTGGAGCTATTGCCTTCT		

## References

1. **Damer CK, Bayeva M, Kim PS, Ho LK, Eberhardt ES, Socec CI, Lee JS, Bruce EA, Goldman-Yassen AE, Naliboff LC.** 2007. Copine A is required for cytokinesis, contractile vacuole function, and development in Dictyostelium. *Eukaryot Cell* **6**:430-442.
2. **Lucas J, Bilzer A, Moll L, Zundorf I, Dingermann T, Eichinger L, Siol O, Winckler T.** 2009. The carboxy-terminal domain of Dictyostelium C-module-binding factor is an independent gene regulatory entity. *PLoS One* **4**:e5012.
3. **Duodu S, Larsson P, Sjodin A, Soto E, Forsman M, Colquhoun DJ.** 2012. Real-time PCR assays targeting unique DNA sequences of fish-pathogenic *Francisella noatunensis* subspecies noatunensis and orientalis. *Dis Aquat Organ* **101**:225-234.
4. **Brudal E, Winther-Larsen HC, Colquhoun DJ, Duodu S.** 2013. Evaluation of reference genes for reverse transcription quantitative PCR analyses of fish-pathogenic *Francisella* strains exposed to different growth conditions. *BMC Res Notes* **6**:76.