

Supplementary information

From crystal structure of α -conotoxin GIC in complex with Ac-AChBP to molecular determinants of its high selectivity for $\alpha 3\beta 2$ nAChR

Bo Lin^{1,#}, Manyu Xu^{2,#}, Xiaopeng Zhu¹, Yong Wu¹, Xi Liu², Dongting Zhangsun¹,
Yuanyan Hu¹, Shi-Hua Xiang⁴, Igor E. Kasheverov³, Victor I. Tsetlin³, Xinquan
Wang^{2,*} and Sulan Luo^{1,*}

¹Key Laboratory of Tropical Biological Resources, Ministry of Education, Key Lab for Marine Drugs of Haikou, Hainan University, Haikou Hainan 570228, P. R. China.

²Ministry of Education Key Laboratory of Protein Science, Center for Structural Biology, School of Life Sciences, Tsinghua University, Beijing 100084, P. R. China.

³Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry, Russian Academy of Sciences, Miklukho-Maklaya Street, 16/10 Moscow 117997, Russia.

⁴Nebraska Center for Virology, School of Veterinary Medicine and Biological Sciences, University of Nebraska-Lincoln, Lincoln, NE 68583, USA.

#Equal contribution

***Corresponding authors:**

Sulan Luo, Ph.D.

Key Laboratory of Tropical Biological Resources, Ministry of Education, Hainan University, Haikou Hainan 570228, P. R. China

Phone: (86) 898 66289538; Fax: (86) 898 66276720.

E-mail: luosulan2003@163.com

Xinquan Wang, Ph.D.

Ministry of Education Key Laboratory of Protein Science, Center for Structural Biology, School of Life Sciences, Tsinghua University, Beijing 100084, P. R. China.

E-mail: xinquanwang@mail.tsinghua.edu.cn

Running Title: Co-crystal structure of α -conotoxin GIC and acetylcholine binding proteins

Table S1|Crystal diffraction data collection and structural refinement statistics

Data collection	
Beamline	SSRF BL17U
Wavelength	0.9796 Å
Space group	P2 ₁ 2 ₁ 2 ₁
Cell dimensions	
<i>a</i> , <i>b</i> , <i>c</i> (Å)	78.6, 84.9, 208.6
α , β , γ (°)	90.0, 90.0, 90.0
Resolution (Å)	50-2.1
<i>R</i> _{merge} (%)	8.8 (67.7)
<i>I</i> / σ <i>I</i>	18.7 (3.5)
Completeness (%)	96.9 (93.5)
Redundancy	6.7 (6.9)
Refinement	
Resolution (Å)	43.5-2.1
No. Reflections	85409
<i>R</i> _{work} / <i>R</i> _{free} (%)	19.7/23.7
No. atoms	
Protein	8299
Ligand/ion	545
Water	521
B factor (Å²)	
Protein	41.1
Ligand/ion	45.3
Water	42.5
r.m.s. deviations	
Bond lengths (Å)	0.008
Bond angles (°)	1.104

Table S2 | Contacts between *Ac-AChBP* and α -conotoxins ($d < 4 \text{ \AA}$)

<i>Ac-AChBP</i>	GIC^a	PnIA(A10L, D14K)^b	TxA(A10L)^c	ImI^d	BuA
<i>Principal side</i>					
Lys-23				Arg-11	
Tyr-91	His-5	Leu-5, Pro-7	Arg-5, Pro-6, Pro-7	Arg-7	Pro-7
Ser-144		Pro-7	Pro-7	Arg-7	Pro-7
Trp-145	Pro-6, Ala-7	Pro-6, Pro-7	Pro-6, Pro-7	Pro-6, Arg-7	Pro-6, Pro-7
Val-146	Ala-7	Leu-10, Asn-11	Pro-7, Leu-10	Arg-7	Pro-7
Tyr-147			Pro-7	Arg-7	Pro-7
Ser-148	Asn-11	Asn-11	Asn-11		
Glu-151		Asn-11			
Tyr-186	Gly-1, Cys-2, His-5, Cys-8	Gly-1, Cys-2, Leu-5, Cys-8	Gly-1, Cys-2, Arg-5	Gly-1,Cys-2, Asp-5	Gly-1, Cys-2, Thr-5
Cys-188	Cys-2	Cys-2, Tyr-15	Cys-2	Cys-2	Cys-2
Cys-189	Asn-12	Cys-2, Asn-12	Cys-2, Asn-12	Cys-2	Cys-2, Tyr-12
Glu-191	Asn-11, Asn-12	Asn-12	Asn-11, Asn-12	Arg-11	Leu-11, Tyr-12
Tyr-193	His-5, Ala-7, Cys-8, Asn-11, Asn-12	Leu-5, Pro-7, Cys-8, Asn-11, Asn-12	Arg-5, Pro-7, Cys-8, Asn-11, Asn-12	Arg-7, Cys-8	Pro-7, Cys-8, Tyr-12
Ile-194				Arg-7	
Asp-195			Arg-5		
<i>Complementary side</i>					
Thr-34	Ser-4		Cys-3		Cys-3

Tyr-53	Ser-4, Pro-6	Pro-6	Ser-4, Pro-6	Ser-4	Ser-4
Gln-55	Ala-9, Cys-16		Ile-9	Cys-3	Cys-3, Ala-9, Cys-13
Arg-57	Gln-13,	Pro-13	Pro-13		Cys-13
Asp-75				Trp-10	
Arg-77	Asn-11	Asn-11	Asn-11	Trp-10	
Ile-104		Leu-10			
Val-106	Gly-10, Gln-13	Leu-10	Leu-10	Trp-10	Val-10
Thr-108	Gln-13			Trp-10	
Asp-110		Lys-14			
Ser-112	Gln-13	Lys-14			
Met-114	Ala-9,Gly-10, Gln-13	Ala-9, Leu-10, Pro-13	Ile-9, Leu-10, Pro-13	Trp-10	Val-10, Leu-11
Ile-116	Pro-6, Ala-9	Pro-6, Ala-9, Leu-10,	Leu-10	Pro-6, Ala-9	Ala-9, Val-10
Asp-157	Cys-16				
Asp-162	Ser-4	Ser-4	Cys-3	Ser-4	Cys-3
Ser-164	Ser-4	Ser-4	Ser-4	Gly-1, Ser-4	Ser-4
Ser-165	Ser-4		Ser-4		Ser-4

Table S3 | Affinity (as K_i or IC_{50} s, nM) of four α -conotoxins for *Ac-AChBP*, *AChBP* from *Lymnaea stagnalis* (*Ls-AChBP*) and different human or rat nAChR subtypes

α -conotoxins	<i>Ac-AChB</i>	<i>Ls-AChBP</i>	<i>hα3β2</i>	<i>hα3β4</i>	<i>hα7</i>	<i>rα3β2</i>	<i>rα7</i>
P							
GIC	29 ± 2^a	220 ± 10^a	1 ^b	755 ^b			
ImI	33 ± 5^c	4140 ± 400^c	41 ^d	3390 ^d	132 ^e	$>5000^f$	69 ^g
PnIA(A10L,D14K)	28 ± 6^h	13 ± 2^h			260 ^h		
TxA(A10L)		1.1 ⁱ				2.0 ⁱ	39 ⁱ

Data from ^a this work, ^b ref. ¹¹, ^c ref. ⁹, ^d ref. ²¹, ^e ref. ²⁸, ^f ref. ²², ^g ref. ³⁰, ^h ref. ⁷ and ⁱ ref. ¹⁰.

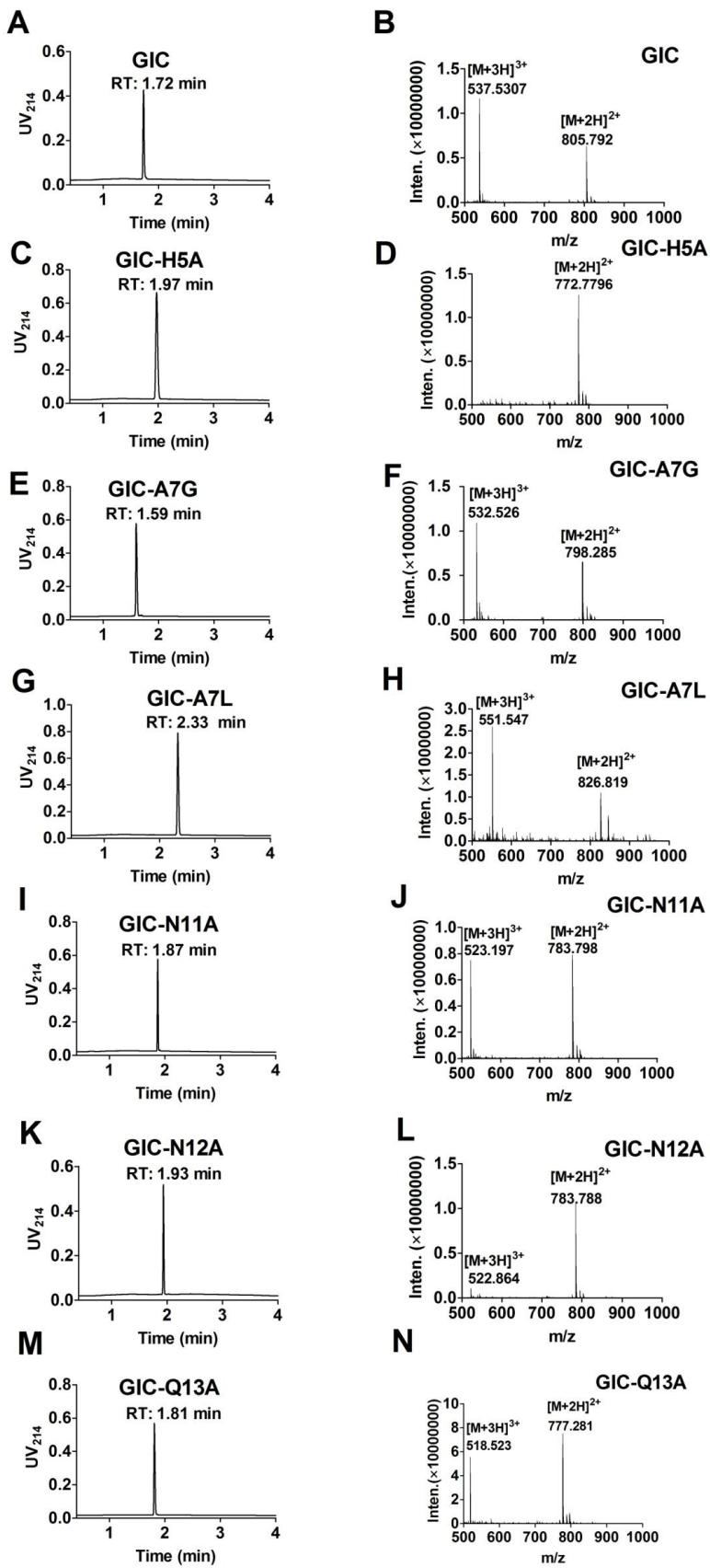


Figure S1| HPLC chromatograms with a retention time and ESI-MS data of

α -conotoxin GIC and its analogues. The peptides were analyzed on a waters ACQUITY UPLC BEH C18 column (2.1×50 mm, $1.7 \mu\text{m}$) using a linear gradient of 5% Buffer B to 40% Buffer B over 3.5 min with flow rate of 0.5 ml/min at temperature of 40°C , where A = 0.1% trifluoroacetic acid (TFA) and B = 0.75% TFA, 90% acetonitrile, and the remainder water. Absorbance was monitored at 214 nm.

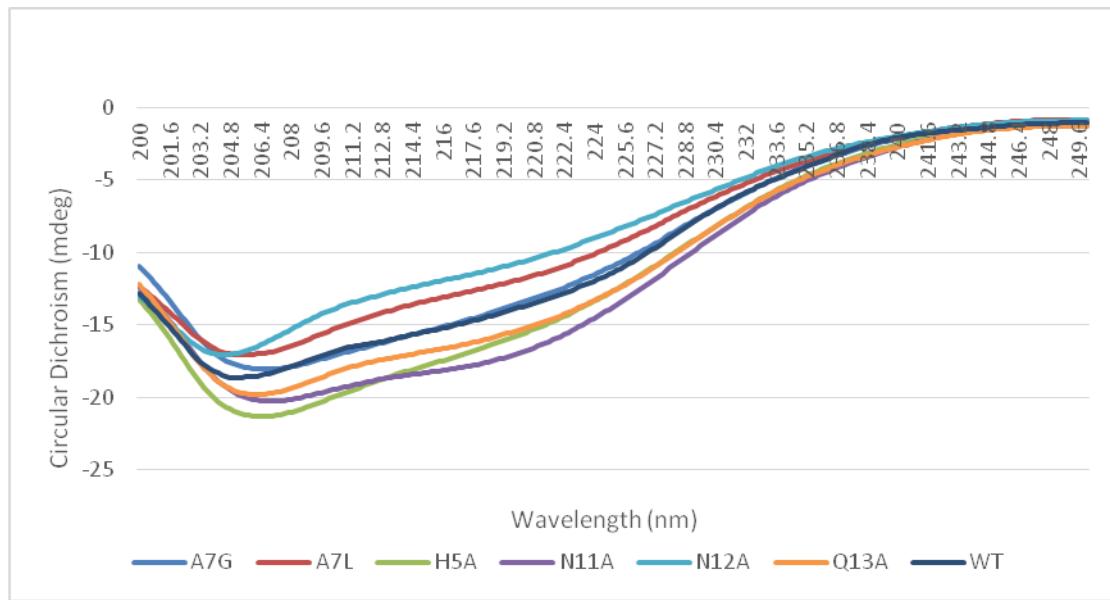


Figure S2| Circular dichroism of α -conotoxin GIC and its analogues. CD spectra were recorded on a JASCO J-715 Spectropolarimeter. Spectra was recorded at room temperature under nitrogen atmosphere. Peptides were dissolved to $100 \mu\text{M}$. Spectra were recorded over a 200-250 nm range at 25°C using an average of 3 scans (scan speed of 200 nm/min).

	20	40	60	
Ac-AChBP	--QANL---MRLKSDLFN---	RSPMYPG--PT-KDDPLTVTLGFTLQDIVKADSSSTNEV		48
Ls-AChBP	LDRADI-----LYN---	IRQTSRPDV IPTQRDRPVA SVS VSLKF INI LEVNEITNEV		48
Human α 3	-----SEAEHRLFERLFED	-YNEIIIRP--VANVSDPVI IHFEVSM SQLVKVDEVNQIM		50
Human α 4	HVE TRA HAEERLLKKLFSG	-YNKWSRP--VANISDVLVLVRFG LSI AQLIDVDEKNQMM		55
Human β 2	-----TDTEERLVEHLLDPSR	YNKLIRP--ATNGSELVTVQLMVSLAQLI SVHEREQIM		52
Human β 4	RV---ANAEEKLMDDLNKTRYNNLIRP	--ATSSSQLISIKLQLSLAQLI SVNEREQIM		54
	80	100		120
Ac-AChBP	DLVYYYEQQ RWKLN SLMWDPN EYGNITDFRTSAADIWTPDITAY	S-S TRPVQVLSPQIAVV		107
Ls-AChBP	DVVFWQQT WSDRTLAWNSSHSPDQVSVP ISS--LWVPDLAA	Y N-AISKPEVLTPQLAR	V	105
Human α 3	ETNLWLKQIWNDYKLKWNP SDYGGAEFMRVPAQKIWKPDIVLY	NNAVGDFQVDDKTKA	LL	110
Human α 4	TTNVWVKQEWHDYKLRWD PAD YENVTSIRIPSEL IWRPDI	VLYNNADGDFAVTHLTKA	HL	115
Human β 2	TTNVWLTQEWEDYRLTWKPEEFDNMKKV RLP SKHIWLPDV	VLYNNADGMYEVSFYNSNA	VV	112
Human β 4	TTNVWLKQEWTDYRLTWNSSRYEGVNILRIPAKRIWLPDI	VLYNNADGTYEVSVYTNLIV		114
	140	160		180
Ac-AChBP	THDG SVMFIPAQR LSFMC--DPTGV DSEEGATCAV KFGS	WVYSGFEIDLKTDTDQV	DLSS	165
Ls-AChBP	VSDGEVLYMPSI RQRFSC--DVGVDTESGATCRIKIGS	WT HHSREISVDP TTENSDDSE		163
Human α 3	KYTGEV TWIPPAIFKSSCKI DVTYFPFDY-QNCTMKG	SWSYDKAKIDLVLIGSSMNLKD		169
Human α 4	FHDGRVQWTPPAIYKSSCSIDVTFFPFDQ-QNCTMKG	SWTYDKAKIDLVNMHSRVDQLD		174
Human β 2	SYDG SIFIWLP PPAIYKSACKIEVKHFPFDQ-QNCTMKG	RSW TYDRTEIDLVLKSEVASLD	D	171
Human β 4	RSNGS V LWLPPA IYKSACKIEVKYFPFDQ-QNCTLKFR	SWTYDHTEIDMVLMTPTASMD		173
	200	220		240
Ac-AChBP	YYAS-SKYEILSATQTRQVQHY	SCCP EPYIDVN L VVKFRER	-	205
Ls-AChBP	YFSQYSRFEILDVTQKKNSVTY	SCCP EAYEDVEV S LNFRKK	-	204
Human α 3	YWES-GEWAIIKAPGYKHDIKY	NCCCEEIYPDITYSLYI RRLPLFYTI	NL IIPCLLISFLT	228
Human α 4	FWES-GEWVIVDAVGTYNT	RKYEC CAAEIYPDITYAFVIRRLPLFYTI	NL IIPCLLISCLT	233
Human β 2	FTPS-GEWDIVALPGRRNENPDD	--STYVDI TYDFI I RRKPLFYTI	NL IIPCVLITS LA	227
Human β 4	FTPS-GEWDIVALPGRR TVNPQD	--PSYVDV T YDFI I KRKPLFYTI	NL IIPCVLTTLLA	229

Figure S3 | Sequence alignment of Ac-AChBP, Ls-AChBP, and human α 3, human α 4, human β 2, human β 4 of nAChR subunits. Each subunit sequence numbering is shown at right side. Residues of the principal binding side that interact with α -conotoxin GIC are shown in light green; residues of the complementary binding side are in light blue. The Arg residues which could be a potential reason of decreased GIC affinity for the Ls-AChBP, human α 4 β 2 nAChR and α 3 β 4 nAChR are typed in red.